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## Synergistic activity of antibiotics combined with ivermectin to kill body lice

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### ABSTRACT

Ivermectin and doxycycline have been found to be independently effective in killing body lice. In this study, 450 body lice were artificially fed on a Parafilm™ membrane with human blood associated with antibiotics (doxycycline, erythromycin, rifampicin and azithromycin) alone and in combination with ivermectin. Fluorescence in situ hybridisation and spectral deconvolution were performed to evaluate bacterial transcriptional activity following antibiotic intake by the lice. In the first series, a lethal effect of antibiotics on lice was observed compared with the control group at 18 days (log-rank test,  $P \leq 10^{-3}$ ), with a significant difference between groups in the production of nits ( $P = 0.019$ , Kruskal–Wallis test). A high lethal effect of ivermectin alone (50 ng/mL) was observed compared with the control group (log-rank test,  $P \leq 10^{-3}$ ). Fluorescence of bacteriocytes in lice treated with 20 µg/mL doxycycline was lower than in untreated lice ( $P < 0.0001$ , Kruskal–Wallis test). In the second series with antibiotic–ivermectin combinations, a synergistic lethal effect on treated lice (log-rank test,  $P < 10^{-6}$ ) was observed compared with the control group at 18 days, associated with a significant decrease in the production of nits ( $P \leq 0.001$ , Kruskal–Wallis test). Additionally, survival of lice in the combination treatment groups compared with ivermectin alone was significant (log-rank test,  $P = 0.0008$ ). These data demonstrate that the synergistic effect of combinations of antibiotics and ivermectin could be used to achieve complete eradication of lice and to avoid selection of a resistant louse population.

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### 1. Introduction

The body louse *Pediculus humanus humanus* is a strictly human parasite, living and multiplying in clothing; their infestation is associated with cold weather and a lack of hygiene [1]. Its prevalence reflects the socioeconomic level of the society [2]. In developed countries, infestation is more common among the homeless population and varies from 7% to 22% [3]. Body lice are extremely contagious and pose a serious public health problem. They are vectors of the pathogens *Rickettsia prowazekii*, *Bartonella quintana* and *Borrelia recurrentis*, which are responsible for epidemic typhus, trench fever and relapsing fever, respectively [1,4], and they may be considered a plague [5].

For decades, several antibiotics have proved to be effective in the treatment of louse-borne diseases. A single dose of 200 mg doxycycline can cure patients suffering from *R. prowazekii*-induced typhus [6], and 500 mg of tetracycline or erythromycin is optimal therapy for the treatment of *B. recurrentis* infections [7]. Azithromycin has been recommended for the treatment of *B. quintana* infections [8]. In addition, rifampicin was shown to be effective against some intracellular bacteria [9]. These results indicate the potential of antibiotics for the management of louse-borne diseases. Recently, we demonstrated the in vitro efficacy of doxycycline at different doses on body lice and found that doxycycline has a direct effect on endosymbionts of lice through the mycetome switch [10].

Several strategies for eradication of these parasites have been used to avoid the spread and/or outbreak of these diseases. However, some products have already shown their limits. The resistance of lice to lindane in Europe, Africa and Asia [11] and to DDT in Korea and Japan [12,13] has been reported. Pyrethroids have been widely used, however resistance to these compounds has emerged and

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spread over the last decade. In the head louse, resistance to permethrin was first reported in France in 1994 and throughout the world thereafter [14]. Ivermectin is a synthetic derivative of the antiparasitic class of compounds known as avermectins [15]. Resistance to ivermectin and related drugs is an increasing problem for parasite control [16]. Previous studies have associated ATP-binding cassette (ABC) transporters with drug resistance in nematodes [17] and arthropods [18]. However, resistance to ivermectin has also been documented in insects and mites of plants [19] and in nematodes [16]. Resistance to ivermectin has been documented in *Rhipicephalus (Boophilus) microplus* ticks in Mexico [20]. The occurrence of resistance to macrocyclic lactones has been reported in other arthropods [21]. Used in scabies, clinical and in vitro resistance have been documented in two patients who received 30 and 58 doses of ivermectin, respectively [22].

Today, post-therapeutic re-infestation of the vector of diseases (lice) is common and still remains a real challenge, thus there is a need to develop new therapeutic strategies to avoid these re-infestations. It was observed that avermectin B1a could act synergistically with the antibiotic meticillin to kill meticillin-resistant *Staphylococcus aureus* (MRSA), rendering the bacteria once again vulnerable to meticillin [23]. The hypothesis that the bacterial symbiont *Candidatus Riesia pediculicola* is a possible target for the development of louse control strategies [24] was recently confirmed in our previous work [10]. In this study we aimed to demonstrate the in vitro combination of ivermectin with several antibiotics as a synergistic association to achieve complete eradication of lice and to avoid the selection of a resistant population of lice.

## 2. Materials and methods

### 2.1. Louse feeding system in vitro

A long-established inbred line of the human body louse (*P. h. corporis*, Orlando strain), which had been bred on rabbits in a laboratory pet shop by our URMITE team in Marseille, France, was used as a model in the experiments in this study [25]. A Hemotek™ feeding system (Discovery Workshops, Accrington, UK) with a Parafilm™ membrane (Chicago, IL), a PS5A/220 Power Unit (Hemotek™ 5W1 System; Discovery Workshops) and six heating units was used (Fig. 1). A square of synthetic feeding membrane measuring ca. 3 cm × 3 cm was stretched around the aperture of the blood meal reservoir and was secured with an O-ring (Fig. 1a). The feeding area was sufficient for 100 lice (Fig. 1b). The blood meal reservoir was removable for easy cleaning and autoclaving if necessary. The system does not use animal hosts and enables modification of the meal by introduction of chemicals or infectious agents and accurate adjustment of the temperature through heating units (Fig. 1c) connected directly to the power unit (Fig. 1d). Lice were fed every day at 37 °C for 1 h (Fig. 1e).

### 2.2. Preparation of blood meal supplemented with antibiotics and ivermectin

Human blood group A rhesus positive (A<sup>+</sup>) [code 74883 from the French Blood Service (EFS)] was used. Ethical approval for the use of human blood in vitro was obtained from the Laboratory Research Ethics Board of Molecular Hematology, EFS. Blood was distributed into 5 mL tubes and was kept at –20 °C before use. The blood was warmed in a water bath at 37 °C for 30 min before use. Drugs used were in injectable and oral form as follows: doxycycline 20 mg/mL (Vibraveineuse® batch A262408; Laboratoires SERB, Paris, France); erythromycin 1 g (Erythrocin® batch 24370TB22; Amdipharm Ltd., Dublin, Ireland); rifampicin 600 mg (Rifadine®

batch A3468; Sanofi-Aventis, Paris, France); azithromycin 600 mg (Azadose® batch 490004; Pfizer Holding, Paris, France); and ivermectin 10 mg/mL (Ivomec® batch BM295/12; Merial, Lyon, France). For each group of lice, 15 µL of uncombined antibiotic diluted at concentrations (doxycycline 20 µg/mL, erythromycin 4 µg/mL, rifampicin 10 µg/mL and azithromycin 8 µg/mL) previously established for the first series of experiments was added to a volume of 1.485 mL of haemolysed blood. In the second series for each group of lice, 15 µL of antibiotic (doxycycline 20 µg/mL, erythromycin 4 µg/mL, rifampicin 10 µg/mL and azithromycin 8 µg/mL) plus 15 µL of ivermectin diluted at a concentration (50 ng/mL) previously established was added to a volume of 1.470 mL of haemolysed blood. The final volume of 1.5 mL obtained for each concentration was introduced into different reservoirs. The reservoir was then attached to the feeder by screwing it onto a nylon stud on the heat transfer plate at the bottom of the feeder.

### 2.3. Monitoring of lice and nits

In total, 450 adult lice [excluding 10 lice sacrificed as live controls in fluorescence in situ hybridisation (FISH)] were used in the experiments. Lice were divided into groups of 45, were placed on a piece of black cloth and were kept in jars in an oven at 29 °C and 90% humidity as breeding lice. The first series of experiments was performed without combination of drugs and the second was based on the antibiotic–ivermectin combinations. A group of lice treated with ivermectin alone was monitored in parallel with the combination series. The control group was fed exclusively with blood without drugs throughout the experiments. Lice were placed on the membrane daily for 1 h of blood meal feeding. Every day, survival was determined by assessing the number of nits in each group and was recorded on a data collection sheet before the next meal (Supplementary Tables S1 and S2). All of the experiments were repeated three times for reproducibility.

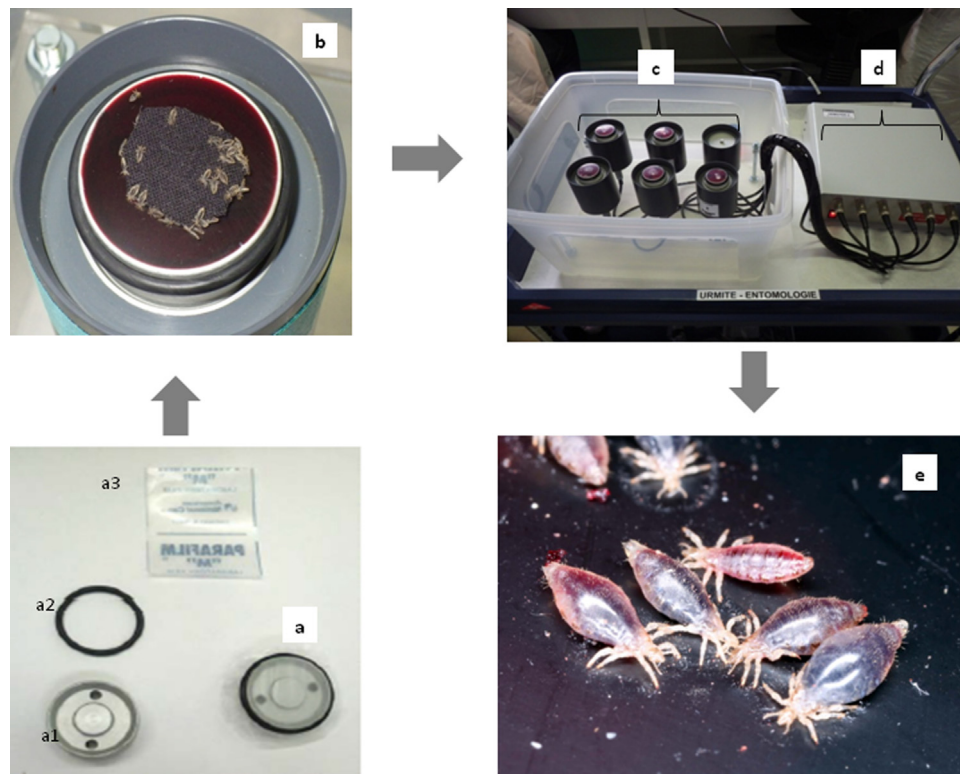
Supplementary Tables S1 and S2 related to this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.ijantimicag.2016.01.001>.

### 2.4. Fluorescence in situ hybridisation (FISH)

For the FISH technique, the aim was to highlight the effect of the antibiotic (especially doxycycline) on the louse endosymbiont as previously demonstrated [10]. Thus, ten specimens were tested in each untreated group (living controls and dead controls) and in the doxycycline 20 µg/mL group (Table 1). Following fixation, samples were decolourised in 6% H<sub>2</sub>O<sub>2</sub> in ethanol for 2 h and were then placed in hybridisation buffer [20 mM Tris–HCl (pH 8.0), 0.9 M NaCl, 0.01% sodium dodecyl sulphate and 30% formamide] containing 10 pmol of fluorescent probe/mL based on the *Candidatus Riesia pediculicola* 16S rRNA sequence: Cy3–Lice1255R (5′–Cy3–TTGGCTCGTCTTACGAGT–3′) [10].

### 2.5. Light and confocal microscopy

The spectral deconvolution technique was used to differentiate autofluorescence caused by chitin from clean fluorescence. First, the louse was marked and mounted between slide and cover slip and imaging was performed as follows using a Leica SP5 AOBs® confocal resonant scanner (Leica Microsystems, Mannheim, Germany) in Lambda mode. The sample was scanned on a total band of 160 nm and a slit width of 5 nm taking care to never saturate the signal. Fluorescence measurement was defined as the extraction of curves (one for fluorescence and one for autofluorescence). After creation of the reference curves, all acquisitions were made with exactly the same settings for gain, offset, laser power and objective. For each acquisition, 32 pictures were collected. Once the references were



**Fig. 1.** The Hemotek™ 5W1 membrane feeding system for lice: (a) the device's meal reservoir (a1, blood meal reservoir; a2, plastic ring; and a3, Parafilm™); (b) assembling the device; (c) the heating units; (d) the PS5 power unit; and (e) blood meal feeding of lice on artificial membrane.

**Table 1**  
Measurement of fluorescence intensity in three populations of lice.

Group	Day	Total area	Surface marked	Intensity ratio
Doxycycline 20 µg/mL	1	16,217	1205	7.4
	2	4808	325	6.7
	3	29,970	1385	5.9
	4	32,994	2179	6.6
	5	19,730	1190	6
	6	29,211	1537	5.2
	7	51,047	2468	4.8
	8	20,479	1216	4.6
	9	37,185	1231	3.3
	10	16,231	395	2.4
Dead con- trols	1	8977	1462	16.2
	2	9523	1275	13.3
	3	9488	1358	14.3
	4	13,279	1755	13.2
	5	12,391	1518	12.2
	6	9873	1280	12.9
	7	9173	1051	11.4
	8	10,506	1163	11
	9	9923	986	9.9
	10	12,675	1269	10
Living con- trols	1	12,861	1943	15.1
	2	12,496	1605	12.8
	3	11,058	1467	13.2
	4	7558	1249	16.5
	5	10,913	1134	10.3
	6	7945	1156	14.5
	7	8962	1580	17.6
	8	12,381	1397	11.2
	9	9914	1290	13
	10	13,091	1898	14.4

determined and assigned, the samples were spectrally deconvolved and the signals were separated by assigning them a distinct colour.

## 2.6. Calculation of intensity ratios

MetaMorph v.4.6 (Molecular Devices, Sunnyvale, CA) and a generated Excel table (Microsoft Corp., Redmond, WA) were used to calculate the intensity ratios.

## 2.7. Statistical analysis

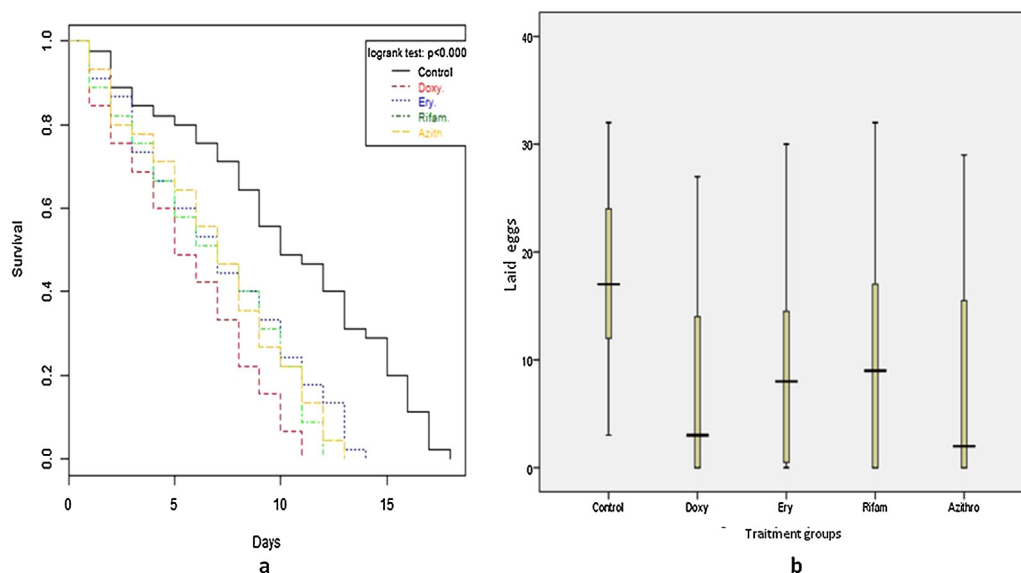
Data were stored in Excel. Analyses were performed with R software v.3.1.1 (R Foundation for Statistical Computing, Vienna, Austria). Kaplan–Meier estimations and log-rank test were used for survival analysis. Analysis of Kruskal–Wallis non-parametric test was used for mean comparisons. Bonferroni correction was applied for subgroup tests. The statistical tests represent bilateral situations, and  $P$ -values of  $\leq 0.05$  were considered significant.

## 3. Results

Feeding lice on artificial membranes enabled them to be treated with antibiotics and ivermectin. The blood feeding rate was >95% in all treatment arms (Fig. 1e).

### 3.1. Effect of antibiotics on the survival of lice and on nit production

To determine the effect of antibiotics, in the first series of experiments each group of lice treated with either doxycycline 20 µg/mL, erythromycin 4 µg/mL, rifampicin 10 µg/mL or azithromycin 8 µg/mL as well as the controls were followed until the end. Survival of lice in the control group was significantly different from survival in the treated groups (log-rank test,  $P \leq 10^{-3}$ )



**Fig. 2.** Evolution trends in the antibiotic treatment groups. (a) Kaplan–Meier curves illustrating survival of lice treated with antibiotics in comparison with the control group. (b) Mean ( $\pm$ 95% CI) number of eggs laid per day in the doxycycline-, erythromycin-, rifampicin- and azithromycin-treated groups and the control group. CI, confidence interval; Doxy, doxycycline; Ery, erythromycin; Rifam, rifampicin; Azithro/Azithro, azithromycin.

(Fig. 2a). Moreover, a decrease in the production of nits was observed in the treated groups compared with the control group (Fig. 2b), with mean [95% confidence interval (CI)] number of eggs laid per day of 7.5 (2.5–12.5), 9.8 (4.3–15.3), 10 (4.6–15.6), 7.7 (2.2–13.2) and 17.8 (13.3–22.3) in the doxycycline-, erythromycin-, rifampicin- and azithromycin-treated groups and the control group, respectively. The difference in mean nit production was significant between the groups ( $P=0.019$ , Kruskal–Wallis test).

### 3.2. Direct effect of doxycycline on the lice mycetome analysed through FISH

To show that doxycycline affects the louse's endosymbionts, we focused on a new analysis based on spectral deconvolution to calculate the ratios between the two signals (Table 1). By using the function extraction curve computation of reply in emission and positioning the reference areas on distinctive signals, two curves were extracted: one for specific fluorescence of the bacterium and one for autofluorescence of the louse. Thus, the results show that bacterial fluorescence in the group treated with doxycycline 20  $\mu$ g/mL (Fig. 3a) was lower than in the untreated groups (living and dead controls) (Fig. 3b and c) ( $P<0.0001$ , Kruskal–Wallis test).

### 3.3. Effect of ivermectin alone on the survival of lice and production of nits

To determine the antiparasitic action of ivermectin alone, lice treated with 50 ng/mL ivermectin were followed until the end of the experiment. A high mortality rate was found compared with the control group (log-rank test,  $P\leq 10^{-3}$ ) at 18 days (Fig. 4a). A significant difference between groups was observed in mean nit production ( $P=0.001$ , Kruskal–Wallis test), with mean (95% CI) number of eggs laid per day of 5.2 (0.6–9) and 17.8 (13.3–22.3), respectively (Fig. 4b).

### 3.4. Synergistic effect of antibiotics and ivermectin on survival of lice and production of nits

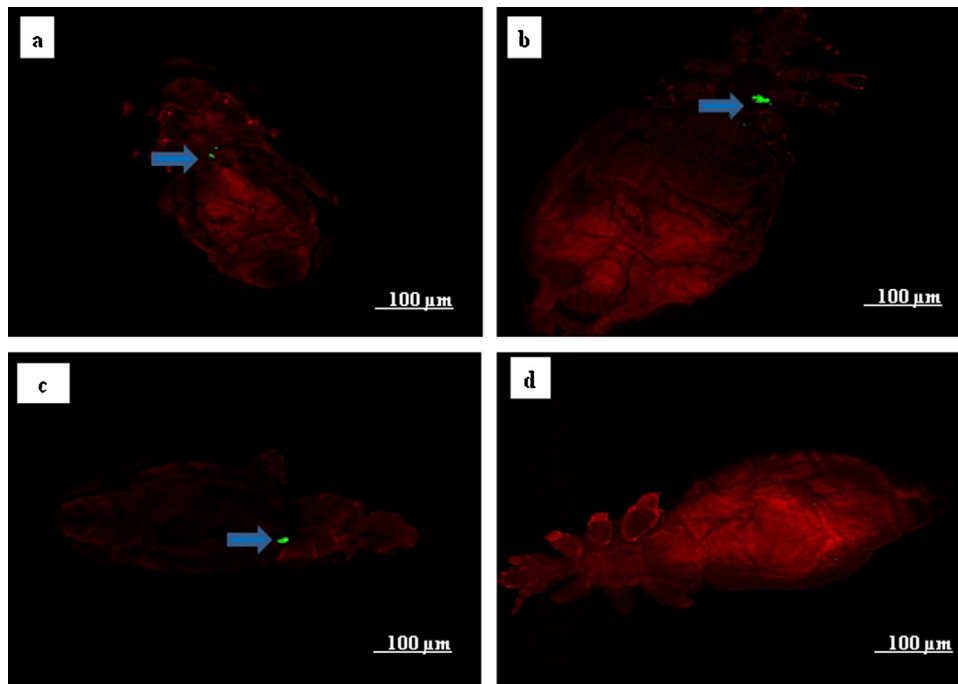
To determine the antiparasitic synergistic action of ivermectin combined with antibiotics, in a second series of experiments five groups of lice were tested for their survival kinetics. In

the combination groups (doxycycline 20  $\mu$ g/mL+ivermectin 50 ng/mL, erythromycin 4  $\mu$ g/mL+ivermectin 50 ng/mL, rifampicin 10  $\mu$ g/mL+ivermectin 50 ng/mL or azithromycin 8  $\mu$ g/mL+ivermectin 50 ng/mL), lice were more rapidly killed compared with the control group, exhibiting potent synergistic action (log-rank test,  $P<10^{-6}$ ) (Fig. 5a). Moreover, a significant decrease in the production of nits was observed in the combination groups compared with the control group (Fig. 5b), with mean (95% CI) eggs per day of 2.4 (–0.9 to 5.8), 4.4 (–0.1 to 9.0), 4.0 (–0.6 to 8.6), 3.6 (–0.5 to 7.7) and 17.8 (13.3–22.3), respectively. Thus, the difference in mean production of nits between groups was significant ( $P\leq 0.001$ , Kruskal–Wallis test). Moreover, overall survival of lice in the combination treatment groups compared with ivermectin alone was also significant (log-rank test,  $P=0.0008$ ) (Fig. 6a). Therefore, the difference was significant when comparing the combination groups with the ivermectin and control groups (Fig. 6b).

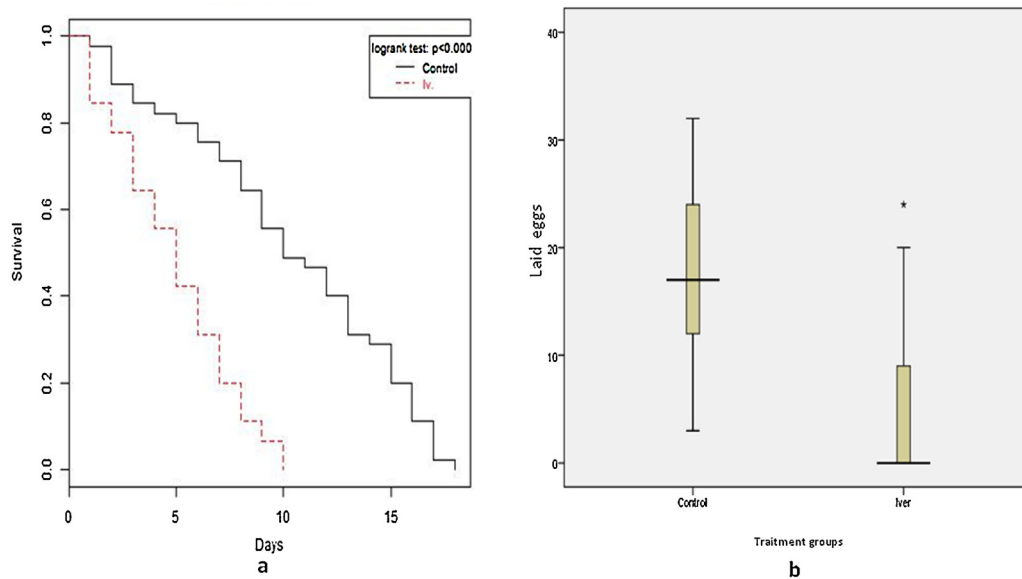
## 4. Discussion

For decades, complete eradication of human lice has remained challenging. In the first in vitro series of experiments, the lethal effect of the antibiotics doxycycline, erythromycin, rifampicin and azithromycin on lice was observed compared with the control group at 18 days, associated with a decrease in the production of nits. Of the four antibiotics tested, doxycycline appeared to be more effective than rifampicin, azithromycin and erythromycin, with a variable survival time according to treatment group (Fig. 2a; Supplementary Table S1). Recently, the in vitro efficacy of doxycycline at different doses on body lice was demonstrated [10]. In light of these results, we can suggest that the use of antibiotics (doxycycline, rifampicin, azithromycin and erythromycin) alone or in combination could be an efficient way to develop new drugs and to overcome drug resistance in lice.

Several decades ago it was demonstrated that body lice depend on obligate endosymbionts to supplement their nutritionally-deficient blood diet [26]. However, they maintain organs called mycetomes that house the primary endosymbiont [27]. Thus, through FISH analysis we were able to demonstrate that damage to the symbiont bacteria caused the death of their host as confirmed



**Fig. 3.** Images of lice by confocal microscopy through the technique of spectral deconvolution: (a) louse treated with doxycycline 20 µg/mL taken at Day 10 showing lower bacterial fluorescence; (b) living control showing higher bacterial fluorescence; (c) dead control taken at Day 10 showing lower bacterial fluorescence; and (d) louse tested with negative probe showing no bacterial fluorescence.

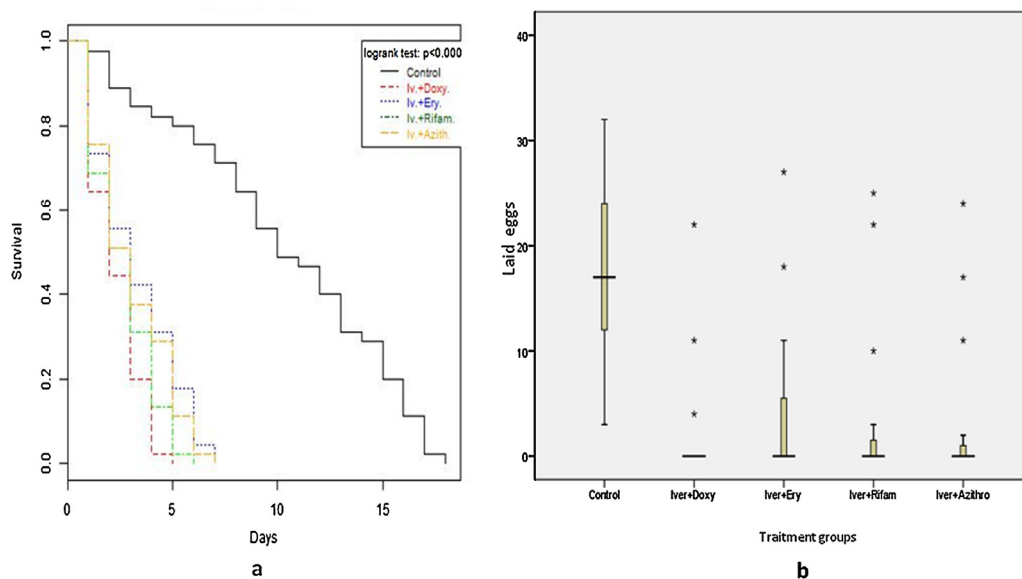


**Fig. 4.** Evolution trends in the ivermectin alone treatment group. (a) Kaplan–Meier curves illustrating survival of lice treated with ivermectin (Iv) in comparison with the control group. (b) Mean ( $\pm 95\%$  CI) number of eggs laid per day in the ivermectin (Iver)-treated group and the control group. CI, confidence interval.

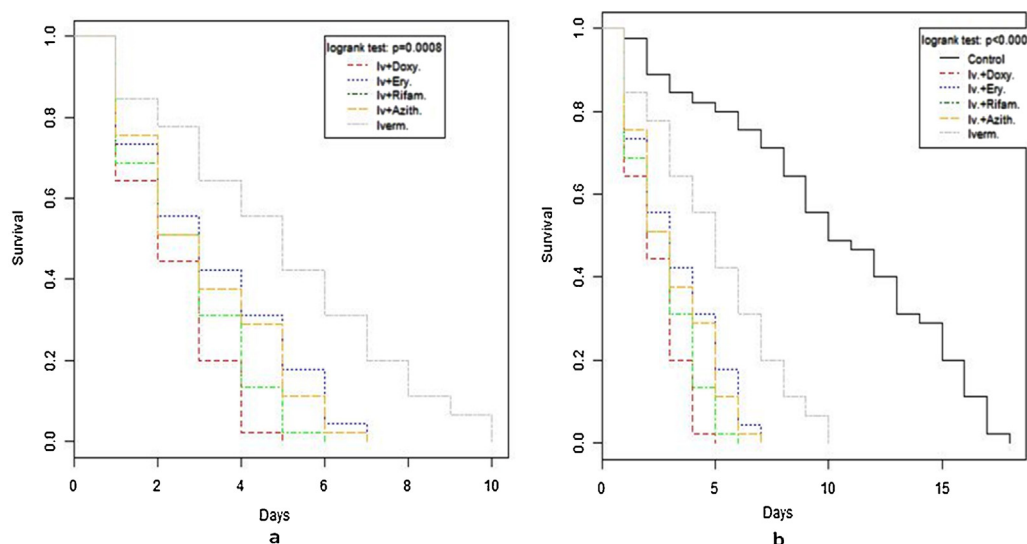
by a decrease in bacterial fluorescence (Fig. 3a). These results corroborate our previous work showing that doxycycline may affect the endosymbionts of lice through the mycetome switch, thus causing death of the louse [10].

Despite the introduction of some products such as organophosphates, pyrethrins and pyrethroids, insecticide resistance remains an increasing problem in many insect vectors of disease [14], especially among lice. Expression levels of ABC transporters have also been associated with drug resistance in arthropods and nematodes. Overexpression of these ABC transporters is associated with ivermectin resistance in *Haemonchus contortus* [16], *Caenorhabditis elegans* [17], *P. h. humanus* [28], *Sarcoptes scabiei* [29] and

*R. microplus* [18]. In a cohort of homeless subjects in Marseilles (France), a prevalence of body lice of 85% was observed [30]. This infestation was reduced temporarily to 19% with three doses of oral ivermectin administered at 7-day intervals. However, treatment failure or early recrudescence is common following ivermectin alone, and multiple doses are usually required to achieve a cure [31]. Moreover, in their randomised trial Chosidow et al. suggest that ivermectin could be an alternative treatment [32], but its transient effect was also demonstrated in some studies [3]. The findings of the current study show the transient effect of ivermectin alone on lice with a monitoring time of 10 days, associated with a slow decrease in the production of nits (Fig. 4; Supplementary Table S2).



**Fig. 5.** Evolution trends in the antibiotic–ivermectin combination treatment groups. (a) Kaplan–Meier curves illustrating survival of lice treated with the antibiotic–ivermectin combinations in comparison with the control group. (b) Mean ( $\pm 95\%$  CI) number of eggs laid per day in the combination treatment groups and the control group. CI, confidence interval; Iv/Iver, ivermectin; Doxy, doxycycline; Ery, erythromycin; Rifam, rifampicin; Azith/Azithro, azithromycin.



**Fig. 6.** Evolution trends in the antibiotic–ivermectin combination treatment groups and the ivermectin alone treatment group. (a) Kaplan–Meier curves illustrating survival of lice treated with antibiotic–ivermectin combinations and with ivermectin alone. (b) Kaplan–Meier curves illustrating the comparison of overall survival of lice in the antibiotic–ivermectin combination groups, the ivermectin alone group and the control group. Iv/Iverm, ivermectin; Doxy, doxycycline; Ery, erythromycin; Rifam, rifampicin; Azith, azithromycin.

Thus, in light of these various therapeutic failures, we may conclude that ivermectin could be useful with other drugs of different classes especially antibiotics, in the control of body lice infestation.

The efficacy of drug combination with ivermectin has been demonstrated in nematodes [33] and ectoparasites [34]. For ivermectin, it was suggested that head lice may be more susceptible than body lice; this could be justified by the model of Mumcuoglu et al., who observed high mortality of nymphs and female body lice fed on rabbits treated with 200  $\mu\text{g}/\text{kg}$  ivermectin for 3 days, followed by a sharp decline reaching the level of the controls on Day 6 [35]. In light of the various therapeutic failures described in lice, we decided test antibiotic combinations with ivermectin. In the second series of in vitro experiments, we were able to demonstrate the antiparasitic synergistic activity of a combination of several drugs. The monitoring time ranged from 5 days to 7 days in the

combination group, whilst that of ivermectin alone was 10 days (Supplementary Table S2). Interestingly, the difference in overall survival of lice in the combination treatment groups compared with ivermectin alone was also significant. Thus, these results highlight the efficacy of drug combinations compared with ivermectin alone. In the literature, compounds including antibiotics have been shown to increase intracellular concentrations of macrocyclic lactones [36]. In addition, it was shown that drug combinations including ivermectin provide antifilarial activity with ancillary benefits on intestinal helminths and ectoparasites such as chiggers and lice [34]. When doxycycline was combined with macrocyclic lactones in adult worms, ivermectin efficacy was ca. 80% compared with 9% when treatment was performed with doxycycline alone [37]. Hoerauf et al. recommend concurrent administration of ivermectin with doxycycline in the treatment of *Onchocerca* worms carrying

*Wolbachia* endobacteria [38], and later Coulibaly et al. found a significant reduction of circulating antigen levels of *Wuchereria bancrofti* in groups treated with a combination of doxycycline plus ivermectin for 12 months [39]. The effect of this combination has also been confirmed in dogs naturally infected with *Dirofilaria immitis* [40]. Thus, the current results are in agreement with the literature, especially that of Guo et al., which identifies for the first time that avermectin (macrocyclic lactone derivative including ivermectin) could act synergistically with meticillin (an antibiotic) to kill bacteria (MRSA) [23]. In addition, the current study demonstrates the first synergistic association of these drugs in vitro against body lice. We believe that the synergistic effect of combinations of antibiotics plus ivermectin could be used to achieve complete eradication of lice and to avoid the selection of a resistant population of lice.

## 5. Conclusion

The present study demonstrates a synergistic effect of antibiotics combined with ivermectin to even more effectively control lice. Thus, it would be interesting to develop this model in vivo in the control of lice infestation in humans.

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**Competing interests:** None declared.

**Ethical approval:** Ethical approval for the use of human blood in vitro was obtained from the Laboratory Research Ethics Board of Molecular Hematology, French Blood Service (EFS).

**Patent:** Title: 'Composed and combination of compounds for the treatment of lice'. Year of submission: 2014.

## References

- [1] Raoult D, Roux V. The body louse as a vector of reemerging human diseases. *Clin Infect Dis* 1999;29:888–911.
- [2] Meinking TL, Taplin D. Infestations: pediculosis. *Curr Probl Dermatol* 1996;24:157–63.
- [3] Brouqui P, Stein A, Dupont HT, Gallian P, Badiaga S, Rolain JM, et al. Ectoparasitism and vector-borne diseases in 930 homeless people from Marseilles. *Medicine (Baltimore)* 2005;84:61–8.
- [4] Cutler SJ, Abdissa A, Trape JF. New concepts for the old challenge of African relapsing fever borreliosis. *Clin Microbiol Infect* 2009;15:400–6.
- [5] Piarroux R, Abedi AA, Shako JC, Kebela B, Karhemere S, Diatta G, et al. Plague epidemics and lice, Democratic Republic of the Congo. *Emerg Infect Dis* 2013;19:505–6.
- [6] Raoult D, Ndiokubwayo JB, Tissot-Dupont H, Roux V, Faugere B, Abegbinni R, et al. Outbreak of epidemic typhus associated with trench fever in Burundi. *Lancet* 1998;352:353–8.
- [7] Butler T, Jones PK, Wallace CK. *Borrelia recurrentis* infection: single-dose antibiotic regimens and management of the Jarisch–Herxheimer reaction. *J Infect Dis* 1978;137:573–7.
- [8] Ohl ME, Spach DH. *Bartonella quintana* and urban trench fever. *Clin Infect Dis* 2000;31:131–5.
- [9] Raoult D, Drancourt M. Antimicrobial therapy of rickettsial diseases. *Antimicrob Agents Chemother* 1991;35:2457–62.
- [10] Sangaré AK, Boutellis A, Drali R, Audoly G, Weber P, Rolain JM, et al. Doxycycline kills human lice through its activity on their bacterial symbiont. *Int J Antimicrob Agents* 2015;45:675–6.
- [11] Gratz NG. Treatment resistance in louse control. In: Orkin M, Maibach H, editors. *Cutaneous infestations and insect bites*. New York, NY: Marcel Dekker; 1985. p. 219–30.
- [12] Hurlbut HS, Altman RM, Nibley Jr C. DDT resistance in Korean body lice. *Science* 1952;115:11–2.
- [13] Kitaoka M. DDT-resistant louse in Tokyo. *Jpn J Med Sci Biol* 1952;5:75–88.
- [14] Hemingway J, Ranson H. Insecticide resistance in insect vectors of human disease. *Annu Rev Entomol* 2000;45:371–91.
- [15] Dourmishiev AL, Dourmishiev LA, Schwartz RA. Ivermectin: pharmacology and application in dermatology. *Int J Dermatol* 2005;44:981–8.
- [16] Xu M, Molento M, Blackhall W, Ribeiro P, Beech R, Prichard R. Ivermectin resistance in nematodes may be caused by alteration of P-glycoprotein homolog. *Mol Biochem Parasitol* 1998;91:327–35.
- [17] James CE, Davey MW. Increased expression of ABC transport proteins is associated with ivermectin resistance in the model nematode *Caenorhabditis elegans*. *Int J Parasitol* 2009;39:213–20.
- [18] Pohl PC, Klafke GM, Carvalho DD, Martins JR, Daffre S, da Silva Vaz Jr I, et al. ABC transporter efflux pumps: a defense mechanism against ivermectin in *Rhipicephalus (Boophilus) microplus*. *Int J Parasitol* 2011;41:1323–33.
- [19] Lasota JA, Dybas RA. Avermectins, a novel class of compounds: implications for use in arthropod pest control. *Annu Rev Entomol* 1991;36:91–117.
- [20] Perez-Cogollo LC, Rodriguez-Vivas RI, Ramirez-Cruz GT, Miller RJ. First report of the cattle tick *Rhipicephalus microplus* resistant to ivermectin in Mexico. *Vet Parasitol* 2010;168:165–9.
- [21] Scott JG. Cross-resistance to the biological insecticide abamectin in pyrethroid-resistant house flies. *Pestic Biochem Physiol* 1989;34:27–31.
- [22] Currie BJ, Harumal P, McKinnon M, Walton SF. First documentation of in vivo and in vitro ivermectin resistance in *Sarcoptes scabiei*. *Clin Infect Dis* 2004;39:e8–12.
- [23] Guo H, Ren B, Dai H, Dai S, Zhang Y, Liu Y, et al. Reversal of meticillin resistance in *Staphylococcus aureus* by the anthelmintic avermectin. *Int J Antimicrob Agents* 2014;44:274–6.
- [24] Kirkness EF, Haas BJ, Sun W, Braig HR, Perotti MA, Clark JM, et al. Genome sequences of the human body louse and its primary endosymbiont provide insights into the permanent parasitic lifestyle. *Proc Natl Acad Sci U S A* 2010;107:12168–73.
- [25] Culpepper GH. Rearing and maintaining a laboratory colony of body lice on rabbits. *Am J Trop Med Hyg* 1948;28:499–504.
- [26] Buchner P. Symbiosis in luminous animals. In: Buchner P, editor. *Endosymbiosis of animals with plant microorganisms*. New York, NY: Interscience Publishers; 1965. p. 543–71.
- [27] Perotti MA, Allen JM, Reed DL, Braig HR. Host–symbiont interactions of the primary endosymbiont of human head and body lice. *FASEB J* 2007;21:1058–66.
- [28] Yoon KS, Strycharz JP, Baek JH, Sun W, Kim JH, Kang JS, et al. Brief exposures of human body lice to sublethal amounts of ivermectin over-transcribes detoxification genes involved in tolerance. *Insect Mol Biol* 2011;20:687–99.
- [29] Mounsey KE, Pasay CJ, Arlian LG, Morgan MS, Holt DC, Currie BJ, et al. Increased transcription of glutathione S-transferases in acaricide exposed scabies mites. *Parasit Vectors* 2010;3:43.
- [30] Foucault C, Ranque S, Badiaga S, Rovey C, Raoult D, Brouqui P. Oral ivermectin in the treatment of body lice. *J Infect Dis* 2006;193:474–6.
- [31] Huffam SE, Currie BJ. Ivermectin for *Sarcoptes scabiei* hyperinfestation. *Int J Infect Dis* 1998;2:152–4.
- [32] Chosidow O, Giraudeau B, Cottrell J, Izri A, Hofmann R, Mann SG, et al. Oral ivermectin versus malathion lotion for difficult-to-treat head lice. *N Engl J Med* 2010;362:896–905.
- [33] Passeri B, Vismarra A, Cricri G, Bazzocchi C, Kramer L, Bacci C. The adulticide effect of a combination of doxycycline and ivermectin in *Dirofilaria immitis*-experimentally infected dogs is associated with reduction in local T regulatory cell populations. *Vet Parasitol* 2014;205:208–10.
- [34] Munirathinam A, Sunish IP, Rajendran R, Tyagi BK. Impact of ivermectin drug combinations on *Pediculus humanus capitis* infestation in primary schoolchildren of south Indian rural villages. *Int J Dermatol* 2009;48:1201–5.
- [35] Mumcuoglu KY, Miller J, Rosen LJ, Galun R. Systemic activity of ivermectin on the human body louse (Anoplura: Pediculidae). *J Med Entomol* 1990;27:72–5.
- [36] Dupuy J, Lespine A, Sutra JF, Alvinerie M. Fumagillin, a new P-glycoprotein-interfering agent able to modulate moxidectin efflux in rat hepatocytes. *J Vet Pharmacol Ther* 2006;29:489–94.
- [37] Menozzi A, Bertini S, Turin L, Serventi P, Kramer L, Bazzocchi C. Doxycycline levels and anti-*Wolbachia* antibodies in sera from dogs experimentally infected with *Dirofilaria immitis* and treated with a combination of ivermectin/doxycycline. *Vet Parasitol* 2015;209:281–4.
- [38] Hoerauf A, Mand S, Volkmann L, Büttner M, Marfo-Debrekyei Y, Taylor M, et al. Doxycycline in the treatment of human onchocerciasis: kinetics of *Wolbachia* endobacteria reduction and of inhibition of embryogenesis in female *Onchocerca* worms. *Microbes Infect* 2003;5:261–73.
- [39] Coulibaly YI, Dembele B, Diallo AA, Lipner EM, Doumbia SS, Coulibaly SY, et al. A randomized trial of doxycycline for *Mansonella perstans* infection. *N Engl J Med* 2009;361:1448–58.
- [40] Grandi G, Quintavalla C, Mavropoulou A, Genchi M, Gnudi G, Bertoni G, et al. A combination of doxycycline and ivermectin is adulticidal in dogs with naturally acquired heartworm disease (*Dirofilaria immitis*). *Vet Parasitol* 2010;169:347–51.