Yeast of the Cryptococcus genus are widely distributed in the biosphere, ranging from terrestrial to marine habitats (1). Sequence tag-based analysis of deep-sea microeukaryotes occurring in deep-sea habitats revealed Cryptococcus as highly abundant taxa in cold methane seep (2), the deep-sea floor (3, 4), and the deep subseafloor (5, 6). A recent RNA-based approach suggested that this phylotype was also metabolically active in deep sedimentary ecosystems (5).

Here, we describe the draft genome sequence of Cryptococcus sp. strain Mo29 (= UB0CC 208024), a putative producer of original bioactive compounds. High-quality genomic DNA was extracted using the hexadecyltrimethylammonium bromide (CTAB) and Genomic-tip (Qiagen) methods. Genomic DNA of Cryptococcus sp. Mo29 was used to generate shotgun and mate-pair libraries with insert sizes of approximately 350 bp and 8 kb, respectively. A shotgun library was made using the TruSeq DNA PCR-free sample preparation kit and a gel-plus mate-paired-end library was generated with the Nextera mate-pair sample preparation kit. Genome sequencing was performed using Illumina HiSeq 2500 sequencing technologies.

The shotgun library produced 41,481,610 reads. The mate-pair library produced 15,799,276 reads. After quality filtering of reads with more than 90% of bases with base quality greater than or equal to Q20, a total of 28,246,332 shotgun reads (2,852.48 Mb) and 7,870,684 mate-pair reads (732 Mb) were retained. The shotgun library produced 41,481,610 reads. The mate-pair library produced 15,799,276 reads. After quality filtering of reads with more than 90% of bases with base quality greater than or equal to Q20, a total of 28,246,332 shotgun reads (2,852.48 Mb) and 7,870,684 mate-pair reads (732 Mb) were retained. The ALLPATHS-LG whole-genome shotgun assembler (7) was used for the creation of the de novo genome assembly from these short reads. The assembly contained a total of 174 scaffolds with an average read length of 144,951 bp. The N50 was 820 kb, and the maximum contig length was 1,690 kb. The total sequence length of the resulting draft genome was 27,528,793 bp with an overall GC content of 50.09%. A total of 174,473 bp were repeats, which constituted 0.63% of the assembled genome size as predicted by repeat masker tool (http://www.repeatmasker.org/). There are 10,009 genes in this fungal genome, as predicted by Augustus 3.0 (8). Our BLAST2GO-based annotation analyses (9) have revealed 57.5% annotated genes, while 42.5% remained unannotated.

The genome analysis of secondary metabolite biosynthesis gene clusters using antiSMASH 3.0 software (10) highlighted the presence of four biosynthetic gene clusters, which included one type III polyketide synthase and one terpene synthase and two clusters that were unknown (also called as others in AntiSMASH annotation). Analysis of the sequence with the CAZy database (11) identified 860 genes with activity involving carbohydrates, including 396 glycoside hydrolases, 130 glycosyltransferases, 31 polysaccharide lyases, 63 carbohydrate esterases, 99 carbohydrate-binding modules, and 141 auxiliary activities. Particularly noteworthy is the finding that Cryptococcus sp. Mo29 is equipped with all necessary enzymes for the deconstruction of terrestrial plant cell walls, including cellulases, xylanases, pectinases and ligninases. The finding of numerous genes associated with synthesis of secondary metabolites and CAZyme in the genome sequence of Cryptococcus sp. Mo29 suggests that further investigation will result in the discovery of useful gene products that may be exploited for biotechnological application.

**Nucleotide sequence accession numbers.** The nucleotide sequence of the Cryptococcus sp. UB0CC 208024 (Mo29) genome is deposited in DDBJ/EMBL/GenBank under accession numbers FKKD01000001 to FKKD01000687. This paper describes the first version of the genome.

**ACKNOWLEDGMENTS**

We thank Macrogen Inc. (South Korea) for genome sequencing and assembly.

This study was supported by a grant from the UE project MaGuMBA (Marine microorganisms: Cultivation Methods for Improv-
ing their Biotechnological Applications, FP7, grant agreement 311975, Brussels, Belgium).

**FUNDING INFORMATION**

This work, including the efforts of Vanessa Rédou, was funded by EC Seventh Framework Programme (FP7) (311975).

**REFERENCES**


