



HAL
open science

Efflux Pump Blockers in Gram-Negative Bacteria: The New Generation of Hydantoin Based-Modulators to Improve Antibiotic Activity

Ewa Otrebska-Machaj, Jacqueline Chevalier, Jadwiga Handzlik, Ewa Szymńska, Jakub Schabikowski, Jean-Michel Bolla, Gérard G Boyer, Katarzyna Kięc-Kononowicz, Jean-Marie J Pagès, Sandrine Alibert

► To cite this version:

Ewa Otrebska-Machaj, Jacqueline Chevalier, Jadwiga Handzlik, Ewa Szymńska, Jakub Schabikowski, et al.. Efflux Pump Blockers in Gram-Negative Bacteria: The New Generation of Hydantoin Based-Modulators to Improve Antibiotic Activity. *Frontiers in Microbiology*, 2016, 7, pp.622. 10.3389/fmicb.2016.00622 . hal-01463003

HAL Id: hal-01463003

<https://amu.hal.science/hal-01463003>

Submitted on 16 Feb 2017

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Efflux Pump Blockers in Gram-Negative Bacteria: The New Generation of Hydantoin Based-Modulators to Improve Antibiotic Activity

Ewa Otrębska-Machaj^{1,2}, Jacqueline Chevalier², Jadwiga Handzlik¹, Ewa Szymańska¹, Jakub Schabikowski¹, Gérard Boyer², Jean-Michel Bolla², Katarzyna Kieć-Kononowicz¹, Jean-Marie Pagès^{2*} and Sandrine Alibert²

OPEN ACCESS

Edited by:

Ivan Rychlik,
Veterinary Research Institute,
Czech Republic

Reviewed by:

Helen Zgurskaya,
University of Oklahoma, USA
Alessandra Polissi,
Università degli Studi di
Milano-Bicocca, Italy
Kunihiko Nishino,
The Institute of Scientific and
Industrial Research, Osaka University,
Japan

*Correspondence:

Jean-Marie Pagès
jean-marie.pages@univ-amu.fr

Specialty section:

This article was submitted to
Antimicrobials, Resistance
and Chemotherapy,
a section of the journal
Frontiers in Microbiology

Received: 07 January 2016

Accepted: 15 April 2016

Published: 03 May 2016

Citation:

Otrębska-Machaj E, Chevalier J,
Handzlik J, Szymańska E,
Schabikowski J, Boyer G, Bolla J-M,
Kieć-Kononowicz K, Pagès J-M
and Alibert S (2016) Efflux Pump
Blockers in Gram-Negative Bacteria:
The New Generation of Hydantoin
Based-Modulators to Improve
Antibiotic Activity.
Front. Microbiol. 7:622.
doi: 10.3389/fmicb.2016.00622

¹ Department of Technology and Biotechnology of Drugs, Medical College, Jagiellonian University, Krakow, Poland,

² UMR-MD1, Aix Marseille Université/IRBA, Facultés de Médecine et de Pharmacie, Marseille, France

Multidrug resistant (MDR) bacteria are an increasing health problem with the shortage of new active antibiotic agents. Among effective mechanisms that contribute to the spread of MDR Gram-negative bacteria are drug efflux pumps that expel clinically important antibiotic classes out of the cell. Drug pumps are attractive targets to restore the susceptibility toward the expelled antibiotics by impairing their efflux activity. Arylhydantoin derivatives were investigated for their potentiation of activities of selected antibiotics described as efflux substrates in *Enterobacter aerogenes* expressing or not AcrAB pump. Several compounds increased the bacterial susceptibility toward nalidixic acid, chloramphenicol and sparfloxacin and were further pharmacomodulated to obtain a better activity against the AcrAB producing bacteria.

Keywords: antibiotics, AcrAB pump, efflux pump blocker, hydantoin, multidrug resistance

INTRODUCTION

Multidrug resistance (MDR) described in Gram-negative bacteria is continuously emerging as a prominent worldwide health concern (Chopra et al., 2008; Laxminarayan et al., 2008; Gandhi et al., 2010). One of the most contributing mechanisms is the overexpression of efflux pumps that are involved in bacterial survival, colonization and virulence (Delmar et al., 2014; Blair et al., 2015; Li et al., 2015; Venter et al., 2015; Davin-Regli et al., 2016). Several efflux pump superfamilies, e.g., major-facilitator (MF), multi-drug and toxic efflux (MATE), ATP-binding cassette (ABC), small multidrug resistance (SMR), resistance-nodulation-division (RND) transporters have been classified^{1,2} and extensively described in well-documented reviews: they differ by their functional structure and organization, their subcellular location inside the bacterium, their energy source (e.g., membrane potential for RND or ATP for ABC) and the involvement of a coupled antiport during the antibiotic expulsion (e.g., proton for AcrAB pump; Delmar et al., 2014; Blair et al., 2015; Li et al., 2015). The overexpression of Gram-negative efflux pumps, especially those belonging to the

¹<http://www.membranetransport.org/>

²<http://www.tcd.org/>

RND family, is now well-described in resistant isolates (Nikaido and Pagès, 2012; Li et al., 2015). This contributes to the acquisition of additional mechanisms of resistance including the mutation in antibiotic targets (e.g., mutation in gyrase/topoisomerase for quinolone) or the production of enzymes that degrade antibiotics (e.g., β -lactamases) and this can be associated or not with the alteration of the outer membrane permeability (Davin-Regli et al., 2008; Pagès et al., 2008). Regarding resistant clinical isolates of Gram-negative bacteria, the archetype of the drug active transporter system is the AcrAB-TolC/MexAB-OprM efflux pumps (Nikaido and Pagès, 2012; Li et al., 2015). The structures of components of efflux systems belonging to RND group, have been solved by X-ray crystallography and models of the pump assembly have been obtained (Yao et al., 2010; Du et al., 2014). The structure and function of the RND efflux pumps must be molecularly deciphered thus allowing the rational design and the synthesis of new compounds to combat MDR. The broad selectivity of efflux pumps makes difficult the identification of precise pharmacophoric groups at the drug surface. However, efflux pumps are attractive target by blocking this efflux mechanism in order to restore the intracellular concentration of antibacterial agents (Bolla et al., 2011; Ruggerone et al., 2013; Dreier and Ruggerone, 2015; Opperman and Nguyen, 2015). Recent computer docking analyses have produced some information about the involvement of certain amino acid residues, but clearly more chemical and biological information are needed to improve models (Schulz et al., 2010; Fischer et al., 2014). This is a 'key' point not only regarding the mode of action and dynamics of the process but also regarding the clinical impact of the design of new antibacterial agents: this last aspect is illustrated by the β -lactamases inhibitors currently used today (Bolla et al., 2011; Chen et al., 2013; Pucci and Bush, 2013).

To enhance the activity of old antibiotics by targeting resistance mechanisms in clinical resistant isolates, they can be combined with adjuvant molecules such as chemosensitizers (e.g., membrane permeabilizer or efflux inhibitor; Jones, 2010; Bolla et al., 2011). In addition, these types of transporter inhibitors may impair the activity of efflux pumps and thus reduce bacterial colonization and virulence (Venter et al., 2015; Davin-Regli et al., 2016). It must be noted that the efficacy of the inhibitors depends on their affinity for transporter binding sites (compared to the antibiotic) and their internal concentration close to the efflux pump. Consequently, due to these parameters associated with penetration rate and affinity for pump sites, some discrepancies can be observed in the level of the internally accumulated antibiotics depending on the bacterial backgrounds (Kaščáková et al., 2012; Cinquin et al., 2015).

Recently, two generations of hydantoin derivatives have been identified as AcrAB-TolC inhibitors with the *Enterobacter aerogenes* CM 64 strain that overproduces AcrAB (Handzlik et al., 2011). Compounds showing chemosensitizing effect on nalidixic acid activity were the starting point for new pharmacomodulations carried out in this study to obtain a new generation of products with an improved activity.

Moreover, tests were extended to other chemically unrelated antibiotics, chloramphenicol and sparflaxacin, for which the antibacterial activity decreased together with the emergence of multidrug-resistant strains. Several chemical derivatives were synthesized to define pharmacophoric groups important for restoring the activity of antibiotics in AcrAB active bacteria.

MATERIALS AND METHODS

Bacterial Strains

All generations of hydantoin derivatives were tested against two strains of *E. aerogenes*: the reference strain, ATCC 13048 (basal efflux producer) and its derivative strain, CM 64, a chloramphenicol-selected resistant strain that over-produces the AcrAB-TolC EPs (Ghisalberti et al., 2005). In addition, two isogenic strains EA289 (a clinical MDR strain that overproduced the AcrAB pump) and its EA294 derivative (an *acrAB* knockout strain) were used (Pradel and Pagès, 2002). These two strains contain additional mechanisms of antibiotic resistance such as β -lactamases, targets mutations, etc. (Malléa et al., 1998; Pradel and Pagès, 2002; Kaščáková et al., 2012).

Chemicals

Hydantoin derivatives used in the pharmacological assays were obtained by chemical synthesis. The new generations IIIA and B were obtained using 3–4-step synthesis (Handzlik et al., 2014; Matys et al., 2015; **Figure 1** and Supplementary data). Purity and identity of new compounds were confirmed using spectral analysis (H-NMR, IR), elemental analysis and melting point measurements. Phenylalanine-Arginine β -naphthylamide (PA β N, dihydrochloride, Sigma) previously described as efflux pump inhibitor was used as reference (Bolla et al., 2011; Misra et al., 2015).

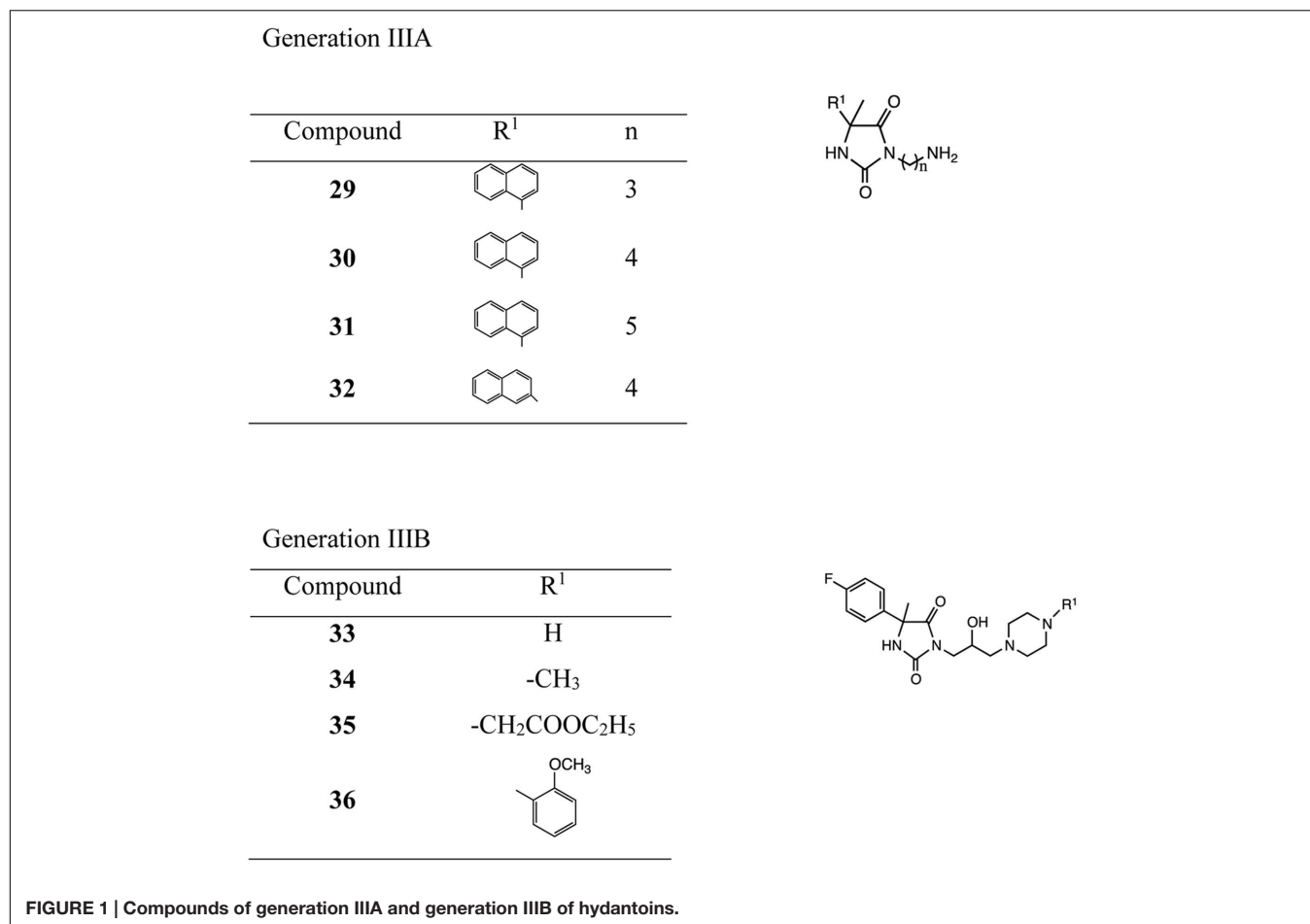
Compound Susceptibility Assays

Susceptibilities of ATCC 13048, CM 64, EA294, and EA289 were determined by using the twofold standard microbroth dilution method (microplates and automatic analyses Tecan[®]; CLSI³). Approximately, 10⁵ CFU (colony forming unit) were inoculated in 200 μ l of Mueller-Hinton II broth (MH II broth cation adjusted, Becton, Dickinson & Company) containing twofold serial dilutions of the targeted molecule. Experiments were performed in triplicate for each compound and each antibiotic. Results were estimated visually after 18 h incubation at 37°C (Philippe et al., 2015).

Antibiotic Susceptibility Potentiating Assays

To assay the possible chemosensitizing activity of compounds, serial dilutions of antibiotics, nalidixic acid (NAL, Sigma), chloramphenicol (CHL, Sigma), doxycycline (DOX, hyclate, Sigma), erythromycin (ERY, lactobionate, AMDIPHARM) and

³<http://clsi.org/>



sparfloxacin (SPX, Sigma), were incubated in the absence or in the presence of compounds. The antibiotics, NAL, CHL, and SPX are substrates of the AcrAB-TolC efflux pump as demonstrated by the increased MIC values obtained in CM 64 strain overexpressing the AcrAB-TolC pump (Table 1)

TABLE 1 | Susceptibility (MIC) of the *Enterobacter aerogenes* reference strain ATCC 13048, the derivative strain CM 64 overexpressing AcrAB-TolC pump, Ea289 overproducing the AcrAB pump and its derivative Ea294 (an *acrAB* knockout strain) to the different compounds belonging to various generations of hydantoin and to nalidixic acid (NAL), chloramphenicol (CHL), sparfloxacin (SPX), doxycycline (DOX), and erythromycin (ERY).

Compound	MIC [mM] ATCC 13048	MIC [mM] CM 64	MIC [mM] Ea294	MIC [mM] Ea289
29–36	>2	>2	>1	>1
NAL	0.034	0.55 (16)*	4.4	>17.6
CHL	0.012	0.79 (66)	0.2	3.2
SPX	0.00015	0.0025 (17)	>2	>2
DOX	0.002	0.07 (35)	0.002	0.07
ERY	0.35	0.7 (2)	0.087	0.35
PAβN	5	5 (1)	0.0625	3.5

*MIC ratio: efflux overproducer/basal producer strains.

compared to the reference one ATCC 13048. Thus we are able to hypothesize that an efflux blocker may reduce the antibiotic MIC in efflux producer strain. Generations IIIA and IIIB of hydantoin derivatives were tested at a concentration of 0.5 mM according to the intrinsic antibacterial activity of each compound (corresponding to the value of MIC/4). To facilitate the comparison of activity and the performance of a rational SAR analysis, they were additionally tested at the concentration corresponding to that of the best first generation of chemosensitizers (0.0625 mM; Handzlik et al., 2011). PAβN, the reference inhibitor for AcrAB pump, was used at 0.050 mM. Control experiments were carried out without compounds. Experiments were performed in triplicate for each antibiotic, each strain and each condition (without and with compound). The results were assessed after 18 h at 37°C and were presented by using the activity gain parameter A, the ratio of the MIC of the antibiotic (determined in the absence of compound) to its MIC in the presence of the compound.

Determination of the FIC Index

To determine the fractional inhibitory concentration index (FIC_i), a two dimensional checkerboard with twofold dilutions of each compound was set up for the study (Berenbaum, 1978;

TABLE 2 | Effect of the hydantoin derivatives on the susceptibility level of *E. aerogenes* ATCC 13048 and CM 64 strains to nalidixic acid (NAL), chloramphenicol (CHL), and sparfloxacin (SPX).

Compound	Concentration [mM]	A _{NAL}		A _{CHL}		A _{SPX}	
		ATCC 13048	CM 64	ATCC 13048	CM 64	ATCC 13048	CM 64
29	0.0625	1	1	1	1	2	2
30	"	2	2	2	1	1	1
31	"	2	2	2	1	1	2
32	"	2	4	1	1	1	2
33	"	1	1	1	1	1	1
34	"	1	0.5	0.5	1	0.5	1
35	"	1	1	0.5	1	0.5	0.5
36	"	1	1	1	1	0.5	0.5
29	0.5	8	4	2	4	4	2
30	"	8	4	4	4	8	4
31	"	16	4	4	4	4	4
32	"	16	32	4	32	8	8
33	"	2	2	1	2	2	1
34	"	2	1	1	1	2	2
35	"	2	1	2	1	2	1
36	"	2	1	2	1	2	1
PAβN	0.05	64	128	2	64	8	32

"A" corresponds to the antibacterial activity gain obtained in the presence of the respective compound and evaluated for each antibiotic (A = MIC without compound/MIC with compound).

Allam et al., 2014). For the first clear well in each row of the microplate containing an antimicrobial agent, the FIC was calculated as follows: FIC of compound A (FIC A) = MIC of compound A in combination with B/MIC of compound A alone, and FIC of compound B (FIC B) = MIC of compound B in combination with A/MIC of compound B alone. The FIC_i was calculated as the sum of the FIC of each compound. The nature of the interaction was classified as follows: synergy FIC_i ≤ 0.5; additivity 0.5 < FIC_i ≤ 1; indifference 1 < FIC_i ≤ 2; and antagonism FIC_i > 2 (European Committee for Antimicrobial Susceptibility Testing (EUCAST) of the European Society of Clinical Microbiology and Infectious Diseases (ESCMID), 2000). For each combination, an isobologram, which graphically illustrates the interaction effect, was constructed: FIC A was showed in the abscissa and FIC B in the ordinate, the profile of the corresponding curve reflects the nature of the interaction.

RESULTS

Antibacterial Activity

It is important that putative inhibitors that could be used as "adjuvant molecule" for antibiotics, do not display a high intrinsic antibacterial activity (Davin-Regli et al., 2008; Bolla et al., 2011). The determination of the antibacterial activity for each compound was performed and presented in **Table 1**. Many of the compounds did not inhibit the growth of bacteria even at the highest tested concentration as the previous generations of hydantoin derivatives (Handzlik et al., 2011). The majority of compounds exhibited a MIC > 2 mM in ATCC13048

and in CM64 strains and MIC ≥ 1 mM in Ea289 and Ea294.

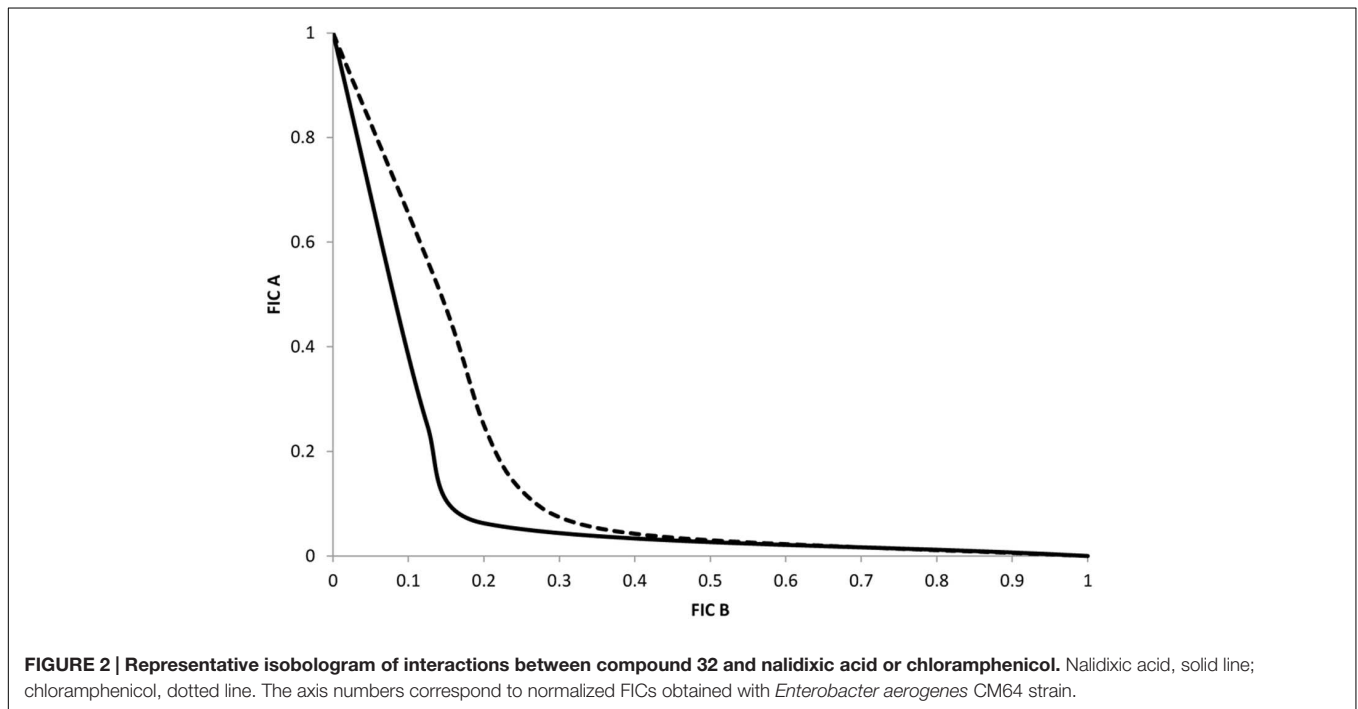
Influence on Antibiotic Susceptibility in ATCC 13048 and CM 64 Strains Effect on Nalidixic Acid Susceptibility

Table 2 presents the chemosensitizing effect of compounds on ATCC 13048 and CM 64 susceptibility to nalidixic acid (NAL). The gain on antibiotic activity (A) was calculated for each compound and each antibiotic. Regarding the CM 64 strain, a moderate or a weak effect was observed on the MIC when compounds were used at 0.0625 mM compared to PAβN. Compounds of generations IIIA (29–32) and IIIB (33–36) were used at a concentration of 0.0625 mM and 0.5 mM due to their higher MIC. A higher concentration of compounds of the generation IIIB did not improve the antibiotic activity whereas we observed a significant decrease of the antibiotic MIC, from 4- to 32-fold in CM 64 (see A_{NAL} in **Table 2**), in the case of compounds of generation IIIA (29–32). These compounds decreased the MIC of NAL from 8- to 16-fold comparing to the PAβN effect (64-fold) in the reference ATCC 13048 strain (see A_{NAL} in **Table 2**). In the case of PAβN and derivatives 32 the difference in activity in both tested strains was not significant (only a twofold stronger activity in the strain overexpressing the AcrAB pump than in the reference one) whereas compounds 29–31 showed a better action on antibiotic activity in the reference strain. It must be noted that the compound 32 used at increased concentration exhibits an activity profile similar to PAβN, which has been shown to be an efficient efflux inhibitor at low concentrations (Nikaido and Pagès, 2012; Misra et al., 2015).

TABLE 3 | Effect of the compounds of generation IIIA (29–32) depends on AcrAB context.

Compound	Ea294			Ea289			CM 64	
	A _{DOX}	A _{ERY}	A _{NAL}	A _{DOX}	A _{ERY}	A _{NAL}	A _{DOX}	A _{ERY}
29	2	1	2	8	1	1	8	1
30	4	4	4	16	1	2	8	1
31	4	8	8	16	2	2	8	1
32	2	2	4	32	4	>2	32	2

"A" corresponds to the antibacterial activity gain obtained in the presence of the respective compound and evaluated for each antibiotic ($A = \text{MIC without compound} / \text{MIC with compound}$). DOX, doxycycline; ERY, erythromycin; NAL, nalidixic acid.



Effect on Chloramphenicol Susceptibility

The results are presented in **Table 2**. In the case of the CM 64 strain, compounds tested at the concentration of 0.0625 mM showed a weak chemosensitizing effect on CHL antibacterial activity compared to PA β N. Compounds of generation IIIA (29–32) examined at a concentration of 0.5 mM decreased the chloramphenicol MIC from 2- to 4-fold in the reference strain and from 4- to 32-fold in the strain overexpressing efflux pump (see A_{CHL} in **Table 2**). We observed a noticeable increase in CHL susceptibility in the AcrAB overproducer CM 64, a 32-fold gain in the susceptibility with the compound 32 compared to compounds 29–31. In this assay, the chemosensitizing effect of compound 32 can be compared to PA β N which was much more active in the strain overexpressing the AcrAB pump than in the parental strain ATCC 13048.

Effect on Sparfloxacin Susceptibility

Table 2 presents the effects of hydantoin on sparfloxacin (SPX) susceptibility. In CM 64, the chemosensitizing effect of low concentrated hydantoin on SPX was as weak as we observed in

the case of CHL. Compounds of generation IIIA (29–32) tested at the highest concentration 0.5 mM caused a 4–8-fold decrease in the MIC in the ATCC 13048 strain and a 2–8-fold decrease in the MIC in CM 64 one. Comparing the results obtained for the active compounds (29–32) to the results obtained for PA β N (see A_{SPX} in **Table 2**) we observed that the action of hydantoin with SPX was less efficient than PA β N, which was more active in the CM64 strain overexpressing the AcrAB pump than in the reference strain ATCC13048. This may suggest a different conformational site for the two molecules, either for recognition or for binding step, in the AcrB monomer (Delmar et al., 2014; Du et al., 2014; Yamaguchi et al., 2015), inside the pump or a different mode of action on the resistance mechanism.

Effect of Compounds 29–32 on the Resistance Level in MDR *E. aerogenes* Strains

In order to evaluate the chemosensitizing effect of the compounds 29–32 (the most effective molecules) on the MDR

background, the Ea289 and Ea294 strains were assayed. It is important to mention that these strains contain various resistance mechanisms (Malléa et al., 1998; Pradel and Pagès, 2002) such as target mutations (e.g., mutations in QRDR region of gyrase that increase quinolone resistance) and expression of modifying enzymes (e.g., β -lactamases that contribute to β -lactam resistance). **Table 3** presents the activity gain parameter A tested with doxycycline (DOX), erythromycin (ERY), and nalidixic acid (NAL) in parental and AcrAB-derivative strain context. It is interesting to note that **32** exhibited significant restoring antibiotic activity with DOX in Ea289 and very weak action in Ea294 that is devoid of the AcrAB efflux pump components. In contrast, a weak chemosensitizing activity was observed with ERY, this effect could be caused by the presence of additional resistance mechanisms for the macrolide antibiotic class as previously reported (Chollet et al., 2004). Regarding the effect on NAL and sparfloxacin susceptibility, the mutations in the quinolone target (DNA gyrase) previously reported in Ea289, can explain the weak effect observed toward these strains (Pradel and Pagès, 2002; Kaščáková et al., 2012).

Determination of the FIC Index for Compound 32

To precise the type of interaction (synergistic, additive, or indifferent) between compound **32** and selected antibiotics, we carried out analysis based on the FIC_i as previously described. Combinations of compound **32** with NAL and CHL respectively were performed in CM 64 strain. This strain overexpresses the AcrAB pump and does not contain target mutation that can impair the effect of compounds on the restoration of antibiotic activity. The nature of the association was determined from the FIC_i average obtained from each combination and the representation was performed for each combination (**Figure 2**). In two cases, the isobologram curve obtained was concave indicating a synergy association between antibiotic and compound **32**. The synergistic association corresponds to an average of calculated FIC_i of about 0.32 for NAL and 0.44 for CHL respectively. These curves fitted well with the **Table 2** data.

DISCUSSION

The aim of this study was to identify compounds that are able to modulate the AcrAB pump activity and restore antibiotic activity on efflux producing strains.

The various molecules combined with NAL and CHL increased the susceptibility of the CM 64 strain and of the reference strain ATCC 13048 to antibiotics. On the one hand, we found compounds that showed stronger activity in the ATCC 13048 strain than in the CM 64 strain (**29–31** tested with NAL, **29, 30** tested with SPX). These findings could indicate in this case that tested compounds are not highly selective toward the AcrAB-TolC pump involved in the efflux of antibiotics. This suggests that they act not

only on the basal AcrAB expressing strain but also that they may recognize other bacterial targets. In contrast, PA β N combined with CHL and SPX as well as the 3-aminobutyl-5- β -naphthylhydantoin **32** tested with CHL exhibited a difference between the AcrAB overproducer strain and the parental one. This time these results suggest a significant selectivity of chemosensitizers for the AcrAB pump expressed in bacterial strains. Taking into account the influence of concentration of 3-aminoalkyl-5-naphthylhydantoins (generation IIIA) on the antibiotic activity, we observed that the higher is the concentration, the stronger is the chemosensitizing effect, not only on the CM 64 strain but also on ATCC 13048. These outcomes could suggest additional mechanisms of action besides the effect on the AcrAB-TolC efflux pump.

The comparison of the results in Ea289 and Ea294 (**Table 3**) indicates the correlation of the restoring activity of antibiotics with the presence of the AcrAB efflux pump. They also illustrate the capability of compound **32** to increase the antibacterial effect of DOX in these strains. The low effect of **32** on the ERY susceptibility when assayed in Ea289 and Ea294 can be due to additional pump, other than AcrB, active in these clinical derivative strains capable to expel this class of drugs (Chollet et al., 2004).

The different structural features of tested compounds allow showing how the nature and the position of their diverse molecular fragments modulate on the one hand their selectivity regarding AcrAB pump and on the other hand their chemosensitizing activity. The tested compounds exhibiting the common hydantoin scaffold were modified using two of diverse substituents at position 5 as well as amine substituents and a selection of linkers to bind an amine to the hydantoin core. The location of an amine-alkyl fragment at position N1 or N3 to an aryl fragment at position 5 was modified for comparing with the activity of the first generations of hydantoin synthesized (Handzlik et al., 2011). In the case of **32**, the hydrophilic primary amine fragment is focused by terminate location at the end of longer 3-hydantoin substitution, opposite to the 5- β -naphthyl one.

The most active compounds (**29–32**) share the same pharmacophore profile than PA β N suggesting a similar physicochemical outline for an identical target. The amphiphilic nature of the Generation IIIA of optimized derivatives seems to be crucial to inhibit antibiotic resistance mediated by efflux pump and open new ways for generating original active compounds able to inhibit pump activity. With the recent published data regarding piperazine arylideneimidazolone derivatives as potential efflux inhibitors in *Escherichia coli* cells (Bohnert et al., 2016), the compound **32** characterized in this study will be used for pharmacophoric modulations in order to develop more potent inhibitors.

AUTHOR CONTRIBUTIONS

JH, KK-K, J-MP, and SA designed research; EO-M, JC, ES, and SA performed research; JS, GB, and J-MB contributed new

reagents or analytic tools; EO-M, JH, KK-K, J-MP, and SA analyzed data; JH, KK-K, J-MP, and SA wrote the paper.

FUNDING

This work was supported by French Embassy in Poland, 757/N-POLONIUM/2010/0, by grant ANR-11-BS07-019-01 “IBEF” Agence Nationale de la Recherche (ANR, France) and partly supported by Polish statutory project K/ZDS/005593 and Aix-Marseille University and IRBA.

REFERENCES

- Allam, A., Maigre, L., Alimi, M., Alves de Sousa, R., Hessani, A., Galardon, E., et al. (2014). New peptides with metal binding abilities and their use as drug carriers. *Bioconjug. Chem.* 25, 1811–1819. doi: 10.1021/bc500317u
- Berenbaum, M. C. (1978). A method for testing for synergy with any number of agents. *J. Infect. Dis.* 137, 122–130. doi: 10.1093/infdis/137.2.122
- Blair, J. M., Webber, M. A., Baylay, A. J., Ogbolu, D. O., and Piddock, L. J. (2015). Molecular mechanisms of antibiotic resistance. *Nat. Rev. Microbiol.* 13, 42–51. doi: 10.1038/nrmicro3380
- Bohnert, J. A., Schuster, S., Kern, W. V., Karcz, T., Olejarz, A., Kaczor, A., et al. (2016). Novel piperazine arylideneimidazolones inhibit the AcrAB-TolC pump in *Escherichia coli* and simultaneously act as fluorescent membrane probes in a combined real-time influx and efflux assay. *Antimicrob. Agents Chemother.* 60, 1974–1983. doi: 10.1128/AAC.01995-15
- Bolla, J. M., Alibert-Franco, S., Handzlik, J., Chevalier, J., Mahamoud, A., Boyer, G., et al. (2011). Strategies for bypassing the membrane barrier in multidrug resistant Gram-negative bacteria. *FEBS Lett.* 585, 1682–1690. doi: 10.1016/j.febslet.2011.04.054
- Chen, J., Shang, X., Hu, F., Lao, X., Gao, X., Zheng, H., et al. (2013). β -Lactamase inhibitors: an update. *Mini Rev. Med. Chem.* 13, 1846–1861. doi: 10.2174/13895575113139990074
- Chollet, R., Chevalier, J., Bryskier, A., and Pagès, J. M. (2004). The AcrAB-TolC pump is involved in macrolide resistance but not in telithromycin efflux in *Enterobacter aerogenes* and *Escherichia coli*. *Antimicrob. Agents Chemother.* 48, 3621–3624. doi: 10.1128/AAC.48.9.3621-3624.2004
- Chopra, I., Schofield, C., Everett, M., O'Neill, A., Miller, K., Wilcox, M., et al. (2008). Treatment of health-care-associated infections caused by Gram-negative bacteria: a consensus statement. *Lancet Infect. Dis.* 8, 133–139. doi: 10.1016/S1473-3099(08)70018-5
- Cinquin, B., Maigre, L., Pinet, E., Chevalier, J., Stavenger, R. A., Mills, S., et al. (2015). Microspectrometric insights on the uptake of antibiotics at the single bacterial cell level. *Sci Rep.* 5, 17968. doi: 10.1038/srep17968
- Davin-Regli, A., Bolla, J. M., James, C. E., Lavigne, J. P., Chevalier, J., Garnotel, E., et al. (2008). Membrane permeability and regulation of drug ‘influx and efflux’ in enterobacterial pathogens. *Curr. Drug Targets.* 9, 750–759. doi: 10.2174/138945008785747824
- Davin-Regli, A., Masi, M., Bialek, S., Nicolas-Chanoine, M. H., and Pagès, J.-M. (2016). “Antimicrobial resistance and drug efflux pumps in *Enterobacter* and *Klebsiella*,” in *Efflux-Mediated Drug Resistance in Bacteria: Mechanisms, Regulation and Clinical Implications*, eds X.-Z. Li, C. A. Elkins, and H. I. Zgurskaya (Berlin: Springer).
- Delmar, J. A., Su, C. C., and Yu, E. W. (2014). Bacterial multidrug efflux transporters. *Annu. Rev. Biophys.* 43, 93–117. doi: 10.1146/annurev-biophys-051013-022855
- Dreier, J., and Ruggerone, P. (2015). Interaction of antibacterial compounds with RND efflux pumps in *Pseudomonas aeruginosa*. *Front Microbiol.* 6:660. doi: 10.3389/fmicb.2015.00660
- Du, D., Wang, Z., James, N. R., Voss, J. E., Klimont, E., Ohene-Agyei, T., et al. (2014). Structure of the AcrAB-TolC multidrug efflux pump. *Nature* 509, 512–515. doi: 10.1038/nature13205
- European Committee for Antimicrobial Susceptibility Testing (EUCAST) of the European Society of Clinical Microbiology and Infectious Diseases (ESCMID) (2000). Terminology relating to methods for the determination of susceptibility of bacteria to antimicrobial agents. *Clin. Microbiol. Infect.* 6, 503–508. doi: 10.1046/j.1469-0691.2000.00149.x
- Fischer, N., Raunest, M., Schmidt, T. H., Koch, D. C., and Kandt, C. (2014). Efflux pump-mediated antibiotics resistance: insights from computational structural biology. *Interdiscip. Sci.* 6, 1–12.
- Gandhi, T. N., DePestel, D. D., Collins, C. D., Nagel, J. J., and Washer, L. L. (2010). Managing antimicrobial resistance in intensive care units. *Crit. Care Med.* 38, 315–323. doi: 10.1097/CCM.0b013e3181e6a2a4
- Ghisalberti, D., Masi, M., Pagès, J. M., and Chevalier, J. (2005). Chloramphenicol and expression of multidrug efflux pump in *Enterobacter aerogenes*. *Biochem. Biophys. Res. Commun.* 328, 1113–1118. doi: 10.1016/j.bbrc.2005.01.069
- Handzlik, J., Bojarski, A. J., Satała, G., Kubacka, M., Sadek, B., Ashoor, A., et al. (2014). SAR-Studies on the importance of aromatic ring topologies in search for selective 5-HT7 receptor ligands among phenylpiperazine hydantoin derivatives. *Eur. J. Med. Chem.* 78, 324–339. doi: 10.1016/j.ejmech.2014.01.065
- Handzlik, J., Szymańska, E., Chevalier, J., Otrębska, E., Kiec-Kononowicz, K., Pagès, J. M., et al. (2011). Amine-alkyl derivatives of hydantoin: new tool to combat resistant bacteria. *Eur. J. Med. Chem.* 46, 5807–5816. doi: 10.1016/j.ejmech.2011.09.032
- Jones, D. (2010). News and analysis: the antibacterial lead discovery challenge. *Nat. Rev. Drug Discov.* 9, 751–752. doi: 10.1038/nrd3289
- Kašćáková, S., Maigre, L., Chevalier, J., Réfrégiers, M., and Pagès, J. M. (2012). Antibiotic transport in resistant bacteria: synchrotron UV fluorescence microscopy to determine antibiotic accumulation with single cell resolution. *PLoS ONE* 7:e38624. doi: 10.1371/journal.pone.0038624
- Laxminarayan, R., Duse, A., Watal, C., Zaidi, A. K. M., Wertheim, H. F. L., Sumpradit, N., et al. (2008). Antibiotic resistance - the need for global solution. *Lancet Infect. Dis.* 13, 1057–1098. doi: 10.1016/S1473-3099(13)70318-9
- Li, X. Z., Plésiat, P., and Nikaido, H. (2015). The challenge of efflux-mediated antibiotic resistance in Gram-negative bacteria. *Clin. Microbiol. Rev.* 28, 337–418. doi: 10.1128/CMR.00117-14
- Malléa, M., Chevalier, J., Bornet, C., Eyraud, A., Pagès, J.-M., and Davin-Regli, A. (1998). Porin alteration and active efflux: two in vivo drug resistance strategies used by *Enterobacter aerogenes*. *Microbiology* 144, 3003–3009. doi: 10.1099/00221287-144-11-3003
- Matys, A., Podlewska, S., Witek, K., Witek, J., Bojarski, A. J., Schabikowski, J., et al. (2015). Imidazolidine-4-one derivatives in the search for novel chemosensitizers of *Staphylococcus aureus* MRSA: synthesis, biological evaluation and molecular modeling studies. *Eur. J. Med. Chem.* 101, 313–325. doi: 10.1016/j.ejmech.2015.06.013
- Misra, R., Morrison, K. D., Cho, H. J., and Khuu, T. (2015). Importance of real-time assays to distinguish multidrug efflux pump-inhibiting and outer membrane-stabilizing activities in *Escherichia coli*. *J. Bacteriol.* 197, 2479–2488. doi: 10.1128/JB.02456-14
- Nikaido, H., and Pagès, J. M. (2012). Broad-specificity efflux pumps and their role in multidrug resistance of Gram-negative bacteria. *FEMS Microbiol. Rev.* 36, 340–363. doi: 10.1111/j.1574-6976.2011.00290.x
- Opperman, T. J., and Nguyen, S. T. (2015). Recent advances toward a molecular mechanism of efflux pump inhibition. *Front Microbiol.* 6:421. doi: 10.3389/fmicb.2015.00421

ACKNOWLEDGMENT

We appreciate greatly fruitful discussions and advices from Dr. Anne Davin-Regli.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fmicb.2016.00622>

- Pagès, J. M., James, C. E., and Winterhalter, M. (2008). The porin and the permeating antibiotic: a selective diffusion barrier in Gram-negative bacteria. *Nat. Rev. Microbiol.* 6, 893–903. doi: 10.1038/nrmicro1994
- Philippe, N., Maigre, L., Santini, S., Pinet, E., Claverie, J. M., Davin-Regli, A., et al. (2015). In vivo evolution of bacterial resistance in two cases of *Enterobacter aerogenes* infections during treatment with imipenem. *PLoS One*. 10:e0138828. doi: 10.1371/journal.pone.0138828
- Pradel, E., and Pagès, J. M. (2002). The AcrAB-TolC efflux pump contributes to multidrug resistance in the nosocomial pathogen *Enterobacter aerogenes*. *Antimicrob. Agents Chemother.* 46, 2640–2643. doi: 10.1128/AAC.46.8.2640-2643.2002
- Pucci, M. J., and Bush, K. (2013). Investigational antimicrobial agents of 2013. *Clin. Microbiol. Rev.* 26, 792–821. doi: 10.1128/CMR.00033-13
- Ruggerone, P., Murakami, S., Pos, K. M., and Vargiu, A. V. (2013). RND efflux pumps: structural information translated into function and inhibition mechanisms. *Curr. Top. Med. Chem.* 13, 3079–3100. doi: 10.2174/15680266113136660220
- Schulz, R., Vargiu, A. V., Collu, F., Kleinekathoefer, U., and Ruggerone, P. (2010). Functional rotation of the transporter AcrB: insights into drug extrusion from simulations. *PLoS Comput. Biol.* 6:e1000806. doi: 10.1371/journal.pcbi.1000806
- Venter, H., Mowla, R., Ohene-Agyei, T., and Ma, S. (2015). RND-type drug efflux pumps from Gram-negative bacteria: molecular mechanism and inhibition. *Front Microbiol.* 6:377. doi: 10.3389/fmicb.2015.00377
- Yamaguchi, A., Nakashima, R., and Sakurai, K. (2015). Structural basis of RND-type multidrug exporters. *Front Microbiol.* 6:327. doi: 10.3389/fmicb.2015.00327
- Yao, X. Q., Kenzaki, H., Murakami, S., and Takada, S. (2010). Drug export and allosteric coupling in a multidrug transporter revealed by molecular simulations. *Nat. Commun.* 1, 1–8. doi: 10.1038/ncomms1116
- Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.
- Copyright © 2016 Otrębska-Machaj, Chevalier, Handzlik, Szymańska, Schabikowski, Boyer, Bolla, Kieć-Kononowicz, Pagès and Alibert. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.