

Contemporary variations of immune responsiveness during range expansion of two invasive rodents in Senegal

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Christophe Diagne, Emmanuelle Gilot-Fromont, Stéphane Cornet, Laëtitia Husse, Souleymane Doucouré, et al.. Contemporary variations of immune responsiveness during range expansion of two invasive rodents in Senegal. Oikos, 2016, 126 (3), pp.435 - 446. 10.1111/oik.03470 . hal-01463805

HAL Id: hal-01463805 https://amu.hal.science/hal-01463805

Submitted on 10 Feb 2017 $\,$

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Biological invasions provide unique opportunities for studying life history trait changes over 2 contemporary time scales. As spatial spread may be related to changes in parasite 3 communities, several hypotheses (such as the evolution of increased competitive ability 4 5 (EICA) or EICA-refined hypotheses) suggest immune changes in invasive species along invasion gradients. Although native hosts may be subject to similar changes in parasite 6 7 selection pressures, their immune responses have been rarely investigated in invasion contexts. In this study, we evaluated immunevariations for invasive house mice Mus 8 9 musculusdomesticus, invasive black ratsRattusrattusand native rodents Mastomyserythroleucus and Ma. natalensis along well-characterised invasion gradients in 10 11 Senegal. We focused on antibody-mediated (natural antibodies and complement) and 12 inflammatory (haptoglobin) responses. One invasion route was considered for each invasive 13 species, and environmental conditions were recorded. Natural-antibody mediated responses increased between sites of long-established invasion and recently invaded sites only in house 14 15 mice. Both invasive species exhibited higher inflammatory responses at the invasion front than in sites of long-established invasion. The immune responses of native species did not change 16 17 with the presence of invasive species. These patterns of immune variations do not support the EICA and EICA refined hypotheses, and they rather suggest a higher risk of exposure to 18 parasites on the invasion front. Altogether, these results provide a first basis to further assess 19 the role of immune changes in invasion success. 20

21 INTRODUCTION

Biological invasions, i.e., the successful establishment and spread of species outside their native range, are increasingly frequent worldwide mostly due to human activities. They often have detrimental consequences for the communities invaded (Kolar and Lodge 2001). Despite the increasing burgeoning interest in invasion science these last decades, several gaps exist in our knowledge and understanding of the factors and forces driving invasion success of nonnative species in new areas (Facon et al. 2006; Lowry et al. 2013).

Parasitism is one factor likely to promote invasion success by influencing a range of host 28 community interactions (Dunn and Hatcher 2015). As such, invasive species may lose their 29 30 natural enemies, including micro- and macro-parasites, in their non-native ranges, with positive outcomes on invader fitness and demography (enemy release hypothesis, Colautti et 31 al. 2004). Alternatively, invaders may introduce exotic parasites with detrimental effects on 32 33 the survival, fecundity and/or regulation of indigenous host populations and provide opportunities for diseases to emerge (spill-over or novel weapon hypothesis, Strauss et al. 34 35 2012). They may also acquire local parasites from their new environment, amplifying the impact of some of them at the expense of native species, with effects at both the individual 36 and population scales (spillback hypothesis, Kelly et al. 2009). As immunity is costly in terms 37 38 of energy and immunopathology(Klasing 2004, Raberg et al. 2002), these changes in parasite pressure have led to some predictions regarding potential variations of invaders' immune 39 defences in the course of invasion. With regards to he enemy release hypothesis, the invaders 40 are expected to reallocate energetic resources from unnecessary defence mechanisms to life 41 42 history traits favouring invasion success, such as dispersal or enhanced reproductive output. This prediction, known as the Evolution of Increased Competitive Ability (EICA) hypothesis 43 (Blossey and Notzold 1995), has been refined to take into account the widespread occurrence 44 of parasitism (Lee and Klasing 2004). In addition to parasites that would not be lost during 45

invasion, invaders could be infected by local parasites (including the possibility of spill-back 46 of parasites carried by native hosts). A drastic reduction of immune responses would make 47 invaders highly susceptible to generalist pathogens encountered in recently invaded host 48 49 communities(Roberts and Janovy 2010). Then, successful invaders should be those that dampen the most expensive and/or least effective immune defences instead favouring less 50 costly and more efficient immune strategies with respect to parasites that are kept from the 51 52 source area or those newly encountered on the invasion front (EICA-refined hypothesis, Lee and Klasing 2004). Such trade-off would allow promoting other life history traits without 53 dampening invader defences. 54

The mechanisms potentially mediating these phenotypic variations include various 55 processes:evolutionary ones as suggested in the name of this hypothesis, but also ecological 56 changes. A major part of immune variation relies on differences in genetic background 57 58 (reviewed in Ardia et al. 2011). Genetic drift and natural selection may therefore be key evolutionary processes by which invaders will evolve on invasion front after few generations 59 (review in Charbonnel and Cosson 2011). Recently, it was also assessed that epigenetic 60 modifications - defined as changes in phenotypes that persist through mitosis and even 61 meiosis, but occur independently of changes in underlying DNA sequence - may also play 62 63 pivotal roles in immune changes related to invasion process (Brown et al. 2015; Na et al. 2016). But alternatively, another important source of immune variation during invasion 64 process could be related to phenotypic plasticity, which is the tendency/ability for phenotypes 65 to change across different environments within generations (reviewed in Gervasi et al. 2015). 66 This mechanism has been widely reported using experimental works modifying 67 environmental conditions (resource quality and availability, cross-fostering) or individual 68 surveys throughout their life, in both vertebrates and invertebrates (Schulenburg et al. 2009). 69

Another complexity resides in the immune system itself. In vertebrates, it is a highly 70 diverse network of organs, cells and molecules that are generally classified into innate and 71 adaptive compartments, both including cellular (e.g., macrophages, lymphocytes) and 72 73 humoral components (e.g., antibodies, peptides), which interact together but may have different costs and benefits for the organism (Klasing 2004). It has been assessed that innate 74 responses associated with local and/or systemic inflammation incur high-energy expenditure 75 and major physiological, behavioural and pathological costs (Sorci and Faivre 2009) 76 77 compared to responses mediated by other effectors, such as both natural (innate) and specific (acquired) antibodies(Raberg et al. 2002, Klasing 2004). This has led to suggest that invasion 78 79 success would therefore be associated with dampened costly systemic inflammatory response and stronger less costly antibody-mediated immunity (Lee and Klasing 2004). Alternatively, 80 because of potential high infection risks encountered on invasion fronts, the invaders' 81 82 capacity to mount rapid, non-specific immune responses could be essential for invasion success. It would be therefore possible to make different predictions than those of Lee and 83 84 Klasing (2004) for immune changes along invasion gradients, such as an increase of immune investment, or the absence of immune trade-offs(Phillips et al. 2010). 85

The impacts that bioinvasions may have on immune responses in native species remain 86 scarcely described and investigated. We can first consider that native species might be 87 exposed to a lower infection risk as a result of dilution effects (the presence of a less 88 competent host decreases the infection prevalence in the native host, Dunn 2009) or of 89 density-dependent effects (the density of native hosts may decrease due to competition with 90 91 invaders, and infection prevalence of parasites transmitted through contacts will consequently decreased too, Keesing et al. 2010). In this context, a decrease in immune investment could be 92 93 selected expected (EICA hypothesis). Nevertheless, parasite spill-over from invasive to native species is frequent (Harris 2009) and can strongly affect native host communities, even 94

leading to local extinction, which favorsthe invasive host (Daszak et al. 2000, Prenter et al. 95 2004). Such a strong impact of exotic infections on native species could result from the lack 96 of efficient immune responses (either in terms of immune effectors or amounts of 97 responsiveness). A dampened immune defense could result from direct competition with 98 invaders through a lowered access to resources or from stress hormones such as 99 glucocorticoids that may ultimately compromise immunity (Martin et al. 2010a). Strong 100 immunopathologic effects affecting natives' fitness may also result from infections with 101 102 exotic parasites (spill-over) or from amplified risk of infections with native parasites (spillback) (Martin et al. 2010b). 103

104 Limited empirical data are currently available concerning the ecoimmunology of invasions (Pedersen and Babayan 2011, White and Perkins 2012). Indeed, few studies have investigated 105 differences in immune phenotypes along invasion gradients for a given invasive species, and 106 even fewer have considered several immune pathways simultaneously (Cornet et al. 2016, but 107 108 see Llewellyn et al. 2012, Brown et al. 2015). Therefore, it remains important to improve our 109 knowledge of such immune variations associated with invasion. It is a necessary pre-requisite 110 before addressing the eco-evolutionary processes dictating invasion success. In this study, we focused on two invasive rodent species, the domestic mouse (Mus musculusdomesticus) and 111 the black rat (*Rattusrattus*). Weanalyzed phenotypic variations in immune responsiveness 112 occurring along their well-known invasion routes in Senegal. These murid rodents are 113 exclusively commensal in the study area (i.e., living in/around human dwellings/man-made 114 structures) and are worldwide significant invasive species (Global Invasive Species Database 115 - http://www.issg.org/database/). Complete syntheses reporting description of their current 116 invasion histories in Senegal (including data of historical inventories, molecular analyses and 117 ecological longitudinal surveys) are provided elsewhere (Dalecky et al. 2015, Konecny et al. 118 2013). Briefly, both species originated from Asia and have expanded their distribution range 119

worldwide, making use of human migration to colonize all the continents. They were first 120 brought to West Africa by European explorers and settlers from the 15th century. Large and 121 likely standing populations of rats and mice are reported in coastal colonial cities from the 122 middle of the 19th century. At the beginning of the 20th century, they began to expand 123 eastwards with the development of inland commercial transport. Their distribution currently 124 covers much of North and Central Senegal for M. m. domesticus and South and Central 125 Senegal for R. rattus. Recent longitudinal surveys (Granjon and Duplantier 2009, Dalecky et 126 127 al. 2015) have documented how the ongoing expansion of both species has resulted in the extirpation of native rodents (mostly Mastomyserythroleucus and Ma. natalensis) from 128 commensal habitats, these latter species being now found almost only in villages at the 129 invasion front and in non-invaded areas. 130

Immune phenotypes were described using effectors involved in natural antibody (NAb)-131 132 mediated and inflammatory immune pathways. They are main components of innate constitutive immunity acting in the earliest phase of immune defence against general 133 134 challenges and new parasites(Rossi et al. 2013). First, NAbs are humoral components of constitutive immunity, providing the first-line of non-cellular protection against antigens 135 (Matson et al. 2005). They serve as recognition molecules capable of opsonising invading 136 137 micro-organisms and initialising the Complement (Cp) enzyme cascade, which ends in cell lysis (Carroll and Prodeus 1998). They are unique among immunoglobulin molecules because 138 their presence does not require prior exposure to exogenous antigens. The assessment of this 139 immune pathway is particularly appealing because NAbs should be less sensitive than 140 acquired antibody responses to short-term variations in environmental conditions, nutritional 141 status, or stress levels (Baumgarth et al. 1999). Moreover, Nab levels in the serum could be 142 positively correlated with the ability to produce antibodies after a challenge (Matson et al. 143 2005). Second, we used haptoglobin (Hp) concentration to assess the inflammatory state of 144

the rodents, as previously performed in wild birds (Martin et al. 2010a). Hp is a 145 multifunctional hepatic acute-phase protein highly released in the blood during inflammation, 146 with strong anti-inflammatory and anti-oxidant properties (Huntoon et al. 2008). It circulates 147 148 at low concentrations in the blood of a range of animal species and its concentration is dependent on health status (Dobryszycka 1997). Hp has been shown to be a distinctive trait of 149 individuals (Matson et al. 2012). Both immune tests used here are simple, highly repeatable 150 151 and non-specific. Moreover they do not require the recapture of animals or their maintenance 152 in animal facilities. For these reasons, they enable to cope with the specific constraints of comparative immunological studies dealing with numerous samples of different non-model 153 154 species caught in natural populations and sacrificed at the time of capture. All assays were performed at both long-established and recently invaded sites, for both invasive species, and 155 for *Mastomys* species found either in sympatry with the invaders at the invasion frontor alone 156 at sites not yet invaded. 157

We specifically addressed the two following questions: 1)do the immune responsiveness 158 159 patterns observed in rats and mice along invasion routes support the EICA or EICA refined 160 hypotheses? We expected a decrease of immune responsiveness (Nabs and Hp) in recently invaded localities compared to long established ones under the EICA hypothesis, or a 161 decrease of costly immune responsiveness (Hp) at the expense of less costly ones (NAbs) 162 under the EICA-refined hypotheses;2) do native species exhibit variations in immune 163 responsiveness associated with the presence of invasive ones ?Up to now, no framework has 164 been developed with regard to this question, what prevents us to make any 165 predictions. Altogether, these results enabled to discuss some of the general predictions of Lee 166 and Klasing's paper (2004), which is one of the reference work when studying the potential 167 links between immunology and invasion. Nevertheless, this study is descriptive and as such, it 168

169 was not designed todisentangle the ecological and evolutionary processes potentially170 underlying the immune patterns observed.

171

172 MATERIALS AND METHODS

173 Ethical Statement

Trapping campaigns within private properties were systematically realized with prior explicit agreement from relevant institutional (CBGP: 34 169 003) and local authorities. All animalrelated procedures were carried out in accordance with relevant requirements of Senegalese and Frenchlegislation and following official ethical guidelines (Sikes et al. 2011).

178 Rodent sampling and blood collection

Field sampling was conducted separately along an invasion route for each invasive species 179 (Figure 1). It was carried out during the dry season, in March-April 2013 for the 'mouse' 180 invasion route and from November 2013 to February 2014 for the 'rat' invasion route. The 181 sampling sites belonged to one of three invasion status categories, defined on the basis of 182 183 historical records and longitudinal surveys (see references in Dalecky et al. 2015, Konecny et al. 2013): (i) sites of long-established invasion, in which invasive populations have been 184 185 recorded for centuries; (ii) sites at the invasion front (= sites recently invaded) in which 186 invasive populations have been established for less than 30 years; and (iii) non-invaded sites in which invasive species have never yet been recorded or trapped. For each category, we 187 sampled three to four sites (Figure 1). We used a standardised sampling protocol for all 188 localities of the three categories of sites considered. It enabledto standardize the potential 189 190 impacts of stress on immune responses due to animal capture and handling. The trapping 191 procedures are described in detail elsewhere (Dalecky et al. 2015). At each site, we set at least 120 traps within houses during one to three successive nights, with the aim to capture 20 adult 192 193 rodents per species. Traps were checked and baited once a day with peanut butter

supplemented with fresh onions. Rodents captured at nightwere retrieved the following 194 morning and then sacrificed by cervical dislocation within the following four hours. They 195 wereweighed to the nearest 0.5 g, sexed and dissected. Identification was based on 196 197 morphometrics (head-body, tail, hind foot and ear lengths) and genetics (ZFY2 gene-based RFLP for identification to the subspecies level for *M. musculus*; cytochrome b gene-based 198 RFLP for species characterisation in the genus Mastomys). As suggested by Granjon and 199 200 Duplantier (2009), rodents were considered to be adults on the basis of body weight and 201 reproductive status. Blood samples were collected by cardiac puncture after the animals were euthanatized. They were kept on ice for 24 hours and then centrifuged. The floating serum 202 203 supernatant fraction was removed, frozen in liquid nitrogen, and then stored in a freezer (-20°C) in Dakar (Senegal). Samples were transferred in dry ice from Dakar to Montpellier 204 (France) in accordance with the regulations enforced in Senegal and France. 205

206 Environmental data

Because environmental variations may drive differences in immune responses between rodent 207 208 populations independently of the invasion status categories, we described relevant climatic and commensal habitat parameters for all sampling sites and included them in further 209 statistical analyses. We focused on these factors and did not include vegetation information 210 211 because house mice and black rats are strictly commensal in Senegal, and because both invasive species expand their range through human trade and transport rather than by 212 individual dispersal in the wild (Dalecky et al. 2015). Means and standard deviations of 213 climatic data collected between 1997 and 2012 were used (data available on 214 http://www.ncdc.noaa.gov/cdo-web/datasets for temperatures, and http://richardis.univ-215 paris1.fr/precip/rainser1.html for rainfall with GPCP-1DD as the source of data). We 216 considered rainfall data in mm (for each year:cumulated annual rainfall, cumulated rainfall 217 218 during the rainy season, minimum and maximum monthly rainfall during the rainy season)

and temperature data in degrees Celsius (for each year: monthly mean, monthly mean minimum and maximum, minimum of monthly mean minimum and maximum of monthly mean maximum). Commensal habitat characteristics were recorded during trapping sessions. In particular, we recorded for each sampled room the material used (sand, banco, cement, sheet metal, fibers) for each part of the building (floor, wall, ceiling), the type of room (dwelling house, shop, storehouse, kitchen) and for each site the inhabited area surface estimated using Google Earth Pro 7.1.

226 Hemagglutination-hemolysis (HA-HL) assay

The goal of this assay was to evaluate components of humoral innate immunity through the 227 228 ability of plasma to agglutinate and lyse foreign cells through NAbs-Cp system. We characterised the serum agglutination of heterologous red blood cells (RBC) due to NAbs 229 (HA assay) and RBC hemolysis due to NAbs-mediated Cp activation (HL assay), with a 230 slightly modified version of the protocol described by Matson et al. (2005) for mammal 231 species. Briefly, we used chicken RBCs as target cells, and serum samples from a rabbit 232 233 immunised against chicken RBCs as positive controls. All the HA/HL assays from an invasion route were carried out using the same RBC suspension. We diluted twofold serially 234 (from 1/2 to 1/128) ten rodent serum samples per plate with 10µl in every well. The plates 235 236 were read blindly following Rossi et al. (2013) and scores were attributed for each sample (column) as the log2 of the last dilution exhibiting each phenomenon (HA, HL). To exclude 237 potential observer effects, all images were scored the same day by the same trained observer; 238 thus correction for observer effect was not necessary in the subsequent analyses. It was 239 possible to assign a half-point score in cases of ambiguity. High scores of HA reflected high 240 241 concentrations of NAbs in the blood. Levels of HL reflected both the NAbs and complement activities. 242

243 Haptoglobin (Hp) assay

Hp was quantified from 10µl of serum with a commercially available colorimetric 244 immunoassay kit ("PHASE" Haptogloblin Assay, TP-801, Tridelta), according to the 245 manufacturer's instructions. Absorbance at 650 nm was determined with a spectrophotometer, 246 247 both before and after the addition of the final reagent triggering the colorimetric reaction. We used the pre-scan absorbance to correct for differences in plasma colour and cloudiness, 248 including the initial redness of the serum(i.e., the initial serum hemolysis) that can hamper the 249 assay if not taken into account (Matson et al. 2012). Serum Hp concentration (mg/ml) was 250 251 estimated by comparing the difference in absorbance (final – pre-scan) to a calibration curve.

252 Statistical analyses

253 In order to assess whether environmental features differed between the three categories of sampled sites, we carried out a two-stage Principal Component Analysis (PCA) with sites as 254 255 observations, independently for climatic features (using 16-years mean values and their respective standard deviations) and commensal habitat characteristics (proportion data). As 256 many indicators appeared highly correlated (r > 0.75), we kept only one variable in each set of 257 258 highly correlated variables, based on both contributions to the axis construction and quality of representation. We then performed a final PCA with remaining variables. We tested 259 statistically whether data were structured according to invasion status categories using a 260 Between/Within-groups Analysis (BWA). Monte-Carlo tests of permutations (999 261 permutations) were applied to analyse the statistical significance of the groups graphically 262 observed on the PCAs. 263

Variations in Hp concentration (log-transformed), HA and HL values were analysed independently, using linear models, in R software v.3.1.0 (R Core Team 2015). The 'mouse' and 'rat' invasion routes were investigated separately. The starting models included a factor combining the species and site status on the invasion route, hereafter referred to as 'specific invasion status' and classified into four categories : A) invasive species alone in sites of long-

established invasion; B) invasive species at recently invaded sites; C) native species at 269 recently invaded sites and D) native species at non-invaded sites. Combining the two factors 270 'species' and 'site category' into a single one (called "specific invasion status" hereafter) 271 272 enabled to avoid confounding effects between host species and invasion on immune responses. Individual factors (sex, body mass and age class) may greatly affect immune 273 system parameters. We therefore included these factors and their 2-ways interactions with 274 specific invasion status as fixed effects. We also added the descriptors of environmental 275 276 variations as potential predictors in the full models. Therefore, coordinates of each site on the two first PCA axes for both climatic and commensal habitat data were included. Finally, we 277 278 had to include several additional factors specific to the different experimental protocols used: (i) a 'plate' factor in the HA model to take into account both the chronology in which the 279 plates were analysed and the involvement of two different experimenters, and (ii) HA scores 280 281 in the HL model as HL directly depends on the presence of antibody-antigen complex revealed by HA. For both the HA and HL models, we also included the initial level of serum 282 283 hemolysis as initial hemolysisis expected to interfere with the interpretation of HA and HL results. A value was then systematically given to the serum prior dilution, ranging from 1 (not 284 red) to 8 (dark red) according to the intensity of the initial serum redness in order to correct 285 for non-visibility and over-interpretation of the phenomena during the plate reading. The 286 Akaike information criterion with correction for samples of finite size (AICc) was used for 287 model selection. Models with all possible combinations of the terms included in the starting 288 289 model were generated with the MuMIn v.1.10.5 R package (dredge function, Barton (2016)). 290 Models with a $\Delta AICc < 2$ with respect to the model with the lowest AICc were selected and the most parsimonious of these models was chosen. The significance of explanatory variables 291 292 and their interactions was determined by deletion testing and log-likelihood ratio tests. The assumptions of each final model were checked graphically, by an analysis of their residuals. 293

Post-hoc tests for multiple comparisons were carried out with Tukey's test (95% family-wiseconfidence level).

296

297 **RESULTS**

We analysed serum samples from 646 individuals belonging to four rodent species captured at 298 23 sites (Table 1, Figure 1). The dominant species in the native rodent communities were Ma. 299 300 erythroleucus along the 'mouse' invasion route and Ma. erythroleucus or Ma. natalensis along the 'rat' invasion route. Mastomysnatalensis was found specifically at the invasion front 301 of R. rattus, coinciding geographically with the limited distribution area of this native species 302 303 in Senegal. Mastomyserythroleucus was occasionally captured in sites of long-established invasion (n = 15) and in non-invaded sites beyond the invasion front in western Senegal (n = 15)304 3) along the 'rat' invasion route. These individuals were too few in number for more detailed 305 analysis. No significant difference was detected in HA ($F_{1,49}$ = 0.18, p = 0.71), HL ($F_{1,49}$ = 306 2.14, p = 0.28) or Hp($F_{1,53} = 1.27$, p = 0.38) levels between *Mastomys* species at sites recently 307 308 invaded by R. rattus. Mastomysnatalensis and Ma.erythroleucuswere therefore considered as 309 a single taxon, native Mastomys sp., in further statistical analyses on the 'rat' invasion route.

310 Environmental data

311 We found significant climatic differences along 'mouse' and 'rat' (Monte-Carlo tests, p < 0.05) invasion routes. Along the 'mouse' invasion route, climate seemed cooler and drier in sites of 312 long-established invasion compared to the other sites. Along the 'rat' invasion route, climate 313 seemed cooler and wetter in sites of long-established invasion compared to the other sites. 314 315 These results are well represented on the second axis of mouse and rat PCAs (Figure S1a, 316 Figure S2a, supplementary materials). The Between/Within analysis on commensal habitat characteristics revealed a significant segregation between invaded and non-invaded sites 317 318 along the 'mouse' invasion route (Figure S1b, supplementary materials), suggesting more

traditional buildings in non-invaded sites. Along the 'rat' invasion route, no significant difference in commensal habitat characteristics was detected (Monte-Carlo test, p > 0.05) with regard to site invasion status (Figure S2b, supplementary materials). However, these results must be taken cautiously as this study was not designed to investigate the influence of environmental characteristics on immune responsiveness.

324 Variation in HA/HL estimates

325 'Mouse' invasion route - The most parsimonious model explaining variation in HA included the climate (component 1: $F_{1,290} = 5.83$, p < 0.0001; component 2: $F_{1,290} = 16.81$, p < 0.0001), 326 plate factor ($F_{1,290} = 11.27$, p = 0.0163), specific invasion status ($F_{3,290} = 12.48$, p < 0.0001, 327 328 Figure 2) and its interaction with sex ($F_{3,290} = 4.75$, p = 0.0029) (Table S1, supplementary materials). Sex accounted for differences in HA levels only at non-invaded sites, with higher 329 values recorded for males than females. Mice from sites of long-established invasion had the 330 lowest HA values. No significant difference in mean HA was detected in Ma. erythroleucus 331 between recently invaded and non-invaded sites (post-hoc Tukey's test, p = 0.9999), nor 332 between Ma. erythroleucus and M. m. domesticus at the invasion front (post-hoc Tukey's test, 333 p = 0.9999). 334

The most parsimonious model explaining variation in HL included the initial degree of serum 335 hemolysis ($F_{1,293} = 99.82$, p < 0.0001), HA score ($F_{1,293} = 50.72$, p < 0.0001), sex($F_{1,293} = 9.87$, 336 p = 0.0018) and specific invasion status ($F_{3,293} = 26.56$, p < 0.0001, Figure 2) (Table S1, 337 supplementary materials). High values of HL were recorded for high values of HA and initial 338 339 serum hemolysis, and for males compared to females. Mice from sites of long-established invasion had lower HL values compared with other invasion statuses. On invasion front, M. 340 341 m. domesticus had lower values of HL than Ma. erythroleucus from recently invaded (posthoc Tukey's test, p < 0.0001) and non-invaded sites (post-hoc Tukey's test, p = 0.0006). No 342

significant difference in mean HL between recently invaded and non-invaded sites was detected in *Ma. erythroleucus* (post-hoc Tukey's test, p = 0.8723).

³⁴⁵ '*Rat' invasion route* - The most parsimonious model explaining variation in HA included the ³⁴⁶ initial degree of serum hemolysis ($F_{1,299} = 12.74$, p = 0.0004), age ($F_{1,299} = 9.40$, p = 0.0024), ³⁴⁷ body mass ($F_{1,299} = 4.93$, p = 0.0270) and climate ($F_{1,299} = 12.91$, p = 0.0004) (Table S2, ³⁴⁸ supplementary materials). Heavier and adult rodents had higher HA levels than lighter and ³⁴⁹ juveniles, respectively. Serum hemolysis was negatively correlated with HA. No significant ³⁵⁰ difference was found according to specific invasion status ($F_{3,299} = 1.41$, p = 0.2399).

The most parsimonious model explaining variations in HL included the initial degree of serum hemolysis ($F_{1,300} = 57.76$, p < 0.0001), HA score ($F_{1,300} = 355.92$, p < 0.0001) and commensal habitat characteristics (component 1: $F_{1,300} = 21.82$, p < 0.0001) (Table S2, supplementary materials). HL values were positively correlated with both HA scores and with initial serum hemolysis. Specific invasion status had no significant effect on HL values ($F_{3,297} = 0.82$, p = 0.4862).

357 Variation in Hp estimates

'Mouse' invasion route - The most parsimonious model explaining variation in Hp 358 concentration included climate (component 1: $F_{1,183}$ = 13.31, p = 0.0003; component 2: $F_{1,183}$ = 359 22.74, p < 0.0001) and specific invasion status ($F_{3,183} = 5.22$, p = 0.0017) (Table S1, 360 supplementary materials; Figure 2). Mice sampled from the invasion front had Hp levels 361 twice those of mice from sites of long-established invasion (post-hoc Tukey's test, p =362 0.0039). No significant difference in mean Hp concentration was found between Ma. 363 *erythroleucus* from recently invaded and non-invaded sites (post-hoc Tukey's test, p =364 0.9307). Furthermore, Hp levels were similar in Ma. erythroleucus and M. m. domesticus 365 sampled in recently invaded sites (post-hoc Tukey's test, p = 0.4349). 366

'Rat' invasion route - The most parsimonious model best explaining variation in Hp included 367 climate ($F_{3,308} = 12.62$, p = 0.0004; Table S2, supplementary materials) only. However, the 368 specific invasion status was included in about half of the models within $\Delta AICc \le 2$ (in seven 369 370 of the 16 selected) with R. rattus from sites of long-established invasion having lower Hp concentrations than rodents trapped in recently invaded sites (R. rattus: post-hoc Tukey's test, 371 p = 0.0362; Mastomys sp.: post-hoc Tukey's test, p = 0.0058; Figure 3). At the invasion front, 372 Hp levels were similar in invasive and native species (post-hoc Tukey's test, p = 0.74). No 373 significant difference was detected in mean Hp between Mastomys sp. from recently invaded 374 and non-invaded sites (post-hoc Tukey's test, p = 0.2163). 375

376

377 **DISCUSSION**

Most studies investigating the role of immunity in the context of biological invasions have 378 379 focused on comparisons between sympatric invasive and native species or between phylogenetically related species with contrasted levels of invasion success (Lee et al. 2006, 380 Martin et al. 2010a). Because differences in immune investment may be due to intrinsic 381 species characteristics, sampling designs focusing on interspecific comparisons may not be 382 entirely appropriate for evaluating immune defences - invasion issues. Comparing several 383 384 invasive (areas in which invaders are well established vs newly colonised areas) and native (non-invaded vs recently invaded areas) populations along a well-defined invasion route 385 appears to be a more relevant approach, as it allows overcoming specific differences and 386 taking into account the invasion history as a spatio-temporal continuum. Moreover, potential 387 immune changes in native communities should be considered as a full issue that may 388 ultimately impact the invasion process. Several predictions have been proposed with regard to 389 390 immunity and invasion (Lee and Klasing 2004, Phillips et al. 2010). Our results did not fully support the major one, which is the EICA-refined hypothesis. Antibody-mediated defences 391

were found to increase along the invasion route, but for *M. m. domesticus* only (Fig. 2; Table S1, Supplementary materials). Moreover, the inflammatory response was found to be stronger in invasive populations at the invasion front than in populations from sites of long-established invasion, for both invasive species (Fig. 2; Fig. 3; Table S1, Table S2, Supplementary materials). Finally, we did not observe any difference in the immune effectors surveyed in native species between invaded and non-invaded localities.

398

399 Effects of methodological biasesand individual characteristics on immune variations

We carefully considered experimental factors in our statistical models as they could have 400 401 biased our results. The initial levels of serum hemolysis and HA scores were found to significantly influence HA/HL results irrespective of the invasion route (Table S1, Table S2, 402 Supplementary materials). Higher values lead to higher HA/HL scores, regardless to the 403 404 species and the invasion status of sites. These results were expected as Cp lysis is activated by the formation of antigen-NAb complexes (Matson et al. 2005). Furthermore, the initial serum 405 406 hemolysis is part of the HL score and may represent the total final HL in extreme cases where 407 NAb-mediated lysis did not occur in a serum. These results highlight the crucial importance to incorporate such 'experimental' factors as potential predictors in the explanatory models. 408

409 Immune responses could also be biased due tostress resulting from capture and handling procedures. However, the immune effectors studied here are not strongly affected bystressors. 410 In particular, it has recently been shown that Nabs and Cpare insensitive to capture and 411 handling stress (Buehler et al. 2008). Besides, Nab production seems to be largely 412 413 independent of internal or external stimuli (Ochsenbein and Zinkernagel 2000). In addition, we minimized the potential biases due to different effects of stress between localities of 414 415 colonization statuses by using a consistent standardized capture protocol among sampling sites. This design aimed at preventing any differences in methodology-related stressors 416

between rodent populations. Finally, a strong argument showing that capture and handling procedures had a very few impact on the immune patterns observed is that native rodents, which are also subject to these methodological stresses, exhibited no immune variation between non-invaded sites and invasion fronts. This result suggests that the invasion front itself is not an area generating specific stress reactions. Altogether, these elements argue in favour of a limited impact of stress on the immune variations observed here.

We found a significant influence of age, body mass and sex on some immune responses that 423 were higher in adults, heavier individuals and males, respectively (Table S1, Table S2, 424 Supplementary materials). Age and body mass influenced significantly NAb levels for both 425 426 native and invasive rodents on the 'rat' invasion route. These findings were congruent with other works (Rossi et al. 2013) and corroborated the common trend of an increased immune 427 capacity with host growth and condition. Our result of male mice showing higher HL levels 428 429 than females corroborates he life-history theory predicting that females should invest more in specific immune pathways than in constitutive immunity, which would thus be expected to be 430 431 downregulated (Lee 2006). Moreover, other factors including ecological, physiological, social 432 and behavioural ones, probably interacting in a complex way, may contribute to the observed patterns of sexual differences observed in rodent immunity. For instance, it has been shown in 433 434 voles that social environment and steroid hormones affect in a complex way sex differences in immune function (Klein et al. 1997). This question therefore remains a challenging area of 435 research in evolutionary ecology. 436

437

438 Impact of environmental conditions on immune patterns

As *M. m. domesticus* and *R. rattus*are likelyto encounternew environments along their
respective invasion route, immune variations observed in these invasive species may be
considered as responses to environmental factors(reviewed in Colautti and Lau 2015). It has

been previously shown that environmental parameters can affect the immune functions of 442 443 rodents (see Beldomenico et al. 2008). Climate and commensal habitat structure may influence rodent immune responses, for instance through resource availability or community 444 445 composition, and were thus considered in our statistical models. Nevertheless, these environmental featurescould not explain alone the patterns observed. Climatic data differed 446 between sites of long- and recently-established invasion for both invasive mice and rats but no 447 variation was observed in HA and HL levels for R. rattus, and climate was not a relevant 448 predictor to explain variations of HL in M. m. domesticus. Likewise, habitat structure did not 449 differ between sites of long- and recently-established invasion along the 'mouse' invasion 450 451 route, although immune responses significantly varied. Conversely, habitat structure differed between non-invaded sites and those recently invaded by the house mouse, but no immune 452 variationwas observed in native populations along this invasion transect. 453

454

455 Absence of immune patterns supporting the EICA and EICA refined hypotheses

Variations in immune responses between long- and recently-established invasive populations did not support the predictions expected under the EICA and EICA refined hypotheses, which are respectively a general decrease of immune responsiveness or opposite changes between energetically costly immune pathways (decrease of responses) and less costly ones (increase of responses). Instead, this study revealed increasedinflammatory and/or humoral responses in expanding invasive populations of mice and rats compared to long established populations (Fig. 2; Fig. 3).

These results questioned the relevance of the physiological and ecological interactions that are at the basis of EICA expectations, namely parasitism and immune costs. The risks of exposure to parasites and subsequent infections are supposed to be an important component of the novel environment encountered during invasion, which may explain these immune variations

along invasion routes. Nevertheless, by contrast with the enemy release emphasized in the 467 EICA hypothesis, invaders may experience novel parasite pressure as they may be exposed 468 and/or infected with novel local parasites when pathobiome communities differ between 469 470 native and invasive species (see Diagne et al. in press; Galan et al. 2016 for examples). The overall infection risk in recently invaded areas could thus increase on invasion front. In 471 particular, the ability to mount strong inflammatory responses may prevent 'naïve host 472 473 syndrome' in invasive species or within the invasion front, a severe adverse effect of parasites 474 on hosts with which they have no co-evolutionary history (Mastitsky et al. 2010). Besides, larger responses for Hp in invaders would be consistent with efforts to attenuate 475 476 inflammation. Indeed, although the secretion of Hp is enhanced in inflammatory states, Hp has strong anti-inflammatory and anti-oxidative properties. During infection, Hp is best 477 characterized as a protein that protects hosts against "all the dangers of the acute phase 478 479 response" (Dobryszycka 1997). Within 24h of infection, circulating Hp concentration increases considerably, to replenish hemoglobin stores and damp down inflammatory 480 481 responses and thus, their detrimental effects. High Hp levels may therefore be of benefit to 482 invaders due to a lessening of the negative impacts of immunity.

The extent to which the presumed costs of immune defences (high for inflammation, low for 483 484 antibody-mediated responses, Lee and Klasing 2004) can be generalized to all biological models and ecological conditions must also be questioned. Immune strategies have been 485 shown to differ significantly between species, or even between races of domesticated species 486 (Mendes et al. 2006). Moreover, the estimation of these costs may be specifically dependent 487 on the response examined for a given pathway, and/or the condition/energetics parameters 488 chosen. However trade-offs between inflammation and humoral responses involving immune 489 490 effectors other than those investigated here cannot be excluded. We must keep in mind that the indicators studied here only reflect one part of the immune capacities to fight infections and 491

may not be generalizable to other effectors. Although it is very difficult to measure immune
costs in the wild, a full study dedicated to this issue should provide crucial information on
immune trade-offs during invasion process.

495

496 Stronger patterns of immune variationsin mouse compared to rat invasive species

Although environmental parameters and interspecific differences might beobvious reasons to 497 498 explain contrasted immune variationsobserved between rats and mice along their respective 499 invasion routes, different abilities in direct competition with native communities may also be proposed. Direct competition is often put forward as an explanation for the replacement of 500 501 native species by exotic rodents (Drake and Hunt 2009). The black rat and house mouse strongly differ in their competitive interactions with native rodents. R.rattus has been shown 502 to be aggressive to intruders, physically eliminating experimentally introduced conspecific 503 504 individuals from an insular population (Smith and Banks 2014). They are much larger than Mastomys species and would therefore be at an advantage in direct competition. On the 505 506 contrary, M. m. domesticus is known to perform poorly in direct competition with several 507 native rodents (Gomez Villafane et al. 2013). Variation in immune phenotype may therefore be a less important strategy for the invasion success of R. rattusthan for M. m. domesticus. 508 Invasion success mediated by immune variations in house mice would account for 509 observations that this species is less parasitized than rats, for zoonoses for instance (Blackwell 510 1981, Meerburg et al. 2009). This hypothesis needs to be tested, and it would also be 511 interesting to investigate whether it is observed in other cases of biological invasions. 512

513

514 No variation of immune response in native species

515 The questions of what happens in native species and whether immune responses change in 516 invaded compared to non-invaded areas remain scarcely explored. In this work, native

Mastomys species exhibited similar immune responses in non-invaded and recently invaded 517 sites, along both the 'mouse' and 'rat' invasion routes (Fig. 2; Fig. 3). This pattern could 518 result from the matching between environmental conditions (especially parasite pressure, 519 520 resource availability, etc.) and the development of adequate immune responses by native rodents that have co-evolved with their natural environment. However, we found significant 521 environmental differences between non-invaded sites and invasion fronts for both invasion 522 523 routes. It is also likely that novel epidemiological conditions are established during invasion 524 as the introduction of invaders directly affects host community composition and may also modify the infection risk of native species (Dunn and Hatcher 2015, Diagne et al. in revision, 525 526 Galan et al. unpublished data). As such, the infection risk may be either amplified by spillover and/or spill-back mechanisms, or decreased when invaders are not competent to maintain 527 and transmit local parasites (dilution effect). Thus, the absence of immune variations in native 528 529 species could reflect their inability to adapt their immune phenotypes.

530

531 In summary, our data provide no support for (i) the predictions of the EICA-refined 532 hypothesis that antibody-mediated immunity should be favoured over inflammation during invader range expansion, and (ii) our expectations on immune changes experienced by native 533 species when co-occurring with invaders. Whether immune changes could occur due to a 534 higher risk of exposition to/infection with novel parasites at the invasion front and a greater 535 ability of invasive species, such as M. m. domesticus in particular, to adjust their immune 536 phenotypes during invasion are hypotheses that need to be confirmed experimentally in the 537 future. Such studies could also enable to assess the respective roles of evolutionary and 538 ecological processes in driving these phenotypic immune variations. 539

Acknowledgments.We thank Laurent Granjon and Jean-Marc Duplantier for their precious help and advice during this work. Jean-Claude Berges and Natacha Volto are thanked for their help with climatic variables. This work was supported by the ANR ENEMI project (ANR-11-JSV7-0006). We acknowledge support from the *Service de Coopérationetd'ActionCulturelle* (French Embassy, Senegal) and the *AgenceUniversitaire de la Francophonie* through Ph.D. scholarships. We are particularly indebted to all the people in Senegal who allowed us to carry out rodent trapping in their homes.

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Data archiving.Data for this study will be available onDryad (DataDryad.Org) after
manuscript is accepted for publication.

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Authorship.CD performed the HAHL analyses and wrote the first draft of the paper. EGF participated in the interpretation and statistical analyses of HAHL data. SC, LH, SD performed the Hp analyses and SC carried out the statistical analyses. OFG participated in the multivariate analyses of environmental data. CB, AD and NC designed the sampling. NC designed the immunological experiments and formulated the hypotheses tested. AD and KB managed the extensive field work. KB, MK, YN, MD, AS, CB, NC and CD performed the field sampling. SP and EA were responsible for sample collection and the database.

Table 1: Sample size for each assay, by invasion status (LI = sites of long-established 696 invasion; IF = invasion front; NI = non-invaded sites), sampled site (code in parentheses) and 697 host species for a) the 'mouse'invasion route (166 Mus musculusdomesticus, 698 145Mastomyserythroleucus) and b) the 'rat' invasion route (196 Rattusrattus, 699 700 88Mastomysnatalensis, 50 Mastomyserythroleucus). Numbers in parentheses indicate sample size for males/females, respectively. '-' indicates that no rodent was trapped or 701 analysed.Sample sizes differ between immune assays because of the limited volume of some 702 703 serum samples prevented to perform both assays.

704 a)

Invesion	Sites	Hemagglutinat	tion-Hemolysis	Haptoglobin		
status		M. m.	Ma.	M. m.	Ma.	
	-	aomesticus	eryinroleucus	aomesticus	eryinroleucus	
LI	Dagathie (DAG)	19(10/9)	-	12(6/6)	-	
	Mbakhana (MBA)	23(13/10)	-	12(8/4)	-	
	Thilene (THL)	20(8/12)	-	13(3/10)	-	
	Ndombo (NDB)	21(10/11)	-	15(7/8)	-	
	Dodel (DOD)	23(9/14)	19(11/8)	10(3/7)	12(9/3)	
IF	Aere Lao (AEL)	21(10/11)	19(12/7)	17(8/9)	11(7/4)	
IF	Dendoudi (DEN)	17(12/5)	23(16/7)	10(6/4)	16(12/4)	
	Lougue (LOU)	22(10/12)	22(15/7)	10(3/7)	14(11/3)	
NI	Doumga Lao (DOL)	-	20(10/10)	-	14(7/7)	
	Lambango (LAM)	-	20(9/11)	-	16(7/9)	
	SaréMaoundé (SAM)	-	9(5/4)	-	5(3/2)	
	DiomandouWalo (DIW)	-	13(9/4)	-	11(7/4)	

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706

b)

Invasion status	Sites	Hemagglutination-Hemolysis			Haptoglobin			
		R. rattus	Ma. erythroleucus	Ma. natalensis	R. rattus	Ma. erythroleucus	Ma. natalensis	
LI	Diakene Wolof (DIK)	24(9/15)	-	-	24(9/15)	-	-	
	Diattacounda (DIT)	27(13/14)	-	-	27(13/14)	-	-	
	Marsassoum (MAR)	26(13/13)	-	-	26(13/13)	-	-	
	Tobor (TOB)	20(6/14)	-	-	20(6/14)	-	-	
IF	BadiNieriko (BAN)	21(8/13)	11(5/6)	-	23(10/13)	12(6/6)	-	
	Bountougoufara (BOU)	31(9/22)	12(7/5)	-	31(9/22)	13(8/5)	-	
	Kedougou (KED)	22(9/13)	-	22(9/13)	22(9/13)	-	22(9/13)	
	Soutouta (SOU)	22(11/11)	9(5/4)	-	23(12/11)	9(5/4)	-	
NI	Bransan (BRA)	-	-	23(10/13)	-	-	23(10/13)	
	Mako (MAK)	-	-	23(11/12)	-	-	26(13/13)	
	Segou (SEG)	-	-	20(7/13)	-	-	21(8/13)	





Figure 1.Sampling sites. Triangles, squares and circles correspond respectively to sites of
long-established invasion (> 100 years), recently invaded sites (< 30 years) and non-invaded
sites. Mouse invasion route (symbols in white): Dagathie (DAG), Mbakhana (MBA), Thilene
(THL), Ndombo (NDB), Dodel (DOD), Aere Lao (AEL), Dendoudi (DEN), Lougue (LOU),
Doumga Lao (DOL), Lambango (LAM), SaréMaoundé (SAM), DiomandouWalo (DIW).
Rat invasion route (symbols in black): Diakene Wolof (DIK), Diattacounda (DIT),

730 Marsassoum (MAR), Tobor (TOB), BadiNieriko (BAN), Bountougoufara (BOU), Kedougou

731 (KED), Soutouta (SOU), Bransan (BRA), Mako (MAK), Segou (SEG).



Figure 2. Effects of specific invasion status on the variation in (i) hemagglutination (HA), (ii) 733 734 hemolysis (HL) and (iii) haptoglobin (Hp) levels in the serum of Mus musculusdomesticus (boxplots in grey) and Mastomyserythroleucus(boxplots in white). The whiskers denote 1.5 735 Inter-Quartile Range.Legend: Invasive alone = M. m. domesticus in sites of long-established 736 invasion; Invasive IF = M. m. domesticus on invasion front (IF); Native IF = Ma. 737 erythroleucuson IF; Native alone = Ma. erythroleucusin non-invaded sites. The letters above 738 739 boxplots denote the significance of differences between specific invasion statuses: boxplots with the same letter above are no significantly different while boxplots with different letters 740 above are significantly different. 741





Figure 3. Effects of specific invasion status on the variation of serum haptoglobin (Hp) concentration in Rattusrattus(boxplots in grey) and Mastomysspp. (boxplots in white). The whiskers denote 1.5 Inter-Quartile Range.Legend: Invasive alone = R. rattusat sites of longestablished invasion; Invasive IF = R. rattusoninvasion front (IF); Native IF = Ma. erythroleucus(n = 34) and Ma. natalensis(n = 22) on IF; Native alone = Ma. natalensisin non-invaded sites. The letters above boxplots denote the significance of differences between specific invasion statuses: boxplots with the same letter above are no significantly different while boxplots with different letters above are significantly different.

Supplementary materials

Table S1: Results of the linear mixed-effect models for hemagglutination (HA),hemolysis
(HL) and haptoglobin (Hp) variations along the *Mus musculusdomesticus*invasion route. 'i:j'

indicates the interaction between the factors 'i' and 'j'. Significant effects are highlighted in bold. AICc: Akaike's information criterion with correction for finite sample size. Δ : difference between the model chosen and the model with the lowest AICc. N: total number of models selectedas Δ AICc< 2 with respect to the model with the lowest AICc.S: number of models selected in which the factor was significant.

Assay	AICc (Δ)	Factors in the model selected	F-value	p-value	S (N)
	1187.4 (1.07)	Plate	11.27	0.0163	8 (8)
		Climate (component 1)	5.83	< 0.0001	7 (8)
Hemagglutination (HA)		Climate (component 2)	16.81	< 0.0001	8 (8)
		Invasion status	12.48	< 0.0001	8 (8)
		Sex	0.0001	0.9931	8 (8)
		Invasion status:Sex	4.75	0.0029	8 (8)
		Initial hemolysis	99.82	< 0.0001	5 (5)
Hamelusis (III.)	1014.8 (1.71)	НА	50.72	< 0.0001	5 (5)
Hemorysis (HL)		Sex	9.87	0.0018	5 (5)
		Invasion status	26.56	< 0.0001	5 (5)
) 464.7 (0.00)	Climate (component 1)	13.31	0.0003	5 (5)
Haptoglobin (Hp)		Climate (component 2)	22.74	< 0.0001	5 (5)
		Invasion status	5.22	0.0017	5 (5)

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Table S2: Results of the linear mixed-effect models for hemagglutination (HA),hemolysis (HL) and haptoglobin (Hp) variations along the *Rattus. rattus*invasion route. 'i:j' indicates the interaction between the factors 'i' and 'j'. Significant effects are highlighted in bold. AICc: Akaike's information criterion with correction for finite sample size. Δ : difference between the model chosen and the model with the lowest AICc. N: total number of models selectedas Δ AICc< 2 with respect to the model with the lowest AICc.S: number of models selected in which the factor was significant.

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Assay	AICc (Δ)	Factors in the model chosen	F-value	p-value	S (N)
) 1210.3 (0.84)	Serum hemolysis	12.74	0.0004	8 (8)
Hemagglutination (HA)		Age	9.40	0.0024	8 (8)
		Body mass	4.93	0.0270	8 (8)
		Climate (component 1)	12.91	0.0004	8 (8)
		Serum hemolysis	57.76	< 0.0001	16 (16)
Hemolysis (HL)	765.6 (1.33)	НА	355.92	< 0.0001	16 (16)
		Serum hemolysis 12.74 0.0004 Age 9.40 0.0024 Body mass 4.93 0.0270 Climate (component 1) 12.91 0.0004 Serum hemolysis 57.76 < 0.0001 1.33) HA 355.92 < 0.0001 Commensal habitat (component 1) 21.82 < 0.0004	< 0.0001	16 (16)	
Haptoglobin (Hp)	2170.0 (1.64)	Climate (component 1)	12.62	0.0004	10 (16)

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Fig. S2.Principal component analysis (PCA) of climatic (a) and commensal habitat (b) data for categories of localities sampled along the house mouse invasion route. PCAs were based on the uncorrelated climatic (temperatures in °C, rainfall in mm, recorded between 1997 and 2012) and commensal habitat (b) variables remaining after a first PCA (left side). Between-within analysis showed significant classes (Monte-Carlo test, p < 0.05; see figures right side). Legend: max rain:maximum monthly rainfall during rainy season (mean per year); mnMnTM:lowest

monthly minimum temperature (mean per year); S-cement: floor in cement; W-banco: wall in
banco; S-sand: floor in sand; R-banco:: ceiling in banco; R-fiber: ceiling in fibers; localities of
long-established invasion (red); localities of invasion front (blue); non-invaded localities (green).

Temperature data were recorded from local weather stations closest to sampled localities and available on http://www.ncdc.noaa.gov/cdo-web/datasets; rainfall data were recorded from satellite products available on http://richardis.univparis1.fr/precip/rainser1.html with GPCP-1DD as date source. Commensal habitat data were recorded directly on the field during rodent sampling.





Fig. S3.Principal component analysis (PCA) of climatic (a) and commensal habitat (b) data for categories of localities sampled along the rat mouse invasion route. PCAs were based on the uncorrelated climatic (temperatures in °C, rainfall in mm, recorded between 1997 and 2012) and commensal habitat (b) variables remaining after a first PCA (left side). Betweenwithin analysis showed significant classes (Monte-Carlo test, p < 0.05; see figure right side)

only for climatic data.Legend: max MMxT: highest daily maximum temperature (mean per year); min MMnT: lowest daily minimum temperature (mean per year); min Rain: minimum
monthly rainfall during rainy season (mean per year); W-cement: wall in cement;S-cement:
floor in cement;W-banco: wall in banco;S-banco: floor in banco;localities of long-established
invasion (red); localities of invasion front (blue); non-invaded localities (green).

Temperature data were recorded from local weather stations closest to sampled localities and available on http://www.ncdc.noaa.gov/cdo-web/datasets; rainfall data were recorded from satellite products available on http://richardis.univparis1.fr/precip/rainser1.html with GPCP-1DD as date source.Commensal habitat data were recorded directly on the field during rodent sampling.

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