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Reliable estimation of the kinetic parameters of redox enzymes by taking into account mass transport towards rotating electrodes in protein film voltammetry experiments

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Abstract

In Protein Film Voltammetry, a redox enzyme is immobilized on a rotating electrode in a configuration allowing fast, direct electron transfer. This technique is used to probe the mechanism of enzymes by quantitatively interpreting the response in current as a function of the experimental conditions. Limitation by mass transport of the substrate towards the electrode may obscure important features and complicate the analysis of the enzymatic response, so much so that the enzyme has high activity. In this work, we derive equations taking into account mass transport of substrate, for the steady-state current generated by an enzyme following Michaelis-Menten kinetics and immobilized onto a hydrodynamic (e.g. rotating) electrode. We use these equations to model the current response of films of CO-dehydrogenase, a metalloenzyme that catalyzes the oxidation of CO, to transient exposures to its gaseous substrate. We show that neglecting transport yields poor fits and overestimated, unreliable, values of K_m (even when using the Koutecky-Levich approximation), whereas taking into account transport yields much better fits and more reliable parameters. We reinterpret previously published data by taking into account transport limitations.

Keywords: protein film voltammetry; mass transport; CO dehydrogenase; Koutecky-Levich; gaseous substrates

1. Introduction

Protein Film Voltammetry (PFV) has been used for two decades to study the mechanism of redox enzymes[1–3]. In this technique, an enzyme is immobilized onto a rotating electrode in such a way that electron transfer is direct and fast. This technique was used to study catalytic signals, i.e. signals resulting from the catalytic oxidation or reduction of the enzyme's substrate at the electrode. The resulting catalytic current, which is proportional to the enzymatic activity, lends itself well to quantitative interpretation: one can monitor its evolution over time to learn about inactivation/activation processes[4–7], or model its dependence on electrode potential to learn about the steps in the catalytic cycle[8–10]. In catalytic PFV, the substrate is consumed at the electrode by the catalytic reaction. To avoid depletion, electrochemists have therefore used rotating disc electrodes, taking care that the electrode is rotated fast enough that the enzymatic response does not depend on the rotation rate, and hence reflects only the properties of the enzyme. Early reports of the oxidation of H₂ at an electrode modified with an hydrogenase showed a strong dependence of the current on the electrode rotation rate, which was modelled using the Koutecky-Levich equation[11]. Heering and coworkers[12], Reda and Hirst[13], and Léger and Bertrand[2] derived exact expressions for the current of an enzyme following Michaelis-Menten kinetics taking into account mass transport.

CO dehydrogenases are enzymes that employ a NiFe₄S₄ active site to catalyze the reversible oxidation of CO to CO₂[14].

They are extremely fast[15], with turnover rates up to 4×10^4 s⁻¹, which makes them desirable for use in biotechnological devices, where they could be combined for instance with hydrogenases to catalyse the water-gas shift reaction[16]. Distinct CODHs have been studied using PFV in the past decade, such as CODH I and II from *Carboxydotherrmus hydrogenoformans* or CODH from *Desulfovibrio vulgaris*[17–20]. However, the high activity of CODHs introduces a complication for their study using PFV, since many of the electrochemical signals we recorded with *Desulfovibrio vulgaris* CODH adsorbed onto pyrolytic graphite edge electrodes showed signs of limitation by transport, even when rotating the electrode at 6 krpm; we thus decided to include transport limitation in our modelling.

In this work, we derive equations slightly different from those of the previous works[13, 2] for the current given by a redox enzyme following Michaelis-Menten kinetics and immobilized on an electrode, taking into account transport limitation. We use this model to interpret the response of films of *Desulfovibrio vulgaris* CODH to transient exposures to CO, and reinterpret data previously published by others[17]. We show that neglecting mass transport leads to unreliable, overestimated values of the K_m parameter. In addition, we show that using the Koutecky-Levich analysis does not improve the reliability of the determined K_m values, but yields to largely underestimated values. Finally, we propose a simple method to model the data.

2. Formalism

In this paper, we consider the enzymatic oxidation of a substrate at a hydrodynamic electrode. We assume that the hydrodynamic flow towards the electrode imposes a flux of substrate

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that takes the following form[21, 22]:

$$J_t = m(c^b - c^0) \quad (1)$$

where J_t is the flux of substrate towards the electrode, c^b is the concentration of substrate in the ‘‘bulk’’ (i.e. far away from the electrode), c^0 is the concentration at the electrode surface, and m is the mass-transfer coefficient[21], which depends on the nature of the flow and on its velocity.

At the electrode, we consider that the substrate is oxidized by an enzymatic reaction with a rate per molecule of enzyme given by:

$$k = \frac{k_{\text{cat}}}{1 + \frac{K_m}{c^0}} = \frac{k_{\text{eff}}c^0}{1 + \frac{c^0}{K_m}} \quad (2)$$

where k_{cat} the maximum catalytic rate, K_m the Michaelis constant for the oxidation of the substrate, and k_{eff} the catalytic efficiency, defined as:

$$k_{\text{eff}} = \frac{k_{\text{cat}}}{K_m} \quad (3)$$

The electrode, covered with the enzyme with a surface concentration Γ , consumes a flux of substrate given by:

$$J_c = \Gamma \frac{k_{\text{eff}} c^0}{1 + \frac{c^0}{K_m}} \quad (4)$$

We now consider three models that differ in the way the transport is taken into account.

In model **(a)**, the transport is assumed to be so fast that c^0 is always equal to c^b . In that case, the electrical current density is given by:

$$j_{(a)} = n F J = n F \Gamma \frac{k_{\text{eff}} c^b}{1 + c^b/K_m} = n F \Gamma \frac{k_{\text{cat}}}{1 + K_m/c^b} \quad (5)$$

in which n is the number of electrons involved in the oxidation reaction, and F the Faraday constant.

In model **(b)**, we assume that the transport can be taken into account using the Koutecky-Levich approximation, so that:

$$1/j_{(b)} = 1/j_{\text{Levich}} + 1/j_{\text{enzyme}} \quad (6)$$

in which j_{Levich} is the so-called Levich current, completely limited by mass-transport (obtained for $c^0 = 0$) and $j_{\text{enzyme}} = j_{(a)}$ is the current given by the system when the transport is infinitely fast (equation (5)). This gives:

$$j_{(b)} = n F \Gamma \frac{k_{\text{eff}}^{\text{app}} c^b}{1 + c^b/K_m^{\text{app}}} \quad (7a)$$

$$k_{\text{eff}}^{\text{app}} = \frac{k_{\text{eff}}}{1 + \mu/m} \quad (7b)$$

$$K_m^{\text{app}} = K_m (1 + \mu/m) \quad (7c)$$

where we define $\mu = \Gamma k_{\text{eff}}$. Model **(b)** is therefore formally equivalent to model **(a)**, with only a different meaning of the values of K_m and k_{eff} .

In the last model, model **(c)**, we make no hypothesis. Instead we determine the concentration c^0 of substrate at the electrode using the conservation of mass. This model gives therefore the true value of the current, unlike models **(a)** and **(b)**

which are approximations. Under steady state conditions, $J_c = J_t$, which gives the following equation for c^0 :

$$m(c^b - c^0) = \Gamma \frac{k_{\text{eff}} c^0}{1 + \frac{c^0}{K_m}} = \frac{\mu}{1 + \frac{c^0}{K_m}} c^0 \quad (8)$$

Computing c^0 and substituting into either (1) or (4) yields the following expression for the electrical current density:

$$j_{(c)} = n F \Gamma \frac{k_{\text{eff}} K_m}{1 + \frac{K_m}{c^0}} \quad (9a)$$

$$c^0 = c^b \frac{\gamma + \sqrt{\gamma^2 + 4 \frac{K_m}{c^b}}}{2} \quad (9b)$$

$$\gamma = 1 - \frac{K_m}{c^b} \left(1 + \frac{\mu}{m}\right) \quad (9c)$$

Similar expressions, using k_{cat} rather than k_{eff} , have been derived before[12, 13, 2]. Expression (9) has the advantage of making it clear which non-dimensional parameters influence the current: c^b/K_m and μ/m .

Below, we compare the dependence of the current on the normalized concentration c^b/K_m for models **(b)** and **(c)**, and we study how the current is affected by the value of μ/m , which quantifies the competition between enzymatic substrate oxidation and transport. If μ/m is greater than 1, then catalysis is faster than transport, and vice-versa for values lower than 1. For $\mu/m = 0$, both model **(b)** and model **(c)** fall back to model **(a)**, which does not take transport into account. Under these conditions, the dependence of the current on the bulk concentration truly reflects the properties of the enzyme.

When c^b tends to infinity, regardless of the model used, the current density reaches a maximum value j_{max} :

$$j_{\text{max}} = n F \Gamma k_{\text{eff}} K_m \quad (10)$$

We used this value to normalize the current density for all models.

Figure 1a shows curves of j/j_{max} against c^b/K_m for several values of μ/m for model **(b)** (equation (7), dotted lines) and for model **(c)** (equation (9), solid lines). All curves show a similar behaviour, with a linear region at low concentration and a saturation at high concentration. The current is not significantly affected by the limitation by transport up to values of μ/m around 0.3, but it deviates significantly for higher values of μ/m . Increasing μ/m increases the concentration at which saturation is reached, and decreases the slope of the initial linear response. Regarding the comparison between models **(b)** and **(c)**, figure 1a shows that, apart from the initial linear region where both models give the same values, the Koutecky-Levich model (model **(b)**) systematically underestimates the current with respect to the exact model (model **(c)**).

The current in the initial linear region can be computed by considering the limit $c^b \rightarrow 0$ in equation (9) (model **(c)**), which gives:

$$j = n F \frac{\Gamma k_{\text{eff}} c^b}{1 + \mu/m} \quad (11)$$

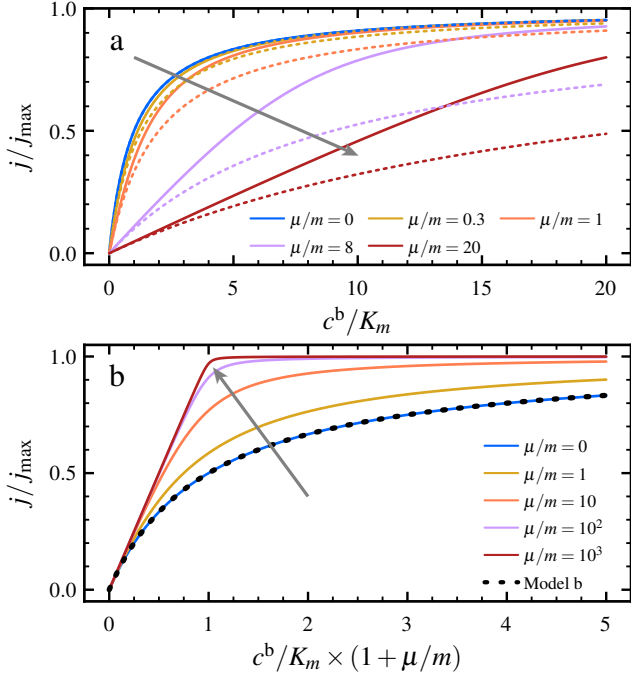


Figure 1: Panel a: normalized current density of model (b) (dotted lines) or model (c) (solid lines) as a function of either the normalized concentration c^b/K_m for different values of the μ/m ratio. Panel b: normalized values of the current density of model (c) as a function $c^b/K_m \times (1 + \mu/m)$ for different values of μ/m . In this representation, model (b) gives a curve independent of μ/m , drawn as thick dark dots. In both panels, the arrow points in the direction of increasing μ/m .

Model (b) also gives the same expression for the current in the linear region.

In figure 1a, we have plotted the normalized current for only a small range of values of μ/m , since increasing this value yields curves with very low initial slopes and saturations at too high concentrations. Equation (11) suggests that plotting the current as a function of $(1 + \mu/m) \times c^b/K_m$ should make it easier to compare curves with very different values of μ/m , as the initial linear regions should all match, as should the plateaus at $j/j_{\max} = 1$. We have done so in figure 1b. In this representation, the curves given by model (b) do not depend on μ/m anymore, and match that of model (c) at $\mu/m = 0$. Figure 1b shows that as the limitation by the transport becomes more important (the μ/m ratio increases), the transition between the initial linear region and the plateau current becomes more abrupt, until, at high values of μ/m , the graph becomes two lines that intersect at $(1 + \mu/m) \times c^b/K_m = 1$. This sharp transition is reproduced neither by model (a), that does not take transport into account, nor by model (b), for which the modelling of transport is too crude; this sharp transition is characteristic of transport limitations.

3. Experimental

Protein purification

Desulfovibrio vulgaris CO dehydrogenase was purified as before[19].

Electrochemistry

Electrochemical experiments were conducted as before[20].

Data modelling

To model experimental data with equation (9), it is more convenient to model the current directly rather than the current density, which is just the current multiplied by A the electrode area. We used QSoas for data fitting, using the following Ruby function:

```
def itrprt(cb, km, nFAM, nFAMu)
  cg = cb - km * (1 + nFAMu/nFAM)
  c0 = 0.5*(cg + (cg**2 + 4*km*cb)**0.5)
  return nFAMu * km/(1 + km/c0)
end
```

To use this function, we saved it as a function.rb text file and loaded it using the command:

```
ruby-run function.rb
```

The fits were run using the following command:

```
fit-arb itrprt(s,km,nFAM,nFAMu)+io /with=s:1,exp
```

The `/with=s:1,exp` specifies that the parameter s varies according to equation (12). km is the K_m parameter, $nFAM$ is the value of $nFAm$ and $nFAMu$ that of $nFA\mu$ (it is not possible to determine m or μ from the fit without knowing A). We use `+io` to account for a slight offset with respect to $I = 0$. Fitting equation (5) to data can be achieved using the following command:

```
fit-arb nFAMu*s/(km+s)+io /with=s:1,exp
```

4. Results

CO dehydrogenase from *Desulfovibrio vulgaris* forms stable and very active films on pyrolytic graphite edge electrodes[20]. Using a rotating disc electrode configuration, we have submitted such films to injections of aliquots of CO in the electrochemical buffer. Upon injection, the concentration of CO in the bulk increases instantly, and then the exchange between the electrochemical buffer and the atmosphere, facilitated by the electrode rotation, results in an exponential decrease of the concentration over time[23, 24]:

$$c^b = \begin{cases} c^{\text{inj}} \exp\left(-\frac{t-t_0}{\tau}\right) & \text{if } t > t_0, \\ 0 & \text{otherwise.} \end{cases} \quad (12)$$

where c^{inj} is the injected concentration, t_0 the time of the injection and τ the time constant of the decrease of CO. This makes it possible to probe the response of CODH to a large range of concentrations of CO in a single experiment.

Figure 2 shows the current monitored at a fixed potential produced by an electrode modified with a film of CODH submitted to a single injection of an aliquot of CO. Initially, there is no substrate, and therefore no current. The injection at $t = t_0 = 0$ results in the appearance of a CO oxidation current that has

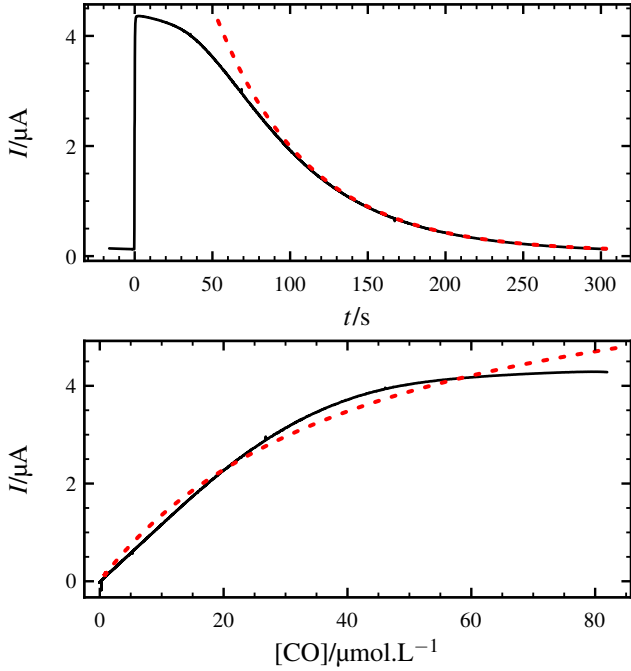


Figure 2: Panel a: current generated by a film of *Dv* CODH submitted to a single injection of $83 \mu\text{M}$ CO at $t = 0$. The red trace is the exponential fit to the tail of the current ($\tau = 60.8 \pm 0.3$ s). Panel b: current as a function of the concentration of CO deduced from the time constant of the fit in panel a. The red trace is a fit of model (a) (equation (5)). Conditions: $T = 25^\circ\text{C}$, pH 7, $\omega = 4$ krpm, electrode potential: -310 mV.

a vaguely sigmoidal shape, with an initial plateau, an inflexion point, and a perfect mono-exponential tail. The latter mirrors the exponential decrease of the concentration of CO, because, as the concentration decreases, it reaches low enough values that the current becomes proportional to the concentration (equation (11)). Therefore, fitting a mono-exponential decay to the tail of the current yields the time constant τ of the decay of the bulk concentration of CO. This makes it possible to determine the whole evolution of the concentration of CO over time, using equation (12), because both c^{inj} and t_0 are known.

We have taken advantage of this possibility to plot the current as a function of the bulk concentration of CO in figure 2b. While the $I = f([\text{CO}])$ curve follows a saturation behaviour, with a large region in which the current depends linearly on the concentration before reaching a plateau at higher concentrations, the transition between the linear region and the plateau is more abrupt than that of model (a), which neglects transport limitation. This is attested by the poor fit of model (a) (equation (5)) to the data (red dashed curve in figure 2b). In fact, the experimental $I = f([\text{CO}])$ trace of figure 2b is much closer in appearance to the $\mu/m = 8$ model (c) trace of figure 1a.

To determine the K_m of the CODH under the experimental conditions used, there are two strategies: (i) one can fit model (c) (equation (9)) to the dependence of the current on the bulk concentration of CO (figure 2b), which was itself deduced by extrapolating the exponential decrease fitted in the tail of the current. (ii) alternatively, one can directly substitute

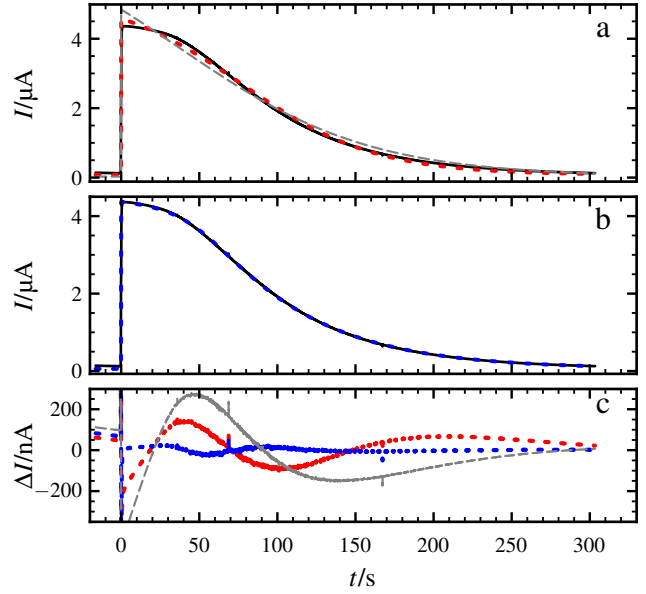


Figure 3: Panels a & b, black trace: same data as in figure 2a. Panel a, red dashed trace: fit of equation (5) (model (a)) with c^b given by (12) to the data; dashed gray trace: fit of the same model, but with the value of τ fixed to that determined by the exponential fit in figure 2a. Panel b, blue dashed trace: fit of equation (9) (model (c)) with c^b given by (12) to the data. Panel c: residuals of the fit, with the same color code as in panels a&b. Conditions as in figure 2. Parameters of the fit ($A = 4.15 \text{ mm}^2$ is the area of the electrode), panel a, red trace: $K_m = 13.7 \pm 0.3 \mu\text{M}$, $nFA\Gamma k_{\text{eff}} = 380 \pm 70 \text{ nA}\cdot\mu\text{M}^{-1}$ (corresponding to $I_{\text{max}} = 5.24 \pm 0.03 \mu\text{A}$), $\tau = 42.7 \pm 0.3$ s; gray trace: $K_m = 45 \pm 1 \mu\text{M}$, $nFA\Gamma k_{\text{eff}} = 164 \pm 1 \text{ nA}\cdot\mu\text{M}^{-1}$ (corresponding to $I_{\text{max}} = 7.4 \pm 0.3 \mu\text{A}$), τ fixed to 60.8 s. Panel b: $K_m = 2.2 \pm 0.1 \mu\text{M}$, $nFA\Gamma k_{\text{eff}} = 2.1 \pm 0.1 \mu\text{A}\cdot\mu\text{M}^{-1}$ (corresponding to $I_{\text{max}} = 4.50 \pm 0.01 \mu\text{A}$), $nFAm = 127 \pm 2 \text{ nA}\cdot\mu\text{M}^{-1}$, $\tau = 60.8 \pm 0.3$ s.

equation (12) into equation (9) and fit the resulting expression to the raw data. As the equation for model (b) is strictly equivalent to that of model (a), fitting with model (a) in fact gives the values of K_m for both models, provided μ/m is determined independently.

We chose option (ii), since it requires only one fit, and therefore is less prone to systematic errors. Moreover, this is also how we proceeded before[23] (but using model (a)). We also followed the same strategy but using model (a) (equation (5)) to the data to assess the extent to which neglecting completely transport affects the quality of the fits and of the determined parameters.

Figure 3a shows a fit of model (a) to the data plotted in figure 2a. The fit is not very good, much more symmetric around the inflexion point than the data really are. Moreover, the exponential decrease is not well reproduced, and the fitted model decreases faster than the real current, which means that the extrapolated concentration of CO is not correct. Imposing the correct value of τ , as determined from the fit in figure 2 yielded even poorer fits (gray lines in figure 3a and c). On the other hand, fitting model (c) to the data (figure 3b) yields a perfect fit that reproduces both the exponential decrease and the noticeable asymmetry around the inflexion point (the value of $\tau = 60.8$ s determined from this fit is the same as that determined directly using a simple mono-exponential decay). The

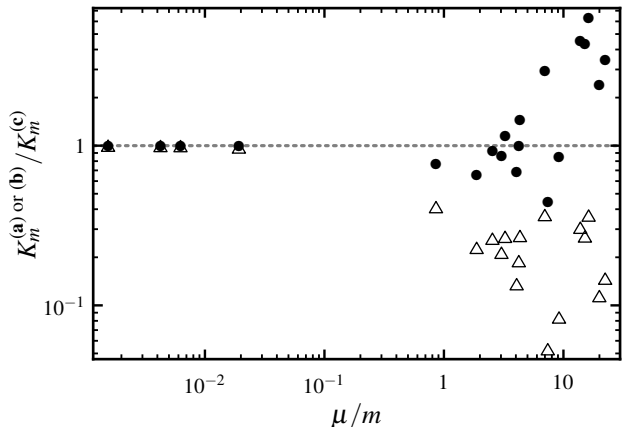


Figure 4: Systematic fits of models (a) and (c) to responses to injections of CO of $D\nu$ CODH films. Full circles: values of $K_m^{(a)}/K_m^{(c)}$ as a function of μ/m (determined from fits of model (c)). Open triangles: values of $K_m^{(b)}/K_m^{(c)}$ as a function of μ/m ; the value of $K_m^{(b)}$ is determined from equation (7c) and the value of μ/m determined from fits of model (c). Conditions variable, pH 7, 25°C, electrode potential from -460 mV to -410 mV.

better quality of the fit¹ the full model is confirmed by the inspection of the residuals in figure 3c, since those for model (c) (in blue) are much smaller and evenly distributed around zero than those of model (a) (in red). From the fitted value of $nFAm$ (A is the electrode area, $A = 0.0415 \text{ cm}^2$), one can compute a value of the mass transport coefficient of $m = 16 \times 10^{-3} \text{ cm}\cdot\text{s}^{-1}$. For a rotating disc electrode, the mass transport coefficient takes the form[21]:

$$m = 0.62D^{2/3}\omega^{1/2}\nu^{-1/6} \quad (13)$$

where D is the diffusion coefficient of the CO, ω the angular velocity of the RDE, and ν is the kinematic viscosity of the solution. As the kinematic viscosity of water of $\nu = 10^{-2} \text{ cm}^2\cdot\text{s}^{-1}$ and the rotation rate is 4 krpm, we obtain a coefficient for the diffusion of CO in the electrochemical buffer of $D_{\text{CO}} = 1.4 \times 10^{-5} \text{ cm}^2\cdot\text{s}^{-1}$, well in the expected range for the diffusion of a small molecule in water, and compatible with the value of $D_{\text{CO}} = 2 \times 10^{-5} \text{ cm}^2\cdot\text{s}^{-1}$ determined at 20°C by Wise and Houghton from the measure of the rate of collapse of small bubbles in water[25].

As model (c) includes (a) and the known limitation by transport, the K_m values determined via model (c) are necessarily closer to the real values. In the above example, the error in the K_m value from fitting model (a) to the data rather than model (c) is large, almost an order of magnitude. This prompted us to study in a systematic fashion the error on K_m resulting from wrongly fitting model (a) to chronoamperograms in which the limitation by transport cannot be neglected. We have run a number of experiments like that in figure 2, under various conditions. For each, we have fitted both model (a) and model (c) to the resulting current traces. We have obtained two distinct values of K_m : $K_m^{(a)}$ and $K_m^{(c)}$. Using the values of μ/m determined from the

fit of model (c), we could also compute $K_m^{(b)}$, the value of K_m deduced from the Koutecky-Levich approximation. Figure 4 shows the ratios $K_m^{(a)}/K_m^{(c)}$ (dots) and $K_m^{(b)}/K_m^{(c)}$ (open triangles) as a function of μ/m , collected over a range of experimental conditions. For low μ/m ratios, all three models are equivalent and give exactly same value for the K_m , as attested by a ratio of 1. However, for $\mu/m > 1$ there is large discrepancy between the “true” values and the values given by models (a) and (b), as attested by the significant deviation from the $y = 1$ horizontal line. Using model (a) or (b) tends to overestimate or underestimate respectively the K_m values.

5. Discussion

The use of very active films of redox enzymes in Protein Film Voltammetry can result in depletion of substrate at the electrode, complicating the analysis of the electrocatalytic current. We have derived an equation for the current given by a redox enzyme that follows Michaelis-Menten kinetics immobilized on a hydrodynamic electrode, under conditions where transport limitation is not neglected (model (c), equation (9)). Our approach differs from the previous ones in that we chose to parametrize the enzymatic activity using the catalytic efficiency k_{eff} rather than the maximum turnover rate. We show that the parameter that governs substrate depletion is the ratio $\Gamma k_{\text{eff}}/m$. Increasing this ratio makes the dependence of current on bulk concentration continuously change from the true Michaelis-Menten enzymatic response when $\Gamma k_{\text{eff}}/m$ is low to a linear response that abruptly saturates to give a plateau when $\Gamma k_{\text{eff}}/m$ is high (figure 1b). Increasing the electroactive coverage or using more active enzymes amplifies the effects of substrate limitation, while increasing the mass-transport coefficient m (by increasing the rotation rate) decreases the effects of substrate limitation. We note that increasing the $\Gamma k_{\text{eff}}/m$ ratio increases the range in which the electrode responds linearly in current to the concentration of substrate, which may be desirable for the construction of sensors with a large operating range.

We showed that using the Koutecky-Levich approximation (model (b), equation (7)) does not change the shape of the $j = f(c^b)$ curve with respect to the model not taking transport limitation into account (model (a), equation (5)); it just changes the interpretation of the values of K_m and k_{eff} determined from the fit, while requiring an independent measure of the μ/m ratio. In particular, the Koutecky-Levich model does not reproduce the sharp transition between the low-concentration linear regime and the saturation typical of mass-transport limited configurations (at high values of μ/m).

We have applied this model to the specific case of a gas processing enzyme, CO-dehydrogenase. Injections of aliquots of buffer saturated with the substrate in a RDE electrochemical cell results in an exponential decrease over time of the substrate concentration which may be exploited to probe the enzymatic response over a large range of substrate concentration in a single experiment[23].

We have taken advantage of this possibility to characterize CO oxidation by *Desulfovibrio vulgaris* CO dehydrogenase.

¹Based on the assumption of a standard deviation of 60 nA in the current, the χ^2 “goodness-of-fit” is 1 for model (c) and 10^{-20} for model (a).

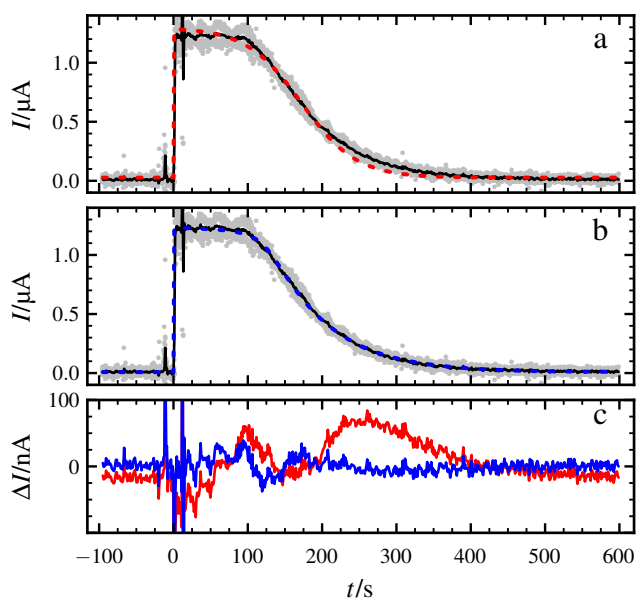


Figure 5: Panels a & b, gray dots: data from figure S2 of supplementary information of ref. 17, black line: same data, smoothed. Panel a, red dashed trace: original modelling of the data in ref. 17, using model (a). The authors did not fit model (a) directly to the raw data, but rather used a linear fit to $\ln(I_{\max}/I - 1)$. Panel b, blue dashed trace: fit of model (c) to the data. Panel c: residuals of the fit, with the same color code as in panels a&b. Parameters of the fit (A , the area of the electrode, is unknown, and we assumed from figure S2ii that the injected concentration of CO is 300 μM), panel a: $K_m = 2.9 \mu\text{M}$, $nFA\Gamma k_{\text{eff}} = 428 \text{ nA}\cdot\mu\text{M}^{-1}$, $\tau = 39 \text{ s}$. Panel b: $K_m = 1.6 \pm 0.4 \mu\text{M}$, $nFA\Gamma k_{\text{eff}} = 575 \pm 2 \text{ nA}\cdot\mu\text{M}^{-1}$, $nFAm = 22 \pm 1 \text{ nA}\cdot\mu\text{M}^{-1}$, $\tau = 75 \pm 2 \text{ s}$. Adapted with permission from ref. 17. Copyright 2007 American Chemical Society.

We have shown that fitting model (a) or model (b) to experimental data yields poor fits that misrepresent the exponential decay of the current in the linear part of the response and fail to reproduce the sharp transition between the initial saturation and the subsequent decrease (figure 3a). On the contrary, fitting model (c) (equation (9)) to experimental data yields very good fits with meaningful values of K_m and correct values of m (figure 3b). As m is actually determined from the fit, this removes the need for the independent determination of m , which would require measuring a diffusion coefficient.

We systematically compared fits of model (a) (neglecting transport) to fits of model (c). We showed that using model (a) yields overestimated and dispersed K_m values when the μ/m parameter is above 1, whereas the fits of model (c) yielded much more reproducible parameter values. Fitting model (b) to the data does not improve reliability, since models (a) and model (b) are strictly equivalent. However, the determination of K_m values from fits of model (b) requires independent determination of the value of m , and yields greatly underestimated values.

Parkin and coworkers have used model (a) to analyse experiments similar to that in figure 2, to determine the value of K_m for *Carboxydotherrnus hydrogenoformans* CODH I [17]. Figure 5 shows the response in current of a film of *C. hydrogenoformans* CODH I to an injection of an aliquot of CO-saturated buffer

(adapted from ref. 17, figure S2). The resulting current is very similar to the data shown in figure 2a, including the asymmetry around the inflexion point that is indicative of mass-transport limitation.

We have reproduced the original data modelling using (a) in figure 5a, and we have fitted model (c) to the data in figure 5b. As can be seen from the residuals (figure 5c), the fit of model (c) to the data is significantly better than the original modelling; moreover, it reproduces the asymmetry of the signal. Model (a) yields a K_m value two times higher than that of model (c). This error is small, considering the very high $\mu/m \approx 20$ ratio. This results from the fortuitous compensation of the natural propensity of model (a) to overestimate K_m values with the two-fold error in the time constant for the decrease of CO.

Parkin et al obtained an estimate of K_m by averaging over 13 experiments. The value of $\mu/m \approx 20$ that we determined from the experimental data in figure 5 suggest that the authors may have observed some variability in the fitted values of K_m , which would have been much smaller had model (c) been used.

To ease the modelling of experimental data, we propose in the Experimental section a simple method using the data analysis program QSoas [26] to analyze data.

6. Conclusion

We have analysed the responses in current of CODH-modified electrodes to transient exposures to CO to determine the K_m value of the enzyme. We show that failing to take into account mass transport yields to unreliable (generally overestimated) values of K_m , whereas using the Koutecky-Levich approximation yields to unreliable, underestimated values of K_m . We derived an equation taking into account the mass transport of substrate, and show that it models perfectly the data, yielding meaningful values of the K_m and of the transport parameters.

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