



## Genetic and phenotypic heterogeneity suggest therapeutic implications in SCN2A-related disorders

Markus Wolff, Katrine M. Johannesen, Ulrike B. S. Hedrich, Silvia Masnada, Guido Rubboli, Elena Gardella, Gaetan Lesca, Dorothée Ville, Mathieu Milh, Laurent Villard, et al.

### ► To cite this version:

Markus Wolff, Katrine M. Johannesen, Ulrike B. S. Hedrich, Silvia Masnada, Guido Rubboli, et al.. Genetic and phenotypic heterogeneity suggest therapeutic implications in SCN2A-related disorders. Brain - A Journal of Neurology , 2017, 140 (5), pp.1316-1336. 10.1093/brain/awx054 . hal-01668653

**HAL Id: hal-01668653**

**<https://amu.hal.science/hal-01668653>**

Submitted on 23 Mar 2018

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Genetic and phenotypic heterogeneity suggest therapeutic implications in SCN2A-related disorders

Markus Wolff,<sup>1,\*</sup> Katrine M. Johannesen,<sup>2,3,\*</sup> Ulrike B. S. Hedrich,<sup>4,\*</sup> Silvia Masnada,<sup>5</sup> Guido Rubboli,<sup>2,6</sup> Elena Gardella,<sup>2,3</sup> Gaetan Lesca,<sup>7,8,9</sup> Dorothée Ville,<sup>10</sup> Mathieu Milh,<sup>11,12</sup> Laurent Villard,<sup>12</sup> Alexandra Afenjar,<sup>13</sup> Sandra Chantot-Bastaraud,<sup>13</sup> Cyril Mignot,<sup>14</sup> Caroline Lardennois,<sup>15</sup> Caroline Nava,<sup>16,17</sup> Niklas Schwarz,<sup>4</sup> Marion Gérard,<sup>18</sup> Laurence Perrin,<sup>19</sup> Diane Doummar,<sup>20</sup> Stéphane Auvin,<sup>21,22</sup> Maria J. Miranda,<sup>23</sup> Maja Hempel,<sup>24</sup> Eva Brilstra,<sup>25</sup> Nine Knoers,<sup>25</sup> Nienke Verbeek,<sup>25</sup> Marjan van Kempen,<sup>25</sup> Kees P. Braun,<sup>26</sup> Grazia Mancini,<sup>27</sup> Saskia Biskup,<sup>28</sup> Konstanze Hörtnagel,<sup>28</sup> Miriam Döcker,<sup>28</sup> Thomas Bast,<sup>29</sup> Tobias Loddenkemper,<sup>30</sup> Lily Wong-Kissel,<sup>31</sup> Friedrich M. Baumeister,<sup>32</sup> Walid Fazeli,<sup>33</sup> Pasquale Striano,<sup>34</sup> Robertino Dilella,<sup>35</sup> Elena Fontana,<sup>36</sup> Federico Zara,<sup>37</sup> Gerhard Kurlmann,<sup>38</sup> Joerg Klepper,<sup>39</sup> Jess G. Thoene,<sup>40</sup> Daniel H. Arndt,<sup>41</sup> Nicolas Deconinck,<sup>42</sup> Thomas Schmitt-Mechelke,<sup>43</sup> Oliver Maier,<sup>44</sup> Hiltrud Muhle,<sup>45</sup> Beverly Wical,<sup>46</sup> Claudio Finetti,<sup>47</sup> Reinhard Brückner,<sup>48</sup> Joachim Pietz,<sup>49</sup> Günther Golla,<sup>50</sup> Dinesh Jillella,<sup>51</sup> Karen M. Linnet,<sup>52</sup> Perrine Charles,<sup>53</sup> Ute Moog,<sup>54</sup> Eve Öglane-Shlik,<sup>55</sup> John F. Mantovani,<sup>56</sup> Kristen Park,<sup>57</sup> Marie Deprez,<sup>58</sup> Damien Lederer,<sup>58</sup> Sandrine Mary,<sup>58</sup> Emmanuel Scalais,<sup>59</sup> Laila Selim,<sup>60</sup> Rudy Van Coster,<sup>61</sup> Lieven Lagae,<sup>62</sup> Marina Nikanorova,<sup>2</sup> Helle Hjalgrim,<sup>2,3</sup> G. Christoph Korenke,<sup>63</sup> Marina Trivisano,<sup>64</sup> Nicola Specchio,<sup>64</sup> Bertien Ceulemans,<sup>65</sup> Thomas Dorn,<sup>66</sup> Katherine L. Helbig,<sup>67</sup> Katia Hardies,<sup>68,69</sup> Hannah Stamberger,<sup>68,69,70</sup> Peter de Jonghe,<sup>68,69,70</sup> Sarah Weckhuysen,<sup>68,69,70</sup> Johannes R. Lemke,<sup>71</sup> Ingeborg Krägeloh-Mann,<sup>1</sup> Ingo Helbig,<sup>45,72</sup> Gerhard Kluger,<sup>73,74,#</sup> Holger Lerche<sup>4,#</sup> and Rikke S. Møller<sup>2,3,#</sup>

\*,# These authors contributed equally to this work.

Mutations in *SCN2A*, a gene encoding the voltage-gated sodium channel Na<sub>v</sub>1.2, have been associated with a spectrum of epilepsies and neurodevelopmental disorders. Here, we report the phenotypes of 71 patients and review 130 previously reported patients. We found that (i) encephalopathies with infantile/childhood onset epilepsies ( $\geq 3$  months of age) occur almost as often as those with an early infantile onset ( $< 3$  months), and are thus more frequent than previously reported; (ii) distinct phenotypes can be seen within the late onset group, including myoclonic-atonic epilepsy (two patients), Lennox-Gastaut not emerging from West syndrome (two patients), and focal epilepsies with an electrical status epilepticus during slow sleep-like EEG pattern (six patients); and (iii) West syndrome constitutes a common phenotype with a major recurring mutation (p.Arg853Gln: two new and four previously reported children). Other known phenotypes include Ohtahara syndrome, epilepsy of infancy with migrating focal seizures, and intellectual disability or autism without epilepsy. To assess the response to antiepileptic therapy, we retrospectively reviewed the treatment regimen and the course of the epilepsy in 66 patients for which well-documented medical information was available. We find that the use of sodium channel blockers was often associated with clinically relevant seizure reduction or seizure

freedom in children with early infantile epilepsies (<3 months), whereas other antiepileptic drugs were less effective. In contrast, sodium channel blockers were rarely effective in epilepsies with later onset ( $\geq 3$  months) and sometimes induced seizure worsening. Regarding the genetic findings, truncating mutations were exclusively seen in patients with late onset epilepsies and lack of response to sodium channel blockers. Functional characterization of four selected missense mutations using whole cell patch-clamping in tsA201 cells—together with data from the literature—suggest that mutations associated with early infantile epilepsy result in increased sodium channel activity with gain-of-function, characterized by slowing of fast inactivation, acceleration of its recovery or increased persistent sodium current. Further, a good response to sodium channel blockers clinically was found to be associated with a relatively small gain-of-function. In contrast, mutations in patients with late-onset forms and an insufficient response to sodium channel blockers were associated with loss-of-function effects, including a depolarizing shift of voltage-dependent activation or a hyperpolarizing shift of channel availability (steady-state inactivation). Our clinical and experimental data suggest a correlation between age at disease onset, response to sodium channel blockers and the functional properties of mutations in children with *SCN2A*-related epilepsy.

- 1 Department of Pediatric Neurology and Developmental Medicine, University Children's Hospital, Tübingen, Germany
- 2 The Danish Epilepsy Centre, Dianalund, Denmark
- 3 Institute for Regional Health Services, University of Southern Denmark, Odense, Denmark
- 4 Department of Neurology and Epileptology, Hertie Institute for Clinical Brain Research, University of Tübingen, Tübingen, Germany
- 5 Department of Brain and Behavior, University of Pavia, Italy
- 6 University of Copenhagen, Copenhagen, Denmark
- 7 Department of Genetics, Lyon University Hospital, Lyon, France
- 8 Claude Bernard Lyon I University, Lyon, France
- 9 Lyon Neuroscience Research Centre, CNRS UMR55292, INSERM U1028, Lyon, France
- 10 Department of Pediatric Neurology and Reference Center for Rare Children Epilepsy and Tuberous Sclerosis, Hôpital Femme Mere Enfant, Centre Hospitalier Universitaire de Lyon, HCL, France
- 11 APHM Service de neurologie pédiatrique, Marseille, France
- 12 Aix Marseille Univ, Inserm, GMGF, UMR-S 910, Marseille, France
- 13 AP-HP, Unité de Génétique Clinique, Hôpital Armand Trousseau, Groupe Hospitalier Universitaire de l'Est Parisien, Paris, France
- 14 AP-HP, Département de Génétique; Centre de Référence Déficiences Intellectuelles de Causes Rares; Groupe de Recherche Clinique UPMC "Déficiences Intellectuelles et Autisme" GH Pitié-Salpêtrière, Paris, France
- 15 Service de Pédiatrie neonatale et Réanimation - Neuropédiatrie, 76000 Rouen, France
- 16 Sorbonne Universités, UPMC Univ Paris 06, UMR S 1127, Inserm U 1127, CNRS UMR 7225, ICM, France
- 17 Department of Genetics, Pitié-Salpêtrière Hospital, AP-HP, F-75013 Paris, France
- 18 Service de Génétique Clinique, CHU Caen, France
- 19 Department of Genetics, Robert Debré Hospital, AP-HP, Paris, France
- 20 AP-HP, Service de Neuropédiatrie, Hôpital Trousseau, Paris, France
- 21 Université Paris Diderot, Sorbonne Paris Cité, INSERM UMR1141, Paris, France
- 22 AP-HP, Hôpital Robert Debré, Service de Neurologie Pédiatrique, Paris, France
- 23 Department of Pediatrics, Herlev University Hospital, Herlev, Denmark
- 24 Institute of Human Genetics, University Medical Center Hamburg-Eppendorf, Hamburg, Germany
- 25 Department of Genetics, University Medical Center Utrecht, Utrecht, The Netherlands
- 26 Department of Pediatric Neurology, Brain Center Rudolf Magnus, University Medical Center Utrecht, The Netherlands
- 27 Department of Clinical Genetics, Erasmus University Medical Center, Rotterdam, The Netherlands
- 28 CeGaT - Center for Genomics and Transcriptomics, Tübingen, Germany
- 29 Epilepsy Center Kork, Kehl, Germany
- 30 Division of Epilepsy and Clinical Neurophysiology, Boston Children's Hospital, Harvard Medical School, Boston MA, USA
- 31 Division of Child and Adolescent Neurology, Department of Neurology, Mayo Clinic, Rochester MN, USA
- 32 Children's Hospital, RoMed Klinikum, Rosenheim, Germany
- 33 Pediatric Neurology, University Hospital Cologne, Germany
- 34 Pediatric Neurology and Muscular Diseases Unit, Department of Neurosciences, Rehabilitation, Ophthalmology, Genetics, and Maternal and Child Health, University of Genoa 'G. Gaslini' Institute, Genova, Italy
- 35 Servizio di Epilettologia e Neurofisiopatologia Pediatrica, UO Neurofisiopatologia, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Milano, Italy
- 36 Centro di Diagnosi e Cura delle Epilessie Infantili, Azienda Ospedaliera -Policlinico Gianbattista Rossi, Verona, Italy
- 37 Laboratory of Neurogenetics and Neuroscience, Department of Neuroscience, "G. Gaslini" Institute, Genova, Italy
- 38 Department of Pediatric Neurology, University Children's Hospital, Münster, Germany
- 39 Children's Hospital, Klinikum Aschaffenburg, Germany
- 40 University of Michigan, Pediatric Genetics, Ann Arbor, MI USA

- 41 Division of Pediatric Neurology and Epilepsy – Beaumont Children’s Hospital, William Beaumont Oakland University School of Medicine, Royal Oak, Michigan, USA
- 42 Department of Neurology, Hôpital Universitaire des Enfants Reine Fabiola, Université Libre de Bruxelles, Brussels, Belgium
- 43 Children’s Hospital Lucerne, Luzerner Kantonsspital, Kinderspital Luzern, CH-6000 Luzern 16, Switzerland
- 44 Department of child neurology, Children’s Hospital, St. Gallen, Switzerland
- 45 Department of Neuropediatrics, University Medical Center Schleswig-Holstein, Christian-Albrechts University, Kiel, Germany
- 46 Gillette Children’s Specialty Healthcare, Saint Paul, MN, USA
- 47 Klinik für Kinder- und Jugendmedizin, Elisabeth-Krankenhaus, Essen, Germany
- 48 Wangen, Germany
- 49 Pediatric Practice University Medical Center for Children and Adolescents, Angelika Lautenschläger Children’s Hospital, Heidelberg, Germany
- 50 Klinik für Kinder- und Jugendmedizin, Klinikum Lippe GmbH, Detmold, Germany
- 51 Division of Epilepsy and Clinical Neurophysiology, Department of Neurology, Boston Children’s Hospital, Boston, MA, USA
- 52 Department of Pediatrics, Aarhus University hospital, Aarhus, Denmark
- 53 Department of Genetics and Cytogenetics, Assistance Publique-Hôpitaux de Paris, Groupe Hospitalier Pitié-Salpêtrière Charles-Foix, Paris, France
- 54 Institute of Genetics, University Hospital, Heidelberg, Germany
- 55 Children’s Clinic, Institute of Clinical Medicine, University of Tartu, Tartu, Estonia
- 56 Department of Pediatrics and Mercy Kids Autism Center, Mercy Children’s Hospital, St. Louis, Missouri, USA
- 57 Department of Pediatrics and Neurology, Children’s Hospital Colorado, University of Colorado School of Medicine, Aurora, CO, USA
- 58 Centre de Génétique Humaine, Institut de Pathologie et Génétique, Gosselies, Belgium
- 59 Pediatric Neurology Unit, Pediatric Department, Centre Hospitalier de Luxembourg, Luxembourg
- 60 Department of Pediatrics, Pediatric Neurology and Neurometabolic Unit, Cairo University Children Hospital, Cairo, Egypt
- 61 Department of Pediatrics, Division of Pediatric Neurology and Metabolism, Ghent University Hospital, Ghent, Belgium
- 62 Department of Development and Regeneration, Section Pediatric Neurology, University Hospital KU Leuven, Leuven, Belgium
- 63 Zentrum für Kinder- und Jugendmedizin (Elisabeth Kinderkrankenhaus), Klinik für Neuropädiatrie u. Angeborene, Stoffwechselerkrankungen, Oldenburg, Germany
- 64 Neurology Unit, Department of Neuroscience, Bambino Gesù Children’s Hospital, IRCCS, Rome, Italy
- 65 Paediatric Neurology University Hospital and University of Antwerp, Wilrijkstraat 10, 2650 Edegem, Belgium
- 66 Swiss Epilepsy Center, Zurich, Switzerland
- 67 Division of Clinical Genomics, Ambry Genetics, Aliso Viejo, California, USA
- 68 Neurogenetics Group, Center for Molecular Neurology, VIB, Antwerp, Belgium
- 69 Laboratory of Neurogenetics, Institute Born-Bunge, University of Antwerp, Antwerp, Belgium
- 70 Division of Neurology, University Hospital Antwerp (UZ), Antwerp, Belgium
- 71 Institute of Human Genetics, University of Leipzig Hospitals and Clinics, Leipzig, Germany
- 72 Division of Neurology, The Children’s Hospital of Philadelphia, Philadelphia, USA
- 73 Neuropediatric Clinic and Clinic for Neurorehabilitation, Epilepsy Center for Children and Adolescents, Schoen Klinik, Vogtareuth, Germany
- 74 PMU Salzburg, Austria

Correspondence to: Markus Wolff,  
 Department of Pediatric Neurology and Developmental Medicine,  
 University Children’s Hospital, Hoppe-Seyler-Str. 1,  
 72076 Tübingen,  
 Germany  
 E-mail: Markus.Wolff@med.uni-tuebingen.de

Correspondence may also be addressed to: Rikke Steensbjerre Møller,  
 The Danish Epilepsy Centre Filadelfia,  
 Kolonivej 1, 4293 Dianalund,  
 Denmark  
 E-mail: rimo@filadelfia.dk

**Keywords:** SCN2A; epilepsy; epilepsy genetics; treatment response; sodium channel blockers

**Abbreviations:** ACTH = adrenocorticotrophic hormone; AED = antiepileptic drug; B(F)NIS = benign (familial) neonatal/infantile seizures; EIMFS = epilepsy of infancy with migrating focal seizures; ESES = electrical status epilepticus during slow sleep; SCB = sodium channel blocker

# Introduction

The *SCN2A* gene encodes the voltage-gated sodium channel  $\text{Na}_v1.2$ , one of the major neuronal sodium channels that play a role in the initiation and conduction of action potentials.  $\text{Na}_v1.2$  is expressed in axon initial segments and nodes of Ranvier of myelinated nerve fibres in early development, and in the adult brain in the axon initial segment and unmyelinated axons (Boiko *et al.*, 2001, 2003; Kaplan *et al.*, 2001; Liao *et al.*, 2010b). Accordingly, *SCN2A* mutations have been mainly shown to affect the early developmental period (Catterall, 2014), but some mutations have also been found as causes of later onset neurological diseases (Kobayashi *et al.*, 2012; Horvath *et al.*, 2016), or a combination of both (Schwarz *et al.*, 2016).

Since the first description of a patient with epilepsy caused by a *SCN2A* mutation and the findings of *SCN2A* mutations in benign (familial) neonatal/infantile seizures [B(F)NIS] (Sugawara *et al.*, 2001; Heron *et al.*, 2002), the phenotypic spectrum has expanded considerably. In particular, severe phenotypes with encephalopathy have been reported, including distinct epileptic syndromes such as Ohtahara syndrome (Nakamura *et al.*, 2013; Allen *et al.*, 2016), epilepsy of infancy with migrating focal seizures (EIMFS) (Howell *et al.*, 2015), infantile spasms (Ogiwara *et al.*, 2009; Wong *et al.*, 2015) or West syndrome (Allen *et al.*, 2013; Nakamura *et al.*, 2013), as well as patients with unclassified severe epilepsy phenotypes. However, *SCN2A* mutations have also been found in patients with intellectual disability and/or autistic features without epilepsy, suggesting the possible involvement of the gene in the aetiology of autism spectrum disorders (Sanders *et al.*, 2012; Li *et al.*, 2016).

To date, the mechanisms for the phenotypic heterogeneity, ranging from benign to very severe clinical presentations, are poorly understood. Differences in functional effects of the mutations may account at least in part for the phenotypic diversity. In addition, the efficacy of anti-epileptic drugs (AEDs), especially of sodium channel blockers (SCBs), could be influenced by the way in which specific *SCN2A* mutations affect  $\text{Na}_v1.2$  activity.

Therefore, we aimed to assess systematically the phenotypic spectrum and treatment effects in a large cohort of *SCN2A*-related disorders comprising 201 patients, 71 of whom were not reported previously. For some missense mutations that we selected based on specific clinical findings, and supported by previous reports from the literature, we were able to correlate phenotype and treatment responses to the specific biophysical effects of the mutations.

## Materials and methods

### Previously unpublished patients

Seventy-one previously unreported patients with a *SCN2A* mutation were included in this study. Patients were referred through a network of collaborating clinicians and geneticists.

Mutations in *SCN2A* were identified in research or diagnostic laboratories and assumed to be pathogenic, if they were non-synonymous, splice-site altering, nonsense or frameshift changes, predicted damaging by one or more prediction software (PolyPhen-2, SIFT and MutationTaster), seen less than twice in >60 000 controls in the exome aggregation consortium browser (exac.broadinstitute.org), and either occurred *de novo*, or were inherited from an affected parent, an unaffected mosaic parent or previously reported as pathogenic in other patients. Sanger sequencing was used to confirm all mutations and perform segregation analysis. The study was approved by the local ethics committees.

Referring physicians were provided with a standardized phenotyping sheet to assess clinical characteristics, EEG, and neuroimaging findings. Seizures were diagnosed according to the International League Against Epilepsy commission on classification (Berg *et al.*, 2010), and were assigned, whenever possible, to defined epileptic syndromes. Data on cognitive development and neurological features were recorded at age at onset and at last evaluation. Based on the presence and severity of epilepsy, cognitive status and age at onset of epilepsy patients were classified into the following groups: (i) B(F)NIS, defined as neonatal/infantile onset seizures with a seizure offset during infancy/early childhood, and/or autosomal-dominant inheritance, and normal cognitive development; (ii) encephalopathy with early infantile epilepsy, defined as seizure onset before the age of 3 months, and impaired cognitive development; (iii) encephalopathy with infantile/childhood epilepsy, defined as seizure onset at the age of at least 3 months, and impaired cognitive development; and (iv) intellectual disability and/or autism without epilepsy.

Antiepileptic treatment data were retrospectively assessed by standardized questionnaires. The effect on seizures was classified according to the judgement of the treating physicians into seizure freedom, seizure reduction, no effect or seizure worsening. Particular attention was given to the effects of SCBs, defined as AEDs that reduce the activity of sodium channels by stabilizing an inactivated state. SCBs included phenytoin, carbamazepine, oxcarbazepine, lacosamide, lamotrigine and zonisamide. To provide a general overview in our retrospective analysis, we specifically assessed whether patients were on an SCB by the time that seizure reduction, seizure freedom, or aggravation of seizures occurred.

### Frequency of *SCN2A*-related disorders

To estimate the frequency of *SCN2A* mutations causing the reported phenotypes in the general population, we used the electronic population databases of National Statistics at the Statens Serum Institute (Denmark) to calculate the birth cohort from 2007 to 2014. The Danish Epilepsy Centre is the only tertiary hospital in Denmark specialized in the treatment of epilepsy, and the majority of patients with presumed genetic epilepsy are referred to this centre for genetic testing.

### Literature review

We searched PubMed using the term ‘*SCN2A*’ and included all relevant patient-related information in our *SCN2A* dataset.



Last search date was 1 June 2016. Papers not available in English, Italian or Danish were excluded. Cases with deletions and duplications spanning *SCN2A* as well as neighbouring genes were excluded. For patients with little or no clinical information, we listed the phenotype mentioned in the respective publication.

## Mutagenesis

To engineer the mutations into the adult splice variant of the human Na<sub>v</sub>1.2 channel, site-directed mutagenesis was performed using Quickchange<sup>®</sup> II XL (Agilent Technologies; primers are available upon request) as described previously (Schwarz *et al.*, 2016). Transfection of the  $\alpha$ -subunit together with pCLH-h $\beta$ 1-EGFP and pCLH-h $\beta$ 2-CD8 in tsA201 cells using Mirus TransIT<sup>®</sup>-LT1 reagent was performed in a standard way as described previously (Liao *et al.*, 2010a; Lauxmann *et al.*, 2013; Schwarz *et al.*, 2016).

## Electrophysiology

Standard whole-cell patch clamp recordings were performed using an Axopatch 200B amplifier, a Digidata 1320A digitizer and pCLAMP 8 data acquisition software (Axon Instruments), as described before (Schwarz *et al.*, 2016). Borosilicate glass pipettes had a final tip resistance of 1–2 M $\Omega$  when filled with internal recording solution containing (in mM): 130 CsF, 5 NaCl, 2 MgCl<sub>2</sub>, 5 EGTA, HEPES (pH 7.4, 290 mOsm). The bath solution contained (in mM): 140 NaCl, 4 KCl, 1 MgCl<sub>2</sub>, 2 CaCl<sub>2</sub>, 5 HEPES, 4 dextrose (pH 7.4, 300 mOsm). We carefully checked that the maximal voltage error due to residual series resistance after up to 95% compensation was always <5 mV. Voltage clamp protocols to study channel kinetics were performed as described previously (Schwarz *et al.*, 2016) and are provided in detail in the [Supplementary material](#).

## Data and statistical analysis

Traces were displayed off-line with Clampfit software of pClamp 10.0 (Axon Instruments). Graphics were generated using a combination of Microsoft Excel, and Origin (version 9.1; OriginLab Inc., USA) software, statistics were performed using SigmaStat 3.1 (Systat Software GmbH, Germany). All data were tested for normal distribution. For statistical evaluation, ANOVA on ranks (Kruskal-Wallis-test) with Dunn's *post hoc* test for not normally distributed data or one-way ANOVA (Bonferroni *post hoc* test) was used when datasets were normally distributed. All data are shown as means  $\pm$  standard error of the mean (SEM), *n* gives the number of cells. We applied the  $\chi^2$  test to estimate the significance of the differences in AED treatment effects in the two groups of epilepsy with encephalopathy with early and late onset.

## Results

In the present study, we report 71 unpublished patients with pathogenic *SCN2A* mutations and review the phenotypes of 130 previously reported ones (see [Tables 1–4](#),

[Supplementary Table 1](#) and [Supplementary Fig. 1](#)). The distribution of phenotypes of the 66 previously unpublished patients with epilepsy and of the total number of patients are displayed in [Fig. 1](#).

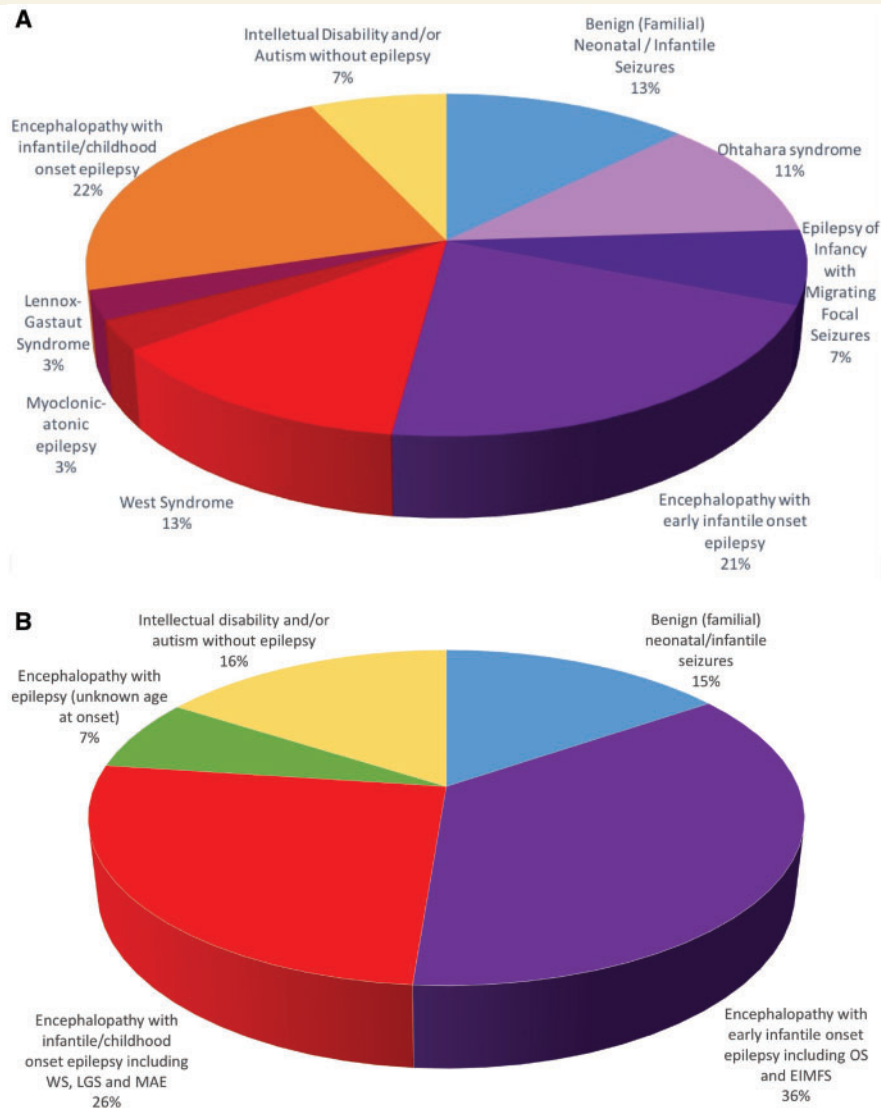
## Phenotypic features

### Benign (familial) neonatal/infantile seizures

We identified nine unpublished patients ([Table 1](#)) and 24 probands from the literature with B(F)NIS due to a *SCN2A* mutation, as well as 109 mutation-positive family members (Berkovic *et al.*, 2004; Striano *et al.*, 2006; Herlenius *et al.*, 2007; Heron *et al.*, 2010; Liao *et al.*, 2010b; Lemke *et al.*, 2012; Lauxmann *et al.*, 2013; Zara *et al.*, 2013; Grinton *et al.*, 2015; Johannesen *et al.*, 2016; Schwarz *et al.*, 2016). The mutations occurred *de novo* in 6/33 of the probands. Age at seizure onset ranged from the first day of life to 23 months. Approximately half of the children had seizure onset within the first month of life. Seizure types were predominantly focal clonic, tonic, and generalized tonic-clonic, frequently occurring in clusters. Interictal EEG showed mostly focal or multifocal spikes, but was normal in some cases. All children became seizure-free at a median age of 5 months (range 5 days to 2 years), and remained seizure-free with normal cognitive development until last follow-up at a median age of 5.5 years (range 7 months to 34 years, data available from 28 cases). A single proband developed a second epilepsy phenotype during later childhood with marked activation of multifocal spikes during sleep and partial cognitive deterioration, resembling electrical status epilepticus during slow sleep (ESES), and two had isolated seizures until the age of 2 and 14 years, respectively. Five children with two recurrent mutations (A263V and R1882G) exhibited episodic ataxia later in life (Liao *et al.*, 2010a; Johannesen *et al.*, 2016; Schwarz *et al.*, 2016).

### Encephalopathy with early infantile epilepsy

Twenty-eight new patients ([Table 2](#)) and 43 previously published ones (Ogiwara *et al.*, 2009; Dhamija *et al.*, 2013; Nakamura *et al.*, 2013; Touma *et al.*, 2013; Baasch *et al.*, 2014; Martin *et al.*, 2014; Matalon *et al.*, 2014; Zerem *et al.*, 2014; Fukasawa *et al.*, 2015; Howell *et al.*, 2015; Allen *et al.*, 2016; Trump *et al.*, 2016) had epilepsy onset within the first 3 months of life. Thirty-one had an identifiable epilepsy syndrome, i.e. Ohtahara syndrome (18 cases) or EIMFS (13 cases). The remaining 40 patients had unclassifiable epilepsies. The predominant seizure types in these were focal, tonic, and tonic-clonic seizures or spasms. Initial EEGs showed a suppression burst pattern in 25 cases (18 with Ohtahara syndrome, two with EIMFS, and five with unclassifiable epilepsies), and multifocal spikes in the majority of the remaining cases. Regardless of the epileptic syndrome, all patients fulfilled the criteria of encephalopathy as they had intellectual disability, being severe in 54/71 cases. Six children had autism



**Figure 1** Distribution of patients according to phenotype and age at epilepsy onset. **(A)** Epilepsy phenotypes in the previously unpublished cohort ( $n = 66$ ). 'Patients with Lennox-Gastaut syndrome' refers to patients with Lennox-Gastaut syndrome not evolving from West syndrome. **(B)** Phenotypes in the overall cohort ( $n = 201$ ). LGS = Lennox-Gastaut syndrome; MAE = myoclonic-atonic epilepsy; OS = Ohtahara syndrome; WS = West syndrome.

spectrum disorder. Additional features included muscular hypotonia ( $n = 32$ ), microcephaly ( $n = 15$ ), marked dystonic or choreatic movement disorders ( $n = 8$ ), spasticity ( $n = 3$ ), or dysautonomia ( $n = 5$ ). Seven patients in this subgroup were deceased at time of follow-up, and causes of death included severe infections and status epilepticus.

### Encephalopathy with infantile/childhood epilepsy

This group included 29 unpublished (Table 3) and 29 previously published patients (Haug *et al.*, 2001; Sugawara *et al.*, 2001; Kamiya *et al.*, 2004; Ogiwara *et al.*, 2009; Shi *et al.*, 2009; Kobayashi *et al.*, 2012; Allen *et al.*, 2013; Nakamura *et al.*, 2013; Sundaram *et al.*, 2013; Hackenberg *et al.*, 2014; D'Gama *et al.*, 2015; Howell *et al.*, 2015; Mercimek-Mahmutoglu *et al.*, 2015;

Samanta and Ramakrishnaiah, 2015; Wong *et al.*, 2015; Dimassi *et al.*, 2016; Horvath *et al.*, 2016). Sixteen presented with West syndrome or infantile spasms, which evolved into Lennox-Gastaut syndrome in 5/16 patients. Two cases were diagnosed as Dravet syndrome, two as Lennox-Gastaut syndrome and two as myoclonic-atonic epilepsy. The majority of the remaining patients with seizure onset after 3 months of age had unclassifiable epilepsies mainly with generalized seizure types including generalized tonic-clonic (15/36, occurring in clusters in four), absence ( $n = 12$ ) and myoclonic seizures ( $n = 8$ ). EEG showed mainly generalized spikes and waves or multifocal spikes. Interestingly, four patients with unclassifiable epilepsies (Patients 41, 61, 62 and 65) and one patient (Patient 50) with West syndrome showed an ESES-like marked

**Table 1 Clinical characteristics and treatment response of the previously unpublished patients: B(F)NIS**

Patient	Mutation/ inheritance	Age at seizure onset	Initial seizure type	Other seizure types	EEG	MRI	Cognition onset/ follow-up	Neuro- logical features	Addi- tional features	Age at last follow-up	Seizure outcome (offset: age)	Seizure- free	Treatment effects	No effect
1	A202V/maternal	1 d	BFNS	F, GTC	NAV	N	NA/N	N		4 y 6 m	Sz free (2 m)		PB, PHT, TPM, CBZ	VPA, LEV
2	G828V/de novo	2 d	BNS	F, C, AU (cluster)	MF spikes	N	NA/N	N		14 m	Sz free (2 m)	PHT→OXC		PB, PP, TPM, LEV
3	Q1531K/de novo	3 d	BNS	S, C, GTC	NAV	NA	NA/N	N		14 m	Sz free (5 d)	PB		
4	S863F/de novo	5 d	BNS→ other	GTC	GTCs, NA→MF spikes, ESES-like	N	NA/N	N	ADS	11 y	Sz free (3 m → 10 y)	OXC→ST	LTG, LEV	
5	D343G/de novo	6 d	BNS	GTC (cluster)	F	N	NA/N	N		18 m	Sz free (6 m)	OXC		LEV, PB, PP, CLB
6	F207S/de novo	9 d	BNS	C	TCS	N	NA/N	N		18 m	Sz free (6 m)	PHT	TPM	PB, CLZ, OXC
7	V261M/de novo	9 d	BNS	GTC	N	N	NA/N	N		20 m	Sz free (3 m)	CLB	PHT, LEV	B6, VPA
8	R36G/maternal	16 m	BFIS	F (cluster)	None	NA	N/N	N		3 y	Sz free (2 y)			
9	R36G/maternal	23 m	BFIS	F	Slowing	NA	N/N	N		4 y 5 m	Sz free (NAV)	ZNS		

ADS = attention deficit disorder; AU = autistic; BNS = benign neonatal seizures; BFNS = benign familial infantile seizures; BNS = benign neonatal seizures; BNS = benign familial infantile seizures; C = clonic; F = focal; FD = focal dyscognitive; GTC = generalized tonic-clonic; I = left; m = months; MF = multifocal; N = normal; NA = not applicable; NAV = not available; Par = parietal; SE = status epilepticus; Sz = seizures; TCS = tonic-clonic seizures; y = years; → = change to. Treatment (sodium channel blockers are highlighted in bold): B6 = vitamin B6; CBZ = carbamazepine; CLB = clobazam; CLZ = clonazepam; LTG = lamotrigine; LEV = levetiracetam; OXC = oxcarbazepine; PB = phenobarbital; PHT = phenytoin; PP = pyridoxal phosphate; ST = sulthiame; TPM = topiramate; VPA = valproate; ZNS = zonisamide.

activation of spikes and waves during sleep, prompting AED treatment to reduce spike-wave activity in order to prevent ESES-induced neuropsychological sequelae. Cognition before seizure onset varied from normal to severely delayed, and was reported abnormal during follow-up in all cases, with severe cognitive impairment in 35/58. Autism spectrum disorder was found in 18 children. Additional features included muscular hypotonia ( $n = 16$ ) and marked choreatic or dyskinetic movement disorder in six patients.

### Encephalopathy with unspecified onset of epilepsy

In 10 of the previously published patients (Need *et al.*, 2012; Wang *et al.*, 2012; Carvill *et al.*, 2013; Saitoh *et al.*, 2015; Li *et al.*, 2016), data were limited, and age at seizure onset was not available. One case was classified as Lennox-Gastaut syndrome, the others as encephalopathies with epilepsy that were not further characterized.

### Intellectual disability and/or autism without epilepsy

This subgroup consisted of patients with a verified SCN2A mutation, but no signs of epileptic seizures. Five unpublished (Table 4) and 27 previously published cases fulfilled these criteria (Weiss *et al.*, 2003; Rauch *et al.*, 2012; Sanders *et al.*, 2012; Jiang *et al.*, 2013; Tavassoli *et al.*, 2014; Codina-Sola *et al.*, 2015; D’Gama *et al.*, 2015; Carroll *et al.*, 2016; Li *et al.*, 2016). The unpublished cases, aged 4 to 13 years, exhibited autism with moderate to severe intellectual disability. From the previously published individuals, 14 had autism, nine had intellectual disability without autism, and four had schizophrenia.

### Estimated frequency of SCN2A mutations in the Danish population

Via the electronic population databases of National Statistics at the Statens Serum Institute (Denmark), we calculated the birth cohort from 2006–14, giving 550 261 births. In the same period at least seven Danish children with an SCN2A mutation causing the reported phenotypes were born, making a total minimum frequency of approximately 1/78 608 births.

### Seizure outcome and treatment effects

Antiepileptic treatment effects on seizures were analysed in all unpublished patients with epilepsy for which sufficiently detailed clinical information was available ( $n = 66$ , Tables 1–4). Besides classical AEDs, corticosteroids or adrenocorticotrophic hormones (ACTH) were tried in 19 children, ketogenic diet in 13, vagal nerve stimulation in three, and immunoglobulins in one.



**blue 2 Clinical characteristics and treatment response of the previously unpublished patients: encephalopathy with early onset epilepsy**

Patient	Mutation/ inheritance	Age at Epilepsy seizure syndrome onset	Initial seizure type	Other seizure types	EEG	MRI	Cognition onset / follow-up	Neurological features	Additional features	Age at last follow-up	Seizure Outcome (offset: age)	Treatment effects		
												Seizure- free	Seizure reduction	No effect
0	V423L/de novo	1 d	OS	M, AP	T	SB→MF spikes	NA	Hypotonia	MC	34 m (deceased)	Intractable	BR		B6, PR, MDZ, LEV, PHT, RGB, KD, CBZ, LCM PB, B6, PP, VGB, TPM, LEV
1	E999K/de novo	1 d	OS→other T	M, TCS	M, TCS	SB→beta→f spikes	N	Dystonia	Oculogyric crises	5 y	Sz free (1 m, relapses with low PHT levels)	PHT		PHT, B6, PP, TPM, LEV
2	Q181I/de novo	1 d	OS→other T	GTCs, S, F	SB→Spikes I	N	NA/MD	Hypotonia, unsteady gait	ASD	8 y	Sz free (4 y)	LTG + VPA + LEV	PHT, TPM	VGB
3	M1548V/de novo	1 d	OS→WS	T	GTC	SB→HA→slowing	N	Hypotonia	ASD	18 m	Intractable	TPM		PB, PHT, B6, PP, LEV, VGB, CBZ
4	I237N/de novo	1 d	other	F	(variable onset)	MF spikes, slowing	HM	Hypotonia	Poor eye contact	3 y 9 m	Intractable	VPA, CBZ		VGB, LEV, TPM, LTG, B6, PHT, PB, CLZ
5	V887A/de novo	1 d	OS→VS	M	S	SB→HA	N	Hypotonia		15 m	Sz free (6 m, relapse with low PHT levels)	PHT	TPM	PB, CLZ PB, ST, CS
6	G882R/de novo	1 d	EIMFS	unilateral	T	MF spikes, ictal pattern r / l	N	Hypotonia	MC	10 m	Intractable		PHT, LCM, ZNS	LEV, CS, B6, PP
7	I1640S/de novo	1 d	Other	T	TC r/l	Spikes r	NA	NAV		9 y	Sz free (7 y)	LCM		B6, PB, VPA, LEV
8	K908E/de novo	1 d	Other	M	S	Gen + MF spikes, slowing	At	Nystagmus, hypotonia, dystonia	MC	8 y	Sz free (7 y 6 m)		LTG, TPM	LEV, ZNS
9	R1882Q/ unknown	1 d	Other	NA	F →; C	MF + bil spikes, poly spikes	Subtle CD	N	MC	10 y 6 m	Intractable	CBZ		PB, LEV, OXC, TPM, VPA, CLB, LTG, PHT
0	V1627M/de novo	2 d	EIMFS	T r/l migrating	F, AP	MF spikes→N	N	Hypotonia		14 m	Sz free (2 m)	VGB		LEV, B6, PP, TPM, PB, VPA
21	R856Q/de novo	2 d	OS	T	T	SB→SW	Sinus thrombosis, ischaemia CC	NAV	Connatal sinus thrombosis	3 m (deceased)	Intractable	MDZ		LEV, PB, PHT, B6, VGB, TPM
22	A1500T/de novo	2 d	OS→ other	F, T	C	SB→spikes I te→slowing	HM, At	Spasticity	Poor eye contact, MC	13 y	Sz free (3 m), relapse (12 m), sz free (13 m), relapse (5 y), sz free (10 y)	TPM (3 m), ACTH (13 m)	LTG, VPA	
23	M1545V/de novo	2 d	EIMFS	T, C r/l migrating	F, TCS	SB→MF spikes→N	N	Hypotonia		12 m	Sz free (3 m, relapses with low PHT levels)	PHT→CBZ	LEV	PB, B6, PP, VPA
24	E430A/de novo	2 d	Other	F		MF spikes	N	Hypotonia	Regression, ASD	22 m	Sz free (12m)	PHT		TPM, CLB
25	S1536R/de novo	2 d	Other	T	AP, SE, TCS	MF spikes, occ slowing	N	Hypotonia		2 y 8 m	Sz free (6 m, relapses with low PHT levels)	PHT		TPM, PB, LEV, B6
26	F1597L/de novo	3 d	EIMFS	T r/l migrating	T→S	SB→spikes r/l te, slowing	At, calcifications	Dystonia	Dysautonomia, irritability	3 y	Intractable		PHT, CBZ, LTG, BR	B6, PB, LEV, VGB, KD, ST, VPA, LCM, ZNS
27	V424L/de novo	3 d	Other	F	TCS	SB→MF biocc, slowing	HM	Hypotonia	Poor eye contact	4 y	Sz free (15 m)	CBZ	CS	PB, VPA
28	I891T/de novo	3 d	Other	T		MF spikes, bisynchrony	N	Coordination difficulties	ADS	3 y	Sz free (3 m)	PHT		B6, PP, CBZ, CLZ, MDZ, TPM, VPA, ACTH
29	E999K/de novo	3 d	Other	T r/l		SB→gen SW	At, HM	Dystonia	ASD, nystagmus	22 y	Sz free (3 y)	CBZ		PB
30	G882E/de novo	4 d	EIMFS	F, AU, HC r/l M, C migrating	T, AA,	MF spikes, slowing	At, HM	Dystonia	Dysautonomia	8 y	Intractable		PHT, LCM	VPA, LEV, FB, GBR, CLB, CLZ, MSX, TPM, PB, KD, CS, LI, ME TPM, others
31	R1319Q/ unknown	5 d	Other	M	S	MF spikes, slowing	HM	Hypotonia, limb hypertonia		14 m	Sz free (13 m)	ACTH		
32		6 d	other	S	GTCS, FM	SW fr→MF spikes	N	Hypotonia		8 y	Sz free (4 m)	PHT		

(continued)

able 2 Continued

Patient Mutation/ inheritance	Age at Epilepsy seizure syndrome onset	Initial seizure type	Other seizure types	EEG	MRI	Cognition onset / follow-up	Neurological features	Additional features	Age at last follow-up	Seizure Outcome (offset: age)	Treatment effects			
											Seizure- free	Seizure reduction	No effect	Wor- sening
R1629H/de novo														
33 V423L/de novo	6 d	OS	F	TCS, T r/l SB→MF spikes, slowing	T2H	NA/SD	Spasticity	MC	4 y	Intractable	OXC		CLZ, MDZ, VPA, LEV, VGB, B6 PB, LEV, VGB, CS, ZNS, RUF, VPA, TPM, LTG	
34 A263V/de novo	3 w	other	S	T, GTC Atypical HA→MF spikes, slowing	At, HM	N/SD	Spasticity	MC, scoliosis	13 y (deceased)	Intractable	ACTH,NZR CS,MDZ, CLB, LTG, VPA, CLZ,TPM		PB, PHT, TPM, VNS	
35 F1651C/ mosaic/ de novo	6 w	other	GTC	C, SE MF spikes	N	N/MID	Hypotonia		9 m	Sz free (3 m, relapses with low PHT levels)	PHT		PB, OXC, TPM, LEV, B6, PP	
36 R1319Q/de novo	2 m	other	GTC	FC SW, slowing	HM	N/MD	N	ASD	3 y 10 m	Sz free (8 m)	VGB		LEV	
37 F895S/de novo	2 m	other	S, M	C, GTC SB→MF spikes	N	SD/SD	Hypotonia	Optic atrophy	4 y 8 m	Intractable			VGB, PB, TPM, VPA, CLB	

AA = atypical absences; A = atonic; AB = absences; ADS = autism spectrum disorder; ASD = attention deficit disorder; AU = autistic seizures; At = atrophy; Bif = bifrontal; Bil = bilateral; C = clonic; CC = corpus callosum; CD = cortical dysplasia; Ce = central; DA = drop attacks; ED = epileptiform discharges; F = focal; FD = focal convulsion; fr = frontal; Gen = generalized; GTC = generalized tonic-clonic; HA = hypsarrhythmia; HC = hemidonic; HM = hypomyelination; IS = infantile spasms; L = left; m = months; MID = mild intellectual disability; MD = moderate intellectual disability; M = myoclonic; MC = microcephaly; MF = multifocal; N = normal; NA = not applicable; NAV = not available; NCSE = non-convulsive status epilepticus; Occ = occipital; OS = Ohtahara syndrome; Par = parietal; R = right; S = spasms; SE = status epilepticus; SB = suppression burst; SD = severe intellectual disability; SW = spike and waves; Sz = seizures; T = tonic; T2H = T<sub>2</sub>-hyperintensities; TCS = tonic-clonic seizures; Te = temporal; w = week; y = years; → = change to.

Treatment (sodium channel blockers are highlighted in bold): AZA = acetazolamide; B6 = vitamin B6; BR = bromide; CBZ = carbamazepine; CLB = clobazam; CLZ = clonazepam; CS = corticosteroids; ESM = ethosuximide; FBM = felbamate; GBP = gabapentin; IVIG = intravenous immunoglobulins; KD = ketogenic diet; LCM = lacosamide; LI = lidocaine; LTG = levetiracetam; MDZ = Midazolam; ME = metilexine; MSX = mesuximide; OXC = oxcarbazepine; PB = phenobarbital; PHT = phenytoin; PP = pyridoxal phosphate; RGB = retigabine; RUF = rufinamide; ST = sulthiame; STP = stiripentol; TPM = topiramate; VGB = vigabatrin; VNS = vagal nerve stimulation; VPA = valproate; ZNS = zonisamide.

### Benign (familial) neonatal/infantile seizures

All children became seizure-free at a median age of 3 months (range: 5 days to 2 years). All but one remained seizure-free until last follow-up at a median age of 20 months (range: 14 months to 11 years) ( $n = 9$ , Table 1). Patient 4 developed an ESES-like picture at the age of 8 years, and became seizure-free again at the age of 10 years. Seizures stopped spontaneously in three patients, and with AED treatment in six. The sequence of events strongly suggested that seizure freedom was reached by treatment and not by natural history in those cases. Phenytoin was effective in two patients, oxcarbazepine in two, zonisamide in one and clobazam in one. Initial AED treatment failed in seven cases, and six received more than two AEDs (mean 4.3) before the seizures stopped.

### Encephalopathy with early infantile epilepsy with onset younger than 3 months

Seizure freedom has so far been achieved during the first year of life in 11 children, and during childhood in another six (median observation period after seizure freedom: 2.5 years, range 1 month to 18 years) ( $n = 28$ , Table 2). Effective AEDs in terms of seizure freedom included phenytoin ( $n = 8$ ), ACTH ( $n = 2$ ), and carbamazepine, lacosamide, vigabatrin, topiramate as well as a combination of lamotrigine, valproate and levetiracetam in single cases each. One child with Ohtahara syndrome (Patient 11) and one with EIMFS (Patient 23) became rapidly seizure-free after application of phenytoin at the age of 1 and 3 months, respectively, and the burst suppression pattern on EEG disappeared. In Patient 23, low plasma levels of phenytoin ( $<13$  mg/l) resulted in seizure relapse on several occasions. With a high dosage of phenytoin (15 mg/kg/d, divided in three daily doses) higher phenytoin serum levels were obtained, and seizure freedom was finally achieved. A switch to high dose carbamazepine (45 mg/kg/d) was successful during follow-up. Seizure relapses due to low phenytoin plasma levels were also seen on several occasions in Patient 11 and three other children (Patients 15, 25 and 35) during follow-up. Patient 26 showed prompt resolution of burst suppression pattern and temperature instability with phenytoin, but had ongoing seizures although with markedly reduced frequency. AED-related seizure reduction was evident in another 16 cases, most frequently with topiramate ( $n = 6$ ), phenytoin ( $n = 3$ ), and carbamazepine ( $n = 4$ ). Ineffective AEDs included phenobarbital ( $n = 18$ ), levetiracetam ( $n = 17$ ), topiramate ( $n = 13$ ), and valproate ( $n = 11$ ) (Fig. 2A and Table 2).

### Encephalopathy with infantile/childhood epilepsy with onset at 3 months or older

Ten children became seizure-free during follow-up (median observation period after seizure freedom: 3.5 years, range 1 month to 16 years) ( $n = 29$ , Table 3). Eight of nine patients with West syndrome were resistant to treatment, including steroids or ACTH in six. One child (Patient 46) responded to ACTH, but later developed drug-resistant Lennox-

Gastaut syndrome. Among the patients with other epilepsy phenotypes, different AEDs were effective in single cases each, including levetiracetam ( $n = 2$ ) and valproate ( $n = 2$ ). Seizure reduction occurred most frequently with levetiracetam ( $n = 9$ ), benzodiazepines ( $n = 9$ ), and valproate ( $n = 7$ ). Ineffective AEDs included lamotrigine ( $n = 10$ ), valproate ( $n = 10$ ), phenobarbital ( $n = 9$ ), and topiramate ( $n = 9$ ). Drug-induced aggravation of seizures occurred in seven children with carbamazepine ( $n = 3$ ), oxcarbazepine ( $n = 2$ ), phenytoin, lamotrigine and rufinamide, and remitted after discontinuation of the respective AED (see also Fig. 2B and Table 3). Atypical absences were the predominant seizure types in these. In a boy with myoclonic-astonic epilepsy (Patient 52), the frequency of drop attacks and tonic seizures increased markedly after introduction of oxcarbazepine, and lamotrigine provoked episodes of status epilepticus.

### Response to sodium channel blocker versus non-sodium channel blocker treatment

We tested the significance of effects of SCBs versus non-SCBs on seizure outcome of patients with encephalopathy and epilepsy with onset  $<3$  months and  $\geq 3$  months by applying a  $\chi^2$  test (Supplementary Table 3). Treatment with phenytoin and carbamazepine or with all SCBs considered together showed a significantly better response for patients with onset of epilepsy  $<3$  months than for those with onset  $\geq 3$  months ( $P < 0.01$  and  $P < 0.001$ , respectively). In contrast, patients with epilepsy onset  $\geq 3$  months responded significantly better to non-SCBs ( $P < 0.001$ ).

### Impact of the genetic diagnosis on treatment decisions

In most patients, genetic diagnosis was only made late during follow-up and had no impact on treatment decisions, because patients were already seizure-free or many AEDs had been tried before. In some cases, however, suspicion or confirmation of a mutation in *SCN2A* led to specific treatment trials with SCBs: three children (Patients 15, 23 and 25) with severe early onset epilepsies and one (Patient 2) with *de novo* BNIS became seizure-free with phenytoin, and three children with severe early onset EIMFS (Patients 26 and 30) or Ohtahara syndrome (Patient 33) showed partial responses to some SCBs, whereas other types of AEDs had failed before. In contrast, three children with late onset epilepsies (Patients 39, 40 and 41) showed no effect on SCB trials.

## Genetic findings

Mutations were missense in all children with B(F)NIS or encephalopathies with epilepsy onset  $<3$  months. The majority of the missense mutations (both inherited and *de novo*) affects highly evolutionarily conserved amino acids and no obvious correlation between the position of the mutation and the severity of the associated phenotype was observed. In the subgroup of infantile/childhood

**Table 3 Clinical characteristics and treatment response of the previously unpublished patients: encephalopathy with late onset epilepsy**

Patient	Mutation/ inheritance	Age at seizure onset	Epilepsy syndrome	Initial seizure type	Other seizure types	EEG	MRI	Cognition onset /follow-up	Neurological features	Additional features	Age at last follow-up (offset: age)	Seizure outcome (offset: age)	Treatment effects Seizure- free	Sz reduction	No effect	Wors- ening
38	G899S/de novo	3 m	Other	HC	GTC, FD, AA	Bif. SW	N	NA/SD	N	ASD	7 y	Intractable		TPM, LEV, RUF, PB, B6, VPA, LCM, OXC CLZ, CLB		
39	T227I/de novo	3 m	WS	T, AP	S	HA→post spikes, slowing	N	SD/SD	Hypotonia	MC	2 y	Intractable		LEV, ZNS	B6, VGB, CS, ACTH, TPM, KD, PB, OXC	
40	A733T/de novo	3 m	Other	HC r/l	AA, DA, FD	MF spikes, slowing	N	MD/MD	N		12 y	Intractable		LEV	VPA, ESM, LTG, OXC	
41	R1882P/de novo	4 m	Other	T, C r		MF spikes, SW- status te→ESES- like	N	N/SD	N	Regression, ASD, 6 y aggression	6 y	Sz free (8 m)	LEV		OXC PB, VPA, ST, KD, LTG	
42	L1665F/de novo	4 m	Other	TS, SE	GTC (series)	N	N	N/SD	N	Stereotyped behaviour	6 y 3 m	Sz free (7 m)	CBZ	VPA	PB	
43	L1342P/de novo	6 m	WS	S		HA→MF spikes, slowing	At	SD/SD	Axial hypotonia, limb spasticity	Apnoeas, no eye contact	16 y	Intractable		VGB, CS		
44	L881P/de novo	6 m	WS→LGS	S	T, TC, AA	HA→MF spikes at cerebellar	At, HM,	SD/SD	Spasticity, hemiparesis	Small stature, 18 y early puberty	18 y	Intractable		TPM, MDZ, CLB VPA, LTG, PB, LEV, VGB, KD	RUF	
45	I1281F/de novo	7 m	WS→LGS	S	T, TC	HA→MF spikes, slowing, diffuse SW	At	NA/SD			16 y	Intractable			VNS, VPA, CLZ, RUF, TPM, LTG	
46	R853Q/de novo	8 m	WS→LGS	S	AA, T, F (4 y)	HA→for bil spikes, T2H, at SW, Slowing		N/SD	Dystonia	Preterm 28 w, 6 y 6 m MC, regression	6 y 6 m	Intractable	ACTH	VPA, CLB, TPM	LTG, FBM, LEV	CBZ
47	A1652P/de novo	9 m	WS	S		atypical HA→ slowing	HM	SD/SD	Hypotonia	ASD	3 y	Intractable		VGB, LEV, NZP	VPA, TPM, CS	PHT
48	R1319W/de novo	10 m	WS→LGS	T, S	TCS, A, AA	HA→spikes occ →spikes cue-par r	N	SD/SD	Hypotonia	Diabetes, ASD	10 y	Sz free (7 y)			CBZ, TPM, ESM, ZNS, VPA, FBM, LEV, B6, KD, CS, LTG	
49	E121IK / de novo	11 m	WS→other	S	FC, SE, M	HA→MF spikes, slowing	N	SD/SD	Hypotonia		4 y	Intractable		LEV, LTG, PB, ST, VGB, CS		
50	R853Q/de novo	13 m	WS	S	T, AU, M	HA→MF spikes, slowing ESES-like	N	MD/SD	Hypotonia, choreo- athetosis		8 y	Intractable		VGB, PB, CLB	ST, TPM, LEV, ZNS, ESM, LTG, PHT, OXC, CS, KDAZA	
51	H930Q/de novo	15 m	MAE	TCS	A, MAS, T, AA	slowing occ	N	N/MD	N	ASD	6 y	Sz free (18 m)	LEV + TPM		VPA, OXC, CLB	
52	P1622S/de novo	2 y	MAE	MA, DA	T, AA, M	Spike-slow-waves te-par bil	N	MD/MD	Ataxia	ASD, MC	3 y	Intractable		TPM	LEV, ESM, B6, PB, OXC, CLB, VGB, PF, KD	LTG
53	F612S/de novo	2 y 6 m	Other	TCS	M	fr spikes r/l, fr delta	N	MD/MD	N		3 y	Intractable		CLB, LEV, RUF		
54	c.605 + 1 G > T / de novo	2 y 6 m	Other	TCS	T, A	midline spikes ce, slowing→ bitemp spikes	N	N/SD	Hypotonia	ASD, regression	7 y	Intractable		LEV, CLB, KD	ZNS, LTG, FBM, TPM, RUF, B6, IVIG, VPA, PB, CS	
55	V1528Cfs*7/de novo	3 y	LGS	TCS	T, SE	Spike-slow-waves, Beta (sleep)	N	SD/SD	Spasticity	ASD, MC	9 y	Sz free (6 y)	VPA + LTG	CLZ, ESM		
56	R853Q/unknown	3 y	Other	TS	S	HA	CC	SD/SD	Pyramidal signs, hand dystonia		25 y	Intractable		VPA, ACTH		
57	C1170Vfs*15/de novo	3 y	Other	TCS	M, AB	gen SW, 1 fr SW, high voltage theta	HIS	MD/SD	Hypertonic limbs	ASD	17 y	Intractable		VPA, LTG, Br, CBZ, OXC, LEV, CLB	TPM, STP, ST, KD	
58	G1223R/de novo	3 y	Other	F, M	TCS, M, A, AA, NCSE	gen SW, PSW, slowing	N	N/MD	Ataxia	Regression	19 y	Intractable		VPA, CLZ, CLB, PHT, PB, CBZ, LTG, LEV	TPM, VNS, CS	
59	R1235 <sup>fs</sup> /de novo	3 y 4 m	Other	FC	GTC, DA	gen SW, PSW	HM	MD/MD	N	ASD	14 y	Intractable		LEV, ZNS		
60	A1773V/de novo	3 y 6 m	Other	F		Sharp waves r fr (sleep), slowing	N	MD/SD	N	ASD	4 y	Sz free (4 y)	LEV			
61	K1933M/de novo	4 y	Other	F, TCS	AA	MF spikes, bil N SW→ESES-like	bil N	SD/SD	Ataxia, parkinsonian gait	ASD, hyper-kinetic and aggressive behaviour	17 y	Intractable		ST, VPA, TPM, LTG, LCM CLB, CS	CBZ	

(continued)

Table 3 Continued

Patient	Mutation/ inheritance	Age at seizure onset	Epilepsy syndrome	Initial seizure type	Other seizure types	EEG	MRI	Cognition onset /follow-up	Neurological features	Additional features	Age at last follow-up	Seizure outcome (offset; age)	Treatment effects		Wors- ening
													Seizure- free	Sz reduction	
62	c.698-1G>T, splice site/ <i>de novo</i>	4 y 6 m	Other	F		f spikes→MF; like→N	ESES- N	N/MID	N		10 y	Sz free (9 y)	TPM	others	
63	W1716/ <i>de novo</i>	4 y 7 m	Other	Febrile T TCS (cluster)	AA	Gen SW	N	MD/MD	N	Psychosis (17 y)	25 y	Sz free (9 y)	VPA	PB	CBZ
64	N503K(s <sup>19</sup> / <i>de novo</i> )	6 y	Other			Bl sharp waves	N	SD/SD		ASD	20 y	Sz free (6 y)	VPA		
65	W281/ <i>de novo</i>	7y	Other	F	sec. Gen. TCS	MF spikes→ESES- like	N	MD/MD	Clumsiness		13 y	Sz free (10 y)	ST		
66	S1656F/ <i>de novo</i>	8 y 11 m	LGS	GTC	AA	Bifir SW, slowing	N	MD/MD	Hypotonia, crouched gait	Agitation	12 y	Intractable	LLTG, VPA	TPM	

AA = atypical absences; A = atonic; AB = absences; ADS = attention deficit disorder; ASD = autistic spectrum disorder; AU = autonomic seizures; AP = apneic seizures; At = atrophy; Bifir = bifrontal; Bil = bilateral; C = clonic; CC = corpus callosum; Ce = central; DA = drop attacks; ED = epileptiform discharges; F = focal; FD = focal dyscognitive; FC = febrile convulsion; fr = frontal; Gen = generalized tonic-clonic; HA = hypsarrhythmia; HC = hemiclonic; HIS = hippocampal sclerosis; HM = hypomyelination; IS = infantile spasms; L = left; LGS = Lennox-Gastaut syndrome; MAS = myoclonic-astatic seizures; MAE = myoclonic-astatic epilepsy; m = months; MID = mild intellectual disability; MD = moderate intellectual disability; M = myoclonic; MC = microcephaly; MF = multifocal; N = normal; NA = not applicable; NAV = not available; NCSE = non-convulsive status epilepticus; Occ = occipital; Par = parietal; R = right; S = spasms; SE = status epilepticus; SD = severe intellectual disability; SW = spike and waves; Sz = seizures; T = tonic; T2H = T<sub>2</sub>-hyperintensities; TCS = tonic-clonic seizures; Te = temporal; w = week; WS = West syndrome; y = years; → = change to.

reagent (sodium channel blockers are highlighted in bold): AZA = acetazolamide; B6 = vitamin B6; BR = bromide; CBZ = carbamazepine; CLB = clobazam; CS = corticosteroids; ESM = ethosuximide; FBM = felbamate; GBP = gabapentin; IVIG = intravenous immunoglobulins; KD = ketogenic diet; LCM = lacosamide; LTG = lamotrigine; LEV = levetiracetam; MDZ = midazolam; MSX = mesuximide; OXC = oxcarbazepine; PB = phenobarbital; PHT = phenytoin; P = pyridoxal phosphate; RGB = retigabine; RUF = rufinamide; ST = stiripentol; TPM = topiramate; VGB = vigabatrin; VNS = vagal nerve stimulation; VPA = valproate; ZNS = zonisamide.

epilepsies, mutations were missense in 45, early stop codons in six, frameshift in five (both predicting truncated proteins), and altered splice-sites in two. All non-missense mutations occurred in children with seizure onset beyond the first year of life. In children with intellectual disability or autism without epilepsy, mutations were missense ( $n = 8$ ), frameshift ( $n = 7$ ), nonsense ( $n = 8$ ), splice-site ( $n = 5$ ) or in-frame deletions ( $n = 2$ ). Twenty-eight recurrent mutations were observed (Table 5), and some of them were associated with specific phenotypes.

The A263V variant (*de novo*) has previously been described in patients with benign neonatal/infantile seizures and childhood onset episodic ataxia (Liao *et al.*, 2010a; Johannesen *et al.*, 2016; Schwarz *et al.*, 2016). However, a twin pair with the same mutation was identified to have Ohtahara syndrome, one of them being deceased, and the other having severe intellectual disability (Touma *et al.*, 2013). Furthermore, we detected a new case with this mutation (Patient 34, *de novo*), a male who suffered from severe encephalopathy with early infantile epilepsy. He died at the age of 13 years due to status epilepticus.

The R853Q variant was found in nine independent patients, six of whom were previously published and three unpublished (Patients 46, 50 and 56) (Allen *et al.*, 2013; Nakamura *et al.*, 2013; Samanta and Ramakrishnaiah, 2015; Li *et al.*, 2016). Six of these patients presented with West syndrome at 11–13 months of age, all were severely intellectually disabled, and most had intractable seizures.

The R1319Q variant was previously reported in three different families with BFIS (Berkovic *et al.*, 2004), while we found it twice (Patients 31 and 36) occurring *de novo* in patients with encephalopathy with early infantile epilepsy. In line with the previously reported cases, however, the two patients became seizure-free.

Ten mutations were found at amino acid position 1882, with variable substitutions. R1882G resulted in benign infantile seizures with late onset episodic ataxia in two cases, and was shown to cause a gain-of-function (Schwarz *et al.*, 2016). In contrast, R1882Q (Patient 19) (Carvill *et al.*, 2013; Howell *et al.*, 2015; Trump *et al.*, 2016), R1882L (Baasch *et al.*, 2014) and R1882P (Patient 41) resulted in severe phenotypes with intellectual disability.

## Functional studies

Electrophysiological analyses were performed and compared to the wild-type channel for four of the newly identified Na<sub>v</sub>1.2 missense mutations (for location see Fig. 3A) to correlate their functional effects to the time of onset of disease, the severity of the epilepsy, and the treatment response. We chose the four mutations on the following criteria: two were associated with different types of early infantile encephalopathy with epilepsy (one with a clear clinical response to SCBs, and one with a very severe, pharmacoresistant phenotype), the other two with different types of late onset epilepsies and worsening of the seizures



**Table 4** Clinical characteristics of the previously unpublished patients: intellectual disability and/or autism without epilepsy

Patient	Mutation/ inheritance	EEG	MRI	Cognition onset/ follow-up	Neurological features	Additional features	Age at follow-up
67	K1387Sfs*4/ <i>de novo</i>	NA	NA	MD	N	ASD	5 y 4 m
68	R1435*/ <i>de novo</i>	N	T2H	SD	N	ASD, early puberty	7 y 8 m
69	T1711Lfs*8/ <i>de novo</i>	Slowing	N	SD	Hypotonia	Rett-like, ASD	9 y 8 m
70	G1744E/ <i>de novo</i>	N	N	MD	N	ASD	4 y
71	c.386 + 2T > C/ <i>de novo</i>	NA	NA	MD	NAV	Episodic ataxia, ASD	13 y

ASD = autism spectrum disorder; m = months; MD = moderate intellectual disability; N = normal; NA = not applicable; NAV = not available; SD = severe intellectual disability; T2H = T<sub>2</sub>-hyperintensities; y = years.

upon application of SCBs. In detail, mutation F1597L was selected from a female with severe EIMFS (Patient 26) in whom the EEG suppression burst pattern resolved promptly upon phenytoin treatment. V423L recurred in two children with Ohtahara syndrome (Patients 10 and 33), both showing a peculiar severe phenotype with a high pharmacoresistance including lack of response to one or more SCBs (Table 1). G899S was selected from a child (Patient 38) with intractable infantile/childhood epilepsy with tonic-clonic seizures and absences, and mutation P1622S from a child with intractable myoclonic-atonic epilepsy (Patient 52). In both, aggravation of seizures occurred after introduction of oxcarbazepine and/or lamotrigine.

We found gain-of-function effects for the mutations V423L and F1597L, whereas the mutations G899S and P1622S showed loss-of-function effects (Fig. 3 and Supplementary Table 2). The analysis of the V423L mutation revealed a change in slope of activation (Fig. 3C and Supplementary Table 2) as well as a doubling of the window current and a dramatic increase in the persistent sodium current compared with the wild-type (Fig. 3B, C and F). For F1597L mutant channels, we observed a hyperpolarizing shift of the activation curve (Fig. 3C), fast inactivation time constants were significantly larger for mutant channels (Fig. 3D) and recovery from fast inactivation was accelerated (Fig. 3E and Supplementary Table 2).

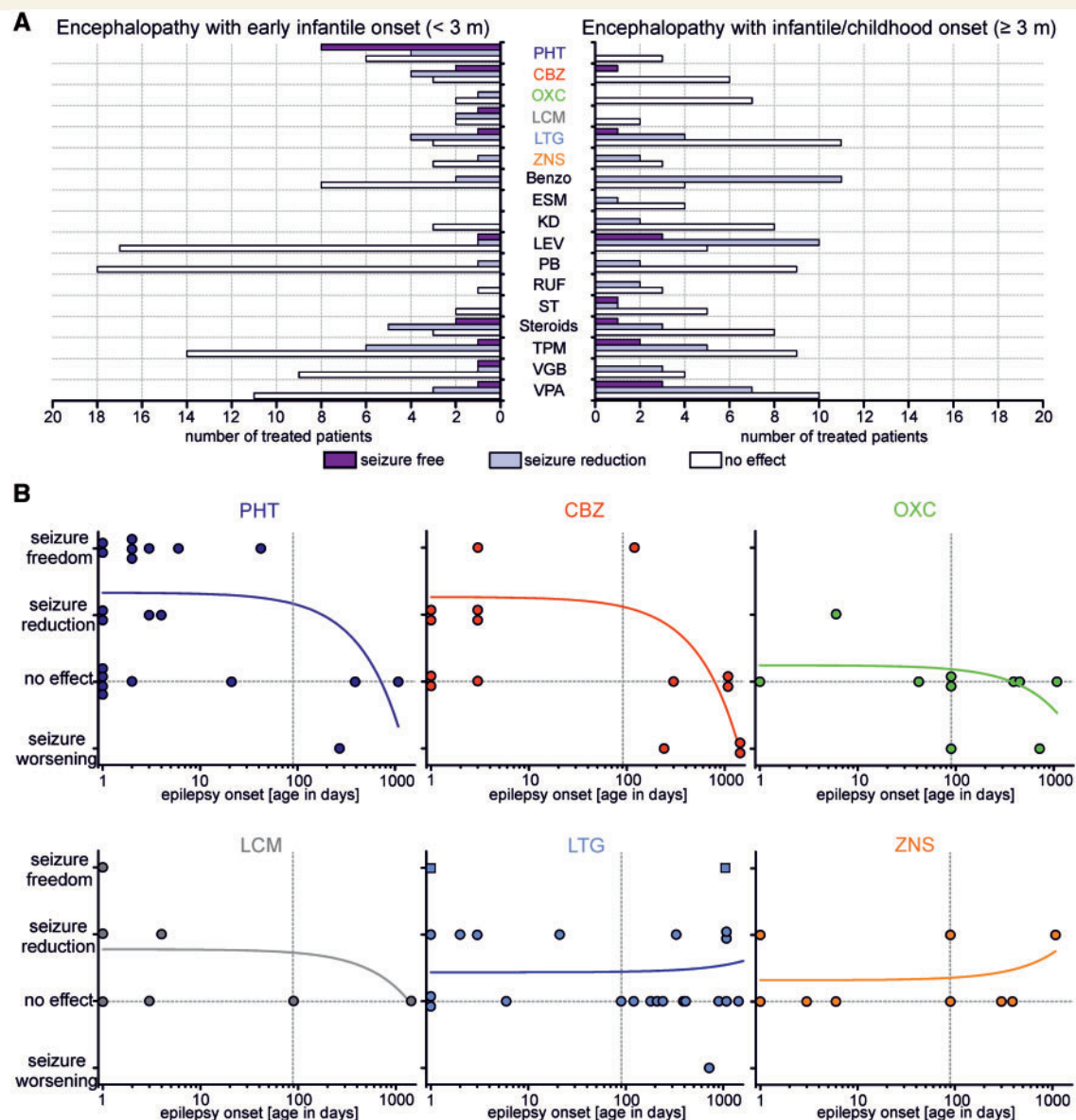
In contrast, electrophysiological analysis of the mutations G899S and P1622S revealed profound loss-of-function changes. The most prominent change for P1622S mutant channels was a significant hyperpolarizing shift of the fast inactivation curve (Fig. 3C). The effect of the G899S mutation on channel kinetics was not as pronounced as for P1622S, consisting of a depolarizing shift and a slope change of steady-state activation (Fig. 3C). Both mutations thus predict a decrease of channel availability and membrane excitability in neurons expressing mutant Na<sub>v</sub>1.2 channels, an effect that would be further enhanced by SCBs. Functional consequences of known SCN2A variants that have been reported in the literature are listed in Supplementary Table 1.

## Discussion

Our study of 71 new patients with SCN2A mutations and additional 130 previously reported cases—including a physiological characterization of some mutations and existing functional data from the literature—reveal evidence that there are two distinct groups among symptomatic SCN2A mutation carriers with epilepsy. The first group is represented by (i) early infantile epilepsy with onset before 3 months of age; (ii) missense mutations with gain-of-function effects of different severity with correlation to the severity of the clinical phenotype; and (iii) a relatively good response to SCBs. The second group is characterized by (i) a later epilepsy onset, i.e. after 3 months of age; (ii) frequent loss-of-function mutations (truncations and splice site mutations, but also missense mutations with loss-of-function effects); and (iii) a relatively poor response to SCBs. The phenotypic spectrum in the early onset group comprises B(F)NIS, Ohtahara syndrome, EIMFS and unclassified encephalopathies, whereas the late onset cases include West syndrome, Lennox-Gastaut syndrome, myoclonic-atonic epilepsy, and focal epilepsies with an ESES-like picture. A third, smaller group is represented by (i) intellectual disability and/or autism without epilepsy; and (ii) frequent loss-of-function mutations (truncations).

In the benign end of the spectrum, both familial and *de novo* mutations are found. These patients are characterized by a normal cognitive development and self-limited epilepsy with cessation of seizures mostly during the first year of life. Seizure semiology shows considerable variation, the typical ‘clustering’ is not always present, and family history may be negative due to the presence of *de novo* mutations. Seizures may be initially difficult to treat: in our cohort of unpublished patients, seizures were initially drug-resistant in 6/9 children (as discussed further below). However, EEG might help to rule out a severe epileptic phenotype, showing typically a normal background activity with or without multifocal spikes, but never a suppression burst pattern. After cessation of neonatal/infantile seizures, five children developed episodic ataxia and pain (Liao *et al.*, 2010a; Johannesen *et al.*, 2016; Schwarz *et al.*, 2016).

The group with encephalopathy and epilepsy is the largest among the SCN2A carriers; 69% of the cohort falls



**Figure 2 Treatment effects on epileptic seizures in patients with SCN2A-related encephalopathies.** (A) Number of treated patients and their seizure outcome according to the judgement of the treating physician (purple = seizure free; light purple = seizure reduction; open bar = no effect) that have been treated with different AEDs. Only effects of AEDs that have been used in more than three patients are shown. (B) Effects of treatments with specific SCBs on seizures as a function of the age of onset of the epilepsy in days. Each dot represents one treatment period with the respective SCB of one patient. Squares in the lamotrigine graph represent a combination of lamotrigine with additional AEDs. Patient 23 has been excluded for carbamazepine, as the seizure freedom correlated with phenytoin treatment. Lines represent linear regressions highlighting the dependence of treatment effects with SCBs on age of onset of the epilepsy. Only for phenytoin the slope was significantly different from 0 (phenytoin:  $P = 0.03$ ; carbamazepine:  $P = 0.06$ ; oxcarbazepine:  $P = 0.43$ ; lacosamide:  $P = 0.50$ ; lamotrigine:  $P = 0.66$ ; zonisamide:  $P = 0.52$ ). benzo = benzodiazepines; CBZ = carbamazepine; ESM = ethosuximide; KD = ketogenic diet; LCM = lacosamide; LTG = lamotrigine; LEV = levetiracetam; OXC = oxcarbazepine; PB = phenobarbital; PHT = phenytoin; RUF = rufinamide; ST = sulthiame; TPM = topiramate; VGB = vigabatrin; VPA = valproate; ZNS = zonisamide.

into this category (Fig. 1). Half of those patients had a seizure onset in the early infantile period (<3 months). In the late onset group, seizure onset was usually before the age of 4 years (only five patients had a later seizure debut).

Fifty-four of the 139 children had an identifiable epileptic syndrome (32%). In the early infantile group, Ohtahara

syndrome ( $n = 18$ ) and EIMFS ( $n = 13$ ) constitute the most important specific phenotypes. Whereas Ohtahara syndrome was the first syndrome to be reported in SCN2A-related encephalopathies, EIMFS was only recently recognized (Howell *et al.*, 2015). In EIMFS, *KCNT1* mutations have been reported as the most common underlying

**Table 5 Recurrent mutations**

Mutation	Frequency % (n cases)	Phenotypes
M136I	1 (2)	EE, EIMFS
V213D	1 (2)	EOEE
V261M	1 (2)	BNS
A263V/T	3.5 (7)	BNS, EE, OS
V423L	1 (2)	OS
V430Q/G/A	1.5 (3)	BFIS, OS
N503K fs	1 (2)	ID
R853Q	4.5 (9)	WS, EE, LGS
R856L/Q	1 (2)	OS, EIMFS
G882R/E	1 (2)	EIMFS
K905N	1 (2)	EE
F928C	1 (2)	EIMFS, EE
R937C	1 (2)	ID
C595*	1 (2)	ASD
E999V/K	3 (6)	EIEE, OS, EOEE
G1013*	1 (2)	ASD
I1021Y*	1 (2)	LGS, EE
E1211K	1 (2)	WS
V1282F	1 (2)	Schizophrenia
R1319Q/W	3 (6)	BFIS, WS, EE
V1326V/D	1 (2)	EIMFS, OS
S1336Y	1.5 (3)	OS
L1342P	2.5 (5)	EOEE, WS
R1435*	1 (2)	ASD
Q1531K	1 (2)	BFNS
T1623N	1 (2)	OS, EE
R1629L/H	1 (2)	EE
R1882G/Q/L	5 (10)	BIS, OS, EE

ASD = autism spectrum disorder (without seizures); BFIS = benign familial infantile seizures; BIS = benign infantile seizures; BNS = benign neonatal seizures; EE = epileptic encephalopathy; EIEE = early infantile epileptic encephalopathy; EIMFS = epilepsy of infancy with migrating focal seizures; EOEE = early onset epileptic encephalopathy; ID = intellectual disability (without seizures); LGS = Lennox-Gastaut syndrome; OS = Ohtahara syndrome; WS = West syndrome.

genetic cause so far (Barcia *et al.*, 2012; Ohba *et al.*, 2015). In *KCNT1*-related EIMFS, the prognosis seems to be uniformly poor, and affected children show severe disability with ongoing seizures during follow-up (Barcia *et al.*, 2012; Ohba *et al.*, 2015). In our *SCN2A* cohort, 2/5 patients with EIMFS became seizure-free at the age of 2 and 12 months with vigabatrin and phenytoin, respectively, and showed mild intellectual disability. Thus, prognosis seems to be more favourable in *SCN2A*-related EIMFS compared to those caused by *KCNT1* mutations.

Epilepsies with onset beyond the early infantile period are increasingly recognized in *SCN2A*-related disorders. In particular, West syndrome has recently emerged as the most important specific phenotype, accounting for 16 patients so far. Of note, one recurring mutation (R853Q), which has been found in nine cases so far, is frequently associated with West syndrome (six cases), and should therefore be considered in the diagnostic work-up of children with West syndrome.

The intellectual disability/autism group without epilepsy is most likely an underestimate of the actual number of

cases, since it is not common practice in all countries/hospitals to perform genetic testing in patients with intellectual disability and/or autism without seizures. This group of patients represents 16% of the cohort.

The relative frequency of the disease groups as represented in Fig. 1A for the newly identified cases with epilepsy and in Fig. 1B for all cases probably contains biases, as (i) benign neonatal-infantile epilepsies were the first entity in which *SCN2A* mutations were detected suggesting a relative over-representation of those and may be also the severe early onset cases in the literature; and (ii) since cases without epilepsy are probably underdiagnosed.

## Genotype

Recurrent mutations are seen both in the benign and severe end of the spectrum (Table 5). However, even with the same mutation, a quite large phenotypic variance is observed. A remarkable phenotype was found for R853Q mutation carriers in whom 6/9 were affected by West syndrome. Furthermore, three of the patients with an A263V mutation showed a BNIS phenotype with late onset episodic ataxia, while three others with the same mutation had more severe phenotypes. Thus, both the mutation itself and other genetic or environmental factors contribute to the individual phenotype.

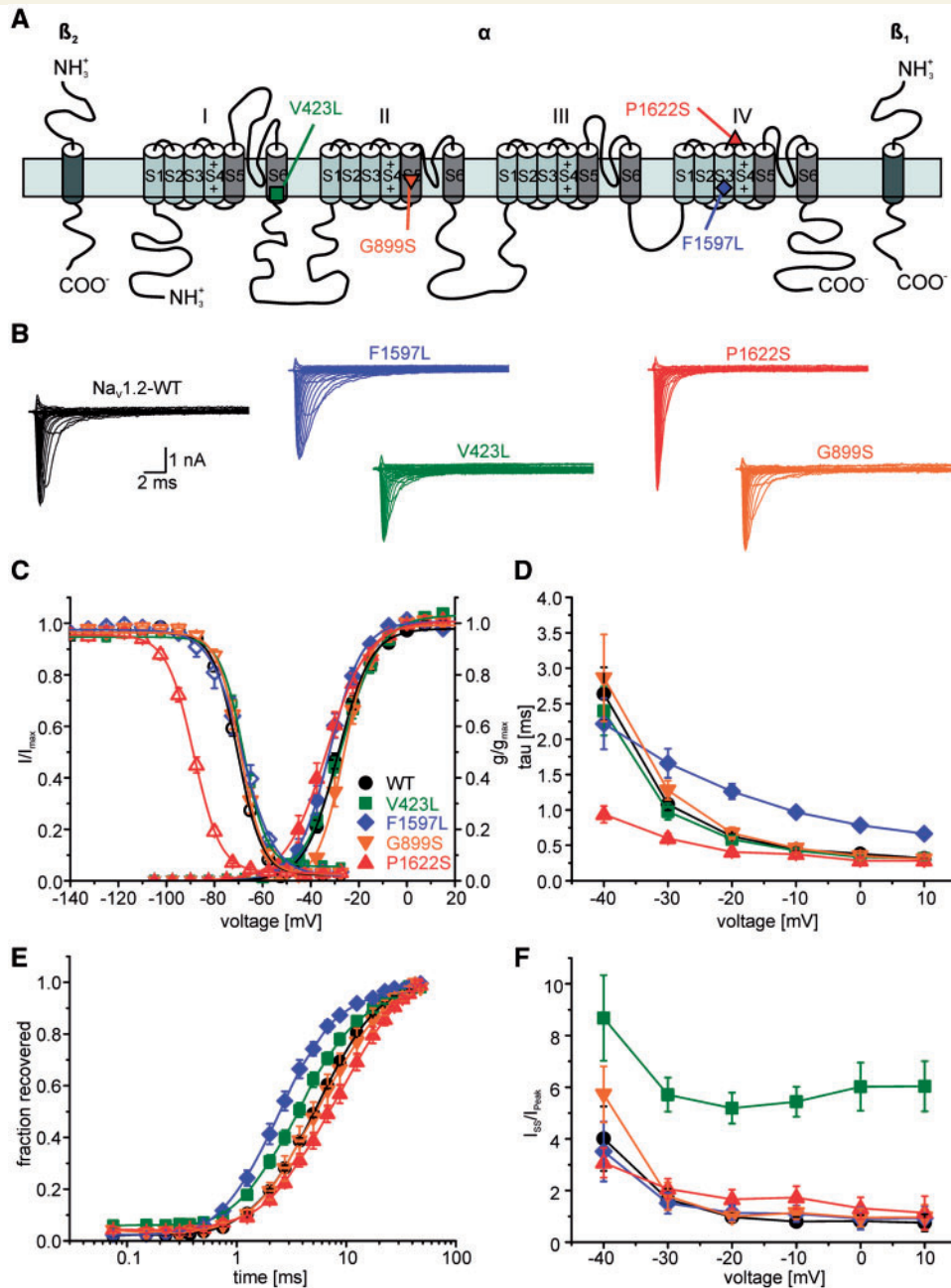
Interestingly, we found only missense mutations in the early onset epilepsies, whereas truncations, splice-site and nonsense mutations were solely seen in the epilepsies with a later onset and in the group of cases without epilepsy. This observation, along with the functional effects described in the results section and discussed below, could have implications for treatment in these two major groups of *SCN2A* patients. Supplementary Table 1 provides an overview of all known *SCN2A* mutations, the associated clinical phenotypes and known functional consequences.

## Prevalence

We estimated the frequency of *SCN2A* mutations causing the reported phenotypes in the Danish population to be 1/78 608. This number will most likely be an underestimate. There is not a strong tradition for a systematic genetic screening of patients with isolated autism or intellectual disability, thus there might be a recruitment bias towards patients with epilepsy.

## Seizure outcome and treatment effects

In B(F)NIS families, seizures are reported to be controlled by AEDs (Berkovic *et al.*, 2004; Striano *et al.*, 2006; Herlenius *et al.*, 2007). In the unpublished cohort, however, many patients were resistant to various AEDs, including phenobarbital, topiramate, levetiracetam and valproate, whereas SCBs (especially oxcarbazepine and phenytoin), were completely or partially effective in 6/9 cases, suggesting a specific effect of SCBs in this subgroup. However, and



**Figure 3 Functional studies reveal pronounced gain-of-function changes for the V423L and F1597L mutations and loss-of-function changes for the P1622S and G899S mutations.** (A) Schematic of the human Na<sub>v</sub>1.2  $\alpha$ -subunit together with  $\beta_1$  and  $\beta_2$  subunits showing the locations of the four functionally studied mutations (V423L green square; F1597L blue diamond; G899S orange inverted triangle; P1622S red triangle). (B) Representative current traces of whole-cell Na<sup>+</sup> currents recorded from tsA201 cells transfected with either Na<sub>v</sub>1.2 wild-type or mutant channels. (C) Voltage dependence of steady state Na<sup>+</sup> channel activation and inactivation revealing a significant depolarizing shift of the activation curve for G899S (loss-of-function) as well as a significant hyperpolarizing shift of the inactivation curve for P1622S (loss-of-function) in comparison with the wild-type. Lines represent fits of Boltzmann functions. (D) Voltage dependence of the fast inactivation time constant for wild-type and mutant channels revealing a slowing of fast inactivation for F1597L and an acceleration for P1622S. (E) The time course of recovery from fast inactivation determined at -100 mV showed significant changes between wild-type and mutant channels. F1597L mutant channels showed a significantly faster recovery (gain-of-function), whereas P1622S mutant channels showed a significant slowing of the recovery from fast inactivation compared to wild-type channels (loss-of-function). Lines represent fits of exponential functions yielding the time constant  $\tau_{rec}$ . (F) Voltage dependence of the persistent sodium current showing a large increase for V423L compared to the wild-type. Current amplitudes recorded at the end of a 70-ms depolarization were normalized to the peak current amplitude (steady state current/initial peak current). Shown are means  $\pm$  SEM.



as known from previous studies, three patients became seizure-free without any AEDs, which is part of the natural history of these patients. Thus, the effect of SCBs in these patients should be interpreted with some caution, although we observed a clear correlation between introduction of the drugs and seizure freedom.

Apart from B(F)NIS, we found a different pattern of seizure outcome and AED effects in our cohort according to the age at seizure onset. Of all patients with encephalopathy and epilepsy onset <3 months, 17/28 (61%) became seizure-free, 10 of them during SCB treatment, mainly with phenytoin ( $n = 8$ ), whereas other standard AEDs (e.g. phenobarbital, levetiracetam) were largely ineffective. Interestingly, in five children recurrent declines of phenytoin plasma levels resulted in prompt seizure relapses that were reversible after adjusting the phenytoin dosage. These observations underline the beneficial effect of phenytoin on seizure activity in these patients.

To date, there are only few reports on treatment response in SCN2A-related epilepsies. In the series of Nakamura *et al.* (2013) on 15 children with early onset seizures, epilepsies were described as intractable in 12 of 15 cases. Phenytoin was tried in five children with seizure onset between 1 day and 6 weeks of age, and showed partial effects in four of them. Zonisamide showed some effects in 4/6 children. Two children became seizure-free with lamotrigine at the age of 6 months and 6 years, respectively. Howell *et al.* (2015) reported improvement of seizure control in 11/15 patients with early seizure onset, phenytoin was reported to be partially effective in seven children with neonatal seizure onset. Sawaishi *et al.* (2002) described a striking effect of lidocaine, a prototypic sodium channel blocker, in a patient with Ohtahara syndrome due to a SCN2A mutation (Sawaishi *et al.*, 2002; Ogiwara *et al.*, 2009).

Taken together, patients with early onset epilepsies were difficult to treat but responded relatively well to SCBs, in particular to phenytoin in appropriate dosages. The high dosages needed to control seizures completely might not have been reached in many patients who did not become seizure-free.

In contrast, only 10/29 (34%) children with infantile/childhood epilepsy became seizure-free, and seizures mostly did not respond to SCBs. In the nine children with West syndrome, seizures and hypsarrhythmia in the EEG were mostly resistant to standard treatment including steroids and ketogenic diet. Nakamura *et al.* (2013) reported on eight children with West syndrome in their series. All were intractable, suggesting that SCN2A-related West syndrome is particularly difficult to treat. However, some case reports describe treatment responses to ACTH (Nakamura *et al.*, 2013), topiramate (Ogiwara *et al.*, 2009; Sundaram *et al.*, 2013; Samanta and Ramakrishnaiah, 2015) and transient effects of prednisolone (Matalon *et al.*, 2014).

Seizure worsening associated with SCB treatment was observed in seven children of our cohort, all of whom had seizure onset beyond the age of 3 months.

Hackenberg *et al.* (2014) reported an increase of seizure frequency related to carbamazepine treatment in a child with seizure onset at 3 months of age. Howell *et al.* (2015) reported the appearance of myoclonus with vigabatrin and lamotrigine in one child. SCBs are known to aggravate seizure activity in epileptic syndromes that are caused by loss-of-function mutations in Na<sub>v</sub>1.1 channels, e.g. Dravet syndrome, putatively because they further reduce sodium channel activity in inhibitory neurons (Brunklaus *et al.*, 2014) expressing Na<sub>v</sub>1.1 as the major sodium channel (Catterall, 2014). As discussed below, a similar mechanism may apply for loss-of-function mutations in Na<sub>v</sub>1.2 channels.

The statistical  $\chi^2$  test confirmed our impression of differential treatment effects of SCBs and non-SCBs in early versus late onset encephalopathies with epilepsy. The late onset cases responded significantly better to non-SCBs than the early onset ones, which may indicate that the epilepsy in the early onset cases is more difficult to treat. This observation even strengthens the finding of a significantly better response to SCBs of the early onset compared to the late onset cases, and suggests that the higher seizure freedom rate of the early onset group (61%, compared to only 34% of the late onset group) is the likely consequence of a specific pharmaco-response to gain-of-function SCN2A mutations (see discussion below on functional effects).

In summary, SCBs seem to have positive effects on seizures in early infantile onset epilepsies, but are not effective or can even worsen seizure activity in epilepsies with onset at 3 months of age or later. However, these conclusions have to be taken with care, as the natural history of these conditions is unknown, our observations are purely retrospective and the exact duration from AED introduction to seizure freedom was not clear in all cases.

## Functional studies and their pharmacological and neurophysiological interpretation

Out of the diversity of SCN2A mutations that have been identified until now, only a small number have been studied functionally. SCN2A mutations can lead to either augmented or reduced Na<sub>v</sub>1.2 activity (Kamiya *et al.*, 2004; Scalmani *et al.*, 2006; Xu *et al.*, 2007; Ogiwara *et al.*, 2009; Liao *et al.*, 2010a, b; Lossin *et al.*, 2012; Rauch *et al.*, 2012; Lauxmann *et al.*, 2013; Sundaram *et al.*, 2013; Codina-Sola *et al.*, 2015; Schwarz *et al.*, 2016). Here, we functionally analysed four additional mutations, two of which were identified in patients suffering from encephalopathy with early infantile onset epilepsy. For these mutations (V423L and F1597L), a clear gain-of-function effect was found (Fig. 3). V423L particularly showed a tremendous increase in persistent sodium current, probably explaining the extremely severe phenotype with highly drug-resistant Ohtahara syndrome in both children affected by this mutation. We hypothesize that this persistent



current was too large to be sufficiently reduced by SCBs in clinically relevant dosages. F1597L caused an accelerated recovery from fast inactivation, and the patient showed an EIMFS phenotype with a prompt response to phenytoin treatment. In contrast, we have shown clear loss-of-function effects for two mutations found in children with infantile/childhood epilepsy (G899S and P1622S). The phenotype consisted of tonic-clonic seizures and absences in one and myoclonic-atonic epilepsy in the other child, which was clearly different from the other two mutations. This difference was equally seen with regard to the treatment response, which showed seizure aggravation upon SCB treatment in both cases.

In previous studies, several disease-causing mutations in *SCN2A* have been functionally analysed. In children with severe early onset epilepsies, gain-of-function mutations were described (Ogiwara *et al.*, 2009; this study, see above). Missense mutations from patients with B(F)NIS have been characterized to cause a gain-of-function (Scalmani *et al.*, 2006; Xu *et al.*, 2007; Liao *et al.*, 2010a, b; Lauxmann *et al.*, 2013; Schwarz *et al.*, 2016) with one of these mutations (A263V) also causing severe epilepsy in some cases (see above). Few studies also indicated some biophysical features indicating a loss-of-function (Scalmani *et al.*, 2006; Misra *et al.*, 2008); however, studies in neurons by Scalmani *et al.* (2006) suggested a net gain-of-function with increased excitability for two such cases so that the main effect seems to be a gain-of-function (R223Q and R1319Q), matching the effective SCB treatment of patients carrying the R1319Q mutation. These distinct effects of the same mutation analysed in neuronal versus non-neuronal cells could be caused by post-translational modifications, phosphorylation, trafficking and protein–protein interactions (such as with  $\beta$ -subunits, which we also used in our study) of the channels which can be quite different in neuronal cells and heterologous expression systems (for review see Shao *et al.*, 2009). It has been shown for mutations in  $\text{Na}_v1.1$  that—besides the  $\beta$ -subunits as the closest interacting partners of  $\alpha$ -subunits—ankyrin, calmodulin or other endogenous proteins can also have essential roles for intracellular trafficking and functional expression (Rusconi *et al.*, 2007; Cestele *et al.*, 2013). Such effects may also apply for the mutations we studied here in tsA201 cells. Furthermore, modifier genes might affect the clinical severity and the variability of phenotypes seen in epilepsy patients with gain-of-function *SCN2A* mutations (Bergren *et al.*, 2005; Hawkins and Kearney, 2016).

In contrast to the observed gain-of-function effects, missense loss-of-function and nonsense mutations have been identified in patients with later onset epilepsies, although two mutations also showed a hyperpolarizing shift of the activation curve as a gain-of-function feature (Kamiya *et al.*, 2004; Ogiwara *et al.*, 2009; Lossin *et al.*, 2012). Finally, autism spectrum disorder without seizures was also associated with loss-of-function mutations (Kamiya *et al.*, 2004; Rauch *et al.*, 2012; Sanders *et al.*, 2012;

Carvill *et al.*, 2013; Codina-Sola *et al.*, 2015; D’Gama *et al.*, 2015; Howell *et al.*, 2015; Carroll *et al.*, 2016; Horvath *et al.*, 2016; Li *et al.*, 2016; Trump *et al.*, 2016) (Supplementary Table 1).

These findings strengthen the hypothesis that there is a link between gain-of-function *SCN2A* mutations, early onset epilepsy, and effectiveness of SCBs on the one hand, and loss-of-function mutations, later onset epilepsy, and ineffectiveness of SCBs on the other hand. Gain-of-function versus loss-of-function mutations affecting the neuronal excitability of different neurons during specific developmental stages might explain the variation in seizure onset and the response to SCBs in early infantile syndromes. Early in development, the  $\text{Na}_v1.2$  channel is highly expressed at nodes of Ranvier and axon initial segments and is partially replaced during development by the  $\text{Na}_v1.6$  channel (Kaplan *et al.*, 2001; Liao *et al.*, 2010b), which could be confirmed in adult human hippocampal brain slices (Liao *et al.*, 2010b).  $\text{Na}_v1.2$  channels are therefore considered to contribute to action potential generation and propagation and influence the axonal firing frequency during early development. Hence, mutations causing gain-of-function effects can alter the characteristic firing patterns of  $\text{Na}_v1.2$ -expressing neurons and cause hyperexcitability, which can be dampened by SCB treatment and thus improve seizure outcome. In contrast, loss-of-function mutations cannot drive  $\text{Na}_v1.2$ -expressing cells into hyperexcitability during early development, and therefore patients may not exhibit early onset seizures. However, loss-of-function mutations seem to lead to severe epileptic phenotypes later in development. In the more mature brain, unmyelinated axons express the  $\text{Na}_v1.2$  channel, such as mossy fibres projecting from the hippocampal dentate gyrus into the CA3 region (Kaplan *et al.*, 2001; Liao *et al.*, 2010b). The most prevalent targets of mossy fibres apart from CA3 pyramidal cells are GABAergic parvalbumin-positive interneurons. Mossy fibres contact inhibitory basket cells ~50 times more frequently than pyramidal cells mediating a powerful feedforward inhibition (Acsady *et al.*, 1998). Therefore, a reduced excitability of dentate granule cells due to  $\text{Na}_v1.2$  loss-of-function mutations can result in CA3 hyperexcitability, which can spread to subsequent hippocampal regions and further. A treatment with SCBs can therefore act in a similar way as hypothesized for loss-of-function  $\text{Na}_v1.1$  mutations. Here, SCBs are predicted to further reduce the activity of inhibitory neurons expressing  $\text{Na}_v1.1$  as major sodium channel. Similarly, SCBs could further reduce the activity of  $\text{Na}_v1.2$ -expressing dentate granule cells, which in turn would activate inhibitory neurons less effectively. As mossy fibres are not the only unmyelinated fibres within the brain, other unmyelinated inhibitory neurons expressing  $\text{Na}_v1.2$  channels could also contribute to neuronal hyperexcitability. Only parvalbumin-positive inhibitory neurons within the cortical layers have been shown to be myelinated (Micheva *et al.*, 2016). Unmyelinated inhibitory neurons expressing  $\text{Na}_v1.2$  channels are controlling the activity of excitatory

neurons and provide feedforward inhibition (Kepecs and Fishell, 2014). In early development, a reduced activity of these inhibitory neurons due to Na<sub>v</sub>1.2 loss-of-function mutations could be of little or no consequence, as excitatory Na<sub>v</sub>1.2-expressing neurons within the cortex also show reduced excitability. As mentioned above, the Na<sub>v</sub>1.2 channel is partially replaced by the Na<sub>v</sub>1.6 channel at nodes of Ranvier and axon initial segments during development, but is still expressed in unmyelinated fibres, such as those from inhibitory neurons. Additionally, the neonatal splice variant of Na<sub>v</sub>1.2 has been shown to have a seizure protective role during early development (Gazina *et al.*, 2015). A reduced excitability of unmyelinated cortical inhibitory neurons later in development could therefore lead to hyperexcitable cortical networks. A disruption of the excitatory/inhibitory balance caused by SCN2A loss-of-function mutations can therefore cause seizures and additionally underlie neuropsychiatric diseases and autism, reflected by a higher prevalence in patients with loss-of-function mutations and late onset phenotypes in our cohort.

In summary, our study reflects the large spectrum of SCN2A-related disorders and identifies SCN2A mutations as one of the most common genetic causes of epilepsy. We have established two distinct groups with seizure onset either before or after 3 months of age, which show phenotypic differences, gain-of-function versus loss-of-function Na<sub>v</sub>1.2 abnormalities and a likely related differential response to treatment with SCBs.

## Acknowledgements

We wish to thank the patients and their families for the participation in this study and Stephan Müller for assistance in performing mutagenesis.

## Funding

K.L.H. is employed by and receives a salary from Ambry Genetics. I.H. was supported by intramural funds of the University of Kiel, by a grant from the German Research Foundation (HE5415/3-1) within the EuroEPINOMICS framework of the European Science Foundation, and additional grants of the German Research Foundation (DFG, HE5415/5-1, HE 5415/6-1), German Ministry for Education and Research (01DH12033, MAR 10/012), and grant by the German chapter of the International League Against Epilepsy (DGfE). H.L. was supported by the EU FP7 project EpiPGX (279062), the EuroEPINOMICS framework of the European Science Foundation (DFG grant Le1030/11-2), by the German Ministry for Education and Research (BMBF) rare disease network IonNeurONet (01GM1105A), as well as the German chapter of the International League Against Epilepsy (DGfE) and the foundation “no epilep”. P.d.J. was supported by ESF/Euro-Epinomics (GA136.11.N and

FWO/ESF-ECRP). H.S. is a PhD fellow of the Fund for Scientific Research Flanders (1125416N).

## References

- Acsady L, Kamondi A, Sik A, Freund T, Buzsaki G. GABAergic cells are the major postsynaptic targets of mossy fibers in the rat hippocampus. *J Neurosci* 1998; 18: 3386–403.
- Allen AS, Berkovic SF, Cossette P, Delanty N, Dlugos D, Eichler EE, et al. *De novo* mutations in epileptic encephalopathies. *Nature* 2013; 501: 217–21.
- Allen NM, Conroy J, Shahwan A, Lynch B, Correa RG, Pena SD, et al. Unexplained early onset epileptic encephalopathy: exome screening and phenotype expansion. *Epilepsia* 2016; 57: e12–7.
- Baasch AL, Huning I, Gilissen C, Klepper J, Veltman JA, Gillessen-Kaesbach G, et al. Exome sequencing identifies a *de novo* SCN2A mutation in a patient with intractable seizures, severe intellectual disability, optic atrophy, muscular hypotonia, and brain abnormalities. *Epilepsia* 2014; 55: e25–9.
- Barcia G, Fleming MR, Deligniere A, Gazula VR, Brown MR, Langouet M, et al. *De novo* gain-of-function KCNT1 channel mutations cause malignant migrating partial seizures of infancy. *Nat Genet* 2012; 44: 1255–9.
- Berg AT, Berkovic SF, Brodie MJ, Buchhalter J, Cross JH, van Emde Boas W, et al. Revised terminology and concepts for organization of seizures and epilepsies: report of the ILAE commission on classification and terminology, 2005–2009. *Epilepsia* 2010; 51: 676–85.
- Berggren SK, Chen S, Galecki A, Kearney JA. Genetic modifiers affecting severity of epilepsy caused by mutation of sodium channel Scn2a. *Mamm Genome* 2005; 16: 683–90.
- Berkovic SF, Heron SE, Giordano L, Marini C, Guerrini R, Kaplan RE, et al. Benign familial neonatal-infantile seizures: characterization of a new sodium channelopathy. *Ann Neurol* 2004; 55: 550–7.
- Boiko T, Rasband MN, Levinson SR, Caldwell JH, Mandel G, Trimmer JS, et al. Compact myelin dictates the differential targeting of two sodium channel isoforms in the same axon. *Neuron* 2001; 30: 91–104.
- Boiko T, Van Wart A, Caldwell JH, Levinson SR, Trimmer JS, Matthews G. Functional specialization of the axon initial segment by isoform-specific sodium channel targeting. *J Neurosci* 2003; 23: 2306–13.
- Brunklaus A, Ellis R, Reavey E, Semsarian C, Zuberi SM. Genotype phenotype associations across the voltage-gated sodium channel family. *J Med Genet* 2014; 51: 650–8.
- Carroll LS, Woolf R, Ibrahim Y, Williams HJ, Dwyer S, Walters J, et al. Mutation screening of SCN2A in schizophrenia and identification of a novel loss-of-function mutation. *Psychiatr Genet* 2016; 26: 60–5.
- Carvill GL, Heavin SB, Yendle SC, McMahon JM, O’Roak BJ, Cook J, et al. Targeted resequencing in epileptic encephalopathies identifies *de novo* mutations in CHD2 and SYNGAP1. *Nat Genet* 2013; 45: 825–30.
- Catterall WA. Sodium channels, inherited epilepsy, and antiepileptic drugs. *Annu Rev Pharmacol Toxicol* 2014; 54: 317–38.
- Cestele S, Schiavon E, Rusconi R, Franceschetti S, Mantegazza M. Nonfunctional Nav1.1 familial hemiplegic migraine mutant transformed into gain of function by partial rescue of folding defects. *Proc Natl Acad Sci USA* 2013; 110: 17546–51.
- Codina-Sola M, Rodriguez-Santiago B, Homs A, Santoyo J, Rigau M, Aznar-Lain G, et al. Integrated analysis of whole-exome sequencing

- and transcriptome profiling in males with autism spectrum disorders. *Mol Autism* 2015; 6: 21.
- D'Gama AM, Pochareddy S, Li M, Jamuar SS, Reiff RE, Lam AT, et al. Targeted DNA sequencing from autism spectrum disorder brains implicates multiple genetic mechanisms. *Neuron* 2015; 88: 910–7.
- Dhamija R, Wirrell E, Falcao G, Kirmani S, Wong-Kisiel LC. Novel *de novo* SCN2A mutation in a child with migrating focal seizures of infancy. *Pediatr Neurol* 2013; 49: 486–8.
- Dimassi S, Labalme A, Ville D, Calender A, Mignot C, Boutry-Kryza N, et al. Whole-exome sequencing improves the diagnosis yield in sporadic infantile spasm syndrome. *Clin Genet* 2016; 89: 198–204.
- Fukasawa T, Kubota T, Negoro T, Saitoh M, Mizuguchi M, Ihara Y, et al. A case of recurrent encephalopathy with SCN2A missense mutation. *Brain Dev* 2015; 37: 631–4.
- Gazina EV, Leaw BT, Richards KL, Wimmer VC, Kim TH, Aumann TD, et al. 'Neonatal' Nav1.2 reduces neuronal excitability and affects seizure susceptibility and behaviour. *Hum Mol Genet* 2015; 24: 1457–68.
- Grinton BE, Heron SE, Pelekanos JT, Zuberi SM, Kivity S, Afawi Z, et al. Familial neonatal seizures in 36 families: clinical and genetic features correlate with outcome. *Epilepsia* 2015; 56: 1071–80.
- Hackenberg A, Baumer A, Sticht H, Schmitt B, Kroell-Seger J, Wille D, et al. Infantile epileptic encephalopathy, transient choreoathetotic movements, and hypersomnia due to a *De Novo* missense mutation in the SCN2A gene. *Neuropediatrics* 2014; 45: 261–4.
- Haug K, Hallmann K, Rebstock J, Dullinger J, Muth S, Haverkamp F, et al. The voltage-gated sodium channel gene SCN2A and idiopathic generalized epilepsy. *Epilepsy Res* 2001; 47: 243–6.
- Hawkins NA, Kearney JA. Hlf is a genetic modifier of epilepsy caused by voltage-gated sodium channel mutations. *Epilepsy Res* 2016; 119: 20–3.
- Herlenius E, Heron SE, Grinton BE, Keay D, Scheffer IE, Mulley JC, et al. SCN2A mutations and benign familial neonatal-infantile seizures: the phenotypic spectrum. *Epilepsia* 2007; 48: 1138–42.
- Heron SE, Crossland KM, Andermann E, Phillips HA, Hall AJ, Bleasel A, et al. Sodium-channel defects in benign familial neonatal-infantile seizures. *Lancet* 2002; 360: 851–2.
- Heron SE, Scheffer IE, Grinton BE, Eyre H, Oliver KL, Bain S, et al. Familial neonatal seizures with intellectual disability caused by a microduplication of chromosome 2q24.3. *Epilepsia* 2010; 51: 1865–9.
- Horvath GA, Demos M, Shyr C, Matthews A, Zhang L, Race S, et al. Secondary neurotransmitter deficiencies in epilepsy caused by voltage-gated sodium channelopathies: a potential treatment target?. *Mol Genet Metab* 2016; 117: 42–8.
- Howell KB, McMahon JM, Carvill GL, Tambunan D, Mackay MT, Rodriguez-Casero V, et al. SCN2A encephalopathy: a major cause of epilepsy of infancy with migrating focal seizures. *Neurology* 2015; 85: 958–66.
- Jiang YH, Yuen RK, Jin X, Wang M, Chen N, Wu X, et al. Detection of clinically relevant genetic variants in autism spectrum disorder by whole-genome sequencing. *Am J Hum Genet* 2013; 93: 249–63.
- Johannessen KM, Miranda MJ, Lerche H, Moller RS. Letter to the editor: confirming neonatal seizure and late onset ataxia in SCN2A Ala263Val. *J Neurol* 2016; 263: 1459–60.
- Kamiya K, Kaneda M, Sugawara T, Mazaki E, Okamura N, Montal M, et al. A nonsense mutation of the sodium channel gene SCN2A in a patient with intractable epilepsy and mental decline. *J Neurosci* 2004; 24: 2690–8.
- Kaplan MR, Cho MH, Ullian EM, Isom LL, Levinson SR, Barres BA. Differential control of clustering of the sodium channels Na(v)1.2 and Na(v)1.6 at developing CNS nodes of Ranvier. *Neuron* 2001; 30: 105–19.
- Kepecs A, Fishell G. Interneuron cell types are fit to function. *Nature* 2014; 505: 318–26.
- Kobayashi K, Ohzono H, Shinohara M, Saitoh M, Ohmori I, Ohtsuka Y, et al. Acute encephalopathy with a novel point mutation in the SCN2A gene. *Epilepsy Res* 2012; 102: 109–12.
- Lauxmann S, Boutry-Kryza N, Rivier C, Mueller S, Hedrich UB, Maljevic S, et al. An SCN2A mutation in a family with infantile seizures from Madagascar reveals an increased subthreshold Na(+) current. *Epilepsia* 2013; 54: e117–21.
- Lemke JR, Riesch E, Scheurenbrand T, Schubach M, Wilhelm C, Steiner I, et al. Targeted next generation sequencing as a diagnostic tool in epileptic disorders. *Epilepsia* 2012; 53: 1387–98.
- Li J, Cai T, Jiang Y, Chen H, He X, Chen C, et al. Genes with *de novo* mutations are shared by four neuropsychiatric disorders discovered from NPdenovo database. *Mol Psychiatry* 2016; 21: 290–7.
- Liao Y, Anttonen AK, Liukkonen E, Gaily E, Maljevic S, Schubert S, et al. SCN2A mutation associated with neonatal epilepsy, late-onset episodic ataxia, myoclonus, and pain. *Neurology* 2010a; 75: 1454–8.
- Liao Y, Deprez L, Maljevic S, Pitsch J, Claes L, Hristova D, et al. Molecular correlates of age-dependent seizures in an inherited neonatal-infantile epilepsy. *Brain* 2010b; 133 (Pt 5): 1403–14.
- Lossin C, Shi X, Rogawski MA, Hirose S. Compromised function in the Na(v)1.2 Dravet syndrome mutation R1312T. *Neurobiol Dis* 2012; 47: 378–84.
- Martin HC, Kim GE, Pagnamenta AT, Murakami Y, Carvill GL, Meyer E, et al. Clinical whole-genome sequencing in severe early-onset epilepsy reveals new genes and improves molecular diagnosis. *Hum Mol Genet* 2014; 23: 3200–11.
- Matalon D, Goldberg E, Medne L, Marsh ED. Confirming an expanded spectrum of SCN2A mutations: a case series. *Epileptic Disord* 2014; 16: 13–8.
- Mercimek-Mahmutoglu S, Patel J, Cordeiro D, Hewson S, Callen D, Donner EJ, et al. Diagnostic yield of genetic testing in epileptic encephalopathy in childhood. *Epilepsia* 2015; 56: 707–16.
- Micheva KD, Wolman D, Mensh BD, Pax E, Buchanan J, Smith SJ, et al. A large fraction of neocortical myelin ensheathes axons of local inhibitory neurons. *Elife* 2016; 5: e15784.
- Misra SN, Kahlig KM, George AL Jr. Impaired NaV1.2 function and reduced cell surface expression in benign familial neonatal-infantile seizures. *Epilepsia* 2008; 49: 1535–45.
- Nakamura K, Kato M, Osaka H, Yamashita S, Nakagawa E, Haginoya K, et al. Clinical spectrum of SCN2A mutations expanding to Ohtahara syndrome. *Neurology* 2013; 81: 992–8.
- Need AC, Shashi V, Hitomi Y, Schoch K, Shianna KV, McDonald MT, et al. Clinical application of exome sequencing in undiagnosed genetic conditions. *J Med Genet* 2012; 49: 353–61.
- Ogiwara I, Ito K, Sawaishi Y, Osaka H, Mazaki E, Inoue I, et al. *De novo* mutations of voltage-gated sodium channel alphaII gene SCN2A in intractable epilepsies. *Neurology* 2009; 73: 1046–53.
- Ohba C, Kato M, Takahashi N, Osaka H, Shiihara T, Tohyama J, et al. *De novo* KCNT1 mutations in early-onset epileptic encephalopathy. *Epilepsia* 2015; 56: e121–8.
- Rauch A, Wieczorek D, Graf E, Wieland T, Ende S, Schwarzmayr T, et al. Range of genetic mutations associated with severe non-syndromic sporadic intellectual disability: an exome sequencing study. *Lancet* 2012; 380: 1674–82.
- Rusconi R, Scalmani P, Cassulini RR, Giunti G, Gambardella A, Franceschetti S, et al. Modulatory proteins can rescue a trafficking defective epileptogenic Nav1.1 Na+ channel mutant. *J Neurosci* 2007; 27: 11037–46.
- Saitoh M, Ishii A, Ihara Y, Hoshino A, Terashima H, Kubota M, et al. Missense mutations in sodium channel SCN1A and SCN2A predispose children to encephalopathy with severe febrile seizures. *Epilepsy Res* 2015; 117: 1–6.
- Samanta D, Ramakrishnaiah R. *De novo* R853Q mutation of SCN2A gene and west syndrome. *Acta Neurol Belg* 2015; 115: 773–6.
- Sanders SJ, Murtha MT, Gupta AR, Murdoch JD, Raubeson MJ, Willsey AJ, et al. *De novo* mutations revealed by whole-exome

- sequencing are strongly associated with autism. *Nature* 2012; 485: 237–41.
- Sawaishi Y, Yano T, Enoki M, Takada G. Lidocaine-dependent early infantile status epilepticus with highly suppressed EEG. *Epilepsia* 2002; 43: 201–4.
- Scalmani P, Rusconi R, Armatura E, Zara F, Avanzini G, Franceschetti S, et al. Effects in neocortical neurons of mutations of the Na(v)1.2 Na<sup>+</sup> channel causing benign familial neonatal-infantile seizures. *J Neurosci* 2006; 26: 10100–9.
- Schwarz N, Hahn A, Bast T, Muller S, Loffler H, Maljevic S, et al. Mutations in the sodium channel gene SCN2A cause neonatal epilepsy with late-onset episodic ataxia. *J Neurol* 2016; 263: 334–43.
- Shao D, Okuse K, Djamgoz MB. Protein-protein interactions involving voltage-gated sodium channels: post-translational regulation, intracellular trafficking and functional expression. *Int J Biochem Cell Biol* 2009; 41: 1471–81.
- Shi X, Yasumoto S, Nakagawa E, Fukasawa T, Uchiya S, Hirose S. Missense mutation of the sodium channel gene SCN2A causes Dravet syndrome. *Brain Dev* 2009; 31: 758–62.
- Striano P, Bordo L, Lispi ML, Specchio N, Minetti C, Vigeveno F, et al. A novel SCN2A mutation in family with benign familial infantile seizures. *Epilepsia* 2006; 47: 218–20.
- Sugawara T, Tsurubuchi Y, Agarwala KL, Ito M, Fukuma G, Mazaki-Miyazaki E, et al. A missense mutation of the Na<sup>+</sup> channel alpha II subunit gene Na(v)1.2 in a patient with febrile and afebrile seizures causes channel dysfunction. *Proc Natl Acad Sci USA* 2001; 98: 6384–9.
- Sundaram SK, Chugani HT, Tiwari VN, Huq AH. SCN2A mutation is associated with infantile spasms and bitemporal glucose hypometabolism. *Pediatr Neurol* 2013; 49: 46–9.
- Tavassoli T, Kolevzon A, Wang AT, Curchack-Lichtin J, Halpern D, Schwartz L, et al. *De novo* SCN2A splice site mutation in a boy with autism spectrum disorder. *BMC Med Genet* 2014; 15: 35.
- Touma M, Joshi M, Connolly MC, Grant PE, Hansen AR, Khwaja O, et al. Whole genome sequencing identifies SCN2A mutation in monozygotic twins with Ohtahara syndrome and unique neuropathologic findings. *Epilepsia* 2013; 54: e81–5.
- Trump N, McTague A, Brittain H, Papandreou A, Meyer E, Ngoh A, et al. Improving diagnosis and broadening the phenotypes in early-onset seizure and severe developmental delay disorders through gene panel analysis. *J Med Genet* 2016; 53: 310–7.
- Wang JW, Shi XY, Kurahashi H, Hwang SK, Ishii A, Higurashi N, et al. Prevalence of SCN1A mutations in children with suspected Dravet syndrome and intractable childhood epilepsy. *Epilepsy Res* 2012; 102: 195–200.
- Weiss LA, Escayg A, Kearney JA, Trudeau M, MacDonald BT, Mori M, et al. Sodium channels SCN1A, SCN2A and SCN3A in familial autism. *Mol Psychiatry* 2003; 8: 186–94.
- Wong VC, Fung CW, Kwong AK. SCN2A mutation in a Chinese boy with infantile spasm—response to modified atkins diet. *Brain Dev* 2015; 37: 729–32.
- Xu R, Thomas EA, Jenkins M, Gazina EV, Chiu C, Heron SE, et al. A childhood epilepsy mutation reveals a role for developmentally regulated splicing of a sodium channel. *Mol Cell Neurosci* 2007; 35: 292–301.
- Zara F, Specchio N, Striano P, Robbiano A, Gennaro E, Paravidino R, et al. Genetic testing in benign familial epilepsies of the first year of life: clinical and diagnostic significance. *Epilepsia* 2013; 54: 425–36.
- Zerem A, Lev D, Blumkin L, Goldberg-Stern H, Michaeli-Yossef Y, Halevy A, et al. Paternal germline mosaicism of a SCN2A mutation results in Ohtahara syndrome in half siblings. *Eur J Paediatr Neurol* 2014; 18: 567–71.