



HAL
open science

Description and analysis of genetic variants in French hereditary breast and ovarian cancer families recorded in the UMD-BRCA1/BRCA2 databases

Sandrine Caputo, Louisa Benboudjema, Olga Sinilnikova, Etienne Rouleau, Christophe Bérout, Rosette Lidereau

► To cite this version:

Sandrine Caputo, Louisa Benboudjema, Olga Sinilnikova, Etienne Rouleau, Christophe Bérout, et al.. Description and analysis of genetic variants in French hereditary breast and ovarian cancer families recorded in the UMD-BRCA1/BRCA2 databases. *Nucleic Acids Research*, 2012, 40 (D1), pp.D992 - D1002. 10.1093/nar/gkr1160 . hal-01681814

HAL Id: hal-01681814

<https://amu.hal.science/hal-01681814v1>

Submitted on 1 Jun 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

Description and analysis of genetic variants in French hereditary breast and ovarian cancer families recorded in the UMD-BRCA1/BRCA2 databases

Sandrine Caputo^{1,*}, Louisa Benboudjema¹, Olga Sinilnikova^{2,3}, Etienne Rouleau¹, Christophe Béroud^{4,5,6}, Rosette Lidereau¹ and the French BRCA GGC Consortium[†]

¹Institut Curie—Hôpital René Huguenin, Service d'Oncogénétique, U735 INSERM—Saint-Cloud,

²Unité Mixte de Génétique Constitutionnelle des Cancers Fréquents, Hospices Civils de Lyon—Centre Léon

Bérard, Lyon, ³INSERM U1052, CNRS UMR5286, Université Lyon 1, Centre de Recherche en Cancérologie de

Lyon, Lyon, ⁴INSERM U827—Montpellier, ⁵CHU Montpellier, Hôpital Arnaud de Villeneuve, Laboratoire de

Génétique Moléculaire—Montpellier and ⁶Université Montpellier1, UFR Médecine—Montpellier, F-34000, France

Received August 15, 2011; Revised October 15, 2011; Accepted November 10, 2011

ABSTRACT

***BRCA1* and *BRCA2* are the two main genes responsible for predisposition to breast and ovarian cancers, as a result of protein-inactivating mono-allelic mutations. It remains to be established whether many of the variants identified in these two genes, so-called unclassified/unknown variants (UVs), contribute to the disease phenotype or are simply neutral variants (or polymorphisms). Given the clinical importance of establishing their status, a nationwide effort to annotate these UVs was launched by laboratories belonging to the French GGC consortium (Groupe Génétique et Cancer), leading to the creation of the UMD-BRCA1/BRCA2 databases (<http://www.umd.be/BRCA1/> and <http://www.umd.be/BRCA2/>). These databases have been endorsed by the French National Cancer Institute (INCa) and are designed to collect all variants detected in France, whether causal, neutral or UV. They differ from other BRCA databases in that they contain co-occurrence data for all variants. Using these data, the GGC French consortium has been able to classify certain UVs also contained in other databases. In this article, we report some novel UVs not contained in the BIC database and explore their impact in**

cancer predisposition based on a structural approach.

INTRODUCTION

BRCA1 and *BRCA2* are the two main genes predisposing individuals to breast and/or ovarian cancer. Most monoallelic germline mutations in these genes are point variants or deletions/insertions of a few nucleotides. Large rearrangements (exon loss or duplication, 5' alterations, etc...) have also been described (1,2). These mutations are situated throughout the entire sequence of the two genes. In France, causal mutations in *BRCA1* and *BRCA2* genes account for about 15% of hereditary breast and ovarian cancers (HBOC) (3).

The pathogenic nature of variants resulting in a premature termination codon (PTC) has been clearly established for those variants leading to nonsense-mediated decay (NMD) (4–7), i.e. *BRCA1* proteins with a PTC located upstream from the last 10 carboxy-terminal codons, and *BRCA2* proteins with a PTC located upstream from the last 110 carboxy-terminal codons. Some specific missense variants can generate a PTC via abnormal splicing, while others clearly alter the function of *BRCA1*, including all those occurring in the RING finger and some located in BRCT domains (8). The pathogenicity of these variants has been inferred from linkage analysis of high-risk families, functional assays, biochemical studies and/or demonstration of abnormal mRNA processing (8–10).

*To whom correspondence should be addressed. Email: sandrine.caputo@curie.net

[†]The members of the BRCA GGC Consortium are listed in the Appendix

The authors wish it to be known that, in their opinion, the first two authors should be regarded as joint First Authors.

The main difficulty encountered in molecular diagnosis of breast and ovarian cancer predisposition relates to the large number of variants with unclear clinical significance. Variants that do not create a PTC represent a significant proportion of DNA variants and are usually called unclassified variants (UVs). UVs mostly consist of missense and intronic variants and in-frame deletions or insertions. Exhaustive collections of UVs could allow cosegregation studies within pedigrees, and these complementary data could help to define a reliable classification. Whether a given UV contributes to the disease phenotype or is merely a neutral variant (or a neutral polymorphism) has direct clinical implications.

Several approaches can be used to determine pathogenicity, including multidisciplinary analysis of cosegregation within pedigrees and degree of family history of the disease, or by indirect measures, including amino acid conservation, severity of amino acid change and evidence from functional assays, outputs from available bioinformatics tools, tumor pathology (11,12), population genetics and comparisons of allele frequencies (13), lack of co-occurrence with causal mutations (14), RNA and protein function studies (15), evolutionary studies of amino acid conservation (16) and a recently proposed combinatorial approach (17). However, such approaches cannot definitively show whether or not a specific variant is causal.

Mutation databases can help to classify variants by providing a unique source of information. The Breast Cancer Information Core Database (<http://research.nhgri.nih.gov/projects/bic/index.shtml>; BIC) is a voluntary international collection of variants identified worldwide (18). Other online databases include LOVD (Leiden Open Variation Database)-literature unclassified variant (<http://chromium.liacs.nl/LOVD2/cancer/home.php>) (19), LOVD-IARC (<http://brca.iarc.fr/LOVD/home.php>), kConFab (Kathleen Cuninghame Foundation Consortium for Research into Familial Breast Cancer) (20), The Singapore Human Mutation and Polymorphism Database (21), and HGMD (The Human Gene Mutation Database at the Institute of Medical Genetics in Cardiff) (22), NGRL/DmuDb (<https://ngrl.manchester.ac.uk/Browser/genes.htm>).

In this report, we present exhaustive databases of French *BRCA1/2* variants (UMD-*BRCA1/BRCA2*: <http://www.umd.be/BRCA1/> and <http://www.umd.be/BRCA2/>) developed with UMD software (23). These databases have been endorsed by the French Cancer National Institute (INCa) and are available online. A curator collects and compiles information from 16 licensed laboratories belonging to the French GGC consortium (Groupe Génétique et Cancer) and located all over France. UMD-*BRCA1/BRCA2* databases centralize all variants (causal, neutral and UVs), allowing identification of co-occurrence data. Data on polymorphisms were not collected. In October 2011, the UMD-*BRCA1/BRCA2* databases contained data from a set of 3743 families for *BRCA1* and 3,650 families for *BRCA2*. In this report, we first analyze *BRCA1/BRCA2* variants in the French population, and then use our co-occurrence data to classify some UVs as either causal or neutral.

Finally, we describe *BRCA1* and *BRCA2* UVs and evaluate their pathogenicity, notably from a structural standpoint (Supplementary Data). Therefore, this database should be a tool to classify variant and promote specific studies.

MATERIALS AND METHODS

Software and search engine

The UMD-*BRCA1/BRCA2* databases were constructed with an updated version of the Universal Mutation Database generic software (<http://www.umd.be>) (23,24), which includes an optimized structure to assist and secure data entry and to allow the input of various clinical or biological data. An offline copy of UMD-*BRCA1/BRCA2* is continuously edited and updated by the curators; edited copies are regularly deposited on the server. These databases are freely available online. Their integrity is ensured by the original (edited) template offline. Notification of errors in the current version would be gratefully received by the curator author. Database design follows general recommendations for LSDB characteristics and their curation as proposed by the Human Genome Variation Society (HGVS) (25).

Website

The Website is divided into six sections, four of which are dedicated to information on the *BRCA1* or *BRCA2* genes and numbering of exons and cDNA sequences, information on the protein and the references, while two sections allow the user to search into the database. Hyperlinks are provided to access the reference sequences and data search capabilities. The web pages also contain links to external information including central biological databases, such as the Human Genome Database (GDB), Pubmed, GeneCard, UniGene, Uniprot, OMIM, HGV, etc.

Probands

About 24 000 probands were managed at the medical genetics departments of French Comprehensive Cancer Care centers and hospitals between January 1995 and October 2011. Eligibility criteria for familial genetic testing are described in the French consensus statement (26). All probands enrolled in this study gave their written informed consent for specific genetic testing. The probands were the index cases of individual families and were screened for *BRCA1* and *BRCA2* variants. Records in the French UMD-*BRCA1/BRCA2* databases are restricted to index cases of families with clearly causal mutations, neutral variants or UVs. A system of five classes has been used for classification (27). An anonymized and unique sample identifier (sample_ID) is generated for each family.

Variant screening

The full coding sequences and exon-intron junctions of the *BRCA1* and *BRCA2* genes were screened for variants, based on prescreening (DGGE, SSCP, PTA,

dHPLC, HRM or EMMA) and sequencing. Several large rearrangements were identified by large cDNA sequencing, MLPA (28), QMPSF (29), qPCR (30), qPCR HRM (31), EMMA (32), bar code screening (33) or dedicated array CGH (34).

UMD-BRCA1/BRCA2 databases

The UMD software includes an automatic procedure to check for the correct description of the sequence variation at the nucleotide level and to generate the variant name at the protein level. All variant types are documented. All variants were subjected to a curation process and named according to the HGVS nomenclature before integration in the databases. Variants previously described either in publications or in other *BRCA1/2* databases were reviewed. Data on 7396 different *BRCA1/BRCA2* families were collected between 1995 and October 2011 (Table 2). Co-occurrence of each UV with causal mutations was recorded in the databases. Data on polymorphism were not collected. Families were recorded only once, independently of the number of members tested. No clinical data were collected.

The present version of *BRCA1* and *BRCA2* is based on GenBank reference sequences NM_007294.2 (mRNA: U14680) and NM_000059.3 (mRNA: U43746), respectively. For *BRCA1*, this entry includes 119 bases of 5'-untranslated sequence with protein translation starting at nucleotide 120. The reference *BRCA2* sequence also includes the 228-base 5'-untranslated region.

Searching tools

The UMD knowledgebase system includes many analytical tools (23,24). In addition to previously developed routines, new tools have been specifically developed for the UMD-BRCA1/BRCA2 for in-depth analysis. The user is able to optimize multicriteria search tools to select records from any field.

Prequeried data can be displayed by several Searching Tools in 'Mutations' such as 'Type and number of mutations' or 'Mutation by exon/intron' (Supplementary Figure S1A). Alternatively, the option 'I found a mutation' provides access to a quick search (nucleotide or amino acid position), 'Free search' provides access to an advanced search and a customized search interface searching several items (sample ID, amino acid position, biological significance, etc). A simple click on hyperlinks provides access to a list of interesting variants (Supplementary Data).

Variant analysis

For co-occurrence analysis, a UV *in trans* with two different causal mutations, was considered to be unlikely to cause disease.

The impact of missense substitutions in the *BRCA1/2* genes was predicted by using three online programs: Align-GVGD (35), SIFT (36) and PolyPhen (37), which predict the effect of missense substitutions on gene

function, as well as the new UMD-Predictor[®] tool included in the UMD package (17).

For structural analysis, we focused on some novel variants not contained in the BIC database and located in a conserved region or in a previously reported functional domain. Regions and domains of *BRCA1* or *BRCA2* are described in Supplementary Data (Supplementary Figure S2).

RESULTS

UMD-BRCA1/BRCA2 databases

In UMD-BRCA1, a total of 1391 distinct *BRCA1* variants comprising 586 causal mutations (42%), 5 likely causal mutations (0.4%), 22 likely neutral variants (1.6%), 74 neutral variants (5%) and 704 UVs (51%) have been identified in 3743 French families (1 index case harbors two different causal *BRCA1* mutations and 81 cases harbor one causal mutation associated with at least one UV) (Tables 1 and 2). In UMD-BRCA2, a total of 1704 distinct *BRCA2* variants were reported in 3650 families, with 567 different causal mutations (33.1%), 17 likely neutral variants (1%), 30 neutral variants (1.8%) and 1092 UVs (64.1%) (1 index case harbors two different causal *BRCA2* mutations and 129 cases harbor one causal mutation and at least one UV) (Tables 1 and 2). Three index cases were double heterozygotes for causal *BRCA1* and *BRCA2* mutations. Most variants were found in a unique family (517 of 704 *BRCA1* UVs and 738 of 1092 *BRCA2* UVs) (Table 1). A total of 3743 *BRCA1* families (2636 causal mutations and 948 UVs) and 3650 *BRCA2* families (1576 causal mutations and 1775 UVs) were found to have at least one mutation (Table 2).

The exon distribution of the UVs, neutral variants and causal mutations in the UMD-BRCA1/BRCA2 databases was also assessed and no statistically significant differences were found between the UMD-BRCA1/BRCA2 and BIC databases (see Supplementary Data). Based on the ratio between UMD-BRCA1/BRCA2 variants and predicted variants, *BRCA1* exons 2, 3, 5, 17, 18, 19 and 21 are the most frequently mutated exons (Supplementary Table S1A). Exons 2, 3, and 5 correspond to the RING-finger, exons 17, 18 and 19 to the first BRCT domain and exon 21 to the second BRCT. In *BRCA2*, exons 2, 3, 18 and 23 are the most frequently mutated exons (Supplementary Table S1B). Exon 18 corresponds to part of HD and OB1 domains and exon 23 corresponds to part of the OB2 domain. Structural data have been previously available for only a small segment of exon 3 (residues 21–39), and no 3D structures have been described or predicted for exon 2.

Characterization of French causal mutations in the UMD-BRCA1/BRCA2 databases

The spectrum of causal mutations in the *BRCA1* and *BRCA2* genes involves the entire sequence (Figure 1). The *BRCA1* exons most frequently mutated are exons 2, 5 and 20 and the *BRCA2* exons most frequently mutated are exons 8, 13 and 23 (Figure 1).

Table 1. The October 2011 issue of UMD-BRCA1/BRCA2 databases

	Causal	Likely causal	UV	Likely neutral	Neutral	Total
<i>BRCA1</i>						
Records	2733 (51.4%)	17 (0.3%)	1200 (22.6%)	190 (3.6%)	1171 (22.1%)	5311
Distinct variants	586	5	704	22	74	1391
Unique variants	311	1	517	5	9	843
<i>BRCA2</i>						
Records	1707 (30.9%)	–	2289 (41.4%)	387 (7%)	1149 (20.7%)	5532
Distinct variants	567	–	1092	17	30	1704
Unique variants	337	–	738	3	0	1078

Causal mutations, UVs and neutral

Table 2. Families and records distribution in the UMD-BRCA1/BRCA2 databases

	Causal mutation	UV	1 causal mutation + x UV	Total of families
<i>BRCA1</i> 5311 entries/3743 families				
Families with 1 variation	2636	948	–	3584
Families with 2 variations	8	50	76	134
Families with 3 variations	–	16	3	19
Families with 4 variations	–	4	2	6
Total of families	2644	1018	81	3743
<i>BRCA2</i> 5532 entries/3650 families				
Families with 1 variation	1576	1775	–	3351
Families with 2 variations	1	141	116	258
Families with 3 variations	–	19	12	31
Families with 4 variations	–	5	1	6
Families with 5 variations	–	2	–	2
Families with 6 variations	–	2	–	2
Total of families	1577	1944	129	3650

Causal mutations and UVs. There are three supplementary families with 1 causal *BRCA1* mutation and 1 causal *BRCA2* mutation.

The most prevalent causal *BRCA1* mutations identified in the French population were: the French founder frame-shift c.3481_3491del (3600del11, exon 11) from North-Eastern France (137 records) (38,39) and c.5266dup (5382insC, exon 20), a founder Ashkenazi Jewish mutation (138 records). This causal mutation was found 48 times in Ile-de-France and 27 times in North-Eastern France. Six other frequent recurrent causal *BRCA1* mutations were observed more than 8 times (Table 3): all had been recorded early in the BIC database (October 2011), except for c.5128G>T (9 records), which was found 8 times in North-Eastern France (39).

We identified eight frequent causal *BRCA2* mutations (Table 3), all of which were contained in the BIC database (August 2011). The most prevalent causal *BRCA2* mutation in the French population is c.2808_2811delACAA, a Western European mutation (41 records) (40). All of these eight recurrent causal *BRCA2* mutations were frequently detected in the Paris region.

In comparison with the BIC database, the UMD database also identified causal mutations from the Ashkenazi Jewish population (26.88% for BIC-BRCA1 versus 6.14% UMD-BRCA1, 9.49% for BIC-BRCA2 versus 0.6% for UMD-BRCA2), but also several frequent causal mutations from other European ethnic populations.

Characterization of French UVs in the UMD-BRCA1/BRCA2 databases

UVs were also detected throughout the two genes. The most frequently mutated exons are 17, 18 and 19 for *BRCA1* and 15, 20 and 26 for *BRCA2* (Figure 1).

As expected, numerous variants (at least 51% of distinct variants in *BRCA1* and 64.1% of distinct variants in *BRCA2*) could not be classified as causal or neutral. These UVs were found in *BRCA1* in 1018 families and in *BRCA2* in 1944 families. In *BRCA1*, 704 distinct UVs were identified in one or several families: 948 families with one *BRCA1* UV only, 81 with a UV associated with a causal mutation and 70 with UVs only (Tables 1 and 2). In *BRCA2*, 1092 distinct UVs were identified: 1775 families with one *BRCA2* UV only, 129 with a UV associated with a causal mutation and 169 with UVs only (Tables 1 and 2).

Most UVs were observed only once (517/704 in *BRCA1* and 738/1092 in *BRCA2*) (Table 1), while some were recurrent, including *BRCA1* c.5075-53C>T (IVS17-53C>T, SNP rs8176258) with 66 entries, c.5277+48_59dup (IVS20+48_59dup, 47 entries) and *BRCA2* c.7008-62A>G (IVS13-62A>G, 69 entries), c.9257-16T>C (IVS24-16T>C, SNP rs11571818, 50 entries) and c.6100C>T (p.Arg 2034Cys, exon 11, SNP rs1799954, 85 entries).

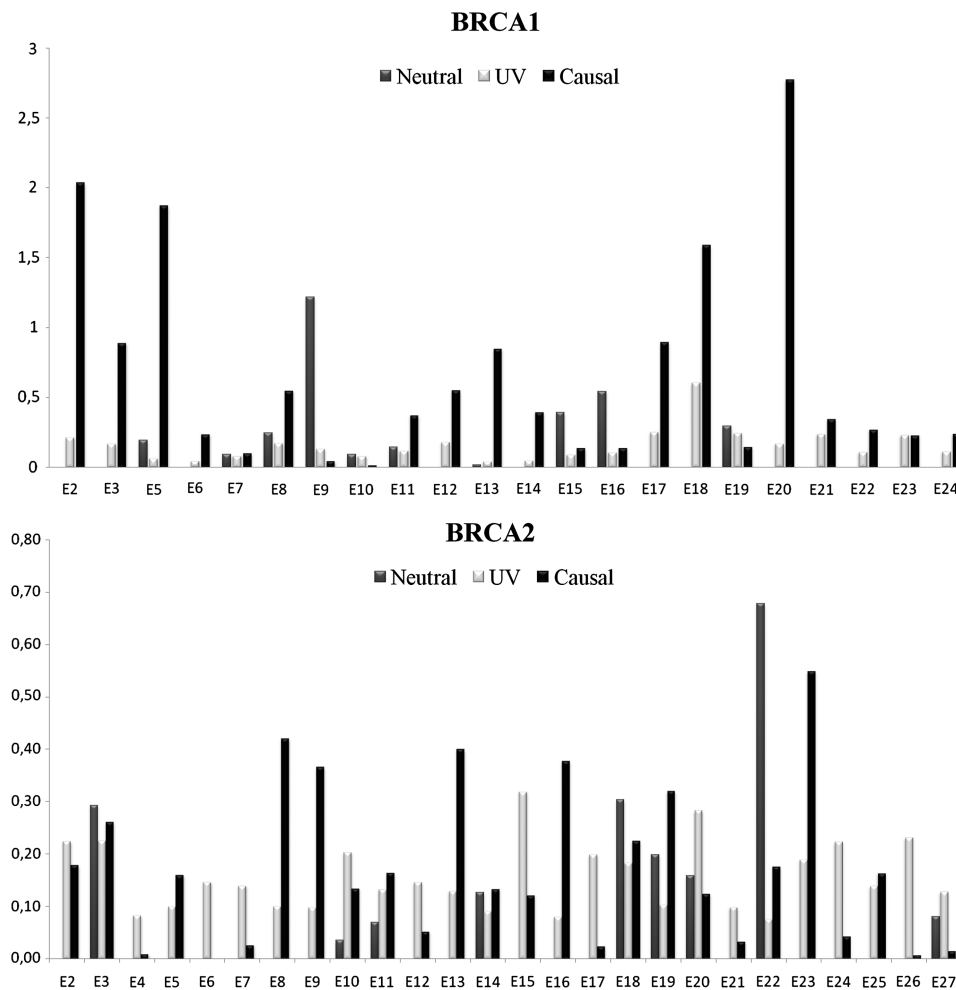


Figure 1. Distribution of all variants in the UMD-BRCA1/BRCA2 databases. The number of entries of all variants is represented per nucleotide and per exon of *BRCA1* or *BRCA2*.

Most UVs identified in this study were either missense changes (45% in *BRCA1* and 53.58% in *BRCA2*) or silent variants (11.5% in *BRCA1* and 15.38% in *BRCA2*) (Figure 2). Small deletions or insertions of various lengths were also observed (2.6% in *BRCA1* and 2.19% in *BRCA2*) (Figures 2), as well as intronic variants (40.9% in *BRCA1* and 15.38% in *BRCA2*) (Figure 2). Intronic variants lie outside intron–exon boundaries and their potential impact on the splicing process is difficult to predict and need specific RNA analysis (Figure 2).

GGC classification of some UVs already reported in other databases

Co-occurrence data on a UV *in trans* of a known causal mutation (the UV and the causal mutation are on opposite chromosomes) can provide a powerful means of ruling out pathogenicity (14). Identification of co-occurrence of a UV and a causal mutation requires further investigations such as cloning to exclude the *in cis* position of these two variants (41). Without these additional analyses, there is still a very weak probability that the two causal mutations are situated *in cis*, which is why the following hypothesis

can be used: if a UV coexists with at least two different causal mutations in two different families, this UV can be considered to be always *in trans* with respect to the causal mutation. To support the co-occurrence results, we also used an integrated approach combining the Align-GVGD, SIFT and PolyPhen programs, published data, and the UMD-Predictor tool, allowing 3 *BRCA2* UVs to be classified as neutral and polymorphism (Table 4). These *BRCA2* UVs, described in the BIC database, were reported in our databases as co-occurring with causal mutations: c.6100C>T [p.Arg2034Cys, exon 11, 137 records, 16 *in trans* (9 distinct causal mutations)], c.7008-62A>G (IVS13-62A>G, 106 records, 6 distinct causal mutations *in trans*), and c.9257-16T>C [IVS24-16T>C, 55 records, 8 *in trans* (4 distinct causal mutations)]. This suggests that these variants do not sufficiently alter *BRCA2* function to predispose to cancer. The status of the *BRCA2* c.6100C>T variant differed between the algorithms: it was predicted to be pathogenic by the UMD-Predictor[®] tool and neutral by SIFT, Align-GVGD, PolyPhen and co-occurrence analysis. Moreover, c.6100C>T has been described as a polymorphism in a Spanish population of cancer families and patients (42). We classified this variant as a neutral

Table 3. Geographical distribution of founder causal mutations in *BRCA1* and *BRCA2* genes

		BRCA1									
		c.68_69delAG exon 2 (185delAG) Ashkenazi Jews	c.3839_3843delCTCAG exon 11 (3958del5ins4) French	c.3481_3491del11 exon 11 (3600del11) North-Eastern France	c.3841C>T exon 11 (3960C>T) France/Belgium/Holland	c.4327C>T exon 13 (4446C>T) French Canadian	c.5030_5033delCTAA exon 17 (5149delCTAA) French	c.5128G>T exon 18 (5247G>T) North-Eastern France	c.5266dup exon 20 (5382insC) Ashkenazi Jews		
IDF		28	12	18	6	17	13	–	–	48	
NW		1	15	4	2	25	3	–	–	11	
North		1	–	2	13	–	7	–	–	5	
NE		5	3	94	4	1	1	8	–	27	
SE		14	–	12	–	1	5	–	–	–	
SW		1	–	3	4	2	3	–	–	11	
Center		5	11	3	2	2	–	1	–	14	
UMD database (3158 records)		56 (1.77%)	41 (1.29%)	137 (4.34%)	31 (0.98%)	48 (1.52%)	32 (1.01%)	9 (0.29%)	–	138 (4.37%)	
BIC database (11329 records)		1980 (17.5%)	3 (0.03%)	58 (0.51%)	12 (0.11%)	126 (1.11%)	17 (0.15%)	0	–	1063 (9.38%)	
		BRCA2									
		c.771_775delTCAAAA exon 9 (999del5) Iceland	c.1310_1313delAAGA exon 10 (1538del4) European	c.2808_2811delACAA exon 11 (3036del4) Western Europe	c.5645C>A exon 11 (5873C>A) East European	c.5946delT exon 11 (6174delT) Ashkenazi Jews	c.6275_6276delTT exon 11 (6503delTT) Dutch	c.6644_6647delACTC exon 11 (6872del4) French	c.9026_9030delATCAT exon 23 (9254del5) Northeast Spanish		
IDF		4	8	21	8	13	8	4	–	14	
NW		–	1	5	–	1	2	1	–	–	
North		–	–	1	–	–	1	1	–	–	
NE		–	3	6	7	2	–	5	–	–	
SE		5	5	3	1	2	3	3	–	7	
SW		–	–	3	1	–	–	3	–	3	
Center		1	3	2	–	1	1	1	–	–	
UMD database (3134 records)		10 (0.32%)	20 (0.64%)	41 (1.3%)	17 (0.54%)	19 (0.6%)	15 (0.48%)	18 (0.57%)	–	25 (0.80%)	
BIC database (11330 records)		14 (0.12%)	10 (0.09%)	105 (0.93%)	26 (0.23%)	1087 (9.49%)	93 (0.82%)	20 (0.18%)	–	10 (0.09%)	

Numbers of families with founder mutations throughout the country (Polymorphisms are excluded); IDF: Paris; NW: Caen, Nantes; North: Lille; NE: Nancy, Reims, Strasbourg; SE: Montpellier, Marseille, Nice; SW: Bordeaux, Toulouse; Center: Lyon, Clermont-Ferrand. * χ^2 test.

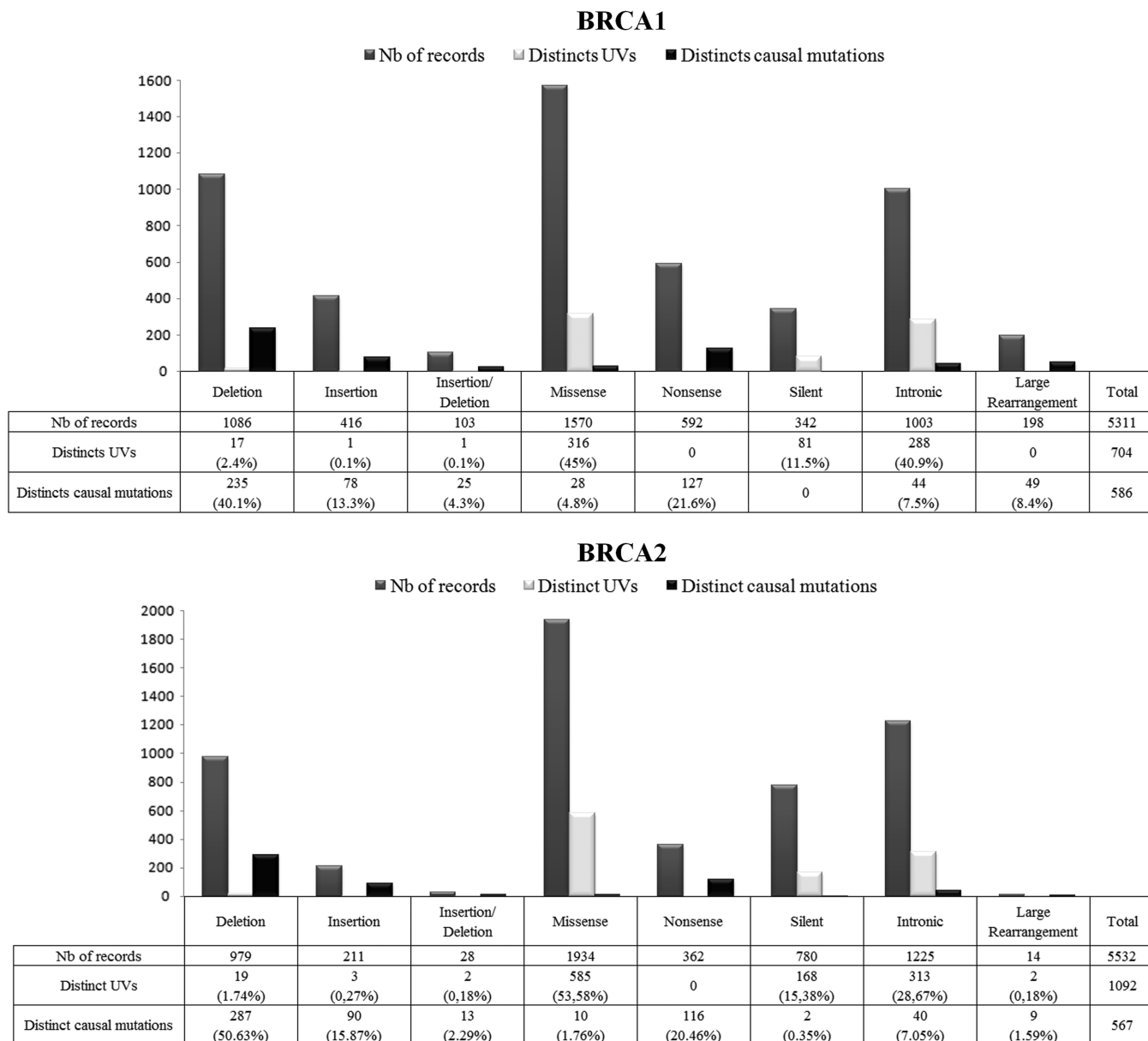


Figure 2. Summary of all allelic variants contained in the UMD-*BRCA1/2* mutation databases. Total number of types of variants in the *BRCA1* and *BRCA2* genes.

variant. Our classification is reported in the UMD-*BRCA1/BRCA2* databases. When it possible, a structural approach is used to help classify the UVs (see example in [Supplementary Data: A structural approach to novel UVs](#)).

DISCUSSION

The UMD-*BRCA1/2* databases provide an overview of the *BRCA1/2* variants in the French population

We examined *BRCA1/2* UVs collected by 16 licensed laboratories located throughout France and belonging to the French GGC consortium ‘Groupe Génétique et Cancer’, which is designed to validate diagnostic procedures and strategies. The 16 licensed laboratories automatically send all results of families tested in France with a

BRCA1/2 variant. Every 3 months, an update is performed by all laboratories. The UMD-*BRCA1/BRCA2* databases have been approved by the French National Cancer Institute (INCa).

Among 24 000 families tested, 2733 *BRCA1* and 1707 *BRCA2* families with one causal mutation were identified, as well as 1018 *BRCA1* and 1,944 *BRCA2* families with at least one UV (Table 2). About 95% of *BRCA1/2* variants (causal mutations and UVs) were observed only once.

Certain exons showed an excess of variants. Interestingly, in *BRCA1*, these exons correspond to the RING-finger domain and the first BRCT domain, while in *BRCA2*, the most frequently mutated exons are not associated with resolved structural domains ([Supplementary Tables S1A and S1B](#)).

Table 4. Variants classified by GGC

Variant (cDNA level) Variant (protein level)	BRCA2		
	c.6100C>T p.Arg2034Cys	c.7008-62A>G –	c.9257-16T>C –
Reported in UMD-BRCA1/BRCA2	137	106	55
Co-occurrence with a causal mutation	16 (9)	6	8 (4)
dbSNP	Rs1 799 954	–	rs11 571 818
Align-GVGD	GV = 257.44, GD = 11.33, C0 (less likely)	ND	ND
PolyPhen	0.423 (benign)	ND	ND
SIFT	0.07 (no deleterious)	ND	ND
UMD-Predictor [®]	82 (pathogenic)	ND	ND
Literature data	Polymorphism	Neutral	Neutral/Polymorphism
Classification BIC database	UV (97)	UV (2)	UV (6)
Classification UMD-BRCA1/BRCA2 database	Polymorphism	Neutral	Neutral

The enriched deleterious group consisted of those substitutions with Align-GVGD scores of $GV < 62$ and $GD > 0$ and SIFT scores ≤ 0.05 . The enriched neutral group consisted of those substitutions with Align-GVGD scores of $GV > 0$ and $GD < 62$ and SIFT scores > 0.1 . PolyPhen results for each variant were classified as benign ($\text{score} \leq 0.5$), possibly damaging ($0.5 < \text{score} < 2$), probably damaging ($\text{score} > 2$), and unknown. UMD predictor[®] results for each missense were classified as polymorphism ($\text{score} \leq 50$), probable polymorphism ($50 \leq \text{score} \leq 64$), probably pathogenic ($65 \leq \text{score} \leq 74$) and pathogenic ($\text{score} > 74$). ND: not determined because algorithms only predict the impact of missense mutations.

As in other populations, some variants are recurrent in the French population (40,43–45). Sixteen recurrent causal mutations in *BRCA1* and *BRCA2* were reported more than eight times each. Some UVs were also recurrent. Three were most frequent in *BRCA2*, namely c.7008-62A>G (106 families), c.9257-16T>C (55 families) and c.6100C>T (137 families). These three variants could be geographic polymorphisms, or specific to the French population. Detection of c.7008-62A>G and c.9257-16T>C could also be variable according to the positions of the primers used to amplify the region and the frequency differs from one database to another due to the selection of PCR primers.

The UMD-BRCA1/BRCA2 databases and pooled data from the French GGC are sufficiently powerful for co-occurrence studies. If homozygous and compound heterozygous variants in *BRCA1* are assumed to be embryonically lethal (46–48), while those in *BRCA2* are responsible for Fanconi anemia (49–51), several variants can be classified as neutral based on a single observation. One patient reported in the UMD-BRCA1/BRCA2 databases harbors two different causal *BRCA1* mutations *in cis* (Bezieau, S. and Delnatte, C. unpublished data). Another patient harbors two different causal *BRCA2* mutations. These two causal mutations result in a breast/ovarian cancer syndrome but also in glioblastoma (Sevenet, N., Longy, M. et al. unpublished data).

A double heterozygote with a causal *BRCA1* mutation and a causal *BRCA2* mutation did not appear to present a more severe phenotype (52). We also identified three families that included double heterozygotes with a causal mutation in *BRCA1* and a causal mutation in *BRCA2*. Coexistence of causal mutations in *BRCA1* and *BRCA2* is unusual in non-Ashkenazi populations. Two of these three families did not harbor the Ashkenazi founder mutations (unpublished data) (52), but these three families presented a breast cancer and other cancers syndrome.

UVs classified by GGC

Most information used to determine the pathogenicity of an UV is indirect, such as segregation data, loss of heterozygosity, absence of the UV in an unaffected subject or *in silico* prediction based on the conservation, position and nature of amino acid change or the impact on the mRNA.

We used co-occurrence data and predictive algorithms to classify three *BRCA2* variants (Table 4). The French consortium uses these data to classify UVs in the UMD-BRCA1/BRCA2 databases. For example, like Goldgar *et al.*, we classified the *BRCA2* c.6100C>T (p.Arg2034Cys) variant as a neutral variant (53). In our database, this c.6100C>T variant was associated with eight distinct causal mutations strongly suggesting that it is not causal. However, the status of the *BRCA2* c.6100C>T variant differed between the algorithms: SIFT score was 0.07, corresponding to borderline pathogenicity (0.05). The UMD Predictor[®] tool contained the score SIFT but also other scores. The final conclusion of neutrality was based on a combination of prediction programs with co-occurrence data. We discuss the contribution of the 3D structure in Supplementary Data.

CONCLUSION

BRCA1 and *BRCA2* are the two genes principally involved in predisposition to breast and ovarian cancer. Classification of UVs of *BRCA1* and *BRCA2* has major implications for hereditary forms of these malignancies. Most variants identified to date cannot yet be definitively classified as causal or neutral. The French UMD-BRCA1/BRCA2 databases, which collect all variants (neutral, causal and UVs), differ from other BRCA databases by the fact that they contain co-occurrence data for all variants, allowing the GGC consortium to classify a number of UVs. In the future, co-occurrence data and structural approaches should help to classify further unclassified *BRCA1/BRCA2* variants.

SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online: Supplementary Tables 1 and 2, Supplementary Figures 1–7, Supplementary References [8,17,23,27,54–61].

ACKNOWLEDGMENTS

The authors thank the French oncogeneticists, the GGC leads by Dr Catherine Noguès and probands for their cooperation.

FUNDING

French National Cancer Institute (INCa) (also to S.C. and L.B.); Development of the UMD tool used to construct the UMD-BRCA1/BRCA2 databases received funding from the European Community's Seventh Framework Program (FP7/2007-2013) under grant agreement no. 200754—the GEN2PHEN project; Association d'Aide à la Recherche Cancérologique de Saint Cloud (ARCs). Funding for open access charge: Institut Curie—Hospital René Huguenin.

Conflict of interest statement. None declared.

REFERENCES

- Mazoyer,S. (2005) Genomic rearrangements in the BRCA1 and BRCA2 genes. *Hum. Mutat.*, **25**, 415–422.
- Sluiter,M.D. and van Rensburg,E.J. (2010) Large genomic rearrangements of the BRCA1 and BRCA2 genes: review of the literature and report of a novel BRCA1 mutation. *Breast Cancer Res. Treat.*, **125**, 325–349.
- INCa. (2010) *Consultations et Laboratoires*. Institut National du Cancer, Boulogne-Billancourt.
- Ware,M.D., DeSilva,D., Sinilnikova,O.M., Stoppa-Lyonnet,D., Tavtigian,S.V. and Mazoyer,S. (2006) Does nonsense-mediated mRNA decay explain the ovarian cancer cluster region of the BRCA2 gene? *Oncogene*, **25**, 323–328.
- Anczukow,O., Ware,M.D., Buisson,M., Zetoune,A.B., Stoppa-Lyonnet,D., Sinilnikova,O.M. and Mazoyer,S. (2008) Does the nonsense-mediated mRNA decay mechanism prevent the synthesis of truncated BRCA1, CHK2, and p53 proteins? *Hum. Mutat.*, **29**, 65–73.
- Caux-Moncoutier,V., Pages-Berhouet,S., Michaux,D., Asselain,B., Castera,L., De Pauw,A., Buecher,B., Gauthier-Villars,M., Stoppa-Lyonnet,D. and Houdayer,C. (2009) Impact of BRCA1 and BRCA2 variants on splicing: clues from an allelic imbalance study. *Eur J Hum Genet.*, **17**, 1471–1480.
- Chen,X., Truong,T.T., Weaver,J., Bove,B.A., Cattie,K., Armstrong,B.A., Daly,M.B. and Godwin,A.K. (2006) Intronic alterations in BRCA1 and BRCA2: effect on mRNA splicing fidelity and expression. *Hum. Mutat.*, **27**, 427–435.
- Carvalho,M.A., Marsillac,S.M., Karchin,R., Manoukian,S., Grist,S., Swaby,R.F., Urmenyi,T.P., Rondinelli,E., Silva,R., Gayol,L. *et al.* (2007) Determination of cancer risk associated with germ line BRCA1 missense variants by functional analysis. *Cancer Res.*, **67**, 1494–1501.
- Sinilnikova,O.M., Mazoyer,S., Bonnardel,C., Lynch,H.T., Narod,S.A. and Lenoir,G.M. (2006) BRCA1 and BRCA2 mutations in breast and ovarian cancer syndrome: reflection on the Creighton University historical series of high risk families. *Fam. Cancer*, **5**, 15–20.
- Morris,J.R., Pangon,L., Boutell,C., Katagiri,T., Keep,N.H. and Solomon,E. (2006) Genetic analysis of BRCA1 ubiquitin ligase activity and its relationship to breast cancer susceptibility. *Hum. Mol. Genet.*, **15**, 599–606.
- Goldgar,D.E., Easton,D.F., Byrnes,G.B., Spurdle,A.B., Iversen,E.S. and Greenblatt,M.S. (2008) Genetic evidence and integration of various data sources for classifying uncertain variants into a single model. *Hum. Mutat.*, **29**, 1265–1272.
- Goldgar,D.E., Easton,D.F., Deffenbaugh,A.M., Monteiro,A.N., Tavtigian,S.V. and Couch,F.J. (2004) Integrated evaluation of DNA sequence variants of unknown clinical significance: application to BRCA1 and BRCA2. *Am. J. Hum. Genet.*, **75**, 535–544.
- Lee,E., McKean-Cowdin,R., Ma,H., Chen,Z., Van Den Berg,D., Henderson,B.E., Bernstein,L. and Ursin,G. (2008) Evaluation of unclassified variants in the breast cancer susceptibility genes BRCA1 and BRCA2 using five methods: results from a population-based study of young breast cancer patients. *Breast Cancer Res.*, **10**, R19.
- Judkins,T., Hendrickson,B.C., Deffenbaugh,A.M., Eliason,K., Leclair,B., Norton,M.J., Ward,B.E., Pruss,D. and Scholl,T. (2005) Application of embryonic lethal or other obvious phenotypes to characterize the clinical significance of genetic variants found in trans with known deleterious mutations. *Cancer Res.*, **65**, 10096–10103.
- Couch,F.J., Rasmussen,L.J., Hofstra,R., Monteiro,A.N., Greenblatt,M.S. and de Wind,N. (2008) Assessment of functional effects of unclassified genetic variants. *Hum. Mutat.*, **29**, 1314–1326.
- Fleming,M.A., Potter,J.D., Ramirez,C.J., Ostrander,G.K. and Ostrander,E.A. (2003) Understanding missense mutations in the BRCA1 gene: an evolutionary approach. *Proc. Natl Acad. Sci. USA*, **100**, 1151–1156.
- Frederic,M.Y., Lalande,M., Boileau,C., Hamroun,D., Claustres,M., Beroud,C. and Collod-Beroud,G. (2009) UMD-predictor, a new prediction tool for nucleotide substitution pathogenicity—application to four genes: FBN1, FBN2, TGFB1, and TGFB2. *Hum. Mutat.*, **30**, 952–959.
- Szabo,C., Masiello,A., Ryan,J.F. and Brody,L.C. (2000) The breast cancer information core: database design, structure, and scope. *Hum. Mutat.*, **16**, 123–131.
- Fokkema,I.F., den Dunnen,J.T. and Taschner,P.E. (2005) LOVD: easy creation of a locus-specific sequence variation database using an “LSDB-in-a-box” approach. *Hum. Mutat.*, **26**, 63–68.
- Osborne,R.H., Hopper,J.L., Kirk,J.A., Chenevix-Trench,G., Thorne,H.J. and Sambrook,J.F. (2000) kConFab: a research resource of Australasian breast cancer families. Kathleen Cuninghame Foundation Consortium for Research into Familial Breast Cancer. *Med. J. Aust.*, **172**, 463–464.
- Tan,E.C., Loh,M., Chuon,D. and Lim,Y.P. (2006) Singapore human mutation/polymorphism database: a country-specific database for mutations and polymorphisms in inherited disorders and candidate gene association studies. *Hum. Mutat.*, **27**, 232–235.
- Cooper,D.N., Stenson,P.D. and Chuzhanova,N.A. (2006) The human gene mutation database (HGMD) and its exploitation in the study of mutational mechanisms. *Curr. Protoc. Bioinformatics*, **Chapter 1**, Unit 1 13.
- Beroud,C., Hamroun,D., Collod-Beroud,G., Boileau,C., Soussi,T. and Claustres,M. (2005) UMD (Universal Mutation Database): 2005 update. *Hum. Mutat.*, **26**, 184–191.
- Beroud,C., Collod-Beroud,G., Boileau,C., Soussi,T. and Junien,C. (2000) UMD (Universal mutation database): a generic software to build and analyze locus-specific databases. *Hum. Mutat.*, **15**, 86–94.
- Cotton,R.G., Auerbach,A.D., Beckmann,J.S., Blumenfeld,O.O., Brookes,A.J., Brown,A.F., Carrera,P., Cox,D.W., Gottlieb,B., Greenblatt,M.S. *et al.* (2008) Recommendations for locus-specific databases and their curation. *Hum. Mutat.*, **29**, 2–5.
- Eisinger,F., Alby,N., Bremond,A., Dauplat,J., Espie,M., Janiaud,P., Kuttann,F., Lebrun,J.P., Lefranc,J.P., Pierret,J. *et al.* (1999) Inserm ad hoc committee: recommendations for the management of women with a genetic risk for developing cancer of the breast and/or the ovary. *Bull. Cancer*, **86**, 307–313.
- Plon,S.E., Eccles,D.M., Easton,D., Foulkes,W.D., Genuardi,M., Greenblatt,M.S., Hogervorst,F.B., Hoogerbrugge,N., Spurdle,A.B.

- and Tavtigian, S.V. (2008) Sequence variant classification and reporting: recommendations for improving the interpretation of cancer susceptibility genetic test results. *Hum. Mutat.*, **29**, 1282–1291.
28. Schouten, J.P., McElgunn, C.J., Waaijer, R., Zwijnenburg, D., Diepvens, F. and Pals, G. (2002) Relative quantification of 40 nucleic acid sequences by multiplex ligation-dependent probe amplification. *Nucleic Acids Res.*, **30**, e57.
 29. Casilli, F., Di Rocco, Z.C., Gad, S., Tournier, I., Stoppa-Lyonnet, D., Frebourg, T. and Tosi, M. (2002) Rapid detection of novel BRCA1 rearrangements in high-risk breast-ovarian cancer families using multiplex PCR of short fluorescent fragments. *Hum. Mutat.*, **20**, 218–226.
 30. Barrois, M., Bieche, I., Mazoyer, S., Champeme, M.H., Bressac-de Paillerets, B. and Lidereau, R. (2004) Real-time PCR-based gene dosage assay for detecting BRCA1 rearrangements in breast-ovarian cancer families. *Clin. Genet.*, **65**, 131–136.
 31. Rouleau, E., Lefol, C., Bourdon, V., Coulet, F., Noguchi, T., Soubrier, F., Bieche, I., Olschwang, S., Sobol, H. and Lidereau, R. (2009) Quantitative PCR high-resolution melting (qPCR-HRM) curve analysis, a new approach to simultaneously screen point mutations and large rearrangements: application to MLH1 germline mutations in Lynch syndrome. *Hum. Mutat.*, **30**, 867–875.
 32. Weber, J., Miserere, S., Champ, J., Looten, R., Stoppa-Lyonnet, D., Viovy, J.L. and Houdayer, C. (2007) High-throughput simultaneous detection of point mutations and large-scale rearrangements by CE. *Electrophoresis*, **28**, 4282–4288.
 33. Gad, S., Aurias, A., Puget, N., Mairal, A., Schurra, C., Montagna, M., Pages, S., Caux, V., Mazoyer, S., Bensimon, A. *et al.* (2001) Color bar coding the BRCA1 gene on combed DNA: a useful strategy for detecting large gene rearrangements. *Genes Chromosomes Cancer*, **31**, 75–84.
 34. Rouleau, E., Lefol, C., Tozlu, S., Andrieu, C., Guy, C., Copigny, F., Nogues, C., Bieche, I. and Lidereau, R. (2007) High-resolution oligonucleotide array-CGH applied to the detection and characterization of large rearrangements in the hereditary breast cancer gene BRCA1. *Clin. Genet.*, **72**, 199–207.
 35. Tavtigian, S.V., Deffenbaugh, A.M., Yin, L., Judkins, T., Scholl, T., Samollow, P.B., de Silva, D., Zharkikh, A. and Thomas, A. (2006) Comprehensive statistical study of 452 BRCA1 missense substitutions with classification of eight recurrent substitutions as neutral. *J. Med. Genet.*, **43**, 295–305.
 36. Ng, P.C. and Henikoff, S. (2003) SIFT: predicting amino acid changes that affect protein function. *Nucleic Acids Res.*, **31**, 3812–3814.
 37. Ramensky, V., Bork, P. and Sunyaev, S. (2002) Human non-synonymous SNPs: server and survey. *Nucleic Acids Res.*, **30**, 3894–3900.
 38. Stoppa-Lyonnet, D., Laurent-Puig, P., Essioux, L., Pages, S., Ithier, G., Ligt, L., Fourquet, A., Salmon, R.J., Clough, K.B., Pouillart, P. *et al.* (1997) BRCA1 sequence variations in 160 individuals referred to a breast/ovarian family cancer clinic. Institut Curie Breast Cancer Group. *Am. J. Hum. Genet.*, **60**, 1021–1030.
 39. Muller, D., Bonaiti-Pellie, C., Abecassis, J., Stoppa-Lyonnet, D. and Fricker, J.P. (2004) BRCA1 testing in breast and/or ovarian cancer families from northeastern France identifies two common mutations with a founder effect. *Fam. Cancer*, **3**, 15–20.
 40. Hall, M.J., Reid, J.E., Burbidge, L.A., Pruss, D., Deffenbaugh, A.M., Frye, C., Wenstrup, R.J., Ward, B.E., Scholl, T.A. and Noll, W.W. (2009) BRCA1 and BRCA2 mutations in women of different ethnicities undergoing testing for hereditary breast-ovarian cancer. *Cancer*, **115**, 2222–2233.
 41. Suzuki, K., Iwata, M., Tsuji, H., Takagi, T., Tamura, A., Ishimoto, G., Ito, S., Matsui, K. and Miyazaki, T. (1997) A de novo recombination in the ABO blood group gene and evidence for the occurrence of recombination products. *Hum. Genet.*, **99**, 454–461.
 42. Diez, O., Osorio, A., Duran, M., Martinez-Ferrandis, J.I., de la Hoya, M., Salazar, R., Vega, A., Campos, B., Rodriguez-Lopez, R., Velasco, E. *et al.* (2003) Analysis of BRCA1 and BRCA2 genes in Spanish breast/ovarian cancer patients: a high proportion of mutations unique to Spain and evidence of founder effects. *Hum. Mutat.*, **22**, 301–312.
 43. Machado, P.M., Brandao, R.D., Cavaco, B.M., Eugenio, J., Bento, S., Nave, M., Rodrigues, P., Fernandes, A. and Vaz, F. (2007) Screening for a BRCA2 rearrangement in high-risk breast/ovarian cancer families: evidence for a founder effect and analysis of the associated phenotypes. *J. Clin. Oncol.*, **25**, 2027–2034.
 44. Petrij-Bosch, A., Peelen, T., van Vliet, M., van Eijk, R., Olmer, R., Drusedau, M., Hogervorst, F.B., Hageman, S., Arts, P.J., Ligtenberg, M.J. *et al.* (1997) BRCA1 genomic deletions are major founder mutations in Dutch breast cancer patients. *Nat. Genet.*, **17**, 341–345.
 45. Palomba, G., Pisano, M., Cossu, A., Budroni, M., Dedola, M.F., Farris, A., Contu, A., Balduin, P., Tanda, F. and Palmieri, G. (2005) Spectrum and prevalence of BRCA1 and BRCA2 germline mutations in Sardinian patients with breast carcinoma through hospital-based screening. *Cancer*, **104**, 1172–1179.
 46. Hohenstein, P., Kielman, M.F., Breukel, C., Bennett, L.M., Wiseman, R., Krimpenfort, P., Cornelisse, C., van Ommen, G.J., Devilee, P. and Fodde, R. (2001) A targeted mouse Brca1 mutation removing the last BRCT repeat results in apoptosis and embryonic lethality at the headfold stage. *Oncogene*, **20**, 2544–2550.
 47. Gowen, L.C., Johnson, B.L., Latour, A.M., Sulik, K.K. and Koller, B.H. (1996) Brca1 deficiency results in early embryonic lethality characterized by neuroepithelial abnormalities. *Nat. Genet.*, **12**, 191–194.
 48. Evers, B. and Jonkers, J. (2006) Mouse models of BRCA1 and BRCA2 deficiency: past lessons, current understanding and future prospects. *Oncogene*, **25**, 5885–5897.
 49. Howlett, N.G., Taniguchi, T., Olson, S., Cox, B., Waisfisz, Q., De Die-Smulders, C., Persky, N., Grompe, M., Joenje, H., Pals, G. *et al.* (2002) Biallelic inactivation of BRCA2 in Fanconi anemia. *Science*, **297**, 606–609.
 50. Alter, B.P., Rosenberg, P.S. and Brody, L.C. (2007) Clinical and molecular features associated with biallelic mutations in FANCD1/BRCA2. *J. Med. Genet.*, **44**, 1–9.
 51. Ludwig, T., Chapman, D.L., Papaioannou, V.E. and Efstratiadis, A. (1997) Targeted mutations of breast cancer susceptibility gene homologs in mice: lethal phenotypes of Brca1, Brca2, Brca1/Brca2, Brca1/p53, and Brca2/p53 nullizygous embryos. *Genes Dev.*, **11**, 1226–1241.
 52. Leegte, B., van der Hout, A.H., Deffenbaugh, A.M., Bakker, M.K., Mulder, I.M., ten Berge, A., Leenders, E.P., Wesseling, J., de Hullu, J., Hoogerbrugge, N. *et al.* (2005) Phenotypic expression of double heterozygosity for BRCA1 and BRCA2 germline mutations. *J. Med. Genet.*, **42**, e20.
 53. Simard, J., Dumont, M., Moisan, A.M., Gaborieau, V., Malouin, H., Durocher, F., Chiquette, J., Plante, M., Avard, D., Bessette, P. *et al.* (2007) Evaluation of BRCA1 and BRCA2 mutation prevalence, risk prediction models and a multistep testing approach in French-Canadian families with high risk of breast and ovarian cancer. *J. Med. Genet.*, **44**, 107–121.
 54. Williams, R.S., Green, R. and Glover, J.N. (2001) Crystal structure of the BRCT repeat region from the breast cancer-associated protein BRCA1. *Nat. Struct. Biol.*, **8**, 838–842.
 55. Shiozaki, E.N., Gu, L., Yan, N. and Shi, Y. (2004) Structure of the BRCT repeats of BRCA1 bound to a BACH1 phosphopeptide: implications for signaling. *Mol. Cell*, **14**, 405–412.
 56. Yang, H., Jeffrey, P.D., Miller, J., Kinnucan, E., Sun, Y., Thoma, N.H., Zheng, N., Chen, P.L., Lee, W.H. and Pavletich, N.P. (2002) BRCA2 function in DNA binding and recombination from a BRCA2-DSS1-ssDNA structure. *Science*, **297**, 1837–1848.
 57. Karchin, R., Monteiro, A.N., Tavtigian, S.V., Carvalho, M.A. and Sali, A. (2007) Functional impact of missense variants in BRCA1 predicted by supervised learning. *PLoS Comput. Biol.*, **3**, e26.
 58. Vallon-Christersson, J., Cayanan, C., Haraldsson, K., Loman, N., Bergthorsson, J.T., Brondum-Nielsen, K., Gerdes, A.M., Moller, P., Kristofferson, U., Olsson, H. *et al.* (2001) Functional analysis of BRCA1 C-terminal missense mutations identified in breast and ovarian cancer families. *Hum. Mol. Genet.*, **10**, 353–360.
 59. Williams, R.S., Chasman, D.I., Hau, D.D., Hui, B., Lau, A.Y. and Glover, J.N. (2003) Detection of protein folding defects caused by

- BRCA1-BRCT truncation and missense mutations. *J. Biol. Chem.*, **278**, 53007–53016.
60. Nikolopoulos,G., Pyrpassopoulos,S., Thanassoulas,A., Klimentzou,P., Zikos,C., Vlassi,M., Vorgias,C.E., Yannoukakos,D. and Nounesis,G. (2007) Thermal unfolding of human BRCA1 BRCT-domain variants. *Biochim. Biophys. Acta*, **1774**, 772–780.
61. Karchin,R., Agarwal,M., Sali,A., Couch,F. and Beattie,M.S. (2008) Classifying variants of undetermined significance in BRCA2 with protein likelihood ratios. *Cancer Inform.*, **6**, 203–216.

APPENDIX

French BRCA GGC consortium: Nicolas Sevenet, Françoise Bonnet, Michel Longy: Institut Bergonié—Bordeaux; Agnès Hardouin, Dominique Vaur, Sophie Krieger: Centre François Baclesse—Caen; Nancy Uhrhammer, Yves-Jean Bignon: Centre Jean Perrin—Clermont-Ferrand; Jean-Philippe Peyrat, Françoise Revillion, Joëlle Fournier: Centre Oscar Lambret—Lille; Audrey Remenieras, Sylvie Mazoyer: Centre de Recherche en Cancérologie de Lyon—Lyon; Mélanie Léone: Hospices Civils de Lyon and Centre Léon

Bérard—Lyon; Hagay Sobol, Tetsuro Noguchi, Violaine Bourdon, Audrey Remenieras: Institut Paoli-Calmettes—Marseille; Jean-Marc Rey: Laboratoire de Biologie Cellulaire et Hormonale (CHU Arnaud de Villeneuve)—Montpellier; Myriam Bronner, Philippe Jonveaux, Christophe Philippe: CHU de Nancy-Brabois—Vandœuvre-lès-Nancy; Capucine Delnatte: CHU—Institut de Biologie—Hôtel Dieu—Nantes; Florence Coulet—Groupe hospitalier Pitié-Salpêtrière, Assistance Publique-Hôpitaux de Paris, Université Pierre et Marie Curie, Laboratoire d'Oncogénétique et Angiogénétique moléculaire—Paris; Laurent Castera, Virginie Caux-Moncoutier, Claude Houdayer, Dominique Stoppa-Lyonnet: Institut Curie—Paris; Chantal Delvincourt, Marie-Claude Gorisse: CHU et Institut Jean Godinot—Reims; Ivan Bièche, Cédric Lefol: Institut Curie—Hôpital René Huguenin, Service d'Oncogénétique—Saint Cloud; Danièle Muller, Joseph Abecassis: Centre Paul Strauss—Strasbourg; Christine Toulas: Institut Claudius Régaud—Toulouse; Marine Guillaud-Bataille, Brigitte Bressac-De Pailleters: Institut Gustave Roussy—Villejuif.