

# Role of the light source position in freely falling hoverflies' stabilization performances

Roman Goulard, Anna Verbe, Jean-Louis Vercher, Stéphane Viollet

## ▶ To cite this version:

Roman Goulard, Anna Verbe, Jean-Louis Vercher, Stéphane Viollet. Role of the light source position in freely falling hoverflies' stabilization performances. Biology Letters, 2018, 14 (5), pp.20180051. 10.1098/rsbl.2018.0051 . hal-01799095

# HAL Id: hal-01799095 https://amu.hal.science/hal-01799095

Submitted on 29 May 2018

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Photo credit: Anna Verbe

# Role of the light source position in freely falling hoverflies ´ stabilization performances

Roman Goulard, Anna Verbe, Jean-Louis Vercher, Stéphane Viollet\*

Aix-Marseille Université, CNRS, ISM UMR 7287, Marseille 13009, France

\*corresponding author: stephane.viollet@univ-amu.fr

## Abstract

The stabilization of plummeting hoverflies was filmed and analysed in terms of their wingbeat initiation times as well as the crash and stabilization rates. The flies experienced nearweightlessness for a period of time which depended on their ability to counteract the free fall by triggering their wingbeats. In this paradigm, hoverflies' flight stabilization strategies were investigated here for the first time under two different positions of the light source (overhead or bottom lighting). The crash rates were higher in bottom lighting conditions than with top lighting. In addition, adding a texture to the walls reduced the crash rates only in the overhead lighting condition. The position of the lighting also significantly affected both the stabilization rates and the time taken by the flies to stabilize, which decreased and increased under bottom lighting conditions, respectively, whereas textured walls increased the stabilization rates under both lighting conditions. These results support the idea that flies may mainly base their flight control strategy on visual cues and particularly, that the light distribution in the visual field may provide reliable, efficient cues for estimating their orientation with respect to an allocentric reference frame. In addition, the finding that the hoverflies' optic flow-based motion detection ability is affected by the position of the light source in their visual field suggests the occurrence of interactions between movement perception and this visual vertical perception process.

keywords: flight stabilization; insect vision; Dorsal Light Response; hoverflies; dipterans

## 1 **Introduction**

Several flying insects including dipterans are known to keep their dorsal surface oriented up-2 wards by holding the brightest part of their environment, which usually shines from above, in 3 a constant position in their visual field [21]. This reflex, which is known as the Dorsal Light 4 Response (DLR) has also been described in detail in fish [5]. The importance of the orientation 5 of an artificial horizon in blowflies' head roll orientation processes has also been previously 6 established, as well as the fact that these insects probably do not use gravity information to 7 perform this task [10], which has been assessed in freely flying hoverflies based on free fall 8 experiments [8], contrary to the well-known negative gravitaxy behaviour observed in walking 9 Drosophila [18, 17]. These results suggest that visual processes predominate over gravity-based 10 ones in the strategies used by flying flies to stabilize their flight, and support the idea that 11 there exists some kind of vertical reference frame in flies' brains based on the DLR. However, 12 although this reflex had been found to be closely linked to the head roll steering mechanism, 13 this situation has been established only in tethered *Calliphora* and *Episyrphus* [12, 7] and has 14 not been studied so far during free flight. 15

16

In the present study, it is therefore proposed to assess the effects of the change in the light 17 source position on freely flying dipterans' stabilization performances. These performances were 18 tested using a free fall procedure under four different visual conditions in which two differently 19 textured lateral walls (uniform and textured) were combined with two different lighting sources 20 (overhead or bottom lighting). The position of the light source was found to be a crucial factor 21 for hoverflies to be able to regain a suitable flight attitude after a free fall. In addition, the 22 results obtained in this study support the idea that the use of lateral visual cues such as 2-D 23 chequerboard patterns generating optic flow (OF) may be involved in hoverflies' attitude and 24 lift control processes [4, 9]. This study clearly shows not only that the light gradient perception 25 and OF-based control processes are interlinked, but also that the static cues consisting of the 26 light source position predominate over the insects' OF-based control processes. 27



Figure 1: **Experimental setup.**(A) The setup used in this study was first presented in [8]. In the present version, a white backlit LED panel was added to illuminate the box from below. (B) Four environments were tested: Contrasting textured lateral walls with Top lighting (CT), Contrasting textured lateral walls with Bottom lighting (CB), Uniform lateral walls with Top lighting (UT), and Uniform lateral walls with Bottom lighting (UB).

## $_{28}$ 2 Methods

#### 29 Animals

Hoverfly pupae (Episyrphus balteatus) were purchased from Katz Biotech AG, Baruth, Ger-30 many. To magnetically maintain the animals in resting position (see in figure 1A), a piece 31 of entomological pin approximately 5 mm long was glued to the dorsal part of the animals 32 thorax, perpendicularly to their longitudinal axis: the pin ( $\approx 5mg$ ) weighed approximately 33 15% of the hoverfly's mass ( $\approx 35mg$ ). The insects' flight ability was checked in the breeding 34 cages throughout the experiments. 39 hoverflies (19 in Halogen/LED experiments and 20 in 35 the control experiments) aged from 3 to 28 days were tested (9 males and 10 females in the 36 Halogen/LED experiments and 2 males and 18 females in the control experiments). 37

#### <sup>38</sup> Experimental procedure

Hoverflies were subjected to free fall conditions in a modified version of the setup previously presented in [8]. In the present version, the box was illuminated alternately from above (Top Lighting: TL) with a white halogen light (Kaiser Studiolight H) and from below (Bottom Lighting: BL) with a white backlit LED panel (Phlox, 50x50cm<sup>2</sup>) featuring a uniformity as high as 95% and two peaks (450nm and one at around 550nm) that match the spectral sensitivity of the hoverfly's (*Erisalis tenax*) photoreceptor cells [15]).

Four different conditions were tested (fig 1B): Contrasting textured lateral walls with Top 45 lighting (CT), Contrasting textured lateral walls with Bottom lighting (CB), Uniform lateral 46 walls with Top lighting (UT) and Uniform lateral walls with Bottom lighting (UB). In addi-47 tion to these conditions, two control experiments were conducted in which the CT condition 48 was compared with a CB condition with white halogen bottom lighting instead of the LED 49 panel to check whether the LED lighting condition affected the hoverflies' performances. The 50 texture on the walls consisted of a randomly generated chequerboard (20x20 squares  $4cm^2$ ) 51 in size). The irradiance was measured in both illumination conditions with an ILT1700 ra-52 diometer (International Light Technologies) under both experimental conditions (with tex-53 tured walls: CT and CB conditions) by orienting the light probe (SED033, visual field 3°) of 54 the radiometer towards either the illuminated side or the opposite side. The irradiance mea-55 sured in the CT condition (direct/indirect measurements) was  $1.12.10^{-8}/3.15.10-9 W.cm^{-2}$ 56 and  $1.10.10^{-9}/2.33.10^{-10}$  W.cm<sup>-2</sup> in the CB condition with the LED light. In the control 57 experiments with two halogen lights, the irradiance was set at the same value (direct measure-58 ments)  $(1.24.10^{-8} W.cm^{-2}).$ 59

A total number of 262 falls were conducted among the four different conditions (see figure 1B), and 91 additional falls were conducted in the control experiments. At each experimental session, a hoverfly was exposed to the four environments consecutively in random order. Each hoverfly could undergo several experimental sessions, but no more than once a day in order to prevent the occurrence of any habituation or fatigue effects. We always checked between experimental sessions whether the hoverflies equipped with their glued pin were able to fly in the breeding cages.

#### 67 Image analysis

The horizontal and vertical 2-D positions of the hoverflies' centre of mass moving over a uniform background were recorded using a custom-made image-processing program running under MATLAB. The fly's speed was calculated from the positions recorded by applying a Savitzky-Golay procedure (order 2, window: 51). Stabilization was determined automatically when the fly reached a positive vertical speed without touching either a wall or the ground.

#### 73 Statistical analysis

Data were analysed statistically using a generalized linear mixed-effects model procedure ('glmer'
in R v3.2.3) and selected using the Akaike Information Criterion ['AIC' [1]].

## 76 **3** Results

As observed previously [8], hoverflies subjected to free falls initiated their flight after approxi-77 mately 100ms in both uniform and contrasting wall texture conditions (p = 0.12; F = 2.4727; 78 Figure 2A). However, top lighting conditions significantly decreased the reaction wingbeat 79 triggering) times (p < 0.01; F = 9.4352). The flies' performances in the CB condition dif-80 fered significantly from those observed in the CT (p < 0.05; z = -2.863) and UT (p < 0.01; 81 z = -3.294) conditions, but the differences in the mean times did not exceed 20ms ( $\overline{\Delta}_{WB}$  (ms): 82 CB = 131.2500; CT = 107.7500; UB = 116.5152; UT = 104.3087). It is worth noting that 83 during our control experiments, the wingbeat triggering times (see Figure 2A) were significantly 84 shorter by around 20ms (p < 0.001; F = 19.6268), but that this did not significantly reduce the 85 difference in the effects observed between CB and CT (p = 0.39; F = 0.7303). 86

Bottom lighting conditions induced a much larger number of touchdowns on the floor (Figure 2B), amounting to approximately 60% of all the trials, than under overhead lighting conditions (p < 0.001;  $Chi^2 = 35.8369$ ), which enabled the hoverflies to avoid crashing in 70-75% (UT) to 90% (CT) of the flights. It is worth noting that the crash rates were quite similar between Halogen-LED and control experiments (Effect of LED light: p = 0.68;  $Chi^2 = 0.1694$ ; interaction with light position effect: p = 0.35;  $Chi^2 = 0.8571$ ) which confirms the validity of using a LED panel to stimulate light-dependent stabilization behaviour. In addition, the crash rates were not significantly affected by the presence of textured walls (p = 0.18;  $Chi^2 = 1.7578$ ), whereas a significant interaction was found to occur between the lighting and texture conditions (p = 0.01;  $Chi^2 = 6.5524$ ). The presence of a 2-D chequerboard pattern on the walls significantly decreased the crash rates under overhead lighting conditions (Post-hoc Tukey contrast, CT vs UT: p < 0.05; z = 2.838) but not under bottom lighting conditions (Post-hoc Tukey contrast, CB vs UB: p = 0.95; z = -0.525).

In the subsequent analysis, stabilized flight was taken to occur whenever the fly adopted 100 a positive vertical speed, corresponding to a rising flight, without subsequently crashing onto 101 the floor (Figure 2C). Since the number of stabilized flights observed in the case of UB was 102 very small (n = 3), no definite conclusions could be reached about the effects of this con-103 dition on the stabilization times, and these data were therefore removed from the statisti-104 cal analysis. The lighting conditions significantly affected the stabilization times (p < 0.05; 105 F = 5.5942): the mean stabilization time was approximately 50ms longer in the CB environ-106 ment than that recorded in the two conditions with overhead lighting ( $\bar{\Delta}_{Stab}$  (ms): CB = 222.75; 107 TL (UT & CT) = 169.0465).108

A large number of stabilized flights occurred with overhead lighting and either textured or uni-109 form lateral walls (Figure 2D). In the BL conditions, hoverflies produced poorer stabilization 110 performances than in the TL conditions (p < 0.001;  $Chi^2 = 46.144$ ), and they were almost un-111 able to prevent themselves from continuing to fall in the uniform environment (UB). Hoverflies 112 surrounded by textured lateral walls, i.e., in conditions CT and CB, achieved better perfor-113 mances than under the same lighting condition with uniform walls, i.e., in conditions UT and 114 UB, respectively  $(p < 0.001; Chi^2 = 23.463)$ . It was also observed that the stabilization rates 115 were similar between Halogen-LED and control experiments (Effect of LED light: p = 0.11; 116  $Chi^2 = 2.5727$ ; interaction with light position: p = 0.27;  $Chi^2 = 1.2127$ ) whereas the effects of 117 light position on stabilization time depended on the kind of light (Effect of LED light: p = 0.49; 118  $Chi^2 = 0.4730$ ; interaction with light position: p < 0.01;  $Chi^2 = 7.52$ ). It can be seen from 119 figure 2C that this effect was mainly observed on the CT condition, where no differences seemed 120 likely to occur. The differences were probably due to the fact that different populations were 121 tested in the control and Halogen/Led experiments. 122

123



Figure 2: (A) Boxplot of the wingbeat triggering times (ms). (B) Bar plot of the crash rates. (C) Boxplot of the stabilization times (ms). (D) Bar plot of the stabilized flight rates. Boxes are composed of 1st, 2nd and 3rd quartiles, and whiskers correspond to extreme data, amounting to no more than 1.5 times the interquartile distance. Significance code, p value: 0 < \* \* \* < 0.001 < \* \* < 0.01 < \* < 0.05

## 124 **Discussion**

In this study, it was attempted for the first time to investigate the impact of the light source 125 position in hoverflies' visual field on their ability to stabilize their flight. In previous studies, 126 it was suggested that the light gradient generated by an artificial horizon may impact insects' 127 attitude perception processes via a mechanism called the Dorsal Light Response [12, 7]. Using 128 a free fall procedure, we reported that hoverflies starting to fly in an unsteady initial state 129 were found to be able to recover stabilized flight efficiently only in situations where the light 130 came from above. In addition to the crucial position of the light source, the optic flow infor-131 mation generated during a free fall may also be used by hoverflies to ultimately avoid crashing 132 [11, 4, 27, 9], but these cues probably do not suffice for stabilization purposes. The light gra-133 dient probably provides hoverflies with a means of estimating their absolute orientation in the 134 environment in order to control their attitude, as found to occur in locusts [6]. This static cue 135 providing a subjective vertical reference value about of the external world [10] would certainly 136 require robust visual processing integrating the lighting information over the whole or most of 137 the spherical field of view. To investigate in greater detail the extent to which the light gradient 138 is actually involved, LED panels might be a useful means of finely controlling the homogeneity 139 of the illumination generated and the amplitude of the light gradient, and generating dynamic 140 changes in the lighting conditions. 141

142

One of the main hypotheses put forward in previous studies on flies' sensorimotor reflexes 143 is that they may depend only on movement perception processes and compensatory reflexes 144 [28]. The inputs originating from both visual structures, the compound eyes and the ocelli 145 [19, 3, 23, 24], and from the halteres [25, 22, 13, 2], which are fused together non-linearly [16], 146 may compensate for a large range of disturbances [26]. A system of this kind is liable, however, 147 to be subject to accumulated errors during flight, resulting in a drift in the attitude control 148 process and eventually in crashing. The results obtained here therefore indicate that the DLR 149 may play an important role by providing a reliable time-invariant vertical reference frame which 150 may be used to complement the insects' motion-based reflexes. However, the initial position 151 of hoverflies with their legs dangling may have decreased the ability of the chordotonal organs 152 (organs acting as pressure sensors [30] linked to postural reflex in insects [14, 20] which were 153

stimulated in this situation only by the legs ' weight) to estimate their orientation with respect
to the gravity experienced prior to the fall and we therefore cannot rule out the latter hypothesis.

In conclusion, this initial study shows that the position of the light source plays an impor-157 tant role in hoverflies' flight stabilization processes. The results presented here suggest that 158 both static (light source, DLR) and movement (optic flow) cues are probably involved [10]. A 159 further question which arises here is how these two visual processes (the DLR and OF-based 160 processes) are fused together to ensure robust flight stabilization under natural conditions, as 161 previously suggested in the case of optomotor responses [29]. Future studies in which conflict-162 ing situations are generated would probably help to understand how these different sensory 163 processes are combined in dipterans' brains. 164

165

## 166 Aknowledgments

We are most thankful to Julien Diperi for his contribution to building the experimental setup, to Marc Boyron for developing the electronics on which all the work presented in this paper was based, and to Jessica Blanc for correcting and improving the English manuscript. We acknowledge support from the Centre National de la Recherche Scientifique (CNRS), Aix-Marseille Universite and the Agence Nationale de la Recherche (ANR) [IRIS project (Intelligent Retina for Innovative Sensing) ANR-12- INSE-0009].

## 173 Ethics

No ethical assessments for animal research purposes or permission to carry out fieldwork were
required for this study.

## 176 Data accessibility

<sup>177</sup> Data are provided in Excel tables in the supplementary files (Data\_total.xls).

## <sup>178</sup> Competing interests

<sup>179</sup> We have no competing interests to declare

# 180 Authors' contributions

RG designed and conducted the experiments, analysed the data, carried out the statistical analyses and drafted the manuscript; AV conducted experiments and helped to draft the manuscript; JLV helped with the statistical analyses and with the drafting of the manuscript. SV designed the experiment, coordinated the study and helped to draft the manuscript. All the authors gave their final approval for publication of the manuscript and agree to be held accountable for the work performed herein.

## 187 Funding

<sup>188</sup> No funding support was involved in this research.

## **References**

- [1] H. Akaike. A New Look at the Statistical Model Identification. *IEEE Transactions on Automatic Control*, 19(6):716–723, 1974.
- [2] M. Dickinson. Haltere-mediated equilibrium reflexes of th fruit fly, drosophila melanogaster.
   Philosophical Transactions of the Royal Society of London B, 354:903–916, 1999.
- [3] M. Egelhaaf, R. Kern, H. G. Krapp, J. Kretzberg, R. Kurtz, and A.-K. Warzecha. Neural encoding of behaviourally relevant visual-motion information in the fly. *Trends in neurosciences*, 25(2):96–102, 2002.
- [4] Fabien Expert and Franck Ruffier. Flying over uneven moving terrain based on optic-flow
   cues without any need for reference frames or accelerometers. *Bioinspiration & Biomimet- ics*, 10(2):026003, 2015.

- [5] G. S. Fraenkel and D. L. Gunn. The orientation of animals: Kineses, taxes and compass
   reactions. Oxford University Press, England, 1961.
- [6] L. J. Goodman. The Role Of Certain Optomotor Reactions In Regulating Stability In
   The Rolling Plane During Flight In The Desert Locust, Schistocerca Gregaria. Journal
   of Experimental Biology, 42:385–407, 1965.
- [7] R. Goulard, A. Julien-Laferriere, J. Fleuriet, J.-L. Vercher, and S. Viollet. Behavioural
   evidence for a visual and proprioceptive control of head roll in hoverflies (*episyrphus baltea- tus*). Journal of Experimental Biology, 218(23):3777–3787, 2015.
- [8] R. Goulard, J.-L. Vercher, and S. Viollet. To crash or not to crash: how do hoverflies cope with free-fall situations and weightlessness? *Journal of Experimental Biology*,
  219(16):2497–2503, 2016.
- [9] R. Goulard, J.-L. Vercher, and S. Viollet. Modeling visual-based pitch, lift and speed control strategies in hoverflies. *PLOS Computational Biology*, 14(1), 2018.
- [10] R. Hengstenberg. Localization and Orientation in Biology and Engineering. Lectures Notes
  in Computer Science, pages 121–133. Springer-Verlag Berlin Heidelberg, 1984.
- [11] R. Hengstenberg. Mechanosensory control of compensatory head roll during flight in the
  blowfly Calliphora erythrocephala. Journal of Comparative Physiology A, 163:151–165,
  1988.
- [12] R. Hengstenberg. Visual Motion and its Role in the Stabilization of Gaze. pages 285–298.
  Elsevier Science, 1993.
- [13] R. Hengstenberg. Controlling the fly's gyroscopes. *Nature*, 392:757–758, 1998.
- [14] E. Horn and H.-G. Lang. Positional Head Reflexes and the Role of the Prosternal Organ
   in the Walking Fly, Calliphora erythrocephala. Journal of Comparative Physiology A,
   126:137–146, 1978.
- [15] GA Horridge, K Mimura, and Y Tsukahara. Fly photoreceptors-ii. spectral and polarized
  light sensitivity in the drone fly eristalis. In *Proc. R. Soc. Lond. B*, volume 190, pages
  225–237. The Royal Society, 1975.

27	[16] S. J. Huston and H. G. Krapp. Nonlinear Integration of Visual and Haltere Inputs in Fly
28	Neck Motor Neurons. The Journal of Neuroscience, 29:13097–13105, 2009.

2

2

- [17] Hidehiko K Inagaki, Azusa Kamikouchi, and Kei Ito. Methods for quantifying simple
  gravity sensing in drosophila melanogaster. *Nature protocols*, 5(1):20, 2010.
- [18] A. Kamikouchi, H.K. Inagaki, T. Effertz, O. Hendrich, A. Fiala, M.C. Göpfert, and K. Ito.
  The neural basis of *Drosophila* gravity-sensing and hearing. *Nature*, 458:165–171, 2009.
- [19] H. G. Krapp and B. Hengstenberg. Estimation of self-motion by optic flow processing in
  single visual interneurons. *Nature*, 384:463–466, 1996.
- [20] D. Kress and M. Egelhaaf. Head and body stabilization in blowflies walking on differently
   structured substrates. *Journal of Experimental Biology*, 215:1523–1532, 2012.
- [21] H. Mittelstaedt. Physiologie des Gleichgewichtssinnes bei Fliegenden Libellen. Zeitschrift
   *für vergleichende Physiologie*, 32:422–463, 1950.
- [22] G. Nalbach. The halteres of the blowfly Calliphora. Journal of Comparative Physiology.
   A, 173:293–300, 1993.
- [23] M. M. Parsons, H. G. Krapp, and S.B. Laughlin. A motion-sensitive neurone responds to
   signals from the two visual systems of the blowfly, the compound eyes and ocelli. *Journal* of Experimental Biology, 209:4464–4474, 2006.
- [24] M. M. Parsons, H. G. Krapp, and Laughlin S.B. Sensor Fusion in Identified Visual In terneurons. *Current Biology*, 20:624–628, 2010.
- [25] D.C. Sandeman and H. Markl. Head Movements in Flies (*Calliphora*) Produced by Deflexion of the Halteres. *Journal of Experimental Biology*, 85:43–60, 1980.
- [26] D. A. Schwyn, F. J. H. Heras, G. Bolliger, M. M. Parsons, H. G. Krapp, and R. J. Tanaka.
   Interplay between Feedback and Feedforward Control in Fly Gaze Stabilization. In 18th
   *IFAC World Congress*, pages 9674–9679, 2011.
- [27] Julien R Serres and Franck Ruffier. Optic flow-based collision-free strategies: From insects
   to robots. Arthropod structure & development, 46(5):703-717, 2017.

- [28] G. K. Taylor and H. G. Krapp. Sensory System and Flight Stability: What does Insects
  Measure and Why? Advances in Insect Physiology, 34:231–316, 2007.
- [29] C. Trischler, R. Kern, and M. Egelhaaf. Chasing behaviour and optomotor following in
   free-flying male blowflies: flight performance and interactions of the underlying control
   systems. Frontiers in Behavioral Neuroscience, 4, 2010.
- [30] John C Tuthill and Rachel I Wilson. Mechanosensation and adaptive motor control in
   insects. *Current Biology*, 26(20):R1022–R1038, 2016.