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Mathieu Santonja, Stephane Greff, Marie L Le Croller, Olivier P Thomas,
Thierry Perez

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1 **TITLE: Distance interaction between marine cave-dwelling sponges and crustaceans**

2

3 **AUTHORS**

4 Mathieu Santonja^{1,2}, Stéphane Greff¹, Marie Le Croller¹, Olivier P. Thomas^{1,3}, Thierry Pérez^{1*}

5

6 **ADDRESSES**

7 **1.** Institut Méditerranéen de Biodiversité et d'Ecologie Marine et Continentale (IMBE), UMR
8 7263 CNRS, IRD, Aix Marseille Université, Avignon Université, Station Marine d'Endoume,
9 rue Batterie des Lions, 13007 Marseille, France.

10 **2.** Univ Rennes, CNRS, ECOBIO - UMR 6553, F-35000 Rennes, France.

11 **3.** Marine Biodiscovery Laboratory, School of Chemistry and Ryan Institute, National
12 University of Ireland, Galway (NUI Galway), University Road, H91 TK33 Galway, Ireland.

13

14 *Corresponding author: thierry.perez@imbe.fr

15

16 **ABSTRACT**

17 Sponges are benthic organisms that are dominant in several ecosystems and known to
18 produce a huge chemical diversity. The putative release of some specialized metabolites in the
19 surrounding seawater is still a matter of debate, but the presence of such compounds in the
20 environment of sponges is thought to influence the behaviour of various mobile organisms and
21 may, thus, contribute to benthic ecosystem structuring and functioning. Underwater
22 Mediterranean caves are characterized by stable environmental conditions and sessile species
23 assemblages dominated by sponges. A two-choice test system was developed to assess the
24 response of two cave-dwelling crustaceans (*Hemimysis margalefi* and *Palaemon serratus*) and
25 two other species living in shallow water environments (*Leptomysis* sp. and *Palaemon elegans*)
26 to various seawater treatments: control- seawater from an exposed coastline, control+ seawater,
27 coming from an underwater cave, and seawater conditioned with four Mediterranean sponges
28 commonly found at the entrance of underwater caves (*Aplysina cavernicola*, *Haliclona fulva*,
29 *Oscarella tuberculata* and *Spongia officinalis*) or their chemical extracts. We tested the
30 swimming behaviour of these crustacean species in three complementary experiments: (i)
31 control seawater vs. cave seawater; (ii) control seawater vs. seawater conditioned with the
32 sponge community, (iii) control seawater vs. seawater containing chemical extracts of the same
33 sponge community.

34 Both cave-dwelling crustaceans were attracted by the seawater conditioned with the
35 sponge community, while *Leptomysis* sp. spent more time in the control seawater and
36 *P. elegans* exhibited indifferent responses. All four crustacean species avoided the seawater
37 containing the sponge extracts. Interestingly, the responses shown by the crustaceans was
38 affected by the time of day. A comparative and untargeted metabolomic approach was applied
39 to the surrounding seawater to identify putative chemomarkers that could explain the
40 crustaceans' behaviours. Among other compounds found in the seawater, a family of

41 metabolites with molecular formulae in accordance with those of oxylipin derivatives is
42 released by sponges and may, therefore, serve as chemical cues acting as kairomones in the
43 homing behaviour of cave-dwelling crustaceans.

44

45 **KEYWORDS**

46 Marine Ecology; Chemical mediation; Porifera; Oxylipins; Mediterranean Sea

47

48 **INTRODUCTION**

49 Distance interactions regulate the structure and functioning of marine ecosystems and
50 species distribution within a complex mosaic of habitats. There is a need for better
51 understanding of these interactions, in particular when attempting to assess the putative effects
52 of human disturbances on system functioning ([van de Koppel et al. 2015](#)). Chemical mediation
53 plays a key role in such distance interactions by influencing the behavioural ecology of mobile
54 organisms ([Hay 2009](#)). Seawater can also be seen as a chemical “seascape” composed of
55 dissolved and/or adsorbed molecules of diverse origins, a good number of which are released
56 by marine organisms and carry crucial signals for the whole ecosystem ([Cassier et al. 2000](#)).
57 Surrounded by this complex chemical environment, mobile animals must decipher a complex
58 cocktail of chemical cues, some of which act as kairomones, and adjust their behavioural
59 response by navigating, selecting habitat, locating food or prey, detecting the presence of
60 conspecifics, or defending against competitors and/or predators ([Heuschele and Selander 2014](#)).
61 This complexity also challenges animals’ abilities to extract the information required to identify
62 the sources of chemical cues which are crucial for their survival ([Derby and Sorensen 2008](#)).
63 For example, fishes can detect natural chemical cues at very low concentrations (parts per
64 billion) and, therefore, need to be able to identify the relevant cues from the higher levels of
65 background noise ([Belanger et al. 2006](#)).

66 During the past decades, special attention has been paid to the functioning of tropical
67 coral reefs. One of the greatest challenges has been to better understand the life cycles of littoral
68 populations of marine species living in fragmented habitats in a vast and complex ocean matrix,
69 species which may have dispersal and larval development phases offshore, followed by the
70 recruitment of juveniles on randomly distributed reefs (Myrberg and Fuiman 2002). Several
71 pioneer studies demonstrated that marine fish and crustacean larvae could distinguish between
72 oceanic and reef waters (Atema et al. 2002; Lecchini et al. 2005; Gerlach et al. 2007; Dixson et
73 al. 2008). These larvae were shown to be capable of selecting a given habitat among the great
74 variety of substrates found in tropical waters (Huijbers et al. 2008; Munday et al. 2009; Lecchini
75 et al. 2010; Devine et al. 2012; Igulu et al. 2013). For example, Huijbers et al. (2008) reported
76 that fish recruits discriminate chemical cues from mangroves, seagrass beds and coral reefs and
77 that a given reef fish actively prefers the odour of its home coral reef. Lecchini et al. (2014)
78 also demonstrated that fish, crustacean and cephalopod larvae were significantly more attracted
79 by chemical cues from living corals than by dead colonies. Further studies have highlighted
80 how environmental disturbances leading to coral reef destructions, such as ocean acidification
81 (Munday et al. 2009; Devine et al. 2012; Leduc et al. 2013), seawater warming (McCormick et
82 al. 2010) or pollution (O'Connor et al. 2016), may affect marine chemical mediation.
83 Additionally, these changes in abiotic conditions may also alter the organism's metabolism and,
84 thus, its ability to produce specialized metabolites and may consequently impact chemical
85 mediation between organisms (Fisher et al. 2006; Munday et al. 2009; Leduc et al. 2013).
86 Surprisingly, even though chemicals are involved in these biotic interactions and in overall
87 ecosystem functioning, few studies have attempted to identify the chemical nature of distance
88 marine chemical cues (Lecchini et al. 2005; Lecchini et al. 2010; Lecchini et al. 2014; Rasher
89 et al. 2015).

90 In comparison to this knowledge gained in tropical reefs, chemical mediation in
91 temperate benthic ecosystems has been poorly studied. Sponges can dominate in terms of
92 biomass and diversity in numerous benthic ecosystems, both in tropical and temperate regions
93 (Diaz and Rutzler 2001; Gerovasileiou and Voultsiadou 2012; Wulff 2012). They are also well
94 known for their ability to produce a great chemical diversity (e.g. Paul et al. 2011). However,
95 there is a lack of knowledge concerning the ecological functions of these chemical compounds
96 which can be released into the surrounding water (Thompson 1985; Walker et al. 1985; Ternon
97 et al. 2016). Today, sponge chemical ecology is mainly based on the assessment of repulsive
98 properties, the so-called “chemical defences”, against predators or competitors for space (Paul
99 et al. 2011; Pawlik 2011). Even though sponges are often considered to be dominant
100 components of benthic ecosystems, the putative role of the portion of their specialized
101 metabolites released into the water column remains unclear and few studies have demonstrated
102 a positive influence of these cues on the recruitment of invertebrate larvae (e.g. Bingham and
103 Young 1991). We hypothesize that the great chemical diversity produced by sponges may
104 construct a true chemical seascape which may be the basis of a still underestimated network of
105 interactions shaping biodiversity.

106 We surmise that underwater caves constitute good ecosystem models in which to test
107 this hypothesis. Sponges are predominant from the semi-dark zone where they coexist with
108 cnidarians and bryozoans to the darkest parts of the cave where they can represent 100 % of the
109 sessile biodiversity (Gerovasileiou and Voultsiadou 2012). In low or no light conditions, other
110 putative producers of chemicals such as macroalgae (Amsler 2008) are absent or are restricted
111 to the entrance of this habitat. Experimenting in underwater caves also avoids, or at least
112 attenuates, variations of abiotic factors present in the open sea such as hydrodynamics, nutrient
113 and resource bioavailability and, in some cases, seawater temperature (Rastorgueff et al. 2011).
114 Underwater caves can thus be considered as *in situ* mesocosms where many ecological

115 processes and interactions are simplified. Finally, considering their very low resilience facing
116 major environmental threats (Gunn et al. 2000), a better understanding of this ecosystem
117 functioning is needed to propose suitable management and conservation plans (Gunn et al.
118 2000; Bussotti et al. 2006; Gerovasileiou and Voultsiadou 2012; Rastorgueff et al. 2015).

119 This study focuses on two dominant components of cave ecosystem functioning:
120 sponges, which are active suspension feeders and which compose the vast majority of the sessile
121 fauna, and swimming crustaceans which act on benthic-pelagic couplings through their
122 migration in and out of caves (Rastorgueff et al. 2015). Our aim was to assess the influence of
123 the chemical seascape of a cave which is most likely primarily determined by sponges on the
124 swimming behaviour of crustaceans (*i.e.* attraction or repulsion). We combined the use of (i)
125 an experimental device to assess the behavioural response of the crustaceans, and (ii) a
126 metabolomics fingerprinting approach to decipher the chemical composition of the various
127 seawater treatments. To conduct our experiments, we selected representatives of the sponge
128 community found near the entrance in semi-dark conditions of Mediterranean underwater
129 caves, and four crustacean species with contrasting ecological habits: two cave-dwelling
130 crustaceans and two crustaceans inhabiting shallow water environments.

131

132 MATERIAL AND METHODS

133 *Study location and species*

134 This study was conducted using the aquaria facilities of the Marine Station of Endoume,
135 Marseille, France. Four characteristic sponge species of the semi-dark community (*Aplysina*
136 *cavernicola* (Vacelet, 1959), *Haliclona fulva* (Topsent, 1893), *Oscarella tuberculata* (Schmidt,
137 1868) and *Spongia officinalis* (Linnaeus, 1759); Supplementary Fig. S1) were collected in a
138 cave in the Riou Archipelago at an average of 20 m depth (43°12'N; 5°22'E) and were
139 maintained alive in aquaria. These species were selected because part of their specialized

140 metabolome had already been described (Garrido et al. 1997; Ivanisevic et al. 2011; Manzo et
141 al. 2011; Nuzzo et al. 2012; Reverter et al. 2016; personal unpublished data). In the same cave,
142 we also collected 70 L of seawater to act as a positive control in our first experiment.

143 The four crustacean species selected to perform the chemosensory assays were two
144 mysid species (*Hemimysis margalefi* Alcaraz, Riera and Gili, 1986 and *Leptomysis* sp.) and two
145 palaemonid species (*Palaemon serratus* (Pennant, 1777) and *Palaemon elegans* Rathke, 1837)
146 (Supplementary Fig. S2). *Hemimysis margalefi* (mean length \pm SD, 2.7 ± 0.1 mm) was collected
147 in the same cave as the sponges, where it usually forms dense swarms. This cave-dwelling
148 mysid undertakes circadian migrations in order to find food outside the cave at night or early
149 morning (Rastorgueff et al. 2011), and we hypothesized that its return towards the cave may be
150 influenced by a chemical seascape composed primarily of sponge chemical cues. *Leptomysis*
151 sp. (8.3 ± 0.3 mm) was collected in shallow waters (8 m depth) adjacent to the Marine Station
152 aquarium seawater pumping site (43°16.80'N, 5°20.95'E). This benthic community is
153 dominated by macrophytes such as *Posidonia oceanica* and is, thus, a totally different chemical
154 seascape which is devoid of any sponge influence. *Palaemon serratus* (13.8 ± 0.3 mm) and
155 *P. elegans* (8.2 ± 0.1 mm) individuals were offspring of gravid females that were captured in
156 the Mediterranean Sea and had been raised in aquaria, so were totally naïve regarding the
157 experimental conditions. *Palaemon serratus* is known to live in semi-dark to dark
158 environments, and we hypothesized that its occurrence in caves may be related to the chemical
159 seascape, whereas *P. elegans* lives in very shallow waters among macroalgae assemblages, so
160 they should not be influenced by a seascape conditioned by sponges. For the duration of their
161 time spent in aquaria, all four crustacean species were fed every 2 days with *Artemia salina*
162 nauplii.

163

164 ***Chemosensory trials***

165 A Y-shaped glass flume providing a choice of two channels was set up according to a
166 protocol developed by [Gerlach et al. \(2007\)](#) to assess the crustacean behavioural response when
167 faced with the choice of different seawater treatments: open flow seawater as the control
168 seawater (CS); cave seawater (+CS), considered here as a positive control; and two types of
169 conditioned seawater, sponge conditioned seawater (SCS) and sponge chemical extracts
170 dissolved in seawater (SCE). Two 10 L tanks were connected to the flume by pipes, creating a
171 constant gravity-driven flow of 50 mL min^{-1} per upstream channel. Prior to the experiments,
172 both flows were measured by adding fluorescein to one tank to ensure that the flows were
173 similar and mixed only in the downstream compartment. The crustaceans were introduced at
174 the downstream end of the apparatus, where they were free to remain or to select and swim
175 towards one of the two channels connected to the tanks ([Supplementary Fig. S3](#)). Each assay
176 was performed with a single specimen which was maintained for a 1-min acclimation period
177 followed by a 5-min testing period. During the selection phase of the test, the position of the
178 crustacean in the downstream compartment or in the right or left channels was regularly
179 recorded at 5-sec intervals. After each test, the flume chamber was emptied and washed with
180 the CS. The seawater treatments were then switched from one channel to the other, providing a
181 control for potential channel preferences that were not associated with the water source. After
182 seawater switching, the entire assay was repeated with a new specimen, with each individual
183 being used in a single assay. In this way, the time (in seconds) spent by the crustacean individual
184 in each of the three compartments of the choice flume (*i.e.* the downstream and the two
185 channels) was recorded.

186 Using the protocol described above, three experiments were conducted with a minimum
187 of 40 individuals per experiment and species, with the exception of *P. elegans*, for which only
188 25 individuals were tested per experiment. Moreover, these experiments were conducted both
189 in the morning (half of the total individuals; 7 h – 11 h UTC) and in the afternoon (half of the

190 total individuals; 14 h – 18 h UTC) in order to assess the daily variability of the crustaceans’
191 response to the seawater treatments. For each experiment, +CS and the two types of conditioned
192 seawater (SCS and SCE) were tested against CS. Because palaemonids were not available for
193 the first experiment (CS vs. +CS), only the two mysids were used. All four crustaceans were
194 used for the second (CS vs. SCS) and the third (CS vs. SCE) experiments.

195 The SCS was expected to contain the sponge community exometabolome (i.e. the
196 metabolites naturally released into the seawater by the four selected sponge species). The SCS
197 was obtained by maintaining living individuals of the four sponge species, measuring about
198 25 cm³ each, in a 10 L aerated-tank for 16 hours. The SCE was expected to contain the global
199 metabolome of the same sponge community, that is to say, all metabolites contained in the
200 tissues of the four selected sponges. This SCE was obtained from the sponges used to obtain
201 the SCS, which were weighed, frozen, ground in a blender, filtered through a 500 µm sieve and
202 then mixed with seawater in order to obtain a homogenous solution. The sponge community
203 used to prepare the SCS and SCE represented a total weight of 108 ± 10 g.

204 To ensure that the modifications of the seawater used to prepare the SCS and SCE was
205 due to the introduction of sponge exometabolomes or extracts only and not by a microbial
206 community present in the pumped open flow seawater, a further CS was prepared for the
207 metabolomic analyses by maintaining this open flow seawater for 16 hours in the same
208 condition as the SCS.

209

210 *Metabolomic analyses*

211 An untargeted metabolomic approach was used to compare the metabolite composition
212 of the different seawater treatments: the control seawater (CS), the cave seawater (+CS) and the
213 two conditioned seawater treatments (SCS and SCE). .

214 Briefly, a volume of 500 mL of seawater samples was collected in the tanks during the
215 two-flume choice experiments and passed successively through 0.45 μm pore-sized nylon
216 filters and octadecyl-bonded silica extraction discs (EmporeTM C₁₈ SPE discs) to obtain
217 particulate and dissolved phases of each seawater. The filters were extracted for particulate
218 phase (or eluted for dissolved phase) with methanol. The extracts were then concentrated to
219 dryness, treated to remove salts, dissolved in methanol and filtered at 0.2 μm before
220 metabolomic analysis. On-line Ultra High-Performance Liquid Chromatography coupled to a
221 High-Resolution Mass Spectrometer (UHPLCHRMS) analyses were performed using a Dionex
222 Ultimate 3000 system equipped with an autosampler and connected to a quadrupole Time of
223 Flight (qToF) mass spectrometer equipped with an electrospray ionization interface (Bruker
224 Impact II) to record mass spectra in the positive and negative modes (see Supplementary
225 Information for detailed methods).

226 Following the UHPLC-HRMS data acquisition, base peak chromatograms (BPC) were
227 exported as line spectra and converted into the netCDF file format to process unit mass
228 resolution data in centroid mode with XCMS (Smith et al. 2006) on R software (3.4.2). The
229 XCMS approach involved peak picking and integration, peak grouping for identification
230 between samples, chromatogram alignment to avoid retention time deviation and, finally, peak
231 filling to avoid missing values in chromatograms. XCMS generated a matrix
232 (ions/retention time \times sample) that was exported on Microsoft Excel. Home-made scripts were
233 used to clean the matrix, removing 1) ions detected in solvent blanks (ratio pools/blanks < 10),
234 2) ions presenting an intensity variation above 30% for pool samples, and 3) ions with correlated
235 intensities for pool samples (threshold = 0.8). Extracted Ion Chromatograms of variable
236 importance in projection (VIP) chemomarkers were drawn to measure their concentrations in
237 the different seawater treatments, taking into account the most important value of ion intensity
238 at each VIP retention time. The heatmap based on data acquired in positive mode was finally

239 selected to tentatively annotate chemomarkers of seawater samples using the molecular
240 formulae provided by Bruker Data-analysis 4.2.

241

242 *Statistical analyses*

243 Paired samples *t* tests (or Wilcoxon tests when normality and homoscedasticity
244 conditions were not met) were performed to compare the time spent in each of the two upstream
245 channels containing the different seawater treatments (CS vs. +CS; CS vs. SCS; CS vs. SCE),
246 in order to determine the channel chosen by each crustacean species. Two-sample *t* tests (or
247 Mann Whitney tests when normality and homoscedasticity conditions were not met) were
248 performed to compare the time spent in the channels filled with CS, +CS, SCS or SCE between
249 morning and afternoon, in order to determine if crustacean choice is affected by the time of day.
250 Principal Component Analyses (PCA) were performed to explore and identify similarities
251 between the chemical fingerprints of the different seawater samples. A permutation test (999
252 permutations, PPLS-DA models, based on a double cross-validation, outer loop = 4 fold cross-
253 validation, and inner loop = 3 fold cross-validation) was performed for the differentiation of the
254 chemical fingerprints of the different seawater samples followed by post hoc permutational
255 pairwise test (Hervé, 2018). The classification error rate (CER, 0 – 1) was calculated to estimate
256 the strength of the generated models. Kruskal Wallis tests, followed by a post-hoc multiple
257 range test (Fisher's Least Significant Difference), were performed to test differences in
258 chemomarker concentrations between seawater treatments.

259 Multivariate analyses (PCA) and heatmaps were performed with MetaboAnalyst 3.5
260 (Xia and Wishart 2016). Univariate analyses were performed with R software (version 3.4.2.)
261 and Statgraphics centurion (version 15.2.11.0).

262

263 **RESULTS**

264 ***Control experiment***

265 When both upstream channels were filled with CS, the four crustaceans showed no
266 significant preference between upstream channels (Supplementary Fig. S4) and remained
267 mainly in the downstream compartment (Supplementary Fig. S5). These results confirmed the
268 equivalence of the two channels for the assay. Moreover, both mysid species exhibited higher
269 locomotor activity than the two palaemonid species, staying twice as long in the upstream
270 channels (Supplementary Fig. S4).

271

272 ***Comparative crustacean responses to the seawater treatments***

273 During the 5-min tests of +CS vs. CS, most crustaceans exhibited stationary swimming
274 behaviour at the intersection of the seawater currents from the two upstream channels before
275 exhibiting their preference. However, occasionally after making their first choice, individuals
276 would switch and explore the opposite channel, showing constant activity while investigating
277 the seawater from both upstream channels. *Hemimysis margalefi* showed a clear preference for
278 the +CS, staying 16 times longer in this channel (Fig. 1) than in the CS. In contrast, *Leptomysis*
279 sp. demonstrated a significant preference for the CS channel, staying 4 times longer in this
280 channel (Fig. 1) than in the +CS. The behaviour of the two mysids varied with the time of day,
281 with *H. margalefi* having a higher preference for the +CS in the morning (Fig. 2). Interestingly,
282 *Leptomysis* sp. had a higher preference for the CS in the afternoon (Fig. 2).

283 In the SCS vs. CS experiment, *Hemimysis margalefi* exhibited a significant preference
284 for the SCS, staying 4 times longer in this channel (Fig. 1) than in the CS. This behaviour varied
285 with the time of day, with a higher preference for the SCS in the morning, when it spent 7 times
286 longer than in the CS channel, whereas this difference was only 2.5 times longer in the afternoon
287 (Fig. 3). *Palaemon serratus* also exhibited a preference for the SCS compared to the CS channel
288 (Fig. 1), but this attraction was less pronounced than for *H. margalefi* and was not affected by

289 the time of day (Fig. 3). Comparatively, *Leptomysis* sp. confirmed its clear preference for the
290 CS channel (Fig. 1) and, in this experiment too, their behaviour varied with the time of day,
291 with no clear preference in the morning, but with a 5-fold longer time spent in the CS channel
292 than in the SCS in the afternoon (Fig. 3). Finally, as was the case for *Leptomysis* sp., *P. elegans*
293 showed no preference in the morning but stayed 4 times longer in the CS than in the SCS
294 channel in the afternoon (Fig. 3).

295

296 In the SCE vs. CS experiment, both *H. margalefi* and *P. serratus* clearly avoided the
297 SCE channel and spent about 15 times longer in the CS channel compared to the SCE channel
298 (Fig. 1). This trend was less marked for *Leptomysis* sp. and *P. elegans*, as these crustaceans
299 stayed in the CS channel only 2 to 3 times longer than they did in the SCE channel (Fig. 1). The
300 two cave-dwelling crustaceans (i.e. *H. margalefi* and *P. serratus*) presented the same
301 behavioural response regardless of the time of day, whereas *Leptomysis* sp. and *P. elegans*
302 exhibited a significant avoidance of SCE in the afternoon only (Fig. 4). Moreover, the
303 swimming behaviour appeared quite different than in the previous experiments. *Hemimysis*
304 *margalefi* and *P. serratus* did not exhibit a stationary phase at the intersection of the seawater
305 currents, but they swam more quickly and presented a faster choice, mostly in favour of the CS
306 channel. In a distinct manner, *Leptomysis* sp. showed a reduced swimming activity and spent
307 most of its time on the bottom of the downstream compartment (Supplementary Fig. S5).

308

309 ***Metabolomic analyses***

310 The comparative untargeted analyses of the CS, +CS, SCS and SCE chemical
311 fingerprints were performed by UHPLC-HRMS in both negative and positive modes and for
312 both dissolved and particulate phases. After XCMS processing and filtering, 156 ions in the
313 negative mode and 144 ions were detected in the positive mode, and these were used to compare

314 the chemical fingerprints of the various seawater treatments (Fig. 5, Fig. S9). As a preliminary
315 result, no change could be detected when comparing the chemical fingerprints of the two CS,
316 i.e. the CS freshly pumped to perform the behavioural experiment or the CS kept for 16 hours
317 as was the conditioned seawater SCS.

318 The chemical fingerprints of the dissolved phases obtained in both negative and positive
319 modes resulted in the separation of +CS on one side and CS, SCS and SCE on the other side
320 along the first axis of the PCA (Fig. 5, Table S1). Along the second axis, the chemical
321 fingerprints recorded in the negative mode resulted in the grouping of SCS and SCE that
322 overlapped opposite to CS (Fig. 5), whereas in the positive mode, SCS grouped with CS
323 opposite to SCE (Fig. 5), but SCS and CS appeared different according to the post hoc
324 permutational pairwise test (Table S1). Chemical fingerprints of particulate phases obtained in
325 both negative and positive modes showed similar patterns with the grouping of CS with SCS
326 separated from both SCE and +CS (Fig. 5).

327 To highlight some chemical markers potentially responsible for the crustaceans'
328 behaviour, heatmaps of the most important features that discriminate groups in both modes and
329 phases were constructed (Fig. 6, and Supplementary Figs. S6, S7 and S8). Many features
330 present in SCE waters in high quantities may help to explain the crustaceans' repulsion. Among
331 them, the brominated derivatives known from *A. cavernicola* were easily identifiable on the
332 basis of their isotopic pattern, exact mass and in comparison with standards (Fig. S10). For
333 example, aeroplysinin-1 (m/z 335.8871, rt 400 s, M338T401 on Fig. S7) was identified mostly
334 in the dissolved phase and arothionin (m/z 812.8406, rt 470 s, M819T471 and M820T472 on
335 Fig. S8, and possible stereoisomer with the same isotopic pattern at rt 500: M817T501 and
336 M819T502, Figs. S8 and S10) was identified mostly in the particulate phase. On the contrary,
337 only a few chemical features were found to possibly explain the attraction of *H. margalefi* by

338 both +CS and SCS. The chemical fingerprints obtained in the negative mode from the dissolved
339 phases were the only ones separating CS from +CS and SCS/SCE (Fig. 5).

340 In the positive mode of the dissolved phase, two sets of features were tentatively
341 annotated. The 1st set of features grouped ions present in both +CS and SCE (Supplementary
342 Table S2, Figs. 6 and 7). Four metabolites, A (C₂₄H₄₄N₄O₄), B (C₃₀H₅₅N₅O₅), C (C₃₆H₆₆N₆O₆)
343 and D (C₄₂H₇₇N₇O₇), were detected in this group and are likely small peptides made of leucine
344 or iso-leucine units. Their concentrations were significantly higher in +CS, whereas the few
345 visible differences between SCS and CS were never statistically significant (Fig. 7). The 2nd set
346 of features grouped ions which were more concentrated in SCS and SCE, present in +CS and
347 overall poorly represented in CS (Supplementary Table S2, Figs. 6 and 7). They were
348 consistently significantly more concentrated in SCS than in other waters. However, although
349 some of them were twice as concentrated in +CS than in CS, these differences were generally
350 not supported statistically except for the metabolite L (C₁₇H₃₄N₂O₃) detected in higher
351 concentration in SCS and +CS compared to CS (Fig. 7). Molecular formulae of the marker
352 metabolites E (C₁₈H₃₆O₆), F (C₁₈H₃₀O₃), G (C₁₈H₃₄O₅) and H (C₁₄H₂₆O₃) were confidently
353 assigned thanks to the high accuracy of the mass and the consistency of the molecular formulae
354 provided by data analysis. These compounds likely belong to the oxylipin family, some
355 derivatives of a family previously found in the marine environment (Qiao et al. 2011; Lamari
356 et al. 2013; Lauritano et al. 2016). Halogenated metabolites harbouring the same number of
357 carbons were also detected, I (C₁₈H₃₁BrN₂O₅), J (C₁₈H₃₄Cl₂O₄) and K (C₁₈H₃₃ClO₄).

358

359 DISCUSSION

360 Distance interactions due to the putative role of chemical mediation in habitat detection
361 by fishes and crustaceans have been previously studied among benthic ecosystems dominated
362 by macroalgae (Lecchini et al. 2007, 2010, 2013; Lecchini et al. 2011) or seagrasses

363 (Nagelkerken et al. 2002; Arvedlund and Takemura 2006; Huijbers et al. 2008; Igulu et al.
364 2013), in coral reefs (Lecchini et al. 2005; Lecchini et al. 2007; Huijbers et al. 2008; Ben-Tzvi
365 et al. 2010; Lecchini et al. 2010; McCormick et al. 2010; Lecchini et al. 2011; Devine et al.
366 2012; Coppock et al. 2013; Igulu et al. 2013; Lecchini et al. 2013; Lecchini et al. 2014) and in
367 mangroves (Huijbers et al. 2008; Igulu et al. 2013). For some model interactions, such as clown
368 fishes and sea anemones, their description demonstrated how these processes are crucial for
369 ecosystem functioning, and how this mediation can be affected by global change (Arvedlund
370 and Nielsen 1996; Arvedlund et al. 1999; Dixson et al. 2008; Munday et al. 2009; Dixson 2012;
371 Dixson et al. 2014; Lecchini et al. 2014).

372 Here, we report for the first time clear behaviours of cave-dwelling crustaceans attracted
373 by cave seawater (+CS) and sponge conditioned seawater (SCS), and the concomitant presence
374 of chemomarkers in these waters. Sponges being dominant in benthic communities such as
375 underwater caves, we hypothesized that sponges and/or their associated microbiota may be
376 responsible for the production of chemical cues that significantly contribute to the network of
377 chemical interactions shaping the ecosystem. Conversely to previous studies in this field, we
378 anticipated that only part of the chemical diversity produced by these sessile invertebrates
379 would contribute to an overall chemical sea-scape conditioning the network of interactions.
380 Indeed, most previous studies on sponge chemical ecology used tissue homogenates or crude
381 extracts (*i.e.* the global metabolome) to highlight their deterrent properties against predators,
382 competitors for space or microorganisms (Bakus and Green 1974; Thompson et al. 1985;
383 Thacker et al. 1998; Kubanek et al. 2002; Pawlik et al. 2002; Pawlik 2011). Our experiment
384 exposing crustaceans to SCE confirmed the strong deterrent effect of seawater containing the
385 entire sponge metabolome. Most likely, deterrent or toxic properties of specialized metabolites
386 present in the sponges are responsible for these behavioural responses. In line with this
387 hypothesis, aerothionin, which is one of the discriminating compounds of the SCE, and other

388 compounds of the same family had already been identified as feeding deterrents in other
389 bioassays and may be responsible for the observed repulsive effects (Thoms et al. 2004;
390 Gochfeld et al. 2012). Interestingly, none of the main specialized metabolites previous known
391 in the four sponges tested were detected in the SCS or the +CS, so they do not appear to belong
392 to what can be called the sponge exometabolome and, thus, do not contribute to the chemical
393 seascape. So far, the exudation of major compounds has been demonstrated in the single case
394 of the sponge *Crambe crambe* (Ternon et al. 2016), but there is crucial need for new
395 experiments to better distinguish between the global metabolome, which is usually extracted
396 from organisms using solvent, and the exometabolome which is the part naturally released by
397 an organism into the surrounding water.

398 Following previous hypotheses by Bingham and Young (1991), our study demonstrates
399 for the first time that seawater conditioned by a sponge assemblage (SCS), potentially
400 containing components of the exometabolome only, exerts an attractant effect on some mobile
401 invertebrate species, leading to a specific behavioural response related to their ecology.
402 Demonstrating such a “positive” effect of sponge compounds would have been impossible with
403 only the rather traditional use of crude extracts or pure major compounds in our experimental
404 plan. This past strategy explains the important knowledge gap that exists concerning processes
405 of chemical mediation within marine benthic ecosystems, in particular where ecosystems are
406 dominated by sponges. Few studies have attempted to demonstrate the putative release of
407 metabolites into the surrounding medium (Walker et al. 1985; Porter and Targett 1988; Duque
408 et al. 2001; Ternon et al. 2016). After characterizing the exudation of chemical cues by the
409 sponge *Crambe crambe*, and assessing the toxic activity of these compounds on invertebrate
410 larvae, Ternon et al. (2016) hypothesized that this process might explain the ecological success
411 of this encrusting sponge. Here, we provide the first demonstration that some sponge
412 metabolites may also be involved in distance interactions, acting as kairomones to shape the

413 behaviour of mobile organisms and, thus, structuring biodiversity. We thus confirm the
414 hypothesis of [Bingham and Young \(1991\)](#) that chemicals released by sponges have the potential
415 to attract organisms. Moreover, we have demonstrated a species-specific response to
416 conditioned seawater, since *H. margalefi* and *P. serratus* only showed a significant attraction
417 for the seawater containing the exometabolome of the sponge community. The opposite
418 response of *Leptomysis* sp. can be explained by the fact that the CS was pumped from heart to
419 the *Leptomysis* sp. Habitat, which is dominated by macrophytes and macroalgae and where
420 sponges are poorly distributed.

421 The responses of our cave-dwelling species support our hypothesis that chemical cues
422 released by the sponges located at the entrance of underwater caves might be a key factor in the
423 homing behaviour of *H. margalefi* and a refuge signal for *P. serratus*. The difference recorded
424 between their behavioural responses is likely related to their life habits. *Hemimysis margalefi*
425 is a cave-dweller living in swarms and is thought to migrate out of the cave for night feeding
426 ([Rastorgueff et al. 2011](#); [Rastorgueff et al. 2015](#)). This circadian movement is presumed to be
427 linked to the decrease in light intensity ([Passelaigue and Bourdillon 1986](#)), at least to explain
428 how these mysids get out of the cave at the end of the day. However, our goal was to provide
429 clues to support the idea that the return to the cave in the night or early morning may be related
430 to the chemical seascape of the cave, through chemical cues potentially produced by sponges
431 guiding these mysids to the entrance of their habitat. We acknowledge that we only conducted
432 our experiments in the morning, and future experiments could be performed in darkness to
433 mimic if the same trend is observed during the night. Interestingly, *H. margalefi* exhibited the
434 same positive attraction when faced with the choice of the natural cave seawater and the sponge
435 conditioned seawater containing exometabolomes. A better understanding of these migrations
436 of cave-dwelling organisms is crucial as they represent important vectors of organic matter
437 transfer from the outside euphotic littoral zone to various locations inside caves, where this

438 matter becomes available to other cave dwellers such as molluscs, crustaceans and teleost fishes
439 ([Rastorgueff et al. 2011](#)). There is likely a strong link between this type of migration pattern,
440 and how it mitigates cave oligotrophy, and the diversity of sessile organisms at the origin of the
441 chemical seascape.

442 *Palaemon serratus* is a crustacean which may occasionally inhabit underwater caves. In
443 this case too, we hypothesize that the sponge seascape might influence the swimming behaviour
444 of this species and contribute to locate its habitat refuge. Interestingly, *P. serratus* spent most
445 of its time in the downstream compartment and exhibited a much less pronounced attraction for
446 the sponge conditioned seawater, compared to *H. margalefi*. In the case of palaemonids, the
447 process of chemical mediation in habitat detection might not be related to gradients of
448 waterborne molecules only. Whereas mysids are swimming crustaceans, palaemonids swim
449 close to and walk on the bottom using their well-developed legs. They continuously explore the
450 bottom, sensing more hydrophobic molecules with leg chemoreceptors. They also use pincers
451 to bring fragments of substrate into contact with chemosensory mouthparts in order to follow
452 hydrophobic cues adherent to the substrate ([Mollo et al. 2014](#)).

453 Variations of the behavioural responses with time of day differed among the species
454 tested. The cave-dwelling mysid seemed to be much more attracted by sponge cues in the
455 mornings, whereas the palaemonid showed no variation, which tends to confirm that their
456 response does not rely on the same ecological process. Daily rhythm of chemosensitivity has
457 already been reported for several arthropods, such as the Tsetse fly *Glossina morsitans*
458 *morsitans* which presents a higher chemosensitivity in early morning and late afternoon when
459 its feeding activity is the greatest ([Van der Goes van Naters et al. 1998](#)). The recent application
460 of underwater electroantennography ([Machon et al. 2016](#)) offers new perspectives of
461 chemosensitivity surveys of our model crustaceans that face various chemical cues at different
462 period of the day. Moreover, the behavioural responses might also be correlated to the

463 expression level of some chemo-receptor such as IR25a ([Zbinden et al. 2017](#)) in order for the
464 crustacean antennules to perceive chemical cues. For technical reason, most of our morning
465 behavioural experiments could only be conducted during daylight hours and we believe that the
466 combination of additional experiments performed in the darkness combined to chemosensitivity
467 surveys might further support our hypothesis on the role of the chemical seascape on diurnal
468 migrations of cave-dwelling crustaceans.

469 Very few studies in chemistry have attempted to identify the structure of the
470 hypothesized marine chemical cues involved in habitat selection ([Lecchini et al. 2005](#); [Derby
471 and Sorensen 2008](#); [Lecchini et al. 2010](#); [Lecchini et al. 2014](#)). Even though we expected to
472 detect some of the previously known compounds of the four sponge models in the SCS, our
473 metabolomics approach shows that none of the specialized metabolites reported so far from the
474 global metabolome of our sponge models could be found in the SCS. Based on their molecular
475 formulae as obtained unambiguously by HRMS, the chemomarkers present in the sponge
476 conditioned seawater, and at low concentration in the cave seawater, correspond to C18-based
477 oxylipins. Further confirmation of these structures is needed, however, using available
478 standards for example. This hypothesis seems realistic as diatom oxylipins are known for their
479 multiple functions, such as sexual pheromones or food-finding cues detectable by grazers ([Fink
480 et al. 2006](#); [Kâ et al. 2014](#)), but also as chemical-alarm cues when facing grazing pressure
481 ([Ianora et al. 2004](#); [Pohnert 2005](#)). Thus, the ecological function of oxylipins may be very
482 broad. The putative oxylipins may be produced by any of the sponge holobionts ([McClintock
483 et al. 2005](#)) and they may be responsible for the homing behaviour of crustaceans. The four
484 peptides naturally found in high concentrations in the cave seawater are also likely produced
485 by sponge holobionts, although in this case their very low concentrations in the sponge
486 conditioned seawater do not provide information on their putative function in the cave chemical
487 seascape.

488 In addition to the organic compounds in the seawater which are detectable by our
489 metabolomics approach, we assume that inorganic compounds may also influence the
490 biological responses of our model organisms. A significant difference in nutrient concentrations
491 between seawater treatments would only result from the fluxes generated by the sponge
492 holobiont catabolism (for a review see [Maldonado et al. 2012](#)), releasing ammonia, nitrate or
493 phosphate for instance during the water conditioning. However, as the sponge volume used in
494 our experiments represented only 1% of the aquarium volume, we can reasonably consider that
495 there was a very low probability of a significant change in nutrient concentration during the
496 preparation of our seawater treatments.

497 Finally, elucidating the mechanisms and strategies used in habitat detection is crucial to
498 predict how individual species or assemblages may respond to the degradation of their
499 environment ([Swihart et al. 2003](#); [Jones et al. 2004](#)). Changes affecting chemical mediation
500 processes are likely to impact subsequent distribution of marine organisms. Sponges are known
501 to be sensitive to global changes, and most of our study species have already suffered mass
502 mortalities related to extreme climate events ([Lejeusne et al. 2010](#)). In a context of increasing
503 frequency of such events ([Giorgi and Lionello 2008](#)), we can expect further alteration of sponge
504 communities to impact distance interactions mediated by sponge chemical cues. Underwater
505 caves are fragmented habitats and their biodiversity patterns are greatly shaped by their
506 connectivity. Therefore, any alteration of the chemical seascape within and surrounding marine
507 caves, directly or indirectly caused by human activities, could have severe consequences on
508 these ecosystems. As has been evidenced in previous studies by [Dixon et al. \(2008, 2011\)](#),
509 connectivity between distinct marine habitats may partly rely on chemical mediations, and not
510 only on currents.

511

512 **AUTHOR CONTRIBUTION STATEMENT**

513 MS and TP designed the experiments. MS and MLC performed the experiments. MS,
514 SG, MLC and OPT analysed the data. MS, SG, MLC, OPT and TP wrote the manuscript.

515

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527

528 **CONFLICT OF INTEREST**

529 The authors declare that they have no conflict of interest.

530

531 **ETHICAL APPROVAL**

532 All applicable international, national, and/or institutional guidelines for the care and use
533 of animals were followed.

534

535 **REFERENCES**

536 Amsler CD (2008) Algal Chemical Ecology. Springer, Berlin

537 Arvedlund M, Nielsen LE (1996) Do the anemonefish *Amphiprion ocellaris* (Pisces:
538 Pomacentridae) imprint themselves to their host sea anemone *Heteractis magnifica*
539 (Anthozoa: Actinidae)? *Ethology* 102:197-211. Doi: 10.1111/j.1439-0310.1996.tb01118.x

540 Arvedlund M, McCormick MI, Fautin DG, Bildsoe M (1999) Host recognition and possible
541 imprinting in the anemonefish *Amphiprion melanopus* (Pisces: Pomacentridae). *Mar Ecol*
542 *Prog Ser* 188:207-218. Doi: 10.3354/meps188207

543 Arvedlund M, Takemura A (2006) The importance of chemical environmental cues for juvenile
544 *Lethrinus nebulosus* Forsskal (Lethrinidae, Teleostei) when settling into their first benthic
545 habitat. *J Exp Mar Biol Ecol* 338:112-122. Doi: 10.1016/j.jembe.2006.07.001

546 Atema J, Kingsford MJ, Gerlach G (2002) Larval reef fish could use odour for detection,
547 retention and orientation to reefs. *Mar Ecol Prog Ser* 241:151-60. Doi: 10.3354/meps241151

548 Bakus GJ, Green G (1974) Toxicity in sponges and holothurians: a geographic pattern. *Science*
549 185:951-953. Doi: 10.1126/science.185.4155.951

550 Belanger RM, Corkum LD, Li WM, Zielinski BS (2006) Olfactory sensory input increases gill
551 ventilation in male round gobies (*Neogobius melanostomus*) during exposure to steroids.
552 *Comp Biochem Physiol* 144:196-202. Doi: 10.1016/j.cbpa.2006.02.027

553 Ben-Tzvi O, Tchernov D, Kiflawi M (2010) Role of coral-derived chemical cues in
554 microhabitat selection by settling *Chromis viridis*. *Mar Ecol Prog Ser* 400:181-187. Doi:
555 10.3354/meps08627

556 Bingham BL, Young CM (1991) Influence of sponges on invertebrate recruitment: a field test
557 of allelopathy. *Mar Biol* 109:19-26. Doi: 10.1007/BF01320227

558 Bussotti S, Terlizzi A, Frascchetti S, Belmonte G, Boero F (2006) Spatial and temporal
559 variability of sessile benthos in shallow Mediterranean marine caves. *Mar Ecol Prog*
560 *Ser* 325:109-119. Doi: 10.3354/meps325109

561 Cassier P, Bohatier J, Descoins C, Nagnan Le Meillour P (2000) Communication chimique et
562 environnement. Belin, Paris

563 Coppock AG, Gardiner NM, Jones GP (2013) Olfactory discrimination in juvenile coral reef
564 fishes: response to conspecifics and corals. *J Exp Mar Biol Ecol* 443:21-26. Doi:
565 10.1016/j.jembe.2013.02.026

566 Derby CD, Sorensen PW (2008) Neural Processing, Perception, and Behavioral Responses to
567 Natural Chemical Stimuli by Fish and Crustaceans. *J Chem Ecol* 34:898-914. Doi:
568 10.1007/s10886-008-9489-0

569 Devine BM, Munday PL, Jones GP (2012) Homing ability of adult cardinalfish is affected by
570 elevated carbon dioxide. *Oecologia* 168:269-276. Doi: 10.1007/s00442-011-2081-2

571 Diaz MC, Rutzler K (2001) Sponges: an essential component of Caribbean coral reefs. *Bull*
572 *Mar Sci* 69:535-546

573 Dixson DL, Jones GP, Munday PL, Planes S, Pratchett MS, Srinivasan M, Syms C, Thorrold
574 SR (2008) Coral reef fish smell leaves to find island homes. *Proc R Soc Lond B* 275:2831-
575 2839. Doi: 10.1098/rspb.2008.0876

576 Dixson DL (2012) Predation risk assessment by larval reef fishes during settlement-site
577 selection. *Coral Reefs* 31:255-261. Doi: 10.1007/s00338-011-0842-3

578 Dixson DL, Abrego D, Hay ME (2014) Chemically mediated behavior of recruiting corals and
579 fishes: a tipping point that may limit reef recovery. *Science* 345:892-897. Doi:
580 10.1126/science.1255057

581 Dixson DL, Jones GP, Munday PL, Pratchett MS, Srinivasan M, Planes S, Thorrold SR (2011)
582 Terrestrial chemical cues help coral reef fish larvae locate settlement habitat surrounding
583 islands. *Ecol Evol* 1:586-595. Doi: 10.1002/ece3.53

584 Duque C, Bonilla A, Bautista E, Zea S (2001) Exudation of low molecular weight compounds
585 (thiobismethane, methyl isocyanide, and methyl isothiocyanate) as a possible chemical
586 defense mechanism in the marine sponge *Ircinia felix*. *Biochem Syst Ecol* 29:459-467

587 Fink P, Von Elert E, Jüttner F (2006) Oxylipins from freshwater diatoms act as attractants for
588 a benthic herbivore. *Archiv für Hydrobiol* 167:561-574. Doi: 10.1127/0003-
589 9136/2006/0167-0561

590 Fisher HS, Wong BBM, Rosenthal GG (2006) Alteration of the chemical environment disrupts
591 communication in a freshwater fish. *Proc R Soc Lond B* 273:1187-1193. Doi:
592 10.1098/rspb.2005.3406

593 Garrido L, Zubía E, Ortega MJ, Salvá J (1997) New furanoterpenoids from the sponge *Spongia*
594 *officinalis*. *J Nat Prod* 60:794-797. Doi: 10.1021/np970160x

595 Gerlach G, Atema J, Kingsford MJ, Black KP, Miller-Sims V (2007) Smelling home can
596 prevent dispersal of reef fish larvae. *Proc Natl Acad Sci USA* 104:858-63. Doi:
597 10.1073/pnas.0606777104

598 Gerovasileiou V, Voultziadou E (2012) Marine caves of the Mediterranean sea: a sponge
599 biodiversity reservoir within a biodiversity hotspot. *Plos One* 7:e39873. Doi:
600 10.1371/journal.pone.0039873

601 Giorgi F, Lionello P (2008) Climate change projections for the Mediterranean region. *Global*
602 *Planet Change* 63:90-104. Doi: 10.1016/j.gloplacha.2007.09.005

603 Gochfeld DJ, Easson CG, Freeman CJ, Thacker RW, Olson JB (2012) Disease and nutrient
604 enrichment as potential stressors on the Caribbean sponge *Aplysina cauliformis* and its
605 bacterial symbionts. *Mar Ecol Prog Ser* 456:101-111.

606 Gunn J, Hardwick P, Wood PJ (2000) The invertebrate community of the Peak-Speedwell cave
607 system, Derbyshire, England - pressures and considerations for conservation management.

608 Aquat Conserv: Mar Fresh Ecosys 10:353-369. Doi: 10.1002/1099-
609 0755(200009/10)10:5<353::AID-AQC413>3.0.CO;2-S

610 Hay ME (2009) Marine chemical ecology: chemical signals and cues structure marine
611 populations, communities, and ecosystems. Annu Rev Mar Sci 1:193-212. Doi:
612 10.1146/annurev.marine.010908.163708

613 Herve, M. (2018). RVAideMemoire: testing and plotting procedures for biostatistics. R
614 Package (03-01-2018).

615 Heuschele J, Selander E (2014) The chemical ecology of copepods. J Plankton Res 36:895-913.
616 Doi: 10.1093/plankt/fbu025

617 Huijbers CM, Mollee EM, Nagelkerken I (2008) Post-larval French grunts (*Haemulon*
618 *flavolineatum*) distinguish between seagrass, mangrove and coral reef water: Implications
619 for recognition of potential nursery habitats. J Exp Mar Biol Ecol 357:134-139. Doi:
620 10.1016/j.jembe.2008.01.004

621 Ianora A, Miralto A, Poulet SA, Carotenuto Y, Buttino I, Romano G, Casotti R, Pohnert G,
622 Wichard T, Collucci-D'Amato L, Terrazzano G, Smetacek V (2004) Aldehyde suppression
623 of copepod recruitment in blooms of a ubiquitous planktonic diatom. Nature 429:403-407.
624 Doi: 10.1038/nature02526

625 Igulu MM, Nagelkerken I, van der Beek M, Schippers M, van Eck R, Mgya YD (2013)
626 Orientation from open water to settlement habitats by coral reef fish: behavioral flexibility
627 in the use of multiple reliable cues. Mar Ecol Prog Ser 493:243-257. Doi:
628 10.3354/meps10542

629 Ivanisevic J, Thomas OP, Lejeusne C, Chevaldonné P, Pérez T (2011) Metabolic fingerprinting
630 as an indicator of biodiversity: towards understanding inter-specific relationships among
631 Homoscleromorpha sponges. Metabolomics 7:289-304. Doi: 10.1007/s11306-010-0239-2

632 Jones GP, McCormick MI, Srinivasan M, Eagle JV (2004) Coral decline threatens biodiversity
633 in marine reserves. Proc Natl Acad Sci USA 101:8251-8253. Doi: 10.1073/pnas.0401277101

634 Kâ S, Carotenuto Y, Romano G, Hwang JS, Buttino I, Ianora A (2014) Impact of the diatom-
635 derived polyunsaturated aldehyde 2-trans,4-trans decadienal on the feeding, survivorship
636 and reproductive success of the calanoid copepod *Temora stylifera*. Mar Environ Res 93:31-
637 37. Doi: 10.1016/j.marenvres.2013.07.009

638 Kubanek J, Whalen KE, Engel S, Kelly SR, Henkel TP, Fenical W, Pawlik JR (2002) Multiple
639 defensive roles for triterpene glycosides from two Caribbean sponges. Oecologia 131:125-
640 136. Doi: 10.1007/s00442-001-0853-9

641 Lamari N, Ruggiero MV, d'Ippolito, G, Kooistra WH, Fontana A, Montresor M (2013)
642 Specificity of lipoxygenase pathways supports species delineation in the marine diatom
643 genus *Pseudo-nitzschia*. Plos One 8:e73281. Doi: 10.1371/journal.pone.0073281

644 Lauritano C, Romano G, Roncalli V, Amoresano A, Fontanarosa C, Bastianini M, Braga F,
645 Carotenuto Y, Ianora A (2016) New oxylipins produced at the end of a diatom bloom and
646 their effects on copepod reproductive success and gene expression levels. Harmful Algae
647 55:221-229. Doi: 10.1016/j.hal.2016.03.015

648 Lecchini D, Shima J, Banaigs B, Galzin R (2005) Larval sensory abilities and mechanisms of
649 habitat selection of a coral reef fish during settlement. Oecologia 143:326-334. Doi:
650 10.1007/s00442-004-1805-y

651 Lecchini D, Osenberg CW, Shima JS, St Marcy CM, Galzin R (2007) Ontogenetic changes in
652 habitat selection during settlement in a coral reef fish ecological determinants and sensory
653 mechanisms. Coral Reefs 26:423-432. Doi: 10.1007/s00338-007-0212-3

654 Lecchini D, Mills SC, Brie C, Maurin R, Banaigs B (2010) Ecological determinants and sensory
655 mechanisms in habitat selection of crustacean postlarvae. Behavior Ecol 21:599-607. Doi:
656 10.1093/beheco/arq029

657 Lecchini D (2011) Visual and chemical cues in habitat selection of sepioid larvae. C R Biol
658 334:911-915

659 Lecchini D, Waqalevu VP, Parmentier E, Radford CA, Banaigs B (2013) Fish larvae prefer
660 coral over algal water cues: implications of coral reef degradation. Mar Ecol Prog Ser
661 475:303-307. Doi: 10.3354/meps10094

662 Lecchini D, Miura T, Lecellier G, Banaigs B, Nakamura Y (2014) Transmission distance of
663 chemical cues from coral habitats: implications for marine larval settlement in context of
664 reef degradation. Mar Biol 161:1677-1686. Doi: 10.1007/s00227-014-2451-5

665 Leduc AOHC, Munday PL, Brown GE, Ferrari MCO (2013) Effects of acidification on
666 olfactory-mediated behaviour in freshwater and marine ecosystems: a synthesis. Philos
667 Trans R Soc B 368:1-14. Doi: 10.1098/rstb.2012.0447

668 Lejeusne C, Chevaldonné P, Pergent-Martini C, Boudouresque CF, Pérez T (2010) Climate
669 change effects on a miniature ocean: the highly diverse, highly impacted Mediterranean Sea.
670 Trends Ecol Evol 25:250-260. Doi: 10.1016/j.tree.2009.10.009

671 Machon J, Ravaux J, Zbinden M, Lucas P (2016) New electroantennography method on a
672 marine shrimp in water. J Exp Biol 219:3696-3700. Doi: 10.1242/jeb.140947

673 Maldonado M, Ribes M, van Duyl FC (2012) Nutrient fluxes through sponges: biology,
674 budgets, and ecological implications. Adv Mar Biol 62:113-182.

675 Manzo E, Ciavatta ML, Villani G, Varcamonti M, Sayem SA, van Soest R, Gavagnin M (2011)
676 Bioactive terpenes from *Spongia officinalis*. J Nat Prod 74:1241-1247. Doi:
677 10.1021/np200226u

678 McClintock JB, Amsler CD, Baker BJ, Van Soest RW (2005) Ecology of Antarctic marine
679 sponges: an overview. Integr Comp Biol 45:359-368. Doi: 10.1093/icb/45.2.359

680 McCormick MI, Moore JAY, Munday PL (2010) Influence of habitat degradation on fish
681 replenishment. Coral Reefs 29:537-546. Doi: 10.1007/s00338-010-0620-7

682 Mollo E, Fontana A, Roussis V, Polese G, Amodeo P, Ghiselin MT (2014) Sensing marine
683 biomolecules: smell, taste, and the evolutionary transition from aquatic to terrestrial life.
684 Front Chem 2:92. Doi: 10.3389/fchem.2014.00092

685 Munday PL, Dixson DL, Donelson JM, Jones GP, Pratchett MS, Devitsina GV, Doving KB
686 (2009) Ocean acidification impairs olfactory discrimination and homing ability of a marine
687 fish. Proc Natl Acad Sci USA 106:1848-1852. Doi: 10.1073/pnas.0809996106

688 Myrberg AA, Fuiman LA (2002) The sensory world of coral reef fishes. In: Sale PF (ed) Coral
689 reef fishes: dynamics and diversity in a complex ecosystem, Academic Press, San Diego, pp
690 123-148

691 Nagelkerken I, Roberts CM, Van Der Velde G, Dorenbosch M, Van Riel MC, De La Moriniere
692 EC, Nienhuis PH (2002) How important are mangroves and seagrass beds for coral-reef fish?
693 The nursery hypothesis tested on an island scale. Mar Ecol Prog Ser 244:299-305. Doi:
694 10.3354/meps244299

695 Nuzzo G, Ciavatta ML, Villani G, Manzo E, Zanfardino A, Varcamonti M, Gavagnin M (2012)
696 Fulvynes, antimicrobial polyoxygenated acetylenes from the Mediterranean sponge
697 *Haliclona fulva*. Tetrahedron 68:754-760. Doi: 10.1016/j.tet.2011.10.068

698 O'Connor JJ, Lecchini D, Beck HJ, Cadiou G, Lecellier G, Booth DJ, Nakamura Y (2016)
699 Sediment pollution impacts sensory ability and performance of settling coral-reef fish.
700 Oecologia 180:11-21. Doi: 10.1007/s00442-015-3367-6

701 Passelaigue F, Bourdillon A (1986) Distribution and circadian migrations of the cavernicolous
702 mysid *Hemimysis speluncola* Ledoyer. Stygologia 2:112-118

703 Paul VJ, Ritson-Williams R, Sharp K (2011) Marine chemical ecology in benthic environments.
704 Nat Prod Rep 28:345-87. Doi: 10.1039/c4np00017j

705 Pawlik JR, Fall GMC, Zea S (2002) Does the odor from sponges of the genus *Ircinia* protect
706 them from fish predators? J Chem Ecol 28:1103-1115. Doi: 10.1023/A:1016221415028

707 Pawlik JR (2011) The chemical ecology of sponges on Caribbean reefs: natural products shape
708 natural systems. *Bioscience* 61:888-898. Doi: 10.1525/bio.2011.61.11.8

709 Pohnert G (2005) Diatom/copepod interactions in plankton: the indirect chemical defense of
710 unicellular algae. *Chem Bio Chem* 6:946-959. Doi: 10.1002/cbic.200400348

711 Qiao MF, Ji NY, Miao FP, Yin XL (2011) Steroids and an oxylipin from an algicolous isolate
712 of *Aspergillus flavus*. *Magn Res Chem* 49:366-369. Doi: 10.1002/mrc.2748

713 Rasher DB, Stout EP, Engel S, Shearer TL, Kubanek J, Hay ME (2015) Marine and terrestrial
714 herbivores display convergent chemical ecology despite 400 million years of independent
715 evolution. *Proc Natl Acad Sci USA* 112:12110-12115. Doi: 10.1073/pnas.1508133112

716 Rastorgueff PA, Harmelin-Vivien M, Richard P, Chevaldonné P (2011) Feeding strategies and
717 resource partitioning mitigate the effects of oligotrophy for marine cave mysids. *Mar Ecol*
718 *Prog Ser* 440:163-176. Doi: 10.3354/meps09347

719 Rastorgueff PA, Bellan-Santini D, Bianchi CN, Bussotti S, Chevaldonné P, Guidetti P, ...
720 Ruitton S (2015) An ecosystem-based approach to evaluate the ecological quality of
721 Mediterranean undersea caves. *Ecol Indic* 54:137-152.

722 Reverter M, Pérez T, Ereskovsky AV, Banaigs B (2016) Secondary Metabolome Variability
723 and Inducible Chemical Defenses in the Mediterranean Sponge *Aplysina cavernicola*. *J*
724 *Chem Ecol* 42:60-70. Doi: 10.1007/s10886-015-0664-9

725 Smith CA, Want EJ, O'Maille G, Abagyan R, Siuzdak G (2006) XCMS: Processing mass
726 spectrometry data for metabolite profiling using nonlinear peak alignment, matching, and
727 identification. *Anal Chem* 78:779-787. Doi: 10.1021/ac051437y

728 Swihart RK, Gehring TM, Kolozsvary MB, Nupp TE (2003) Responses of 'resistant'
729 vertebrates to habitat loss and fragmentation: the importance of niche breadth and range
730 boundaries. *Divers Distrib* 9:1-18. Doi: 10.1046/j.1472-4642.2003.00158.x

731 Ternon E, Zarate L, Chenesseau S, Croué J, Dumollard R, Suzuki MT, Thomas OP (2016)
732 Spherulization as a process for the exudation of chemical cues by the encrusting sponge *C.*
733 *crambe*. Sci Rep 6:29474. Doi: 10.1038/srep29474

734 Thacker RW, Becerro MA, Lumbang WA, Paul VJ (1998) Allelopathic interactions between
735 sponges on a tropical reef. Ecology 79:1740-1750. Doi: 10.1890/0012-
736 9658(1998)079[1740:AIBSOA]2.0.CO;2

737 Thompson JE (1985) Exudation of biologically-active metabolites in the sponge *Aplysina*
738 *fistularis*. I. Biological evidence. Mar Biol 88:23-26. Doi: 10.1007/BF00393040

739 Thompson JE, Walker RP, Faulkner DJ (1985) Screening and bioassays for biologically-active
740 substances from forty marine sponge species from San Diego, California, USA. Mar Biol
741 88:11-21. Doi: 10.1007/BF00393038

742 Thoms C, Wolff M, Padmakumar K, Ebel R, Proksch P (2004) Chemical defense of
743 Mediterranean sponges *Aplysina cavernicola* and *Aplysina aerophoba*. Z Naturforsch C
744 59:113-122

745 van de Koppel J, van der Heide T, Altieri AH, Eriksson BK, Bouma TJ, Olf H, Silliman BR
746 (2015) Long-distance interactions regulate the structure and resilience of coastal ecosystems.
747 Annu Rev Mar Sci 7:139-158. Doi: 10.1146/annurev-marine-010814-015805

748 Van der Goes van Naters WM, Den Otter CJ, Maes FW (1998) Olfactory sensitivity in tsetse
749 flies: a daily rhythm. Chem Senses 23:351-357

750 Walker RP, Thompson JE, Faulkner DJ (1985) Exudation of biologically-active metabolites in
751 the sponge *Aplysina fistularis*. II. Chemical evidence. Mar Biol 88:27-32. Doi:
752 10.1007/BF00393040

753 Wulff J (2012) Ecological Interactions and the Distribution, Abundance, and Diversity of
754 Sponges. Adv Mar Biol 6:273-344. Doi: 10.1016/B978-0-12-387787-1.00003-9

755 Xia J, Wishart DS (2016) Using MetaboAnalyst 3.0 for Comprehensive Metabolomics Data
756 Analysis. *Curr Protoc Bioinform* 55:14.10.1-14.10.91. Doi: 10.1002/cpbi.11
757 Zbinden M, Berthod C, Montagné N, Machon J, Léger N, Chertemps T, ... Ravaux J (2017)
758 Comparative Study of Chemosensory Organs of Shrimp From Hydrothermal Vent and
759 Coastal Environments. *Chem Senses* 42:319-331. Doi: 10.1093/chemse/bjx007
760

761 **Figure legends**

762

763

764 **Fig. 1** Response of the four crustaceans to: (a) underwater cave seawater (+CS) vs. control
765 seawater (CS), (b) sponge conditioned seawater (SCS) vs. CS, and (c) sponge chemical extracts
766 dissolved in seawater (SCE) vs. CS. Results are mean time (\pm SE) in seconds spent by each
767 species in the channel filled with water containing chemical cues (grey bar) and the CS channel
768 (white bar). Paired samples *t* tests (or Wilcoxon tests) were performed to compare the time
769 spent in each of the two upstream channels. Significant differences are indicated with * for $P <$
770 0.05, ** for $P < 0.01$, and *** for $P < 0.001$. HM = *Hemimysis margalefi*, LS = *Leptomysis* sp.,
771 PE = *Palaemon elegans*, PS = *Palaemon serratus*.

772

773 **Fig. 2** Response of *Hemimysis margalefi* and *Leptomysis* sp. to the underwater cave seawater
774 (+CS) vs. control seawater (CS) during the morning and the afternoon. Results are mean time
775 (\pm SE) in seconds spent by each species in the +CS (grey bar) and the CS (white bar) channels.
776 Paired samples *t* tests (or Wilcoxon tests) were performed to compare the time spent in each of
777 the two upstream channels. Two-sample *t* tests (or Mann-Whitney tests) were performed to
778 compare the time spent in the channels between morning and afternoon. Significant differences
779 are indicated with ** for $P < 0.01$, and *** for $P < 0.001$.

780

781 **Fig. 3** Response of the four crustaceans to sponge conditioned seawater (SCS) vs. control
782 seawater (CS) during the morning and the afternoon. Results are mean time (\pm SE) in seconds
783 spent by each species in the SCS (grey bar) and the CS (white bar) channels. Paired samples *t*-
784 tests (or Wilcoxon tests) were performed to compare the time spent in each of the two upstream
785 channels. Two-sample *t* tests (or Mann Whitney tests) were performed to compare the time

786 spent in the channels between morning and afternoon. Significant differences are indicated with
787 the respective symbols * for $P < 0.05$, ** for $P < 0.01$, and *** for $P < 0.001$.

788

789 **Fig. 4** Response of the four crustaceans to sponge chemical extracts dissolved in seawater
790 (SCE) vs. control seawater (CS) during the morning and the afternoon. Results are mean time
791 (\pm SE) in seconds spent by each species in the SCE (grey bar) and the CS (white bar) channels.
792 Paired samples t tests (or Wilcoxon tests) were performed to compare the time spent in each of
793 the two upstream channels. Two-sample t tests (or Mann Whitney tests) were performed to
794 compare the time spent in the channels between morning and afternoon. Significant differences
795 are indicated with * for $P < 0.05$, and *** for $P < 0.001$.

796

797 **Fig. 5** Principal Component Analysis (PCA) based on the chemical features of the different
798 seawater samples analysed by UHPLC-MS in negative (a, c) and positive (b, d) modes, for the
799 dissolved (a, b) and particulate (c, d) phases. A permanova test (PPLS-DA models, 999
800 permutations with double-cross validation) was performed to test differentiation between
801 chemical fingerprints (post hoc permutation pairwise test in Supplementary file S). The
802 classification error rate (CER, 0 – 1) was calculated to estimate the strength of the generated
803 models. CS: control seawater; +CS: cave seawater; SCS: sponge conditioned seawater; SCE:
804 sponge chemical extract; p: particulate phase; d: dissolved phase.

805

806 **Fig. 6** Heatmap based on the variable importance in projection of the features ($VIP > 1$) detected
807 in the metabolic fingerprints of the dissolved phase of the seawater samples analysed by liquid
808 chromatography coupled to mass spectrometry in positive mode. X-axis displays the different
809 seawater treatments, Y-axis displays metabolite features coupling nominal mass (M as atomic
810 mass unit) with chromatographic retention time (T in seconds). Connections of samples and

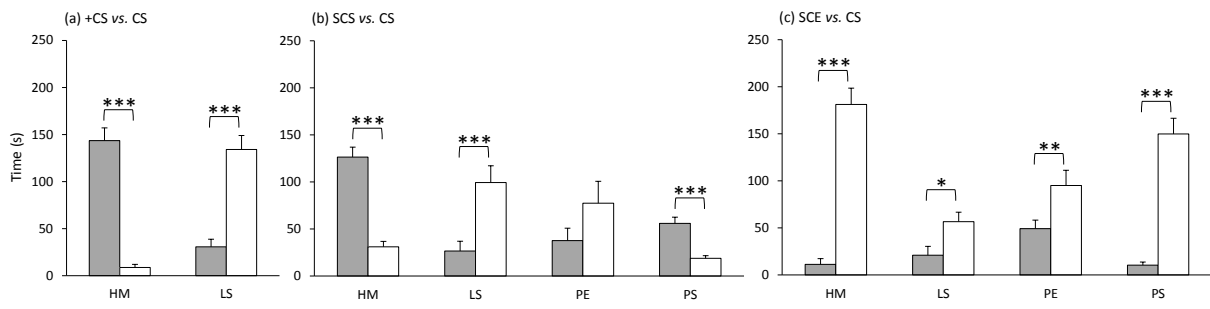
811 metabolic features are based on hierarchical clustering (Euclidian distances). The first group of
812 features gathers ions present in +CS and SCE, the 2nd group of features gathers ions present in
813 SCS and SCE. CS: control seawater; +CS: cave seawater; SCS: sponge conditioned seawater;
814 SCE: sponge chemical extract.

815

816 **Fig. 7** Comparative ion intensities of the features corresponding to the chemical cues A–D (1st
817 group of features that gathers ions present in +CS and SCE) and E–N (2nd group that gathers
818 ions present in SCS and SCE) in the dissolved phase of the seawater samples analysed by liquid
819 chromatography coupled to mass spectrometry in positive mode. The metabolites E, F, G and
820 H likely belong to oxylipin family. Kruskal Wallis tests followed by a post hoc multiple range
821 test (Fisher's Least Significant Difference), $n=4$: significant differences are indicated with
822 $*P < 0.05$, and $**P < 0.01$, and differences between treatments are shown by different letters.
823 CS control seawater; +CS cave seawater; SCS sponge conditioned seawater; SCE sponge
824 chemical extract

825 **Fig. 1**

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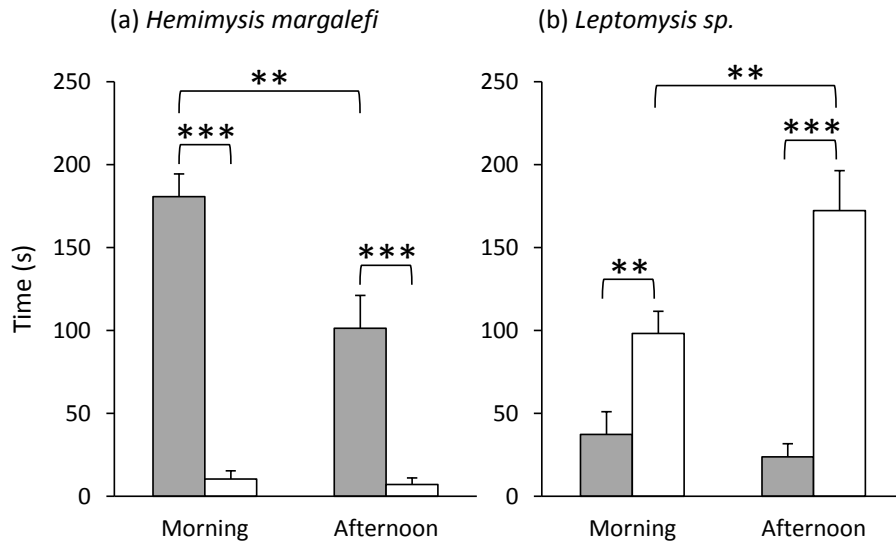


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829 **Fig. 2**

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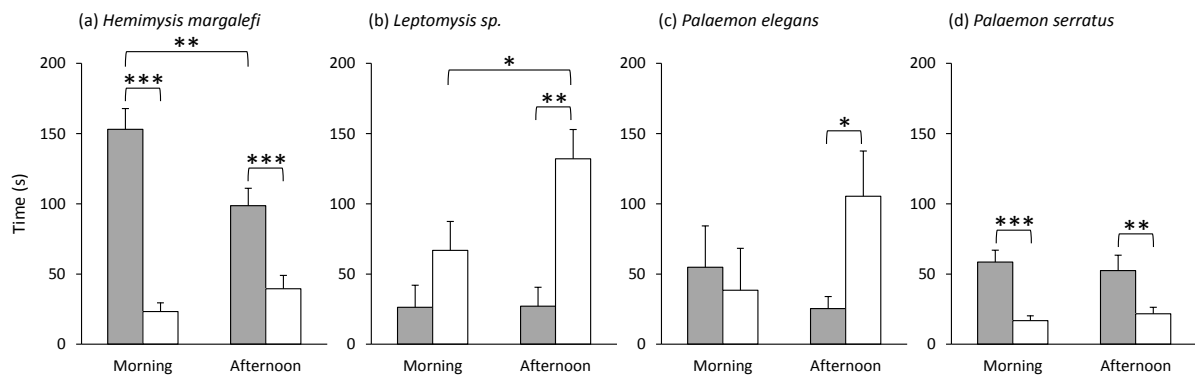


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833 **Fig. 3**

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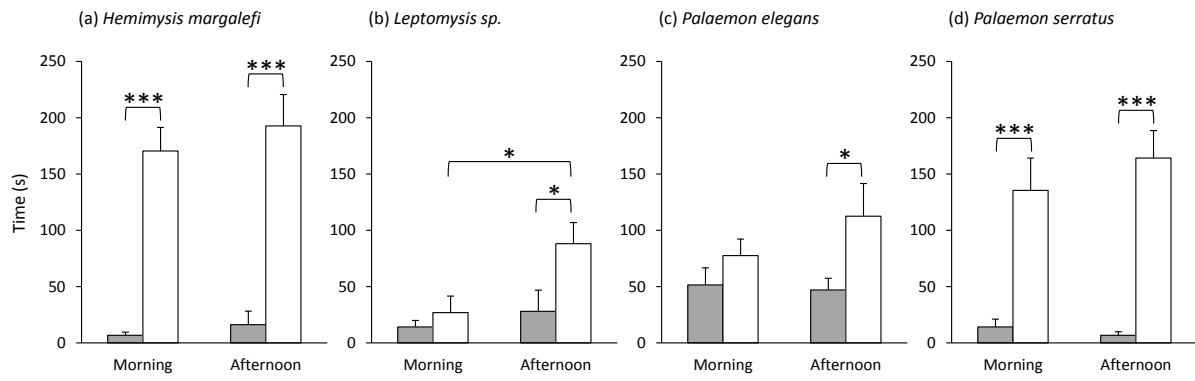


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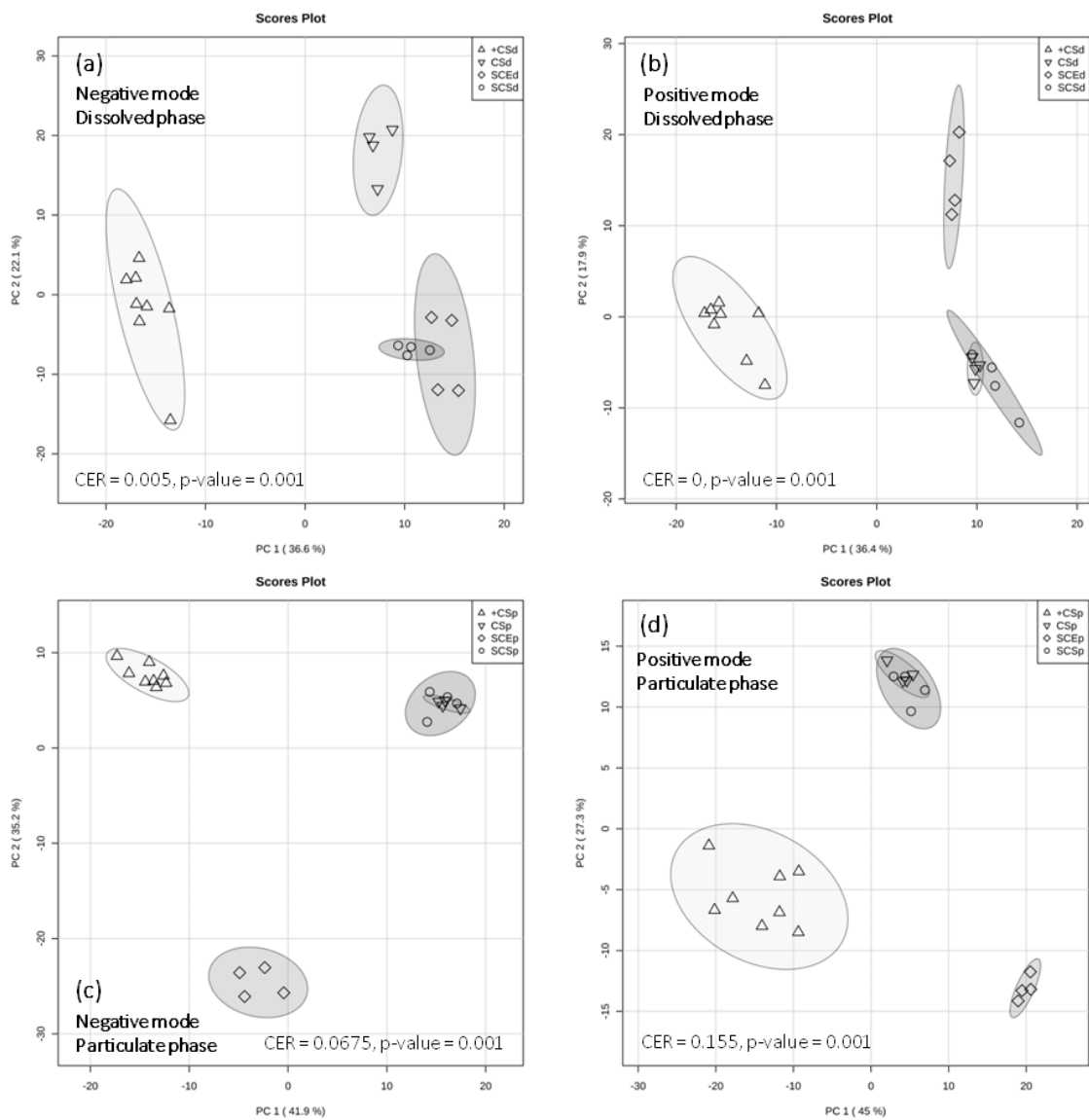
837 **Fig. 4**

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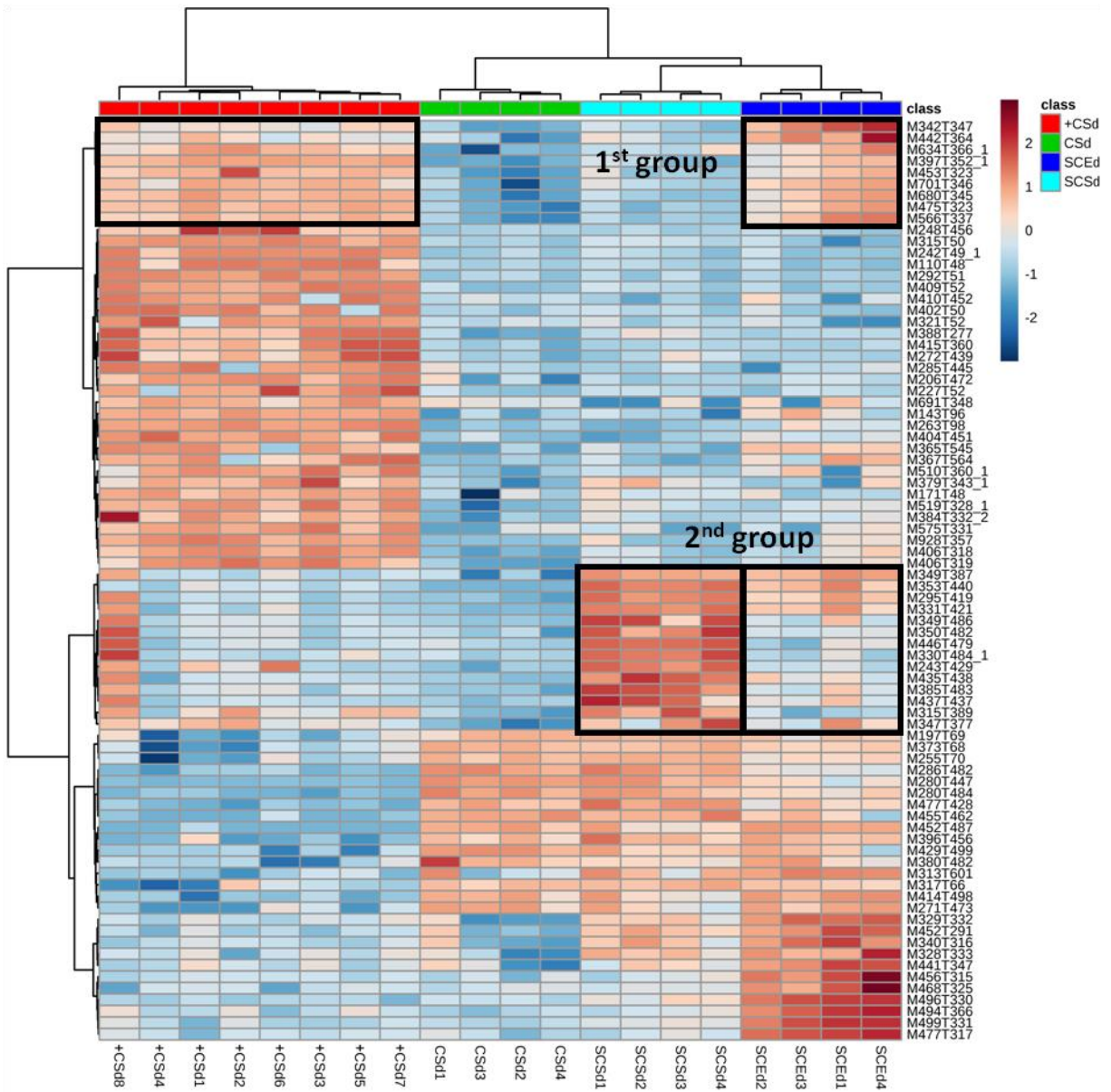
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