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Lisa Bajolle, Isabelle Larocque-Tobler, Adam A Ali, Martin Lavoie, Yves Bergeron, et al.. A chironomid-inferred Holocene temperature record from a shallow Canadian boreal lake: potentials and pitfalls. Journal of Paleolimnology, 2019, 61, pp.69-84. 10.1007/s10933-018-0045-9. hal-01890717

# HAL Id: hal-01890717 https://amu.hal.science/hal-01890717v1

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# HAL Id: hal-01890717 https://hal-amu.archives-ouvertes.fr/hal-01890717

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# A chironomid-inferred Holocene temperature record from a shallow Canadian boreal lake: potentials and pitfalls

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## Abstract

The biodiversity of shallow ( $z_{max} < 5-7$  m) lakes is sensitive to water-level and climate changes, but few such aquatic ecosystems have been studied in the context of quantitative climate reconstruction. Lac Lili (unofficial name) is located in the boreal forest of western Quebec, Canada, and was chosen for its shallowness ( $z_{max} = 1.40$  m) to assess if chironomid assemblages in the sediments could be used to reconstruct Holocene temperature fluctuations quantitatively. Inferred temperatures displayed a decreasing trend from a maximum value ca. 8000–3500 cal year BP, slight warming between ca. 3500 and 3000 cal year BP, followed by cooling to the present. Although chironomid assemblages were influenced by factors other than climate, primarily water depth and changes in macrophyte richness, the reconstructed Holocene temperature pattern was very similar to the known regional climate history. Temperature inferences derived from the chironomid assemblages were, however, warmer than the two reference periods. This deviation was likely a consequence of three factors: (1) shallowness of the lake, which favoured littoral taxa with warmer-than-today temperature optima, (2) the low number of lakes warmer than 16°C in the training set, and (3) the absence of shallow lakes in the training set.

Keywords Chironomidae. Holocene. Paleoclimate reconstructions. Transfer function.

Water level. Boreal forest

### Introduction

Ponds and shallow lakes  $[z_{max} < 5-7 \text{ m}, \text{ and with light penetration to the bottom (Wetzel 2001)}]$ are the most abundant water bodies in the world (Meerhoff and Jeppesen 2009) and they are particularly sensitive to water-level fluctuations and climate changes (Jeppesen et al. 2014; Smol 2016). Water level fluctuations between extreme highs and lows may cause shifts between turbid and clear-water, macrophyte-dominated states (Coops et al. 2003), thus affecting the biota and the lake's ecological functions. Submerged macrophytes play an important role in these lakes, as they influence oxygen conditions and inhibit the resuspension of sediments, which may possibly reduce phytoplankton biomass (Blindow et al. 2002). Shallow eutrophic lakes can develop in two states of equilibria: a clear-water state characterized by high water transparency and abundant submerged vegetation, and a turbid state characterized by high phytoplankton densities and low transparency. A shift from one state to the other can result from water-level changes (Blindow et al. 2002; Scheffer and van Nes 2007; Tarkowska-Kukuryk and Kornijów 2008). Climate projections for the eastern Canada Boreal Shield display an increase in annual mean temperature by 2100, ranging up to 5°C in summer, as well as an increase of 13% in annual precipitation, mainly in winter and spring. with less in summer (Price et al. 2013). Consequently, Mooij et al. (2007) suggested that this change in climate will enhance external nutrient loading, possibly causing a shift from a clear to a turbid state in lacustrine ecosystems. These predictions, however, lack long-term data to study the dynamics among climate (temperature and precipitation) changes, water-level fluctuations, and biotic changes, notably in shallow lakes.

Long-term relationships between shallow lake ecological functions and climate can be obtained from chironomids preserved in lake sediments (Smol 2016). Chironomid (non-biting midges) head capsules are made of chitin that can be preserved for up to 200,000 years (Axford

et al. 2009), and have been used to reconstruct air temperature quantitatively in many areas of the world (Brooks and Birks 2001; Walker and Cwynar 2006; Heiri et al. 2007; Millet et al. 2012), sometimes with high accuracy (difference between instrumental and inferred data =  $0.8^{\circ}$ C on average) (Larocque and Hall 2003; Larocque et al. 2009; Larocque-Tobler et al. 2016; Luoto and Ojala 2017; Zhang et al. 2017). Although the technique can provide accurate results, most of the studied lakes were deep. To our knowledge. quantitative temperature reconstructions using chironomid head capsules have never been conducted in shallow lakes in Eastern Canada and were attempted in only one shallow (depth <5 m) lake in Western Canada (Upiter et al. 2014).

In shallow [<5–7 m (Wetzel 2001)] lakes, chironomid assemblages are composed mainly of littoral taxa influenced primarily by water-level changes (Engels and Cwynar 2011; Engels et al. 2012), turbidity (Greffard et al. 2012), macrophyte species richness (Langdon et al. 2010), and total phosphorus (Brodersen and Lindegaard 1997). Temperature is also an important factor that influences chironomid communities in shallow lakes, but as such communities are composed of littoral taxa, which tend to have warm optima (Korhola et al. 2002), reconstructions based on these assemblages might yield warmer inferences than would otherwise be expected. Nevertheless, because shallow lakes are very sensitive to temperature and precipitation, climate reconstructions from these ecosystems might be valuable.

Our study site is located in western Quebec (Fig. 1), 110 km north of the town of Rouyn-Noranda. "Lac Lili" (Unofficial name: 49°10'25.501"N; 79°22'26.931"O, 320 m a.s.l.) is a shallow lake ( $z_{max} = 1.40$  m) that covers an area of 10 ha and lacks both inlets and outlets. It is located in the black spruce-feather moss bioclimatic domain (Saucier et al. 2009). Forest cover is dominated by trembling aspen (*Populus tremuloides*), black spruce (*Picea mariana* Mill.). and jack pine (*Pinus banksiana* Lamb) (Blouin and Berger 2005). The mean annual temperature recorded at the closest meteorological station [La Sarre: 1961–1990] is 0.8 °C,

with August temperature averages of 15.0 °C (1961–1990) and 15.4°C (1981–2010). Mean annual precipitation is 640 mm, of which 250 mm falls as snow (Environnement Canada 2016).

Lac Lili is located 395 km from Lac Aurélie (49°10'25.501"N; 79°22'26.931"W; black spruce-feather moss bioclimatic domain; 320 m a.s.l; 1 ha. 10 m deep) (Fig. 1), where a chironomid-inferred temperature reconstruction was carried out and was similar to regional and hemispheric temperature records (Bajolle et al. 2018). Multi-site comparison [Lac Lili *versus* Lac Aurélie (Bajolle et al. 2018)] will place the climate reconstruction within a regional context. If the patterns are similar at both sites, it will provide support for regional climate change having been reconstructed accurately, whereas if they diverge. the differences might be a consequence of local effects (e.g. water-level fluctuations. fire regime). The goal of this study was to assess if chironomid assemblages from a shallow lake can be used to infer air temperature quantitatively and reconstruct the general pattern of climate change in this region.

### Materials and methods

Coring, stratigraphy, chronology

Two overlapping sediment cores were taken in March 2014 through the frozen surface at the centre of Lac Lili using a modified Livingstone-type square-rod piston corer ( $100 \times 5$  cm). The water-sediment interface was collected using a Kajak-Brinkhurst (KB) gravity corer (Glew 1991). The extruded cores (310 and 306 cm long) were sectioned into 0.5-cm intervals to achieve high-temporal sampling resolution. Terrestrial plant macrofossils (seeds, needles and bark) were retrieved from seven samples for AMS radiocarbon dating (Table 1). The <sup>14</sup>C dates were calibrated at  $2\sigma$  ranges using the Clam package (Blaauw 2010) of R software version 3.0.1 (R Core Team 2016) based on the non-marine (IntCal13) radiocarbon calibration curve

(Hua et al. 2013; Reimer et al. 2013). The age-depth model (Fig. 2) was obtained using a 'classical' age-depth model with a smoothing spline function (Blaauw 2010). All dates are expressed in calibrated years before present, hereafter cal year BP.

#### Chironomid analyses

Sediment slices of 0.5 cm were sampled at intervals varying between 4, 2, and 1 cm, for a total of 116 samples. Samples were soaked overnight in 10% KOH and then rinsed with water and filtered through a 100- $\mu$ m mesh. The remains were poured into a counting tray and observed under a stereomicroscope at 10× magnification. Each head capsule (whole and half mentum) was individually picked with fine tweezers and mounted on a microscope slide in a drop of glycerine. Slides were observed under a light microscope at 400-1000× magnification following the taxonomic guide of Brooks et al. (2007). A minimum of 50 head capsules is needed for temperature reconstructions (Larocque 2001; Heiri and Lotter 2010). All samples in this study contained at least 92 head capsules.

A chironomid stratigraphy (percentage of each taxon in each sample) was constructed using C2 software (Juggins 2003). Only taxa present in more than six samples are shown in Fig. 3. A detrended correspondence analysis (DCA) was performed on a matrix of square-roottransformed percentages using ade4 and vegan packages from the R v3.2.2 software package (Borcard et al. 2011). The length of the DCA-axis shows whether or not the distribution of the data set along the axis is linear or unimodal (Borcard et al. 2011). A PCA calculated with C2 software was used to study the changes in chironomid assemblages through time on a dispersion matrix of square-root-transformed percentages into which rare taxa with abundances <1% in fewer than two samples were removed. A constrained sum-of-squares cluster analysis (CONISS; Grimm 1987) was used to identify zones in the stratigraphy. with the significance of each zone tested with the broken stick model (Bennett 1996) using the Rioja R package (Juggins et al. 2015). Percentages of littoral and eutrophic taxa were determined (Table 2) based on Brooks et al. (2007).

#### Temperature reconstruction

A new Eastern Canadian model, modified from Larocque et al. (2006) and Larocque (2008), and utilized for climate reconstruction at Lac Aurélie (Bajolle et al. 2018), was used. The training set comprises 75 lakes (no outliers) and 79 taxa with abundances of >1% in more than two lakes. Mean August temperature varied from 3 to 21°C for a temperature gradient of 18°C. WAPLS 999-bootstrap with two components yielded a correlation coefficient ( $r^2_{boot}$ ) of 0.85, an RMSEP of 1.67°C and a maximum bias of 3.05°C. Occurrence of taxa and temperature optima can be found in Bajolle et al. (2018) and in Table 2. Sample specific errors (eSEP) were calculated. To validate the use of the transfer function. an evaluation of modern analogues for each sample was performed using the minimum distance of the fossil sample to the training set assemblages (Overpeck et al. 1985). Fossil samples with a distance above the 20<sup>th</sup> percentile were considered as having "no analogues", and those above the 10<sup>th</sup> were considered to be "poor analogues" (Overpeck et al. 1985). Percentages of taxa colder or warmer than 13.6°C (average of temperature optima of taxa found in Lac Lili) were calculated using the temperature optima obtained from the transfer function (Table 2). This site-specific cut-off is not far from the average temperature optima of the 79 taxa found in the training set (13.2°C). The difference between average temperature inferences and reference values (climate normals: 1961-1990 and 1981–2010) was performed using a t-test.

## Results

Chronology and sedimentation rates

The core was 310 cm long and composed entirely of gyttja. An age of ca. 8407–8522 cal year BP (2 sigma range) was obtained at the bottom of the core (304–306 cm) (Table 1). Three periods were identified with respect to sedimentation rates, based on the age-depth model (Fig. 2). Period 1 (ca. 8600–3500 cal year BP; 310–180 cm) showed a low sedimentation rate (mean: 0.04 cm/yr). During Period 2 (ca. 3500–2700 cal year BP; 180–120 cm), the average sedimentation rate increased quickly to 0.09 cm/yr. Period 3 (ca. 2700 cal yr BP to Present; 120-0 cm) was characterized by a decrease in sedimentation rate (mean: 0.02 cm/year).

Chironomid stratigraphy

In total, 116 samples were analysed. with a total of 11,693 head capsules (hereafter HC) identified. All samples had more than 92 HC, with an average of 100 HC per sample (Fig. 3). Fifty-eight taxa were identified and 42 of them are presented in Fig. 3. The 16 taxa not illustrated occurred in <6 samples at percentages <1.9%, thus did not contribute much to the temperature reconstruction (Table 2).

The chironomid stratigraphy was divided into four statistically significant zones (Lch-1 to Lch-4; Fig. 3). In general, both littoral and warmer-than-13.6°C taxa dominated the chironomid community throughout the core, with average percentages of 80% and 66%. Respectively, whereas colder-than-13.6°C taxa had percentages that ranged between 13 and 50% (average: 34%).

DCA yielded a gradient of 1.6 SD, suggesting that linear regression such as PCA was appropriate. PCA Axis 1 and had eigenvalues of 0.18 and 0.12, respectively.

During zone Lch-1 (ca. 8500–5700 cal year BP; 310–240 cm), littoral taxa were below average and increased after 6300 cal year BP. Eutrophic taxa oscillated around the average. PCA Axis 1 scores were above 0, whereas PCA Axis 2 scores were generally below 0. The dominant taxa were *Tanytarsus glabrescens*-type, *Procladius* spp., *Ablabesmyia* spp., *Polypedilum nubeculosum*-type, *Tanytarsus pallidicornis*-type, *Lauterborniella* spp., *Dicrotendipes nervosus*-type and *Tanytarsus mendax*-type. *Cladotanytarsus mancus*-type and *Psectrocladius sordidellus*-type percentages increased towards the end of the zone. *Pagastiella* spp. and *Allopsectrocladius* spp. had their highest percentages of the record. Taxa with warmerthan-13.6°C optima dominated the assemblages, with percentages >68%.

In zone Lch-2 (ca. 5700–3500 cal year BP; 240–180 cm), percentages of colder-than-13.6°C taxa increased above average (32%), while warmer-than-13.6°C taxa decreased below average. but always remained higher than 48%. PCA Axis 1 scores decreased compared to the previous zone, but remained slightly above 0, whereas PCA Axis 2 scores were mainly above 0. Eutrophic taxa percentages were above average until ca. 5000 cal year BP, then decreased below average until the end of the zone. Percentages of littoral taxa started below average until ca. 5000 cal year BP but increased until the end of the zone. The dominant taxa were the same as in the previous zone, however *Paratanytarsus* spp. started to be recorded consistently and *Tanytarsus* spp. increased towards the end of the zone. *Stempellinella* had higher percentages at the beginning of the zone than in the previous section. but it decreased and occasionally disappeared after ca. 5000 cal yr BP.

In zone Lch-3 (ca. 3500–1200 cal year BP; 180–55 cm), the percentages of littoral taxa fluctuated around the average. Eutrophic taxa percentages started well above average. decreased. and then finally increased again after 2000 cal year BP. PCA Axis 1 scores

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decreased below 0 and PCA Axis 2 scores remained above 0, with a tendency to decrease to the end of the zone. *T. glabrescens*-type percentages decreased sharply compared to zone Lch-2. *Microtendipes pedellus*-type, *Chironomus anthracinus*-type and *Guttipelopia* spp. were recorded consistently. *Stempellinella* spp. and *Stempellina* spp. had higher percentages at the beginning of the zone and then decreased thereafter. Warmer-than-13.6°C and colder-than-13.6°C taxa percentages oscillated around their averages.

Zone Lch-4 (ca. 1200 cal year BP to present; 55–0 cm) had increases in the percentages of *P. septentrionalis*-type, *P. sordidellus*-type, *M. pedellus*-type, *Paratanytarsus* spp., *T. pallidicornis*-type, *T. mendax*-type and a slight increase in *T. glabrescens*-type. *Ablabesmyia* spp. was also recorded more regularly. PCA Axis 1 and Axis 2 scores were below 0. The percentages of eutrophic taxa decreased below average while littoral taxa oscillated. Percentages of colder-than-today taxa were above average (32-50%) whereas warmer-than-13.6°C were below average, but percentages remained higher than 48%.

#### Temperature reconstructions

Eight of the 116 fossil samples had no modern analogues (Fig. 4a) and 85 samples had poor analogues compared to the training set samples. During zone Lch-1 (8500–5800 cal year BP). the average reconstructed temperature was 16.9°C, with a decrease from 19°C (maximum) to 17°C at the end of the zone. In zone Lch-2 (ca. 5700–3500 cal year BP), temperatures had an average of 16.8°C, with a decrease from 17.8°C around 5200 cal year BP to 16.2°C at 3400 cal year BP. Zone Lch-3 (ca. 3500–1200 cal year BP) started with inferences for high temperatures (19.3–18.5°C), followed by a decrease to 16.8°C between ca. 3000 and 2500 cal year BP. An increase (18.3 and 19.6°C) was inferred for the period between 1800 and 1500 cal year BP. The average chironomid-inferred temperature during Zone 3 was 17.9°C. In the last zone (Lch-

4), the temperatures decreased from ca.  $17.5^{\circ}$ C at the beginning of the zone to  $14.8^{\circ}$ C at the end of the zone. The average during this zone was  $16.5^{\circ}$ C.

The temperature anomalies show that throughout the whole record. only six of the inferences were colder than the climate normal of 15.4°C and three were colder than today. with the climate normal of 15°C (Fig. 4b). The average anomaly from 15°C (2.10°C) and from 15.4°C (1.70°C) for the whole record showed that the temperature inferences were generally and significantly ( $p < 2.2 \ 10^{-16}$ ), warmer than today.

### Discussion

#### Chironomid assemblages

As expected for a shallow lake, the chironomid assemblages were dominated by littoral and eutrophic taxa, which tend to have warmer optima (Table 2) and are generally influenced by water levels (Engels and Cwynar 2011; Engels et al. 2012), turbidity (Greffard et al. 2012), macrophyte species richness and abundance (Langdon et al. 2010), and total phosphorus (Brodersen and Lindegaard 1997). In Lac Lili, when the PCA Axis 1 scores decreased below 0, the sum of Chironomini increased (Fig. 5). Korhola et al. (2002) also observed that chironomids with warm temperature optima dominated assemblages in a shallow lake in Finland, and high sums of Chironomini have been linked to water levels and macrophytes species richness and abundance (Langdon et al. 2010). Thus, water levels and macrophytes were possibly the main factors that drove chironomid assemblages in Lac Lili.

The main changes occurred ca. 3500 cal year BP when PCA axis 1 scores changed from positive to negative. Before ca. 3500 cal year BP, the assemblages were dominated by *Tanytarsus glabrescens*-type and were replaced by *Polypedilum nubeculosum*-type and

*Microtendipes pedellus*-type, which are often associated with floating-leaved plants and other macrophytes (Learner et al. 1989; Tarkowska-Kukuryk and Kornijów 2008). Synchronously Tanytarsini-dominated communities shifted to Chironomini-dominated assemblages (Fig. 5), possibly with a change in water level (shallowing) and macrophyte species richness and abundance (Langdon et al. 2010), as illustrated by the strong increase in *Nuphar* (a floating-leaved plant) pollen in Lac Lili (Fig. 5). The timing of this major change in the chironomid assemblages coincides with the start of the Neoglacial, when climate became wetter in this region (Garralla and Gajewski 1992; Viau and Gajewski 2009; Hély et al. 2010; Ali et al. 2012; El-Guellab et al. 2015).

After 1500 cal year BP, the percentages of both *Psectrocladius* types and *Cricotopus* spp. largely increased along with Pentaneurini spp. and *Ablabesmyia* spp. *Psectrocladius* is often associated with macrophytes (Brodersen et al. 2001; Langdon et al. 2008). *Cricotopus. Ablabesmyia* and *Zalutschia* spp. are associated with vegetation (Brooks et al. 2007). Although Chironomini still dominated, they started to decrease, while Tanytarsini started to increase. This change suggests the onset of a new state, perhaps as a response to colder conditions, as suggested by the decrease in taxa with optima warmer than 13.6 °C.

#### Comparison with other regional climate records

PCA axis 1 seems to be related with warmer-than-13.6 °C taxa, but not directly to inferred temperature, suggesting that confounding factors (eutrophication, water-level changes and macrophytes) impacted the chironomid assemblages more than temperature. Nevertheless, the general pattern of temperature changes (i.e. warm middle Holocene to ca. 3500 cal year BP, with a decreasing trend to the present) reconstructed using the chironomids follows the quantitative paleoclimatic reconstructions based on the chironomid record of Lac Aurélie (Fig.

5) and pollen from the Province of Quebec (Viau et al. 2006). A Neoglacial period with cooler conditions started at 3500 cal year BP in this region (Garralla and Gajewski 1992; Viau and Gajewski 2009; Ali et al. 2009; Hély et al. 2010).

The PCA axis 2 scores showed their main changes between ca. 3500 and 3000 cal year BP when chironomid-inferred temperatures were warmer and eutrophic percentages increased. The sedimentation rates during that period also increased (Period 2; Fig. 2). Major changes in sedimentation rates at that time were also recorded by Ali et al. (2008) in western Québec and were interpreted as having been caused by a major flood in the watershed between ca. 3600 and 1900 cal year BP, causing a rapid rise in lake level. Consequently, the increase in sedimentation rate is possibly associated with an increase in precipitation, as suggested by the changes in the chironomid assemblages that show highest percentages of eutrophic taxa. A similar warming (~1°C) period was also reconstructed using chironomids at Lac Aurélie (Fig. 5) and in the pollen record of Viau and Gajewski (2009), but at lower amplitude. The agreement between our results and other regional records suggests that although temperature was not the only factor that affected chironomid assemblages, patterns of climate change could still be adequately reconstructed.

#### Quantitative temperature reconstruction in a shallow boreal lake

The temperature pattern through the Holocene was similar to that in other regional records. The goal of using quantitative methods to reconstruct climate, however, is to obtain accurate inferences, not only a pattern of change. Reconstruction of climate from chironomid assemblages in a shallow study lake may compromise its accuracy. For instance. most inferences were warmer by 1.7 to 2°C than the climate normals (1981–2010, 15.4°C: 1961–1990, 15°C) (Fig. 4). This was not the case for the chironomid-inferred reconstruction in Lac

Aurélie or in pollen records of the region (Viau and Gajewski 2009). which all show periods with colder-than-the-climate normal inferences. Twenty-seven of the taxa have temperature optima warmer-than-13.6°C and nine of fourteen taxa with percentages reaching 20% or higher have warmer-than-13.6°C optima (Fig. 3). Thirteen taxa have temperature optima between 14.9 and 25.3°C. Three of them have percentages of 20% or more and temperature optima between 15.8 and 25.3°C. When using WAPLS. these taxa contribute more to the temperature inference and thus lead to temperature inferences well above the climate normal.

This lower accuracy can potentially be explained by a few factors: (1) littoral taxa dominate the assemblages (average = 80 %), (2) warm-optimum taxa also dominate (average = 66%), (3) the training set has only 11 lakes with mean August temperature above 15.8°C (the actual temperature at Lac Lili) and only five lakes between 15 and 16°C, and (4) the training set function is perhaps inappropriate for shallow lakes. Shallow-water taxa tend to have warmer-than-today temperature optima (Korhola et al. 2002). Thus. it is not surprising that inferences were generally too warm. Lacking a cool hypolimnion, the lake can only support taxa that prefer warmer conditions. This could explain why colder-than-today percentages were always low (average of 32%) in Lac Lili. The training set mainly comprises lakes with water depths greater than 5 m. To increase the accuracy of the chironomid inferences, a larger number of shallow lakes should be added, or preferably, a new transfer function, composed only of shallow lakes. could be developed. Absence of modern analogues in eight samples and poor analogues in 85 samples suggest that fossil samples were dissimilar to the training set, possibly because of the high percentages of littoral taxa in the fossil assemblages. This situation was also observed in a shallow lake used for reconstructing climate in western Canada (Upiter et al. 2014), and the authors concluded that although the pattern of temperature changes was similar to those of the studied region, all inferences were within the error of the model and thus should be viewed with caution. The training set used in this study also possesses few lakes with

mean-August temperatures similar to Lac Lili, and no lakes between 16.8°C and 18.9°C. Inferences above 15°C are thus generally under-estimated (Electronic Supplementary Material Fig. 1). More lakes with mean-August temperatures above 15°C should be sampled to improve the accuracy of the model.

## Conclusions

It has been shown that shallow lakes are impacted by climate change and temperature reconstructions from chironomids, along with other variables in sediments such as diatoms, zooplankton, and pollen, could help decipher the effects that temperature had on lake communities throughout the Holocene. Lac Lili provided an opportunity to determine if shallow lakes could be used to obtain reasonable temperature inferences. which are typically derived from deeper lakes around the world. Our results showed that although chironomids were effective for reconstructing the general pattern of climate change, their ability to provide accurate inferences was compromised and most were above the climate normal. Better accuracy in shallow lakes could be attained if appropriate transfer functions were used; either by increasing the number of shallow lakes in an existing training set, or by developing a transfer function with data exclusively from shallow lakes. In our opinion, the first option is preferable, as lakes that are presently shallow might have been deeper in the past, and if no deep lakes are included in the transfer function, temperature might not be accurately reconstructed, as samples from such lake will have poor analogues. Furthermore, lakes with temperatures between 16 and 19°C should be added to the existing transfer function to increase temperature coverage within the overall range.

### Acknowledgements

This research was funded by the Natural Sciences and Engineering Research Council of Canada (NSERC), the Centre National de la Recherche Scientifique (France), the European IRSES NEWFORESTS program; the MITAC program, the *Institut Universitaire de France* and the *Institut Écologie et Environnement* through the GDRI "Forêts Froides". We thank the French University Institute for its support, the University of Montpellier and Aix-Marseille University. Our thanks to R. Julien, D. Gervais, B. Brossier and M. Girardin for their participation in fieldwork. We greatly appreciate the contribution of the *Ministère des Forêts*. *de la Faune et des Parcs* du Québec (MFFP), especially P. Grondin.

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Laboratory code	Depth (cm)	<sup>14</sup> C age (yr BP)	Calibrated <sup>14</sup> C age ranges (cal yr BP; 2σ-range)*	Material dated
Beta-427033	23-27	$410\pm 30$	430-519 (0.83)	Plant macroremains
			331-358 (0.12)	Plant macroremains
Beta-427032	48-50	$1150\pm30$	978-1145 (0.90)	Plant macroremains
			1158-1170 (0.05)	Plant macroremains
Beta-427031	118-120	$2730\pm30$	2762-2876 (0.95)	Plant macroremains
			2915-2915 (0.01)	Plant macroremains
Beta-427030	178-182	$3180\pm30$	3358-3453 (0.95)	Plant macroremains
Beta-427029	228-232	$4780\pm30$	5468-5589 (0.95)	Plant macroremains
Beta-427028	278-282	$6150\pm30$	6966-7159 (0.94)	Plant macroremains
			6960-6963 (0.01)	Plant macroremains
Beta-427027	304-306	$7670\pm30$	8407-8522 (0.90)	Plant macroremains
			8528-8538 (0.05)	Plant macroremains

Table 1 Radiocarbon dates from terrestrial plant macroremains

\*Calibrated ages: the means of the  $2\sigma$ -ranges with highest probabilities (bold).

**Table 2** Occurrence and maximum percentages of taxa in the sediment of Lac Lili (total of 116 samples). littoral and eutrophic taxa (Brooks et al. 2007). temperature optima yielded by WA from the East Canadian transfer function (Larocque 2008. *modified*). and temperature categories in relation to optima.

Base interfield in sediment oran         Overwrene in the presentation in the presentitic segnet.     Description in the pres	Lac Lili						Optima	Category	
All precended dis spp.446.9X15.8XChironomus anthracinus-type826.1X12.2XChironomus planouss-type636.0X14.8XCladoplanoutracinus-type708.3X13.6XCladounoutraces manuss-type1152.7XX10.4XCladounoutraces manuss-type335.2X11.9XXCristoranous spp.815.914.9XX12.6XCristoranous spp.815.914.9XX12.6XCristoranous spp.112.8XX13.7XXEndechironomus sinpart-type81.9XX13.6XXEndechironomus inpart-type11.0XX13.6XXEndechironomus inpart-type151.1XX14.6XEndechironomus inpart-type151.1XY14.6XEndechironomus inpart-type151.1X23.3XEndochironomus inpart-type161.0XX13.6XEndechironomus inpart-type11.0XX13.6XEndechironomus inpart-type151.1X9.6XEndechironomus inpart-type11.0X13.6XEndechironomus inpart-type11.0X13.6X <td< th=""><th>Taxa identified in sediment core</th><th>Occurrence</th><th></th><th>included</th><th></th><th></th><th>- East Canadian</th><th>Colder than today</th><th>Warmer than</th></td<>	Taxa identified in sediment core	Occurrence		included			- East Canadian	Colder than today	Warmer than
A.         X         1.2.2         X           Chironomus autharcius-type         6.3         6.0         X         14.8         X           Chironomus planonus-type         6.3         6.0         X         14.8         X           Cladoland larenilis-type         70         8.3         X         13.6         X           Cladoland strusts manucus-type-1         115         25.7         X         X         12.2         X           Coryonocurs spp.         58         7.5         10.4         X         11.9         X           Coryonocurs spp.         81         5.9         14.9         X         X         14.5         X           Coryonocurs spp.         4         1.0         X         X         14.5         X           Endochironomus althjennis-type         11         2.8         X         X         14.9         X           Endochironomus impart-type         1         1.0         X         X         14.9         X           Endochironomus impart-type         1         1.0         X         X         14.6         X           Gynonendipes pallens-type         15         1.1         X         X         14.6         X <td>Ablabesmyia spp.</td> <td>110</td> <td>12.8</td> <td></td> <td></td> <td></td> <td>14.2</td> <td></td> <td>Х</td>	Ablabesmyia spp.	110	12.8				14.2		Х
Chironomus plumosus-type       63       60       X       14.8       X         Cladopelma lateralis-type       70       8.3       X       13.6         Cladoianytarsus mancus-type-1       115       25.7       X       X       12.2       X         Corpanderinomus spp.       58       7.5       10.4       X       X       11.9       X         Cricatopus spp.       33       5.2       X       14.9       X       X       14.9       X         Cricatopus spp.       114       12.7       X       X       14.5       X       X         Einfeldia spp.       4       1.0       X       X       12.8       X       X       13.7       X         Eindochironomus albipenuis-type       8       1.9       X       X       13.8       X         Controinomus endems-type       1       1.0       X       X       15.8       X         Giptorentify settemelients-type       1       1.0       X       X       14.6       X         Giptorentify settemelients-type       1       1.1       X       9.6       X       X         Giptorentify settemelients-type       1       1.1       X       15.2	Allopsectrocladius spp.	44	6.9		Х		15.8		Х
Calage-Inductory       70       8.3       X       136         Cladape-Inductory       115       25.7       X       X       12.2       X         Corynoneura spp.       58       7.5       10.4       X         Creizoopas spp.       33       5.2       X       11.9       X         Creizoopas spp.       81       5.9       14.9       X         Creizoonadiges nervosus-type       114       12.7       X       X       13.6       X         Eindechtronomus albipentis-type       11       2.8       X       X       13.7       X         Eindechtronomus albipentis-type       11       2.8       X       X       13.7       X         Eindechtronomus inpur-type       8       1.9       X       X       14.9       X         Eindechtronomus tendens-type       1       1.0       X       X       14.9       X         Eindechtronomus tendens-type       1       1.0       X       X       14.6       X         Gymonetrinceneux       1       1.0       X       X       14.6       X         Gymonetrinceneux       1       1.0       X       X       13.6       X	Chironomus anthracinus-type	82	6.1			Х	12.2	Х	
And       And       X       X       1.2.2       X         Corponeura spp.       58       7.5       10.4       X         Cricatopus spp.       33       5.2       X       11.9       X         Cricatopus spp.       81       5.9       14.9       X         Cricatopus spp.       114       12.7       X       X       14.5       X         Dicrotendipes nervous-type       114       12.7       X       X       14.5       X         Endechironomus albipennis-type       11       10       X       X       13.7       X         Endechironomus impar-type       8       1.9       X       X       14.9       X         Endechironomus inpar-type       1       1.0       X       X       13.8       X         Endechironomus inpar-type       1       1.0       X       X       15.8       X         Endechironomus inpar-type       1       1.0       X       X       15.8       X         Endechironomus inpar-type       1       1.0       X       X       14.6       X         Gorponeutricoenus-spp.       1       1.0       X       X       15.1       X	Chironomus plumosus-type	63	6.0			Х	14.8		Х
Coryonolura spp.       58       7.5       10.4       X         Cricotopus spp.       33       5.2       X       11.9       X         Cryonoluronomus spp.       81       5.9       14.9       X         Dicrotendifies nervosus-type       114       12.7       X       X       14.5       X         Einfeldus spp.       4       1.0       X       X       12.8       X         Endochironomus albipenis-type       11       2.8       X       13.7       X         Endochironomus tendens-type       1       1.0       X       X       15.8       X         Endochironomus tendens-type       1       1.0       X       X       15.4       X         Giptoendifies seellens-type       1       1.1       X       16.6       X         Giptoendifies seellens-type       1       1.1       X       9.6       X         Heterotrissociadius maeri-type       1       1.1       X       9.6       X         Monocludius spp.       2       1.0       X       X       13.5       X         Monocludius spp.       1       1.0       X       12.5       X         Orthocludius spp.       2	Cladopelma lateralis-type	70	8.3		Х		13.6		
Criverious spp.         33         5.2         X         11.9         X           Cryptochirmonus spp.         81         5.9         14.9         X           Dicrostendifies nervosus-type         114         12.7         X         X         14.9         X           Einfeldia spp.         4         1.0         X         X         12.8         X           Einfeldia spp.         4         1.0         X         X         13.7         X           Endochironomus albipennis-type         8         1.9         X         X         14.9         X           Endochironomus tendens-type         1         1.0         X         X         X         15.8         X           Gryptotendironomus tendens-type         15         1.1         X         X         15.1         X           Gloptotendiges severini-type         15         1.1         X         X         15.1         X           Heterotrissocladius maeri-type         1         1.1         X         X         23.3         X           Idueterborniella spp.         1         1.0         X         X         23.3         X           Microtendifipes padellus-type         75         10.3	Cladotanytarsus mancus-type-1	115	25.7		Х	Х	12.2	Х	
Cyperchirmonnus spp.8159149XDicrotendipes nervosus-type114127XX14.5XEinfeldia spp.41.0XX12.8XEndochironomus abipennis-type112.8XX13.7XEndochironomus inpar-type81.9XX14.9XEndochironomus inpar-type11.0XXX14.9XEndochironomus intendens-type11.0XXX15.8XGypothenocladius spp.11.0XXX14.6XGypothenocladius spp.11.1XY16.6XGypothenolipes severini-type11.1X9.6XHeterotrissocladius maeri-type11.1X9.6XHeterotrissocladius maeri-type11.0XX23.3XMoscricotopus spp.21.0XX23.3XMicrotendipes pedellus-type7510.3X13.5XOrnicalian spp.21.0X13.6XParachironomus varus-type271.9X13.6XParachironomus varus-type271.9X13.5XParachironomus varus-type1.11.9X10.7XParachironomus varus-type2513.3X10.7XParacheries nulsiquama-type513.3	Corynoneura spp.	58	7.5				10.4	Х	
Normal Stress         Normal S	Cricotopus spp.	33	5.2		Х		11.9	Х	
Effektion         4         1.0         X         X         1.2.8         X           Endochironomus albipennis-type         11         2.8         X         X         13.7         X           Endochironomus inpar-type         8         1.9         X         X         14.9         X           Endochironomus inpar-type         1         1.0         X         X         X         14.9         X           Endochironomus inpar-type         1         1.0         X         X         X         14.9         X           Endochironomus inpar-type         1         1.0         X         X         X         15.8         X           Gymonentriocnemus- Bropheneroldius spp.         1         1.0         X         X         14.6         X           Glyptotendipes severini-type         15         1.1         X         9.6         X         X           Heterotrissocladius maeri-type         1         1.1         X         9.6         X         X           Microtendipes pedellus-type         75         10.3         X         13.5         X           Onisus spp.         1         1.0         X         15.2         X           Onichochadi	Cryptochironomus spp.	81	5.9				14.9		Х
Endochironomus albipennis-type     11     2.8     X     X     X     1.3.7     X       Endochironomus inpar-type     8     1.9     X     X     14.9     X       Endochironomus inpar-type     1     1.0     X     X     X     15.8     X       Gymonetriocnemus- Bryophenenoladius spp.     1     1.0     X     X     X     15.8     X       Glynotendipes selenis-type     47     5.0     X     X     14.6     X       Glynotendipes selenis-type     15     1.1     X     X     15.1     X       Heterotriscoladius maeri-type     1     1.1     X     9.6     X       Heterotriscoladius smaeri-type     1     1.0     X     X     23.3     X       Heterotriscoladius seque:     1     1.0     X     9.9     X       Lauterborniella spp.     1     1.0     X     13.5     X       Microtendipes pedellus-type     75     10.3     X     13.5     X       Ontincladina spp.     1     1.0     X     13.6     X       Orthocladina spp.     2     1.0     X     13.6     X       Parachironomus varus-type     27     1.9     X     13.5     X	Dicrotendipes nervosus-type	114	12.7		Х	Х	14.5		Х
Endochironomus impar-type         8         1.9         X         X         X         14.9         X           Endochironomus tendens-type         1         1.0         X         X         X         15.8         X           Gymometriocnemus- Bryophaenocladius spp.         1         1.0         X         X         X         15.8         X           Glyptotendipes severini-type         47         5.0         X         X         14.6         X           Glyptotendipes severini-type         15         1.1         X         9.6         X           Heterotrissocladius maeri-type         1         1.1         X         9.6         X           Iderotrissocladius maeri-type         1         1.1         X         9.6         X           Iderotrissocladius maeri-type         1         1.0         X         1.0 <t< td=""><td>Einfeldia spp.</td><td>4</td><td>1.0</td><td>Х</td><td>Х</td><td></td><td>12.8</td><td>Х</td><td></td></t<>	Einfeldia spp.	4	1.0	Х	Х		12.8	Х	
Endochironomus tenden-type         1         1.0         X         X         X         S         1.0         Not in model           Gymnometriconenus- Bryophaenocladius spp.         1         1.0         X         X         X         14.6         X           Gyptonendipes pallens-type         47         5.0         X         X         14.6         X           Glyptonendipes severini-type         15         1.1         X         X         15.1         X           Heterotrissocladius maeri-type         1         1.1         X         9.6         X           Hyporygma spp.         2         1.0         X         X         23.3         X           Lauerborniella spp.         1         1.0         X         9.9         X           Microtendipes pedellus-type         75         10.3         X         13.5         X           Nanocladius spp.         1         1.0         X         15.2         X           Orthocladius spp.         1         1.0         X         13.6         X           Parachironomus varus-type         2         1.0         X         14.5         X           Paradayitarus spp.         92         8.7         X	Endochironomus albipennis-type	11	2.8		х	Х	13.7		Х
Arrow and the server of the s	Endochironomus impar-type	8	1.9		х	Х	14.9		Х
Bryophaenocladius spp.       1       1.0       X       not in model         Glyptotendipes pallens-type       47       5.0       X       X       14.6       X         Glyptotendipes severini-type       15       1.1       X       X       15.1       X         Glyptotendipes severini-type       1       1.1       X       X       15.1       X         Heterotrissocladius maeri-type       1       1.1       X       9.6       X         Hyporygma spp.       2       1.0       X       X       23.3       X         Hyporygma spp.       1       1.0       X       23.3       X         Microtendipes pedellus-type       75       10.3       X       13.5       X         Onisus spp.       1       1.0       X       13.5       X         Orthocladius spp.       2       1.0       X       13.5       X         Orthocladius spp.       1       1.0       X       13.5       X         Orthocladius spp.       2       1.0       X       13.6       X         Pagastiella spp.       2       1.0       X       X       10.7       X         Paradenioreonus varus-type       2	Endochironomus tendens-type	1	1.0	Х	Х	Х	15.8		Х
Glypotendipes pallen-type       47       5.0       X       X       14.6       X         Glypotendipes severini-type       15       1.1       X       X       15.1       X         Glypotendipes severini-type       1       1.1       X       X       15.1       X         Heterotrissocladius maeri-type       1       1.1       X       9.6       X       X         Hyporygma spp.       2       1.0       X       X       23.3       X         Lauterborniella spp.       1       1.0       X       X       9.9       X         Microtendipes pedellus-type       75       10.3       X       13.5       X         Nanocladius spp.       1       1.0       X       12.5       X         Orthocladius spp.       1       1.0       X       15.2       X         Orthocladius spp.       2       1.0       X       13.6       X         Pagastiella spp.       34       3.8       X       14.5       X         Parachironomus varus-type       27       1.9       X       10.7       X         Paratanytarsus spp.       95       13.3       X       12.8       X         Pa	Gymnometriocnemus-	1	1.0	Х			not in model		
Heterotrissocladius maeri-type       1       1.1       X       9.6       X         Hyporygma spp.       2       1.0       X       not in model       X         Lauterborniella spp.       115       16.5       X       X       23.3       X         Mesocricotopus spp.       1       1.0       X       9.9       X       X         Microtendipes pedellus-type       75       10.3       X       13.5       X         Omisus spp.       2       1.0       X       12.5       X         Orthocladius spp.       1       1.0       X       15.2       X         Orthocladius spp.       1       1.0       X       13.6       X         Orthocladius spp.       2       1.0       X       13.6       X         Pagastiella spp.       34       3.8       X       14.5       X         Parachironomus varus-type       27       1.9       X       13.5       X         Paratenytarsus spp.       1       1.9       X       10.7       X         Paratenytarsus spp.       1       1.9       X       not in model       X         Paratenytarsus spp.       1.0       X       X	Glyptotendipes pallens-type	47	5.0		х	Х	14.6		Х
Hyporygna spp.       2       1.0       X       not in model         Lauterborniella spp.       115       16.5       X       X       23.3       X         Mesocricotopus spp.       1       1.0       X       9.9       X         Microtendipes pedellus-type       75       10.3       X       13.5       X         Nanocladius spp.       2       1.0       X       12.5       X         Orthocladius spp.       1       1.0       X       15.2       X         Orthocladius spp.       1       1.0       X       13.6       X         Orthocladius spp.       2       1.0       X       14.5       X         Pagastiella spp.       34       3.8       X       14.5       X         Parachironomus varus-type       27       1.9       X       10.7       X         Paraphaenocladius spp.       1       1.9       X       10.7       X         Paratanytarsus spp.       95       13.3       X       10.7       X         Paratanytarsus spp.       5       1.0       X       Not in model       X         Paratendipes nudisquama-type       5       1.0       X       X       10.9	Glyptotendipes severini-type	15	1.1		х	Х	15.1		Х
Hyporygna spp.       2       1.0       X       not in model         Lauterborniella spp.       115       16.5       X       X       23.3       X         Mesocricotopus spp.       1       1.0       X       9.9       X         Microtendipes pedellus-type       75       10.3       X       13.5       X         Nanocladius spp.       2       1.0       X       12.5       X         Orthocladius spp.       1       1.0       X       15.2       X         Orthocladius spp.       1       1.0       X       13.6       X         Orthocladius spp.       2       1.0       X       14.5       X         Pagastiella spp.       34       3.8       X       14.5       X         Parachironomus varus-type       27       1.9       X       10.7       X         Paraphaenocladius spp.       1       1.9       X       10.7       X         Paratanytarsus spp.       95       13.3       X       10.7       X         Paratanytarsus spp.       5       1.0       X       Not in model       X         Paratendipes nudisquama-type       5       1.0       X       X       10.9	Heterotrissocladius maeri-type	1	1.1	Х			9.6	Х	
Lauerborniella spp.       115       16.5       X       X       23.3       X         Mesocricotopus spp.       1       1.0       X       9.9       X         Microtendijes pedellus-type       75       10.3       X       13.5       X         Nanocladius spp.       2       1.0       X       12.5       X         Orisus spp.       1       1.0       X       15.2       X         Orthocladius spp.       10       1.1       not in model       X       13.6       X         Orthocladius spp.       2       1.0       X       13.6       X       X       14.5       X         Pagastiella spp.       34       3.8       X       10.7       X       X       10.7       X         Parachironomus varus-type       27       1.9       X       13.5       X       X         Parathefferiella spp.       92       8.7       X       10.7       X       X       10.7       X         Parathefferiella spp.       11       1.9       X       not in model       X       X       X       X         Paratendipes nudisquama-type       5       1.0       X       X       12.9       X <td></td> <td>2</td> <td>1.0</td> <td>Х</td> <td></td> <td></td> <td>not in model</td> <td></td> <td></td>		2	1.0	Х			not in model		
Microtendipes pedellus-type         75         10.3         X         13.5         X           Nanocladius spp.         2         1.0         X         12.5         X           Omisus spp.         1         1.0         X         15.2         X           Orthocladinae spp.         10         1.1         not in model         7         13.6         X           Orthocladius spp.         2         1.0         X         13.6         X         14.5         X           Pagastiella spp.         34         3.8         X         14.5         X         X           Parachironomus varus-type         27         1.9         X         13.5         X         X           Paraphaenocladius spp.         92         8.7         X         X         10.7         X           Paraphaenocladius spp.         9         13.3         X         12.8         X           Paraphaenocladius spp.         95         13.3         X         12.8         X           Paratendipes nudisquama-type         5         1.0         X         X         not in model         X           Polypedilum nubifer-type         16         1.9         14.9         X         X <td>Lauterborniella spp.</td> <td>115</td> <td>16.5</td> <td></td> <td>х</td> <td>Х</td> <td>23.3</td> <td></td> <td>Х</td>	Lauterborniella spp.	115	16.5		х	Х	23.3		Х
Microtendipes pedellus-type       75       10.3       X       13.5       X         Nanocladius spp.       2       1.0       X       12.5       X         Omisus spp.       1       1.0       X       15.2       X         Orthocladina spp.       10       1.1       not in model       T       13.6       X         Orthocladius spp.       2       1.0       X       14.5       X         Pagastiella spp.       34       3.8       X       14.5       X         Parachironomus varus-type       27       1.9       X       13.5       X         Paraphaenocladius spp.       1       1.9       X       10.7       X         Paraphaenocladius spp.       1       1.9       X       10.7       X         Paraphaenocladius spp.       1       1.9       X       10.7       X         Paratendipes nudisquama-type       5       1.0       X       X       10.9       X         Polypedilum nubifer-type       16       1.9       14.9       X         Polypedilum nubifer-type       10       2.1       X       X       13.4       X	Mesocricotopus spp.	1	1.0	Х			9.9	Х	
Omisus spp.       1       1.0       X       15.2       X         Orthocladinae spp.       10       1.1       not in model       not in model         Orthocladius spp.       2       1.0       X       13.6       X         Pagastiella spp.       34       3.8       X       14.5       X         Parachironomus varus-type       27       1.9       X       13.5       X         Parachironomus varus-type       27       1.9       X       10.7       X         Parakiefferiella spp.       92       8.7       X       X       10.7       X         Parathaenocladius spp.       1       1.9       X       not in model       X       X         Paratanytarsus spp.       95       13.3       X       12.8       X       X         Paratendipes nudisquama-type       5       1.0       X       X       not in model       X         Polypedilum nubifer-type       16       1.9       14.9       X       X       X       X       X         Polypedilum nubifer-type       10       2.1       X       X       13.4       X       X	Microtendipes pedellus-type	75	10.3		х		13.5	Х	
Omisus spp.       1       1.0       X       15.2       X         Orthocladinae spp.       10       1.1       not in model       not in model         Orthocladius spp.       2       1.0       X       13.6       X         Pagastiella spp.       34       3.8       X       14.5       X         Parachironomus varus-type       27       1.9       X       13.5       X         Parachironomus varus-type       27       1.9       X       10.7       X         Parakiefferiella spp.       92       8.7       X       X       10.7       X         Parathaenocladius spp.       1       1.9       X       not in model       X       X         Paratanytarsus spp.       95       13.3       X       12.8       X       X         Paratendipes nudisquama-type       5       1.0       X       X       not in model       X         Polypedilum nubifer-type       16       1.9       14.9       X       X       X       X       X         Polypedilum nubifer-type       10       2.1       X       X       13.4       X       X	Nanocladius spp.	2	1.0	Х			12.5	Х	
Orthocladius spp.       2       1.0       X       13.6         Pagastiella spp.       34       3.8       X       14.5       X         Parachironomus varus-type       27       1.9       X       13.5       X         Parachironomus varus-type       27       1.9       X       13.5       X         Parachironomus varus-type       27       1.9       X       X       10.7       X         Parakiefferiella spp.       92       8.7       X       X       10.7       X         Paraphaenocladius spp.       1       1.9       X       not in model         Paratendipes nudisquama-type       5       1.0       X       X       not in model         Pentaneurini spp.       16       1.9       14.9       X         Polypedilum nubifer-type       116       23.2       X       X       13.4       X         Polypedilum nubeculosum-type       10       2.1       X       X       18.5       X	Omisus spp.	1	1.0	Х			15.2		Х
Pagastiella spp.       34       3.8       X       14.5       X         Parachironomus varus-type       27       1.9       X       13.5       X         Parachironomus varus-type       27       1.9       X       13.5       X         Parachironomus varus-type       92       8.7       X       X       10.7       X         Parachironomus spp.       92       8.7       X       X       10.7       X         Paraphaenocladius spp.       1       1.9       X       not in model       X         Paratendipes nudisquama-type       5       1.0       X       X       not in model         Pentaneurini spp.       16       1.9       X       not in model       X         Polypedilum nubifer-type       116       23.2       X       X       13.4       X         Polypedilum nubeculosum-type       10       2.1       X       X       18.5       X	Orthocladinae spp.	10	1.1				not in model		
Parachironomus varus-type271.9X13.5XParakiefferiella spp.928.7XX10.7XParaphaenocladius spp.11.9Xnot in modelParatanytarsus spp.9513.3X12.8XParatendipes nudisquama-type51.0XXnot in modelPentaneurini spp.161.914.9XPolypedilum nubifer-type11623.2XX13.4XPolypedilum nubeculosum-type102.1XX18.5X	Orthocladius spp.	2	1.0	Х			13.6		
Parakiefferiella spp.928.7XX10.7XParaphaenocladius spp.11.9Xnot in modelParatanytarsus spp.9513.3X12.8XParatendipes nudisquama-type51.0XXnot in modelPentaneurini spp.161.914.9XPolypedilum nubifer-type11623.2XX13.4XPolypedilum nubeculosum-type102.1XX18.5X	Pagastiella spp.	34	3.8		х		14.5		Х
Paraphaenocladius spp.11.9Xnot in modelParatanytarsus spp.9513.3X12.8XParatendipes nudisquama-type51.0XXnot in modelPentaneurini spp.161.914.9XPolypedilum nubifer-type11623.2XX13.4XPolypedilum nubeculosum-type102.1XX18.5X	Parachironomus varus-type	27	1.9		х		13.5	Х	
Paraphaenocladius spp.11.9Xnot in modelParatanytarsus spp.9513.3X12.8XParatendipes nudisquama-type51.0XXnot in modelPentaneurini spp.161.914.9XPolypedilum nubifer-type11623.2XX13.4XPolypedilum nubeculosum-type102.1XX18.5X	Parakiefferiella spp.	92				Х	10.7		
Paratanytarsus spp.       95       13.3       X       12.8       X         Paratanytarsus spp.       5       1.0       X       X       not in model         Paratendipes nudisquama-type       5       1.0       X       X       not in model         Pentaneurini spp.       16       1.9       14.9       X         Polypedilum nubifer-type       116       23.2       X       X       13.4       X         Polypedilum nubeculosum-type       10       2.1       X       X       18.5       X	Paraphaenocladius spp.			Х			not in model		
Paratendipes nudisquama-type51.0XXnot in modelPentaneurini spp.161.914.9XPolypedilum nubifer-type11623.2XX13.4XPolypedilum nubeculosum-type102.1XX18.5X	Paratanytarsus spp.	95			х			Х	
Pentaneurini spp.161.914.9XPolypedilum nubifer-type11623.2XX13.4XPolypedilum nubeculosum-type102.1XX18.5X	Paratendipes nudisquama-type			Х					
Polypedilum nubifer-type11623.2XX13.4XPolypedilum nubeculosum-type102.1XX18.5X	Pentaneurini spp.								Х
Polypedilum nubeculosum-type 10 2.1 X X 18.5 X					х	Х		Х	
									Х
	Procladius spp.	114	22.1			X	14.5		X

Psectrocladius septentrionalis-type	49	10.0		Х	12.2	Х	
Psectrocladius sordidellus-type	111	18.1		Х	12.4	Х	
Pseudorthocladius spp.	1	1.0	Х		22.6		Х
Sergentia coracina-type	57	4.8			25.9		Х
Stempellina spp.	72	7.9			15.8		Х
Stempelinella spp.	43	6.5			14.6		Х
Stenochironomus spp.	1	1.0	Х		not in model		
Stictochironomus spp.	2	1.0	Х		7.9	Х	
Tanytarsus spp.	110	15.6		Х	12.6	Х	
Tanytarsus chinyensis-type	8	1.9		Х	15.1		Х
Tanytarsus glabrescens-type	114	29.9		Х	15.6		Х
Tanytarsus lugens-type	14	2.1			13.6		
Tanytarsus mendax-type	116	21.4		Х	14.1		Х
Tanytarsus pallidicornis-type	116	24.3		Х	13.9		Х
Tanytarsus with spur on antenna	21	3.9		Х	15.2		Х
Thiennemaniella spp.	13	1.9			not in model		
Thiennemanyia spp.	10	3.1			12	Х	
Zalutschia mucronata-type	1	0.9	Х		13	Х	
Zalutschia zalutschicola-type	4	1.1	Х		14.7		Х
Number of taxa not included in the training set					7		
Maximum temperature (°C)					25.9		
Minimum temperature (°C)					7.9		

## **Figure captions**

Fig. 1 Map of Quebec (Canada) showing the location of study sites and the bioclimatic domains

Fig. 2 Age-depth model and sediment accumulation rates for the Lac Lili core

**Fig. 3** Chironomid stratigraphy of Lac Lili. Chironomid percentage changes for the taxa present in more than six samples, the number of head capsules per sample, percentage of littoral taxa (based on Brooks et al. 2007), percentage of eutrophic taxa (based on Brooks et al. 2007) and the percentage of colder-than-today taxa (based on the East Canadian temperature optima). PCA axis 1 and 2 are the scores of the principal component analysis made on the percentages of chironomid taxa with percentages >1%. Vertical lines displayed in the percentage of littoral taxa and the percentage of colder-than-today taxa are the Holocene average. The vertical lines in the PCA axis 1 and 2 scores are the 0 score. The horizontal lines represent four significant zones identified using CONISS and the broken stick method. The chironomid taxa are ordered by their temperature optima in the training set. The black arrow identifies the 27 taxa with temperature optima equal to or warmer than 13.6°C

**Fig. 4 a** Temperature reconstruction using chironomids preserved in the sediment of Lac Lili. The black line is the temperature reconstruction and the grey dots are the sample-specific errors. The vertical dotted lines are the zones identified in the chironomid stratigraphy. The horizontal grey lines are the average inferred temperatures within the zones. The black squares identify the eight samples with no analogues. **b** Chironomid-inferred temperature anomaly (°C) in relation to  $15^{\circ}$ C (1961–1990) and  $15.4^{\circ}$ C (1981–2010). Inferences colder than reference periods are illustrated by large circles. The horizontal dotted bars are the mean anomaly compared to 15 and  $15.4^{\circ}$ C

Fig 5. Summary of changes in Lac Lili and Lac Aurélie

**Electronic Supplementary Material Fig. S1** Results of the transfer function. **Upper panel**: observed versus inferred temperatures of the lakes in the training set. **Lower panel**: Observed temperature versus residuals. Residuals are the differences between observed and reconstructed temperature. whereas the 1:1 shows the theoretical perfect correspondence between observed and inferred temperatures. From the 1:1 relationship, we can see which samples overestimate or underestimated the true temperature and from the residuals we can see by how much (°C). Lakes warmer than 15°C were generally under-estimated and there were no lakes between 16.8 and 18.9°C



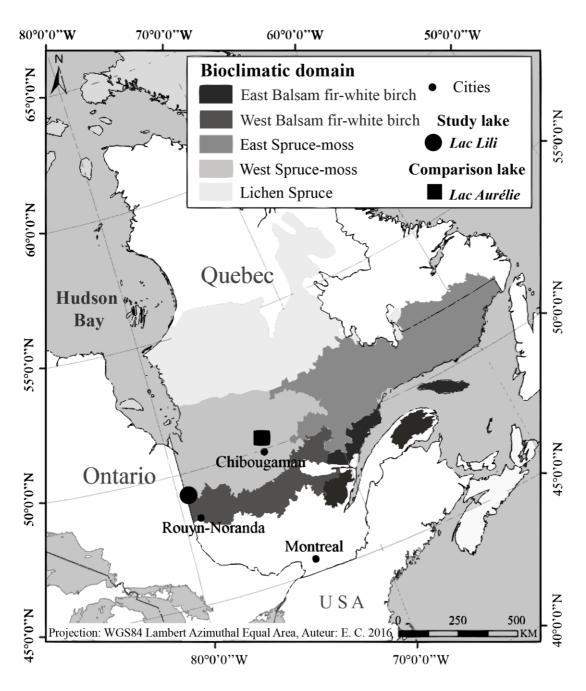


Fig. 1

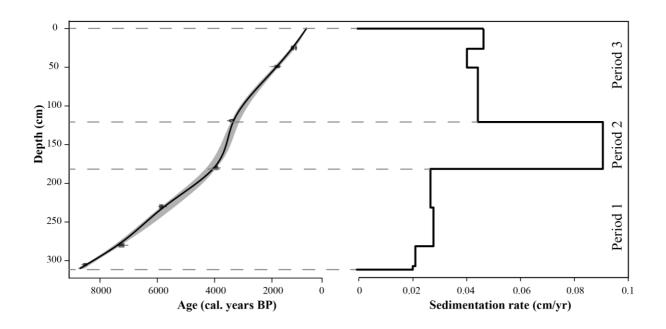
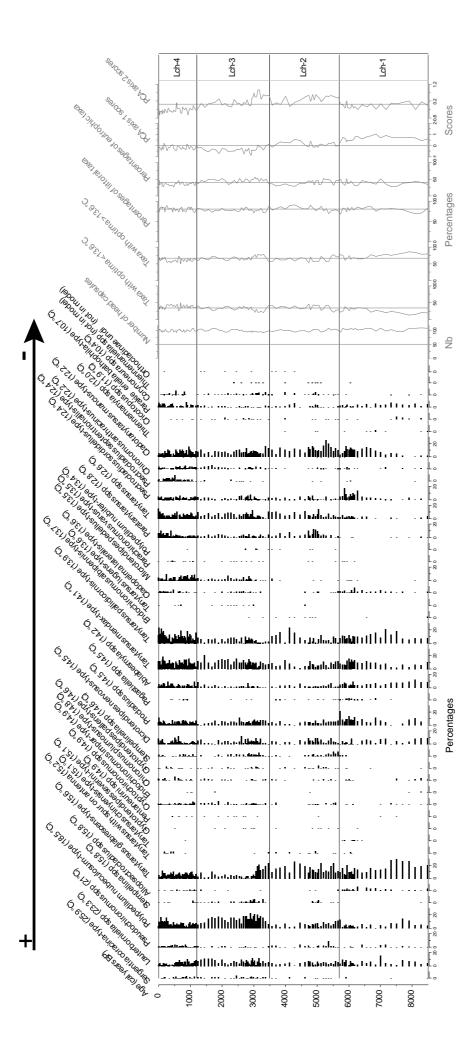


Fig. 2





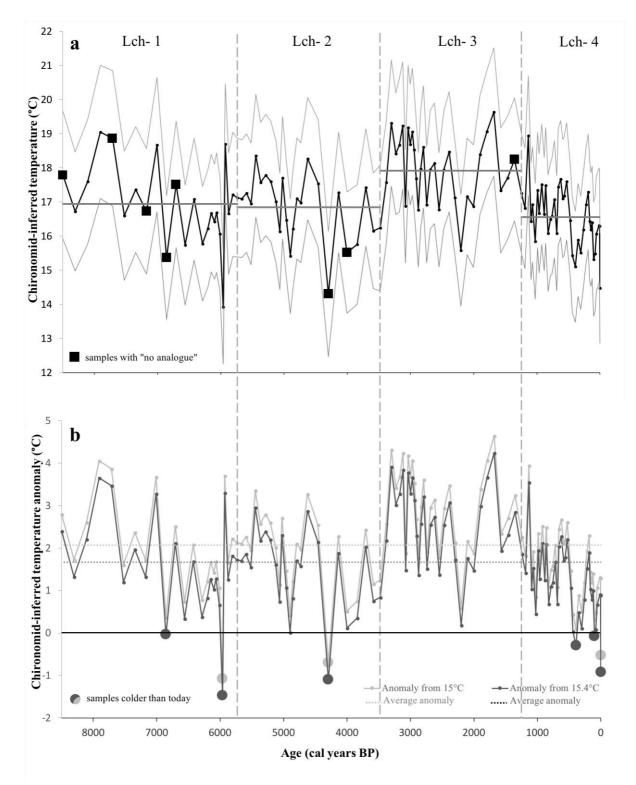


Fig. 4

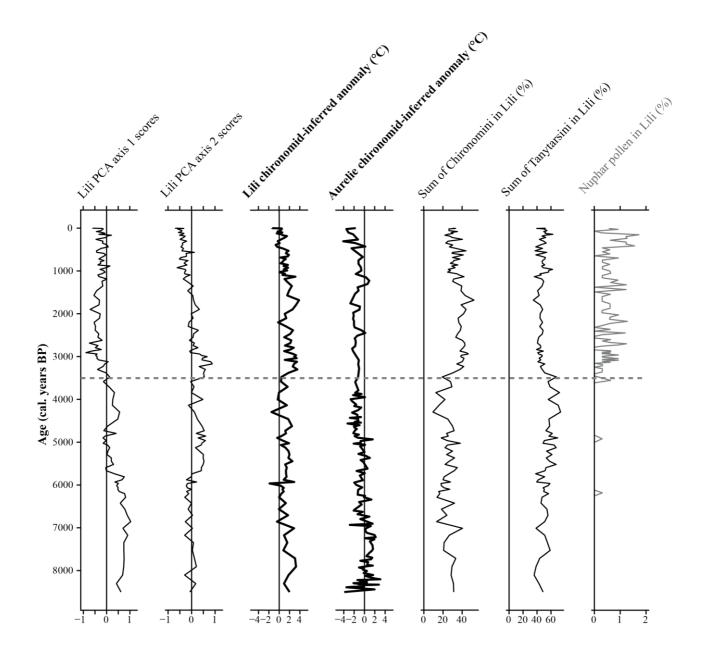


Fig. 4

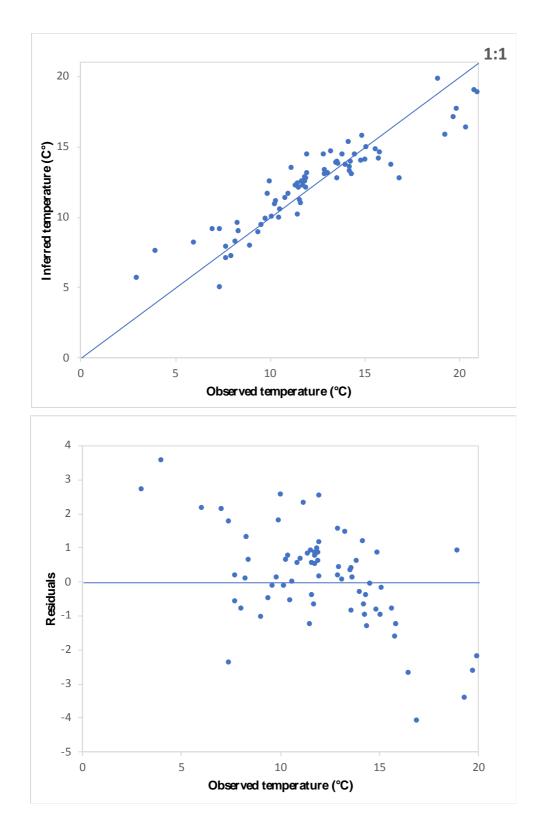


Fig. Supplementary data 1