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### ► To cite this version:

Fabrice Lopez, Guillaume Charbonnier, Yasmina Kermezli, Mohamed Belhocine, Quentin Ferré, et al.. Explore, edit and leverage genomic annotations using Python GTF toolkit. *Bioinformatics*, 2019, 10.1093/bioinformatics/btz116 . hal-02078147

**HAL Id: hal-02078147**

**<https://amu.hal.science/hal-02078147>**

Submitted on 25 Mar 2019

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## Genome analysis

# Explore, edit and leverage genomic annotations using Python GTF toolkit

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Associate Editor: XXXXXXXX

Received on XXXXX; revised on XXXXX; accepted on XXXXX

### Abstract

**Motivation:** While Python has become very popular in bioinformatics, a limited number of libraries exist for fast manipulation of gene coordinates in Ensembl GTF format.

**Results:** We have developed the GTF toolkit Python package (pygtf), which aims at providing easy and powerful manipulation of gene coordinates in GTF format. For optimal performances, the core engine of pygtf is a C dynamic library (libgtf) while the Python API provides usability and readability for developing scripts. Based on this Python package, we have developed the gtftk command line interface that contains 57 sub-commands (v0.9.10) to ease handling of GTF files. These commands may be used to (i) perform basic tasks (e.g. selections, insertions, updates or deletions of features/keys), (ii) select genes/transcripts based on various criteria (e.g. size, exon number, TSS location, intron length, GO terms) or (iii) carry out more advanced operations such as coverage analyses of genomic features using bigWig files to create faceted read-coverage diagrams. In conclusion, the pygtf package greatly simplifies the annotation of GTF files with external information while providing advance tools to perform gene analyses.

**Availability:** pygtf and gtftk have been tested on Linux and MacOSX and are available from <https://github.com/dputhier/pygtf> under the MIT license. The libgtf dynamic library written in C is available from <https://github.com/dputhier/libgtf>

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## 1 Introduction

Several formats exist to store genomic features. The standard BED format stores basic information (chromosome, start, end, name, score and strand) related to generic genomic features (BED6) or composite genomic features (BED12). The GTF/GFF2 format (thereafter referred as GTF) can describe more exhaustively defined genomic features (genes, transcripts, exons...) by taking advantage of the ‘attributes’ column which contains a set of keys/values to store various kinds of annotations. Some composition relationships are implicitly declared in the GTF file making it possible to describe, for instance, the exons of the transcripts corresponding to a gene. This relationship is more explicit in the GFF3 format that can be viewed as a directed acyclic graph with nodes corresponding to features (gene, transcript, exon...) and edges corresponding to part-of relationships. Only few libraries are specifically dedicated to GTFs and most of them propose very focused tasks. The GenomeTools suite is a collection of bioinformatic tools based on the libgenometools C library that handle GTF

and GFF3 formats (Gremme *et al.*, 2013). However, this library extends well beyond these annotation formats and the developing framework may appear rather complicated for naive developers as it requires deep knowledge of C programming language. Regarding R/Bioconductor, the tracklayer provides fast access to the GTF/GFF by providing the user with a GRanges object (Lawrence *et al.*, 2009).

While the Python language has gained lot of popularity among bioinformaticians, only a handful of tools are available for manipulating GTF files. The gffutils package can parse and store GTF/GFF files into SQLite databases. The creation of a subsequent hierarchical models of genomic features while highly useful can be relatively time consuming. We developed the pygtf package with the objective to provide a fast and readable way to load and manipulate GTF files within Python scripts. This package comes with the gtftk command line interface (CLI) that provide various operations to write workflows based on GTF files.

```

from pygtf.gtf_interface import GTF
from pygtf.utils import get_example_file

# Create a GTF instance
cod_pot = get_example_file('mini_real', 'tab')[0]
gn_info = get_example_file('mini_real', 'genome')[0]
gtf = GTF(get_example_file('mini_real', 'gtf.gz'))

# Get a BedTool object containing the TSSs
# of the selected transcripts
tss = gtf.select_by_key('gene_biotype', 'lincRNA')
).select_by_transcript_size(min=200)
).select_by_number_of_exons(min=2)
).add_attr_from_file(feats='transcript',key='transcript_id',
                    has_header=True, new_key='coding_pot',
                    inputfile=cod_pot)
).eval_numeric('coding_pot < 0.2', na_omit='.,?')
).get_tss(name=['transcript_id', 'gene_name', 'gene_id'])
).slop(s=True, l=1000,r=1000, g=gn_info)

```

**Fig. 1.** Use case for the pygtf package. These few lines of codes are used to extract the promoter region ([-1000, 1000] around the TSS) of lincRNAs, with the conditions that the transcripts have size greater than 200nt, at least two exons and a coding potential (assessed by CPAT and joined from an external file) below 0.2. (Wang et al., 2013)

## 2 Implementation

### 2.1 The core libgtf C library

The core of the package is written in C and exposed through a dynamic library called libgtf. The GTF format is represented without hierarchical relationships to maximize performances. More complex operations are carried out by the libgtf Python client.

### 2.2 The pygtf Python package

The GTF class of pygtf comes with a large number of methods. Most of these methods return a new GTF object so that they can be chained intuitively. This object can also produce two additional objects from the gtf library including: a TAB object (representation of a matrix) and a FASTA object (representation of a FASTA file). The GTF object is integrated within the scientific Python ecosystem and can produce *pybedtools.BedTool* objects, *Bio.SeqRecord* generators or a *pandas.DataFrame* (Quinlan, 2014; Cock et al., 2009; McKinney, 2010). A typical use case is proposed in Figure 1 where the transcription start site (TSS) coordinates of lincRNAs are extracted with the conditions that (i) the transcript size is above 200nt, (ii) the number of exons is greater than 2 (iii) and the coding potential (imported from a separated file) is lower than 0.2. The TSSs are then obtained using the *get\_tss()* method returning a *pybedtools.BedTool* object that can be used to extend coordinates by 1000 nucleotides in the 5' and 3' directions. Regarding performances, the human genome annotation in GTF format from Ensembl release 92 (~ 2.7.10<sup>6</sup> lines) is loaded in about 30 seconds while the creation of a hierarchical model using *gffutils* takes about 11 minutes (performed on Intel(R) Xeon(R) CPU E5-2640 v3, 2.60GHz). In addition, the search engine is also highly optimized since it takes 0.6 seconds to select all lincRNAs from the human genome.

### 2.3 The gtf CLI command-line interface

The pygtf package provides a gtf CLI with 57 subcommands. These subcommands can be used to: (i) download GTF files, (ii) edit them,

(iii) mine the GTF files in various ways (e.g. select transcripts by genomic/exonic/intronic size, number of exons, associated GO term...), (iv) annotate the GTF files (e.g. flagging divergent/convergent/overlapping transcripts...), (v) convert them to other formats or (vi) perform epigenomic analyses by producing faceted coverage diagrams through the plotnine Python package (i.e. the recently developed Python port of ggplot2).

## 3 Conclusion

The pygtf package and the associated gtf CLI provides a new way to easily handle gene coordinates with Python. They are regularly updated and users familiar with Python and/or command-line programs should quickly get comfortable and productive with (py)gtf. As the GTF/GFF format is now also used for storing regulatory features and variants, this paves the way for future developments of (py)gtf that could be an interesting framework for the integration of heterogeneous genomic data (Zerbino et al., 2018; Reese et al., 2010).

## Acknowledgements

We thank Jacques van Helden for helpful discussion.

## Funding

G.C. was supported by a fellowship from the “Fondation pour la Recherche Médicale” (FRM). S.S. and D.P. were supported by recurrent funding from INSERM and Aix Marseille Univ and by the Foundation for Cancer Research ARC (ARC PJA 20151203149) and A\*MIDEX (ANR-11-IDEX-0001-02), Plan Cancer 2015 (C15076AS) and Ligue contre le Cancer Equipe Labellisée. Y.K., was supported, by the Franco-Algerian partenariat Hubert Curien (PHC) Tassili (15MDU935).

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