

Supporting Information for

The Amazon River – a major source of organic plastic additives to the tropical North Atlantic?

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SUPPLEMENTARY TEXTS

Text S1: Method details concerning GC-MS and LC-HRMS analysis.

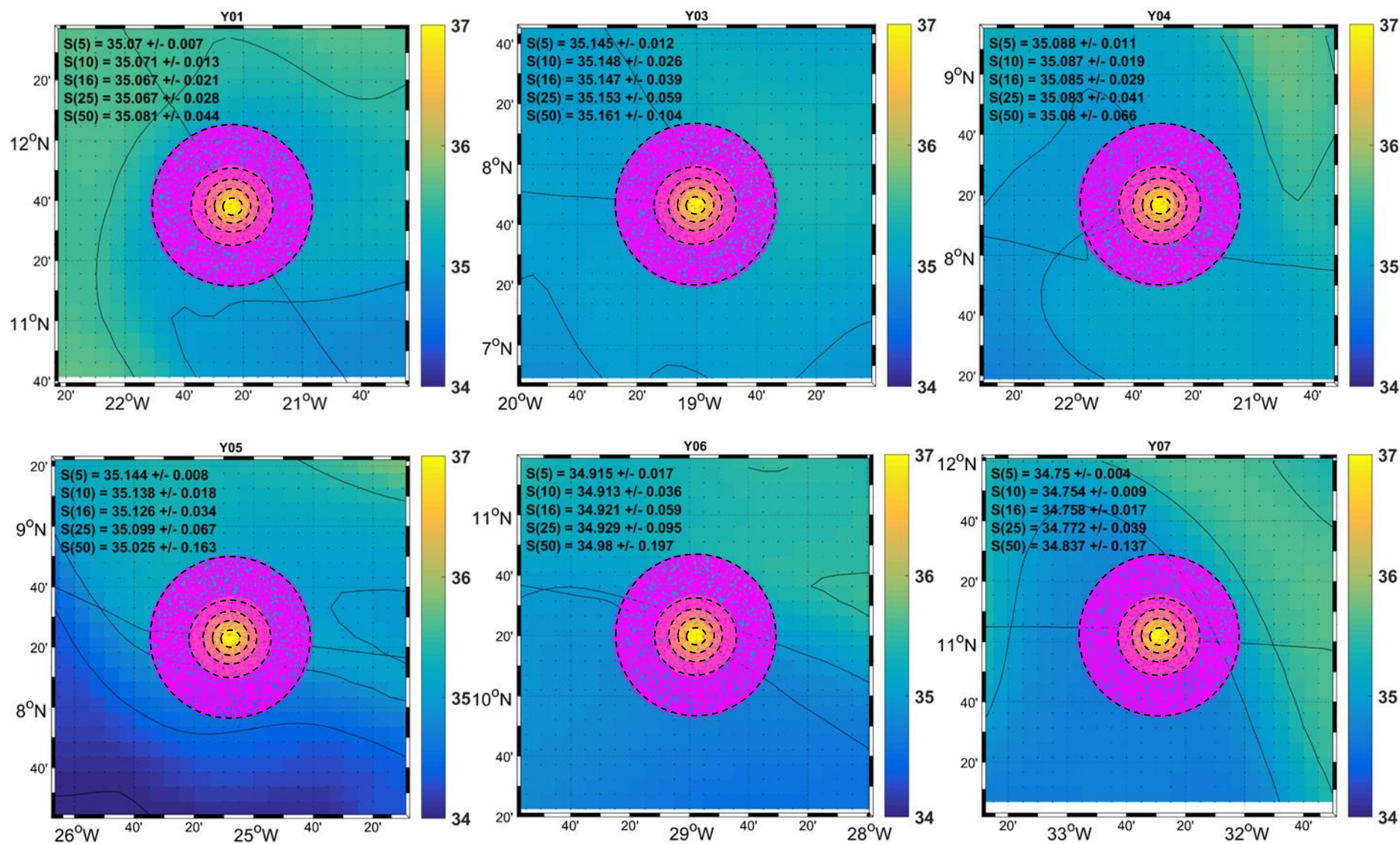
GC-MS analysis (Organophosphate ester analysis)

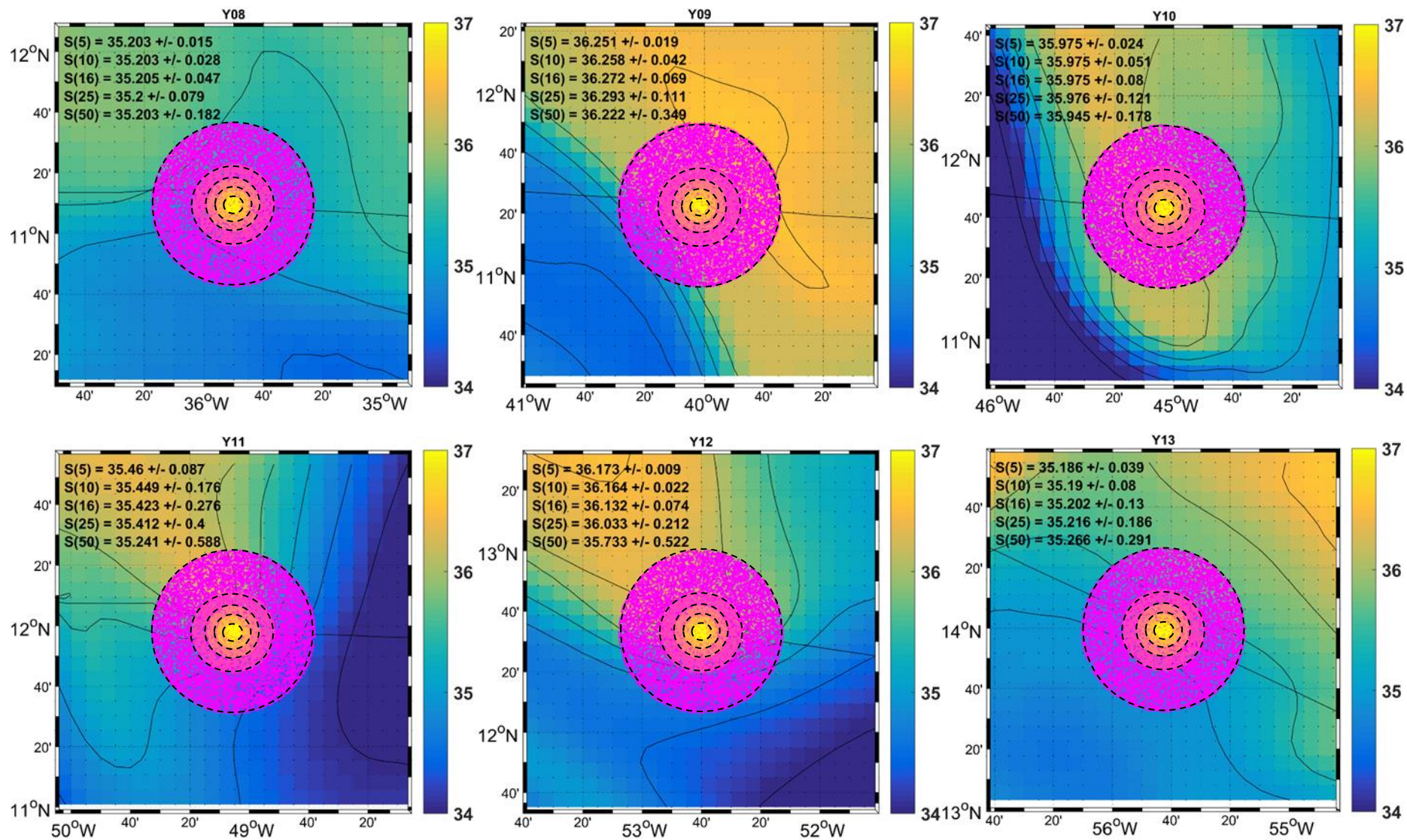
GC analyses were conducted with an Agilent 7820A Series GC coupled with an Agilent 5977E MS, operating in SIM and electron impact (EI, 70 eV) modes. The separation was achieved in a 30 m x 0.25 mm i.d. x 0.25 μm HP-5MS capillary column (Agilent J&W). All target contaminants were quantified by the internal standard (IS) procedure. The injection volume was 2 μL and the helium carrier gas flow was 1 mL min^{-1} . The temperatures of the MS transfer line, the ion source and the quadrupole was set to 300, 230 and 150 $^{\circ}\text{C}$, respectively. The following conditions were applied: injector temperature: 270 $^{\circ}\text{C}$ (splitless) and the oven was programmed from 90 $^{\circ}\text{C}$ to 132 $^{\circ}\text{C}$ at 3 $^{\circ}\text{C min}^{-1}$, to 166 $^{\circ}\text{C}$ at 10 $^{\circ}\text{C min}^{-1}$, to 175 at 1 $^{\circ}\text{C min}^{-1}$ (holding time 2 min), to 232 $^{\circ}\text{C}$ at 2 $^{\circ}\text{C min}^{-1}$, and then to 300 $^{\circ}\text{C}$ at 25 $^{\circ}\text{C min}^{-1}$ (holding time 5 min).

LC-QTOF analysis (bisphenol and perfluorinated compound analysis)

Ten microliters of the extract were injected for simultaneous quantification of BPs and PFCs using LC-electrospray ionization quadrupole time of flight mass spectrometry (LC-ESI-QTOF, Agilent 1290 Infinity LC system coupled with Agilent 6530 Accurate-Mass Q-TOF, Agilent Technologies, Les Ulis, France). The syringe was washed externally with MeOH for 20 s before injection to avoid contamination of the injection port or following samples. Separation was achieved on an Agilent Zorbax Eclipse XDB reversed phase column (50 mm x 2.1 mm, 1.8 μm), with the temperature set at 30 $^{\circ}\text{C}$. Elution was performed with MQ (A) and MeOH (B) under gradient conditions: 0 min 95:5 A/B, 1 min 95:5 A/B, 10 min 0:100 A/B, 15 min 0:100 A/B, 15.1 min 95:5 A/B, 20 min 95:5 A/B. ESI interface was operated in negative mode (3.5 kV capillary voltage), and MS TOF mass acquisition was performed in the range of 50-600 m/z at a rate of 1 spectrum s^{-1} . ESI parameters were set as follows: 300 $^{\circ}\text{C}$ gas temperature, 11 L min^{-1} drying gas, 40 psig nebulizer, 350 $^{\circ}\text{C}$ sheath gas temperature, 11 L min^{-1} sheath gas flow. TOF parameters were: 1500 V nozzle voltage, 175 V fragmentor voltage, 65 V skimmer voltage, 750 V octopole voltage. Ion chromatogram extraction was performed with 10 ppm mass tolerance, based on exact monoisotopic mass and retention time.

SUPPLEMENTARY FIGURES





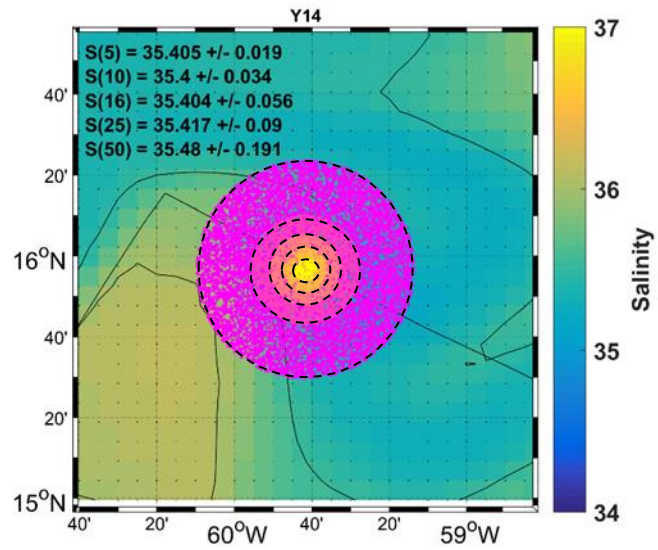


Figure S1: Comparisons between salinity maps of stations Y01-Y14 and different sizes of seeded particle patches (5 km, 10 km, 16 km, 25 km and 50 km). For each patch size, the mean salinity inside the patch and the standard deviation are reported. Black solid lines represent salinity contours for salinity values of 34, 35, 36 and 37. It can be observed that for a patch radius of 50 km, the particles are seeded over a high range of salinities, likely representing different water masses.

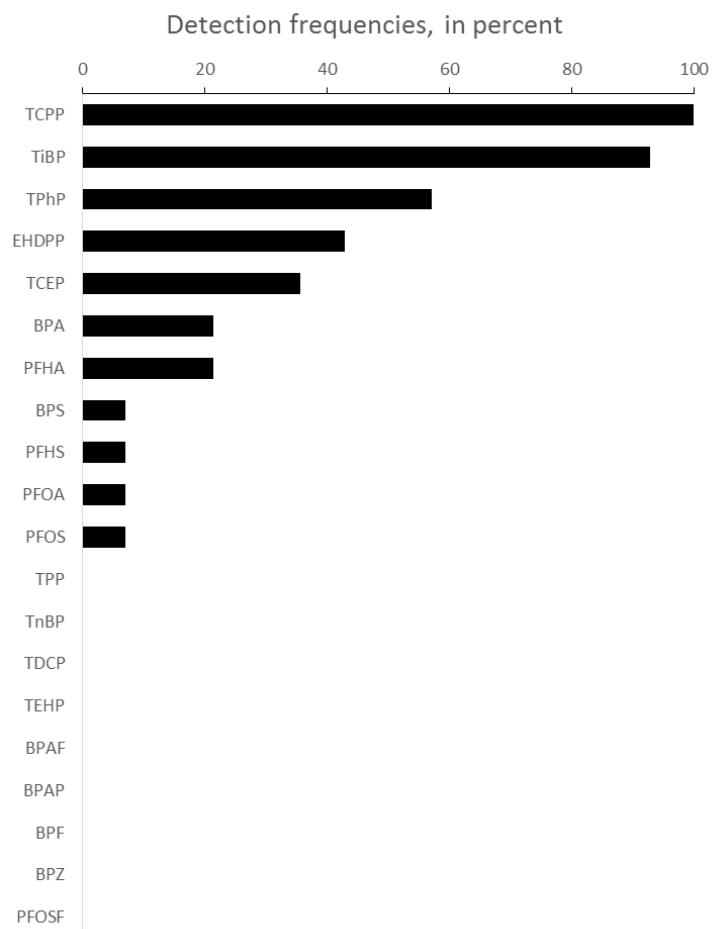
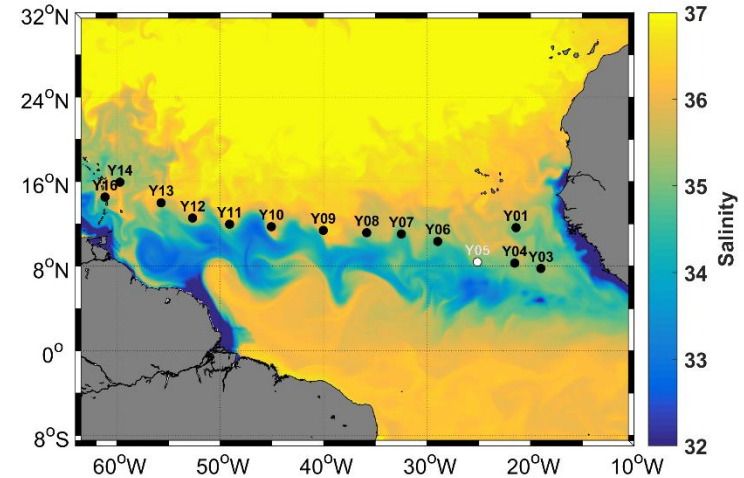
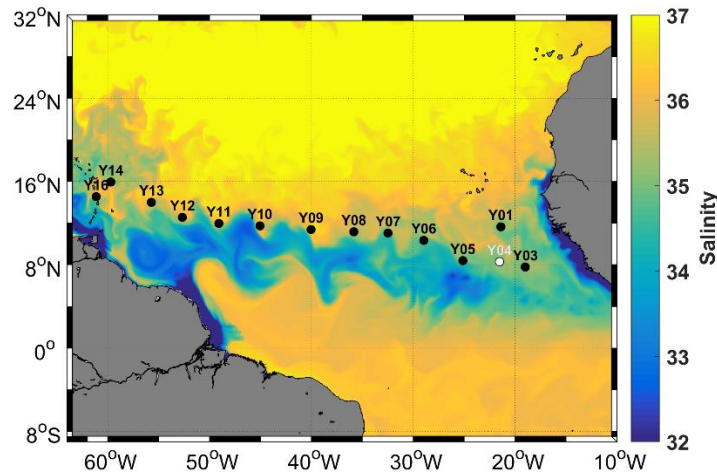
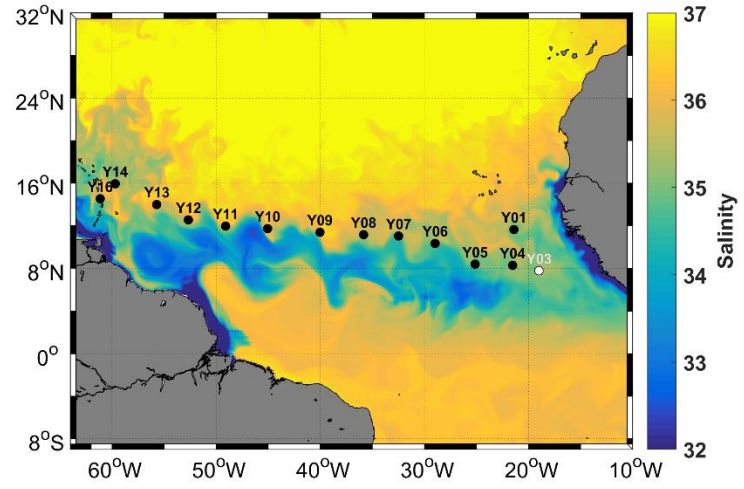
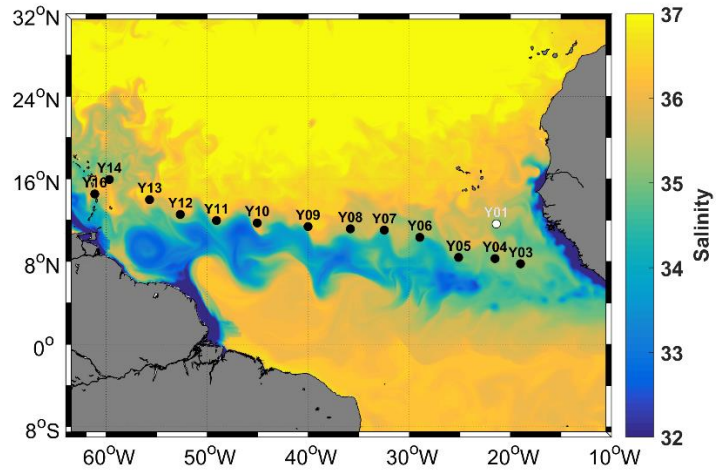
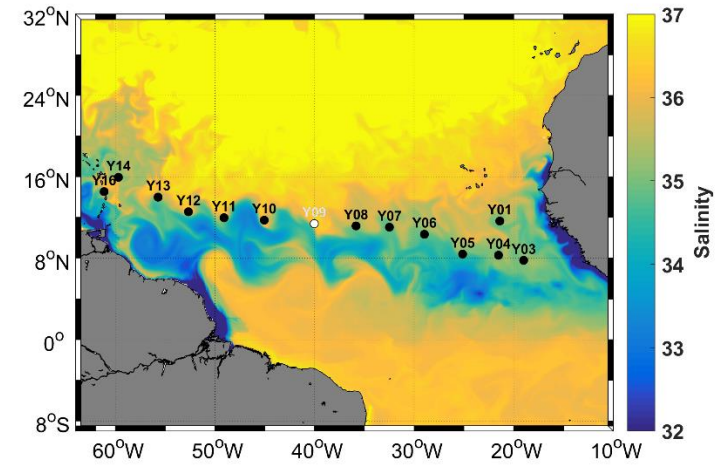
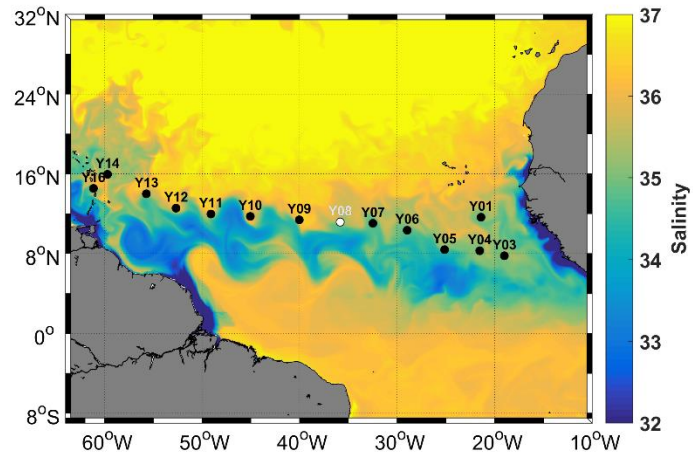
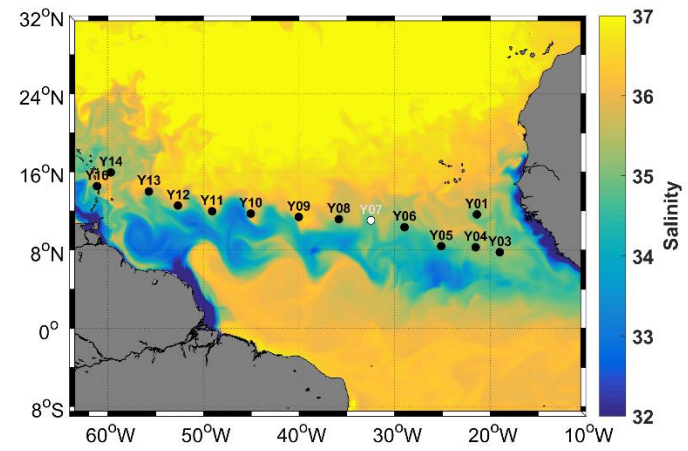
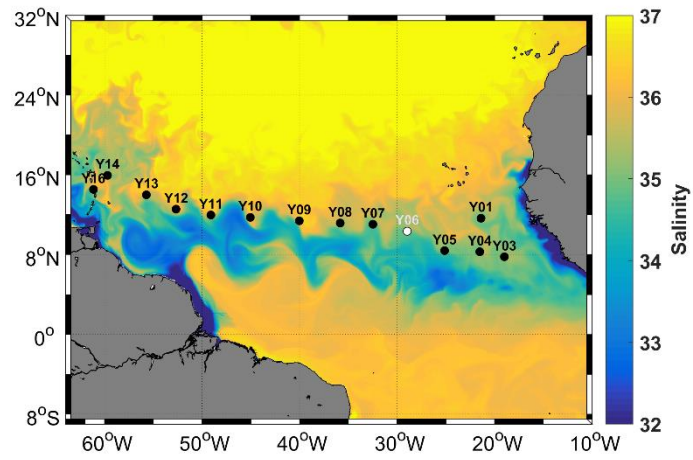
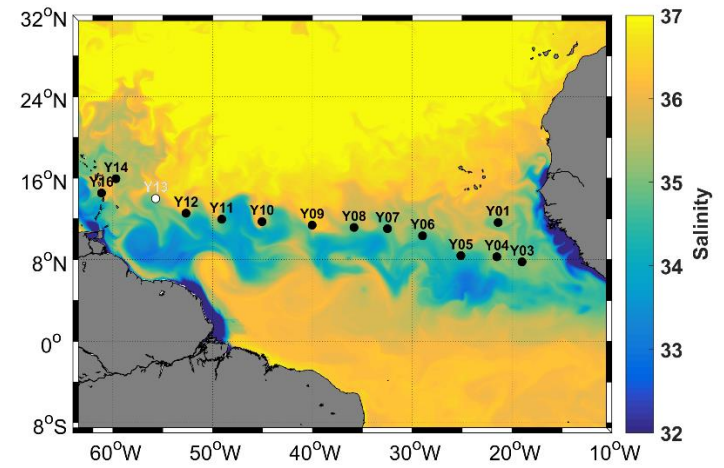
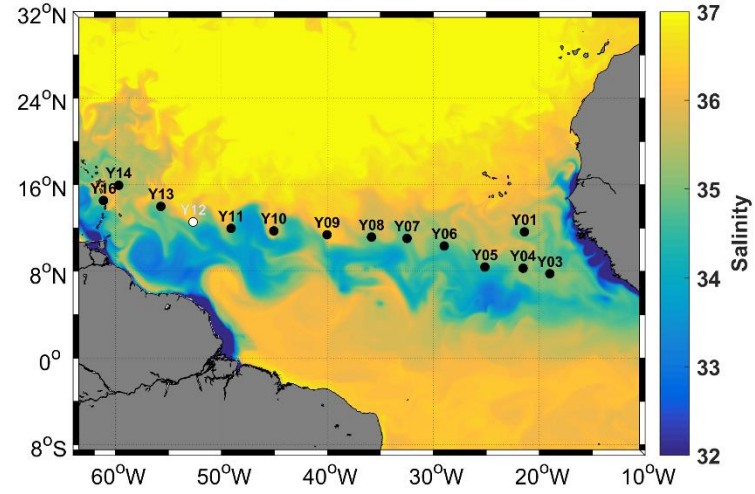
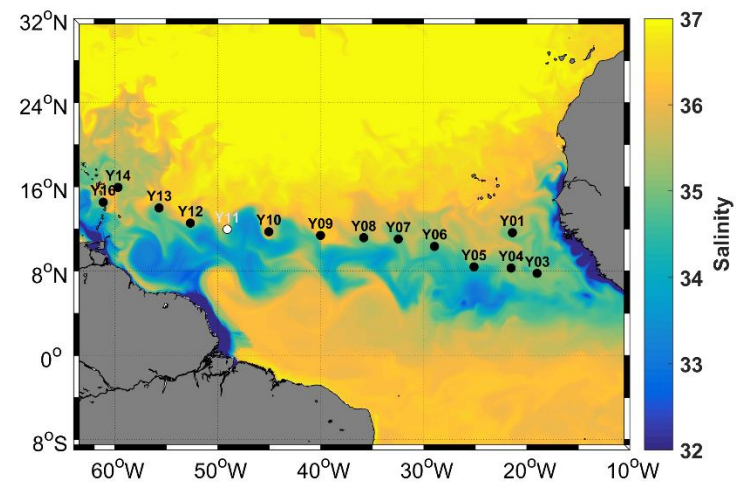
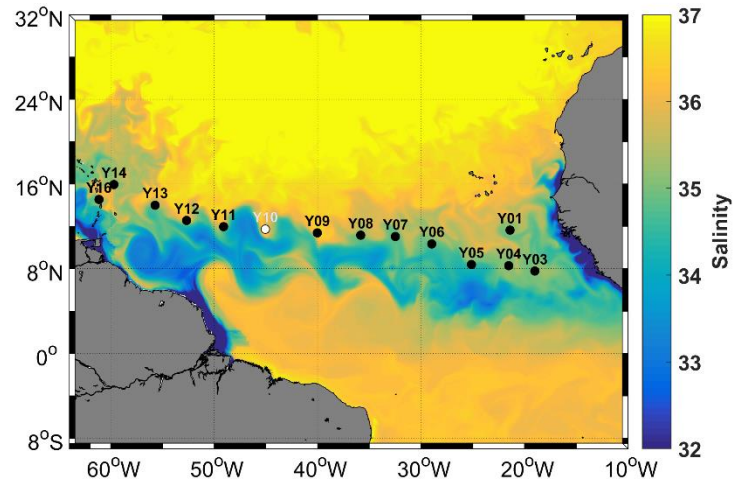


Figure S2: Detection frequencies (in %) of the 20 organic compounds analyzed. Abbreviations: TPP: Tripropylphosphate, TiBP: Tri-iso-butyl phosphate, TnBP: Tri-n-butyl phosphate, TCEP: Tris-(2-chloroethyl) phosphate, TCPP: Tris-(2-chloro, 1-methylethyl) phosphate, TDCP: Tris-(2-chloro-, 1-chloromethylethyl) phosphate, TPhP: Triphenyl phosphate, EHDPP: 2-ethylhexyl-diphenyl phosphate, TEHP: Tris(2-ethylhexyl) phosphate, BPA: Bisphenol A, BPAF: Bisphenol AF, BPAP: Bisphenol AP, BPF: Bisphenol F, BPS: Bisphenol S, BPZ: Bisphenol Z, PFHA: Perfluoro hexanoic acid, PFHS: Perfluorohexane sulfonic acid, PFOA: Perfluorooctanoic acid, PFOS: Perfluorooctane sulfonic acid, PFOSF: Perfluoro octane sulfonyl fluoride.







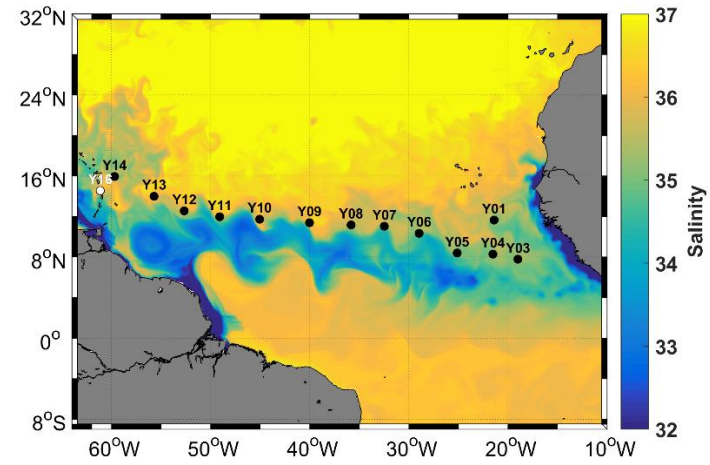
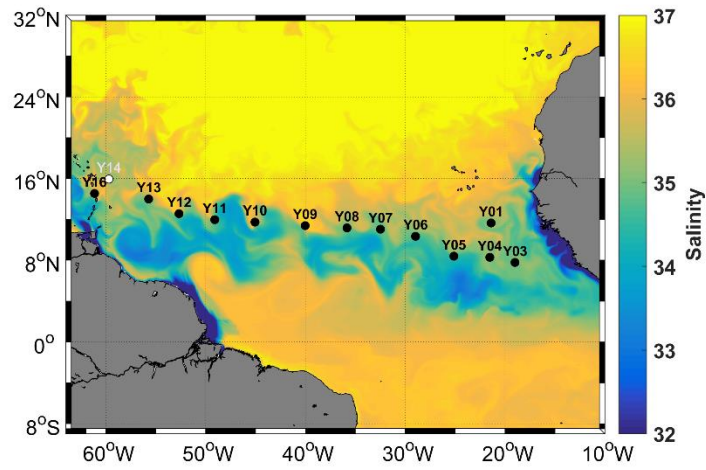


Figure S3: Spatial salinity distribution of the surface waters of the study area during each sampling day. The stations indicated in white are the ones sampled the day from which the MERCATOR salinity data originates.

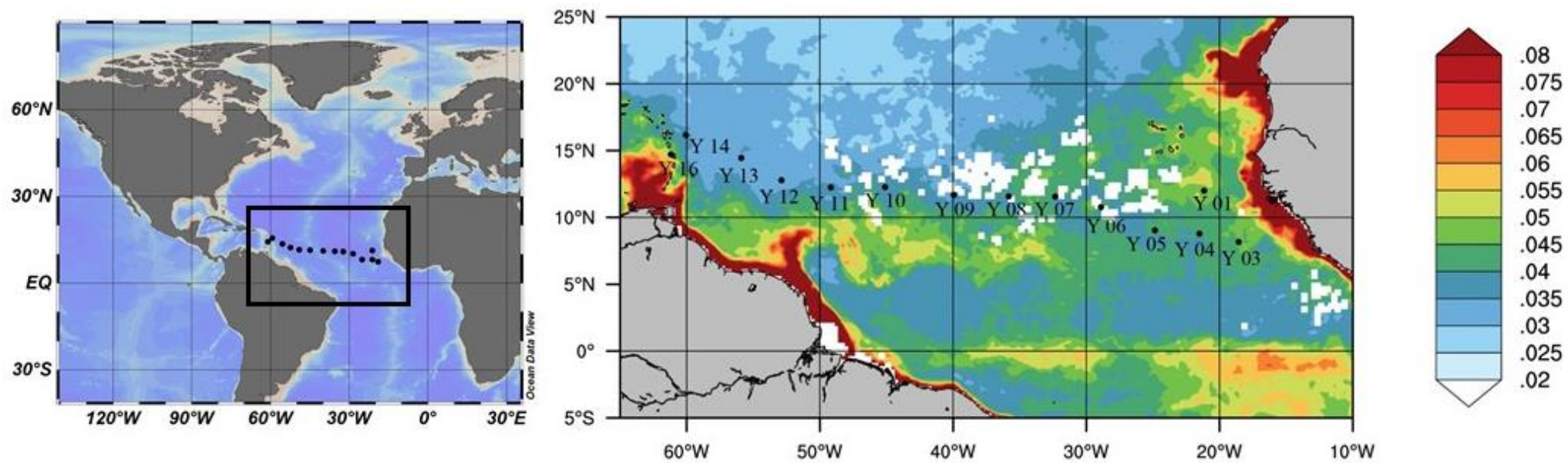
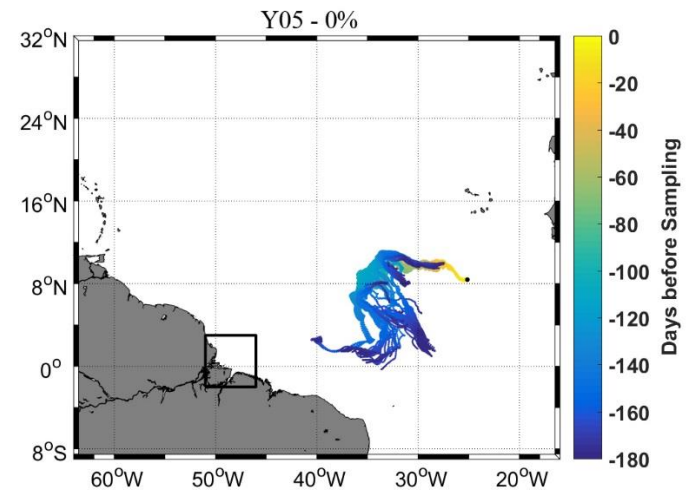
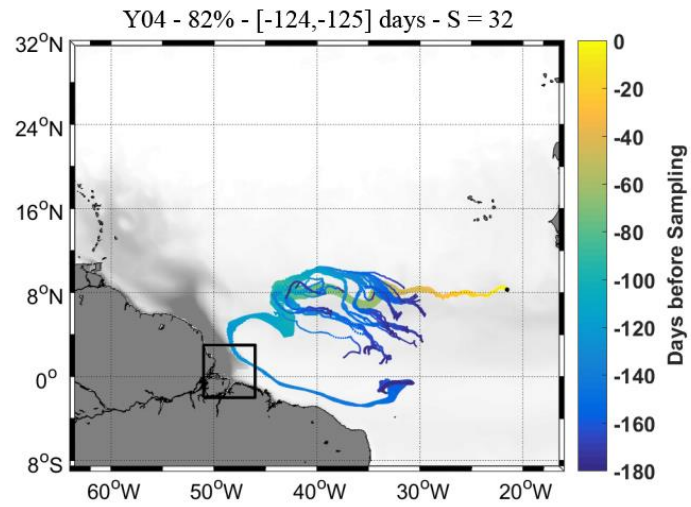
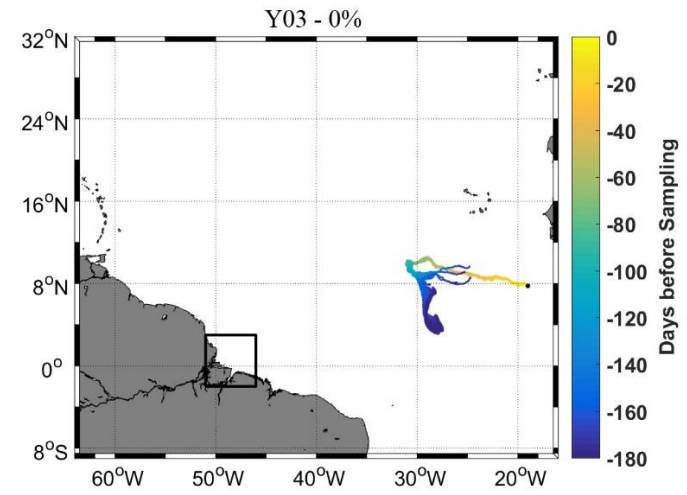
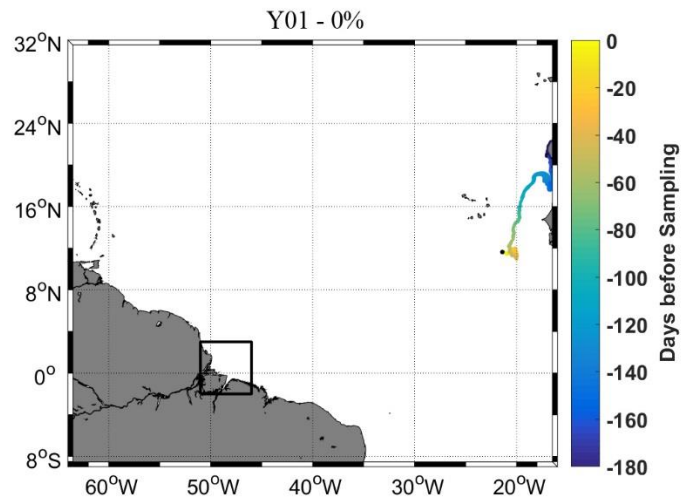
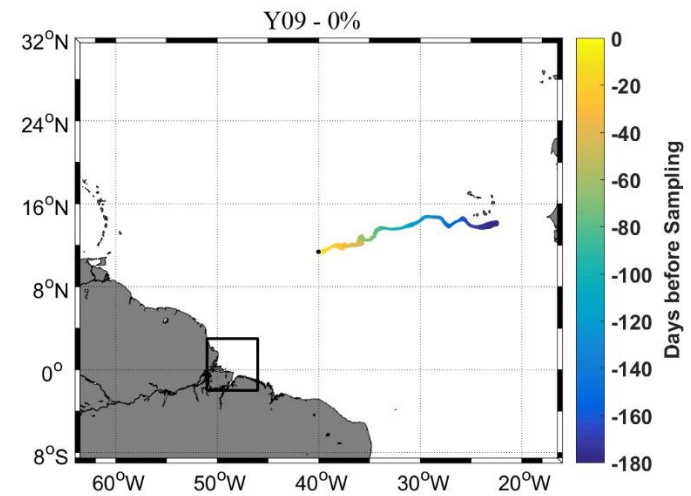
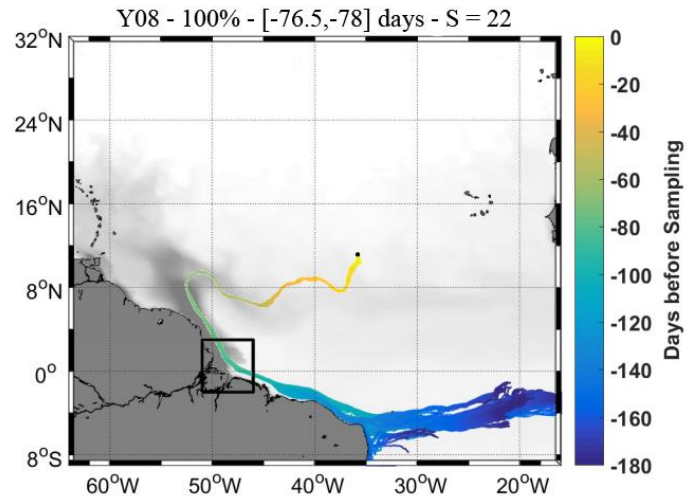
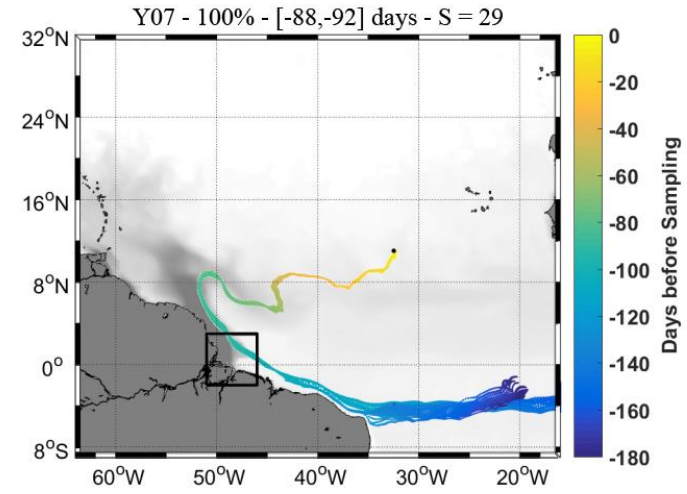
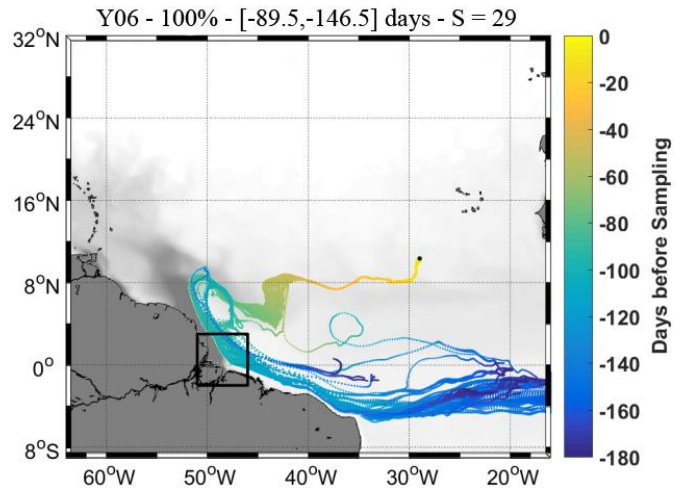
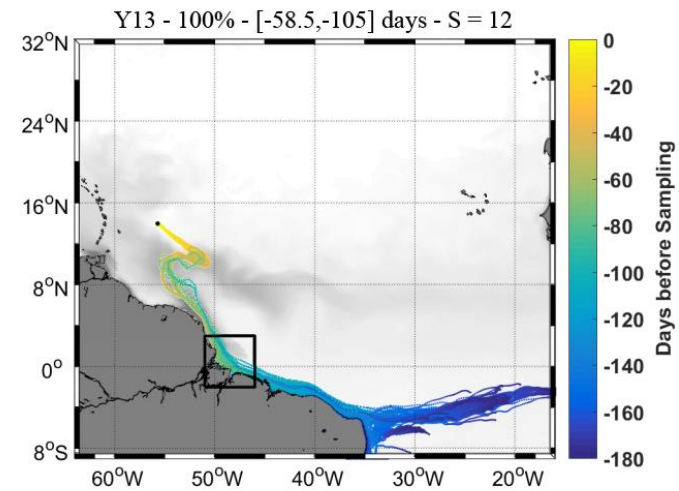
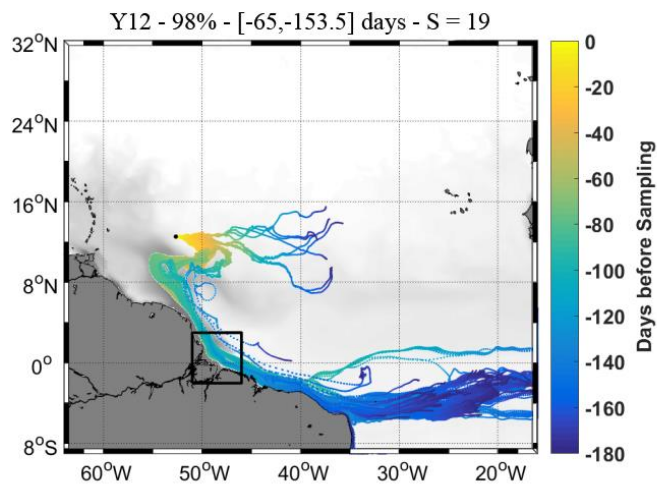
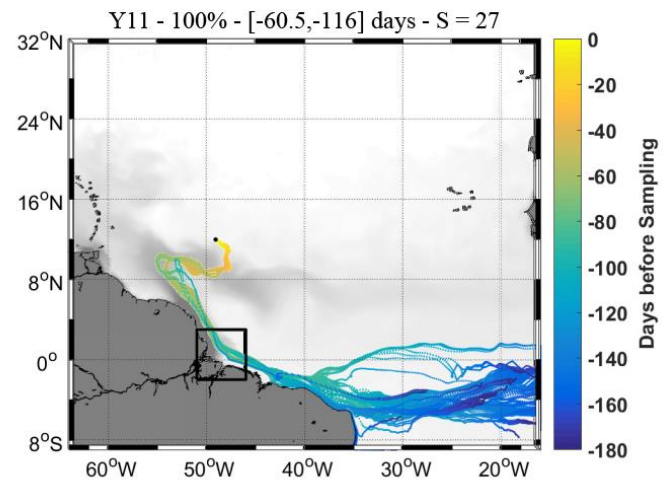
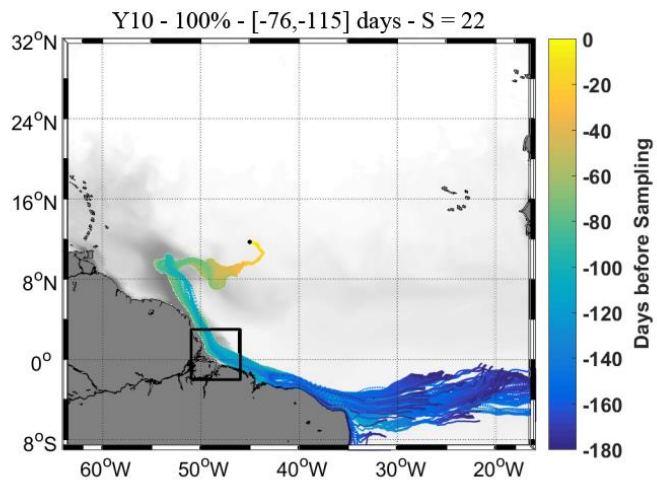


Figure S4: Map showing the position of the sampling area (left) and a zoom (right) showing the sampling stations with the spatial distribution of Kd490 (unit 1/m) in October 2017. Kd490 is a proxy for colored suspended matter and can be used to assess the chlorophyll a distribution.







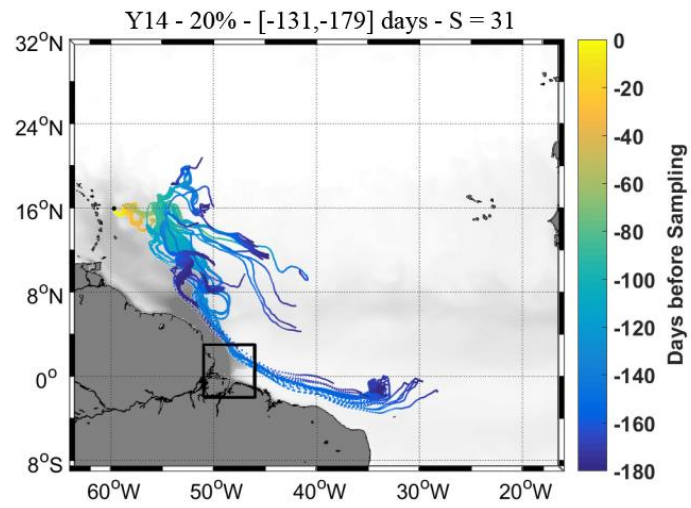
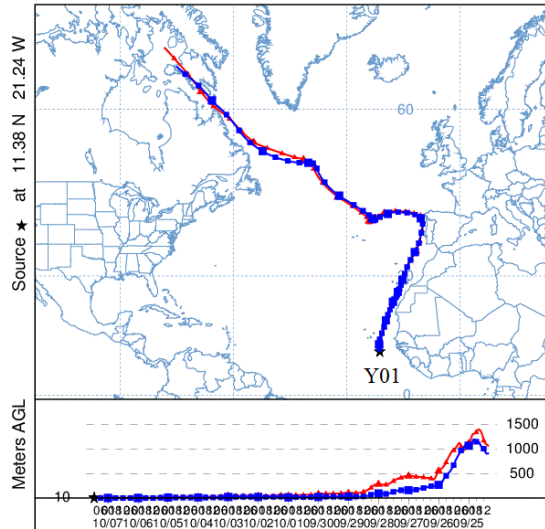
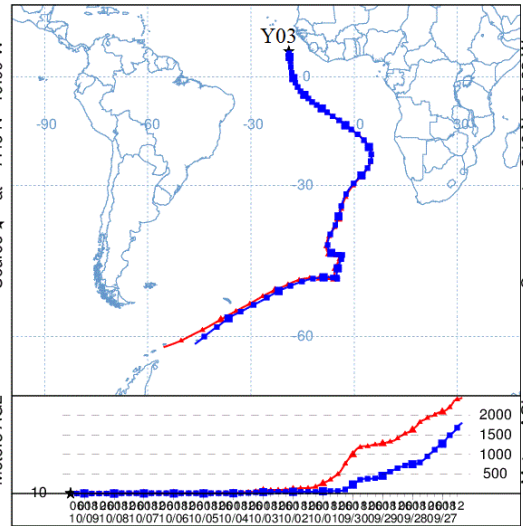


Figure S5: Water mass backwards trajectories of all stations (Y01-Y14, from upper left to lower right) except Y16, which is too close to the coast for reliable simulation results. Each map has a caption indicating the station ID, the percentage of particles that passed through the black box (indicating the Amazon Estuary) and the number of days that passed between the sampling of the water masses and their journey in the black box (indicated in brackets). The minimum of salinity (S) reached by particles during their journey in the black box is indicated as well as the river plume (in grey).

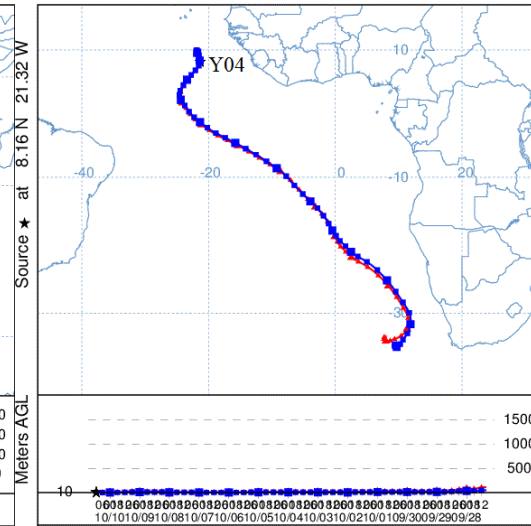
NOAA HYSPLIT MODEL
Backward trajectories ending at 1100 UTC 07 Oct 17
GDAS Meteorological Data



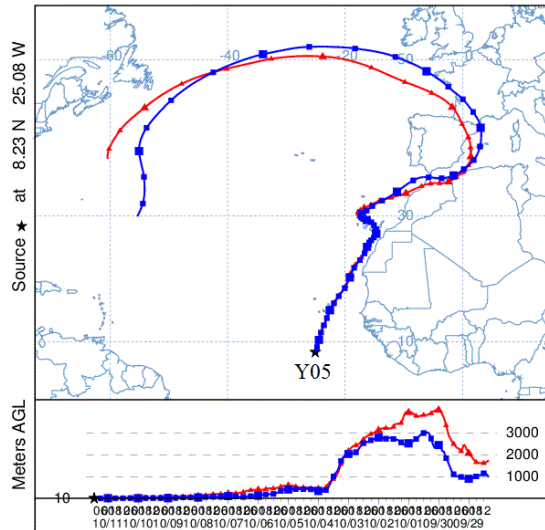
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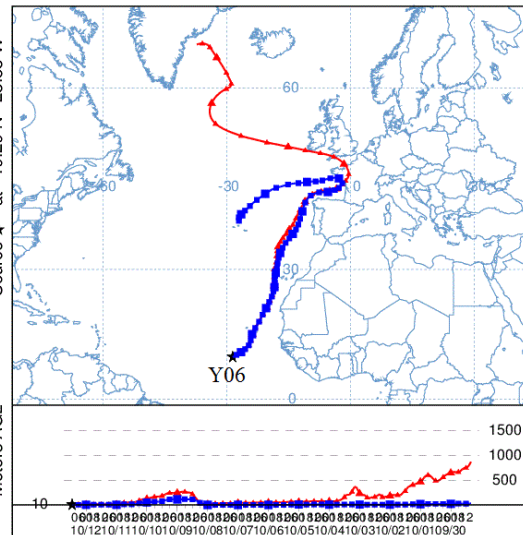
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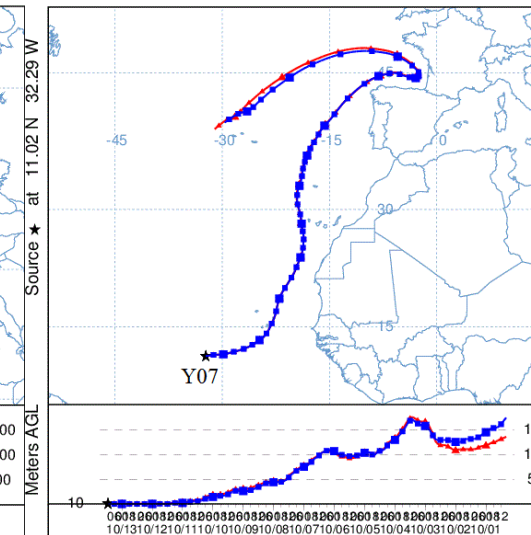
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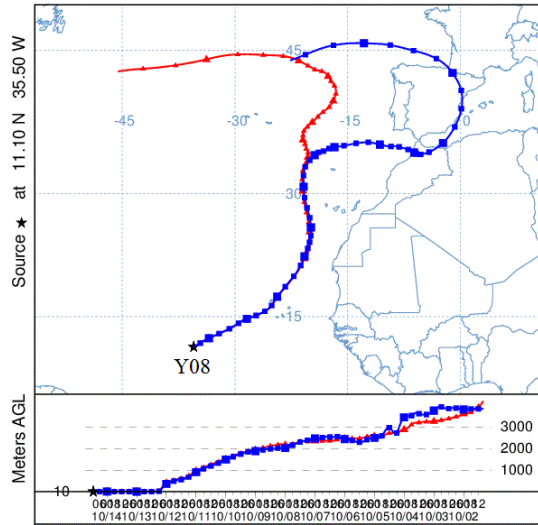
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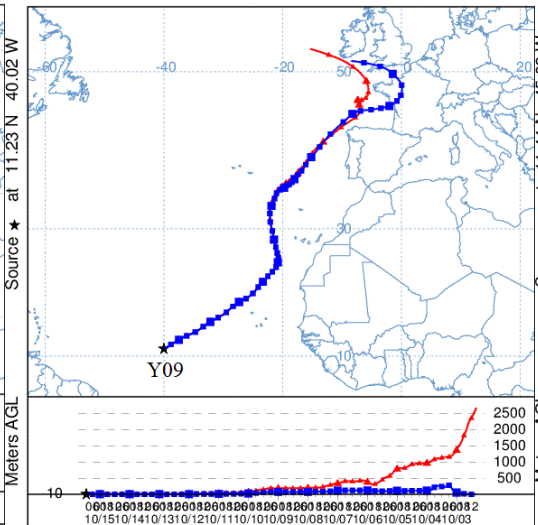
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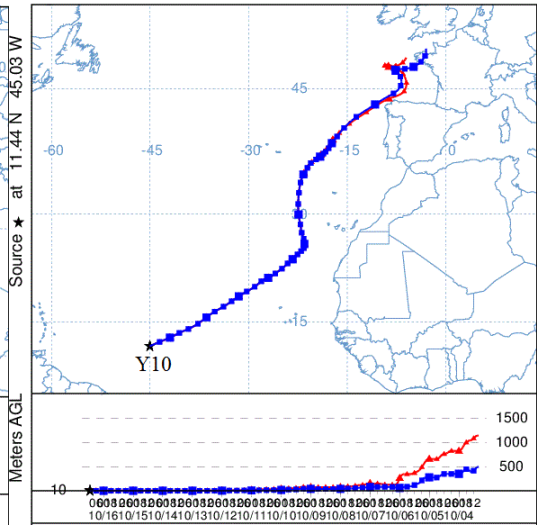
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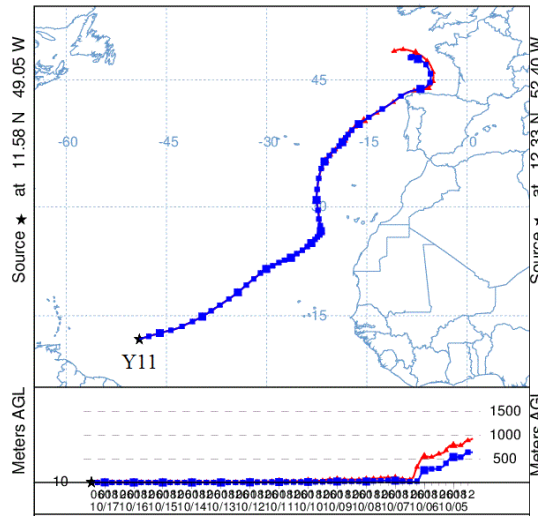
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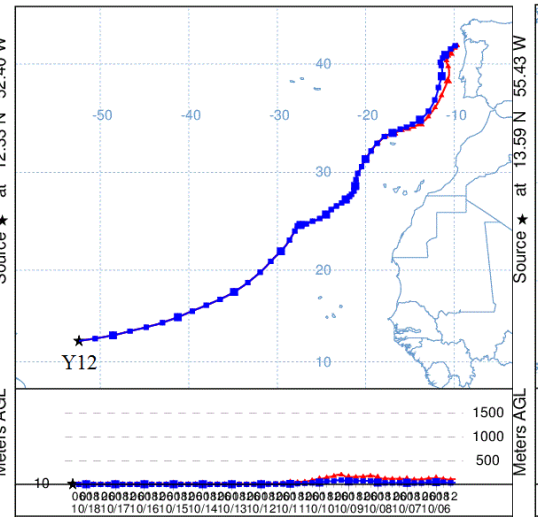
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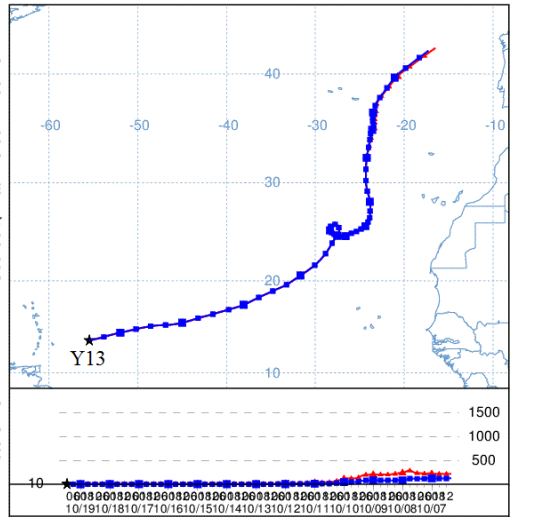
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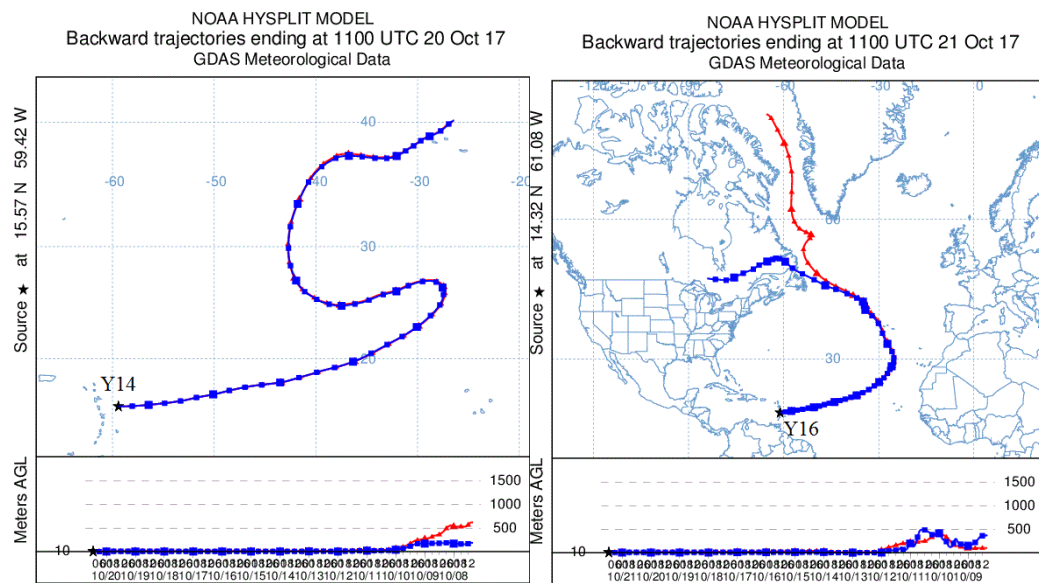


Figure S6: Air mass backward trajectories modelled for each sampling station using the NOAA HYSPLIT Model (<https://www.arl.noaa.gov/hysplit/hysplit/>). Black stars indicate the location of the sampling stations. The model was set to start at 10 m altitude and to run for 315 hours

SUPPLEMENTARY TABLES

Table S1: GPS positions of the sampling stations, collection dates and surface salinity.

Station ID	Latitude	Longitude	Date	Salinity
Y 01	11°38.2081' N	21°23.8295' W	10/07/2017	34.4
Y 03	7°46.3369' N	19°00.1133' W	10/09/2017	34.8
Y 04	8°16.5055' N	21°31.5612' W	10/10/2017	33.8
Y 05	8°23.2678' N	25°07.6840' W	10/11/2017	35.1
Y 06	10°19.9553' N	28°58.0255' W	10/12/2017	34.6
Y 07	11°01.9355' N	32°28.8749' W	10/13/2017	34.3
Y 08	11°09.4697' N	35°50.0678' W	10/14/2017	34.8
Y 09	11°22.7072' N	40°01.6629' W	10/15/2017	34.7
Y 10	11°43.4907' N	45°03.1796' W	10/16/2017	35.0
Y 11	11°57.9510' N	49°05.2901' W	10/17/2017	34.5
Y 12	12°33.1485' N	52°40.1368' W	10/18/2017	34.6
Y 13	13°59.3505' N	55°42.7139' W	10/19/2017	35.1
Y 14	15°56.6274' N	59°41.6454' W	10/20/2017	33.7
Y 16	14°32.1003' N	61°07.9776' W	10/21/2017	33.2

Table S2: Quantification and confirmation ions of OPEs (GC/MS).

Compound	Full name	Q1	Q2	Q3	Q4
TPP	Tripropyl phosphate	99	141	123	183
TiBP	Tri-iso-butyl phosphate	99	155	139	
TnBP	Tri-n-butyl phosphate	99	155	211	
TCEP	Tris-(2-chloroethyl) phosphate	249	251	143	99
TCPP-1	Tris-(2-chloro, 1-methylethyl) phosphate	125	99	157	277
TCPP-2	Tris-(2-chloro, 1-methylethyl) phosphate	125	99	157	277
TDCP	Tris-(2-chloro-, 1-chloromethylethyl) phosphate	191	99	381	209
TPhP	Triphenyl phosphate	326	325	215	170
EHDPP	2-ethylhexyl-diphenyl phosphate	251	250	326	
TEHP	Tris(2-ethylhexyl) phosphate	99	113	112	
TBP- <i>d27</i>	Tri-n-butyl- <i>d27</i> phosphate	103	167	231	
TPhP- <i>d15</i>	Triphenyl- <i>d15</i> phosphate	341	339	223	180
TCPP- <i>d18</i>	Tris-(2-chloro, 1-methylethyl)- <i>d18</i> phosphate	131	164	193	
TDCP- <i>d15</i>	Tris-(2-chloro-, 1-chloromethylethyl)- <i>d15</i> phosphate	197	199	217	394
TPrP- <i>d21</i>	Tri-n-propyl- <i>d21</i> phosphate	103	131	151	199
TCEP- <i>d12</i>	Tris-(2-chloroethyl)- <i>d12</i> phosphate	261	263	148	231
MAL- <i>d7</i>	Malathion- <i>d7</i>	174	131	128	164

Table S3: [M-H]⁻ values for BPs and PFCs quantification by LC-HRMS Agilent LC-QTOF 6530.

Compound	Full name	[M-H] ⁻
BPA	Bisphenol A	227.1078
BPAF	Bisphenol AF	335.0512
BPAP	Bisphenol AP	289.1234
BPF	Bisphenol F	199.0765
BPS	Bisphenol S	249.0227
BPZ	Bisphenol Z	267.1391
BPA- <i>d16</i>	Bisphenol A- <i>d16</i>	243.2082
PFHA	Perfluoro hexanoic acid	312.9728
PFHS	Perfluorohexane sulfonic acid	398.9366
PFOA	Perfluorooctanoic acid	412.9664
PFOS	Perfluorooctane sulfonic acid	498.9302
PFOSF	Perfluorooctane sulfonyl fluoride	500.9259
PFHA- <i>13C</i>	Perfluoro hexanoic acid- <i>13C</i>	317.9896
PFHS- <i>13C</i>	Perfluorohexane sulfonic acid- <i>13C</i>	401.9467
PFOA- <i>13C</i>	Perfluorooctanoic acid- <i>13C</i>	420.9933
PFOS- <i>13C</i>	Perfluorooctane sulfonic acid- <i>13C</i>	506.9571

Table S4: Limits of quantification (LOQ) expressed as absolute amounts (pg) and ng L⁻¹ in seawater.

Compound	LOQ	
	(pg)	(ng L ⁻¹)
TPP (CAS 513-08-6)	5	0.13
TiBP (CAS 126-71-6)	1	0.03
TnBP (CAS 126-73-8)	2	0.05
TCEP (CAS 115-96-8)	10	0.19
TCPP (CAS 13674-84-5)	10	0.25
TDCP (CAS 13674-87-8)	10	0.25
TPhP (CAS 115-86-6)	5	0.10
EHDPP (CAS 1241-94-7)	5	0.13
TEHP (CAS 78-42-2)	10	0.20
TBP- <i>d27</i> (CAS 61196-26-7)	2	0.05
TPhP- <i>d15</i> (CAS 1173020-30-8)	5	0.13
TCPP- <i>d18</i>	30	0.75
TDCP- <i>d15</i> (CAS 1447569-77-8)	11	0.28
BPA (CAS 80-05-7)	10	1.0
BPAF (CAS 14878-61-1)	1	0.1
BPAP (CAS 1571-75-1)	20	2.0
BPF (CAS 620-92-8)	10	1.0
BPS (CAS 80-09-1)	1	0.1
BPZ (CAS 843-55-0)	50	5.0
PFHA (CAS 307-24-4)	80	8.0
PFHS (CAS 3871-99-6)	1	0.1
PFOA (CAS 335-67-1)	20	2.0
PFOS (CAS 1763-23-1)	1	0.1
PFOSF (CAS 307-35-7)	10	1.0

Table S5: Concentrations (in ng L⁻¹) of all 20 molecules analyzed for each sampling station as well as extraction, transport and laboratory blank values. Sampling station data are already blank-corrected. For each sampling station, the average of the concentrations measured in the two replicates is given.

	Extraction Blank	Extraction Blank	Extraction Blank	Transport Blank	Laboratory Blank	Y 01	Y 03	Y 04	Y 05	Y 06	Y 07	Y 08	Y 09	Y 10	Y 11	Y 12	Y 13	Y 14	Y 16	
<i>TPP</i>	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
<i>TiBP</i>	<LOQ	n.d.	n.d.	n.d.	n.d.	3.9	6.5	1.8	2.7	2.0	5.2	6.2	11	33	38	30	17	1.7	<LOQ	
<i>TnBP</i>	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
<i>TCEP</i>	n.d.	n.d.	n.d.	n.d.	n.d.	<LOQ	1.0	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	1.3	n.d.	1.6	1.2	1.6	<LOQ	<LOQ	
<i>TCPP</i>	29	7.9	31	5.4	6.7	203	248	141	142	205	290	577	466	921	1300	966	596	104	74	
<i>TDCP</i>	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
<i>TPhP</i>	<LOQ	<LOQ	<LOQ	n.d.	<LOQ	2.2	6.9	1.5	1.0	<LOQ	<LOQ	<LOQ	1.3	<LOQ	1.5	1.6	1.3	<LOQ	<LOQ	
<i>EHDPP</i>	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	2.4	5.4	2.4	1.2	<LOQ	<LOQ	<LOQ	1.2	1.0	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	
<i>TEHP</i>	n.d.	n.d.	n.d.	15	16	<LOQ	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
Σ₉ OPEs	29	7.9	31	20.4	22.7	211.5	267.8	146.7	146.9	207.0	295.2	583.2	480.8	955.0	1341.1	998.8	615.9	105.7	74.0	
<i>BPA</i>	1.0	1.0	<LOQ	n.d.	n.d.	1.2	n.d.	n.d.	n.d.	n.d.	n.d.	<LOQ	1.8	1.8	<LOQ	<LOQ	<LOQ	n.d.	n.d.	
<i>BPAF</i>	<LOQ	<LOQ	1.0	<LOQ	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	
<i>BPAP</i>	n.d.	<LOQ	<LOQ	<LOQ	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	
<i>BPF</i>	<LOQ	<LOQ	<LOQ	<LOQ	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	
<i>BPS</i>	<LOQ	n.d.	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	<LOQ	n.d.	1.1	<LOQ	n.d.	n.d.	n.d.	n.d.	
<i>BPZ</i>	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	
Σ₆ BPs	1.0	1.0	1.0			1.2							1.8	2.9						
<i>PFHA</i>	2.2	<LOQ	1.0	<LOQ	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	9.7	16	11	n.d.	n.d.	n.d.	
<i>PFHS</i>	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	4.4	n.d.	n.d.	n.d.	n.d.	
<i>PFOA</i>	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	2.9	n.d.	n.d.	n.d.	n.d.	
<i>PFOS</i>	<LOQ	n.d.	<LOQ	<LOQ	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	2.3	n.d.	n.d.	n.d.	n.d.	
<i>PFOSF</i>	n.d.	n.d.	n.d.	<LOQ	<LOQ	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	
Σ₅ PFCs	2.2		1.0											9.7	25.6	11.0				
Total						212.7	267.8	146.7	146.9	207.0	295.2	583.2	482.6	967.6	1366.7	1009.8	615.9	105.7	74.0	

n.d. = not detected; LOQ = limit of quantification

Abbreviations: TPP: Tripropyl phosphate, TiBP: Tri-iso-butyl phosphate, TnBP: Tri-n-butyl phosphate, TCEP: Tris-(2-chloroethyl) phosphate, TCPP: Tris-(2-chloro, 1-methylethyl) phosphate, TDCP: Tris-(2-chloro-, 1-chloromethylethyl) phosphate, TPhP: Triphenyl phosphate, EHDPP: 2-ethylhexyl-diphenyl phosphate, TEHP: Tris(2-ethylhexyl) phosphate, BPA: Bisphenol A, BPAF: Bisphenol AF, BPAP: Bisphenol AP, BPF: Bisphenol F, BPS: Bisphenol S, BPZ: Bisphenol Z, PFHA: Perfluoro hexanoic acid, PFHS: Perfluorohexane sulfonic acid, PFOA: Perfluorooctanoic acid, PFOS: Perfluorooctane sulfonic acid, PFOSF: Perfluorooctane sulfonfyl fluoride.

Table S6: Surrogate recovery rates (in %) for each sampling station and the extraction blanks. Transport and laboratory blanks were not spiked with surrogate standards, which is why no data are available in this case.

	<i>Extraction Blank</i>	<i>Extraction Blank</i>	<i>Extraction Blank</i>	<i>Transport Blank</i>	<i>Laboratory Blank</i>	<i>Y 01</i>	<i>Y 03</i>	<i>Y 04</i>	<i>Y 05</i>	<i>Y 06</i>	<i>Y 07</i>	<i>Y 08</i>	<i>Y 09</i>	<i>Y 10</i>	<i>Y 11</i>	<i>Y 12</i>	<i>Y 13</i>	<i>Y 14</i>	<i>Y 16</i>
<i>D27-TBP</i>	83.0	84.4	77.6	n.a.	n.a.	87.7	74.9	81.6	75.8	64.4	74.4	72.1	79.9	82.7	82.0	87.7	86.2	63.2	82.2
<i>D18-TCPP</i>	63.8	62.4	62.2	n.a.	n.a.	69.5	67.3	66.7	68.5	67.8	74.5	69.4	71.6	73.4	75.5	79.1	67.0	59.8	72.1
<i>D15-TDCP</i>	106.9	89.1	54.4	n.a.	n.a.	76.0	74.9	81.5	69.2	63.3	74.4	71.3	80.4	64.8	71.1	81.9	82.8	67.0	85.1
<i>D15-TPhP</i>	94.4	82.6	63.2	n.a.	n.a.	64.5	62.0	69.9	56.5	48.7	67.4	61.7	73.3	62.0	71.9	101.9	82.8	77.8	83.5

n.a. = not available

Abbreviations: D27-TBP: D27-Tri-n-butyl phosphate, D18-TCPP: D18-Tris (2-chloroisopropyl) phosphate, D15-TDCP: D15-Tris (1,3-dichloro-2-propyl) phosphate, D15-TPhP: D15-Triphenyl phosphate.

Table S7: Sensitivity tests results for radii from 5 to 25 km and densities from 0.5 to 2.5 particles km⁻².

	0.5 particles km ⁻²		1.3 particles km ⁻²		2.5 particles km ⁻²		Sensitivity
	% of particles	travel time (days)	% of particles	travel time (days)	% of particles	travel time (days)	
Y 01							
5 km	0%		0%		0%		0 %
10 km	0%		0%		0%		
16 km	0%		0%		0%		
25 km	0%		0%		0%		
Y 03							
5 km	0%		0%		0%		0%
10 km	0%		0%		0%		
16 km	0%		0%		0%		
25 km	0%		0%		0%		
Y 04							
5 km	100%	[-124,-125]	100%	[-124,-125]	100%	[-124,-125]	40%
10 km	74%	[-124,-125]	82%	[-124,-125]	80%	[-124,-125]	
16 km	67%	[-124,-126]	70%	[-124,-125]	68%	[-124,-128]	
25 km	60%	[-124,-180]	60%	[-124,-126]	61%	[-124,-127]	
Y 05							
5 km	0%		0%		0%		8%
10 km	0%		0%		0%		
16 km	0%		0%		0%		
25 km	7%	[-92,-126]	8%	[-92,-126]	7%	[-92,-126]	
Y 06							
5 km	100%	[-90,-146]	100%	[-90,-113]	100%	[-90,-146]	0%
10 km	100%	[-90,-146]	100%	[-89,-147]	100%	[-83,-151]	
16 km	100%	[-88,-151]	100%	[-88,-147]	100%	[-84,-147]	
25 km	100%	[-88,-147]	100%	[-82,-148]	100%	[-81,-162]	
Y 07							
5 km	100%	[-88,-92]	100%	[-88,-92]	100%	[-88,-92]	0%
10 km	100%	[-88,-92]	100%	[-88,-92]	100%	[-88,-92]	
16 km	100%	[-88,-92]	100%	[-88,-92]	100%	[-88,-92]	
25 km	100%	[-88,-114]	100%	[-88,-117]	100%	[-88,-127]	
Y 08							
5 km	100%	[-77,-78]	100%	[-76,-78]	100%	[-76,-78]	0%
10 km	100%	[-76,-78]	100%	[-76,-78]	100%	[-76,-78]	
16 km	100%	[-76,-78]	100%	[-76,-78]	100%	[-76,-78]	
25 km	100%	[-76,-85]	100%	[-76,-85]	100%	[-76,-85]	
Y 09							

5 km	0%		0%		0%		0%
10 km	0%		0%		0%		0%
16 km	0%		0%		0%		0%
25 km	0%		0%		0%		0%
Y 10							
5 km	100%	[-76,-115]	100%	[-76,-115]	100%	[-76,-115]	
10 km	100%	[-76,-115]	100%	[-76,-115]	100%	[-76,-120]	4%
16 km	100%	[-76,-115]	100%	[-76,-115]	100%	[-76,-115]	
25 km	96%	[-76,-119]	96%	[-76,-119]	96%	[-76,-119]	
Y 11							
5 km	100%	[-61,-65]	100%	[-61,-65]	100%	[-61,-65]	
10 km	100%	[-60,-91]	100%	[-60,-116]	100%	[-60,-116]	3%
16 km	100%	[-60,-115]	100%	[-60,-96]	100%	[-60,-116]	
25 km	97%	[-60,-116]	97%	[-60,-122]	97%	[-60,-120]	
Y 12							
5 km	100%	[-66,-105]	100%	[-65,-111]	100%	[-65,-139]	
10 km	99%	[-65,-140]	98%	[-65,-154]	97%	[-65,-153]	8%
16 km	93%	[-52,-140]	92%	[-52,-153]	92%	[-52,-159]	
25 km	94%	[-37,-159]	95%	[-37,-140]	95%	[-37,-159]	
Y 13							
5 km	100%	[-59,-81]	100%	[-59,-106]	100%	[-59,-107]	
10 km	100%	[-58,-106]	100%	[-58,-105]	100%	[-58,-106]	0%
16 km	100%	[-57,-107]	100%	[-57,-107]	100%	[-57,-107]	
25 km	100%	[-54,-107]	100%	[-54,-107]	100%	[-54,-107]	
Y 14							
5 km	33%	[-138,-141]	30%	[-138,-156]	29%	[-132,-159]	
10 km	23%	[-133,-156]	20%	[-131,-179]	21%	[-131,-174]	26%
16 km	23%	[-78,-157]	26%	[-78,-168]	25%	[-78,-179]	
25 km	46%	[-78,-175]	46%	[-78,-180]	46%	[-78,-179]	