



Synthesis of New β -Lactams Bearing the Biologically Important Morpholine Ring and POM Analyses of Their Antimicrobial and Antimalarial Activities

Oxana B Kazakova, Jean Michel Brunel, Elmira F Khusnutdinova, Sophie Negrel, Gulnara V Giniyatullina, Tatyana V Lopatina, Anastasiya V Petrova

► To cite this version:

Oxana B Kazakova, Jean Michel Brunel, Elmira F Khusnutdinova, Sophie Negrel, Gulnara V Giniyatullina, et al.. Synthesis of New β -Lactams Bearing the Biologically Important Morpholine Ring and POM Analyses of Their Antimicrobial and Antimalarial Activities. Molbank, 2019, 2019 (3), pp.M1078. 10.3390/M1078 . hal-02391762v1

HAL Id: hal-02391762

<https://amu.hal.science/hal-02391762v1>

Submitted on 3 Dec 2019 (v1), last revised 29 May 2020 (v2)

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

A Ring-Modified Triterpenoids and Their Spermidine-Aldimines with Strong Antibacterial Activity

Oxana B. Kazakova ^{1,*}, Elmira F. Khusnutdinova ¹, Sophie Negrel ², Gulnara V. Giniyatullina ¹, Tatyana V. Lopatina ¹, Anastasiya V. Petrova ¹ and Jean Michel Brunel ^{2,*}

¹ Ufa Institute of Chemistry of the Ufa Federal Research Centre of the Russian Academy of Sciences, Ufa 450054, 71 pr. Oktyabrya, Russian Federation; ElmaH@inbox.ru (E.F.K.), gulnaravlg@gmail.com (G.V.G.) ltvioch@yandex.ru (T.V.L.), pnastya08@mail.ru (A.V.P.)

² Aix Marseille Université, INSERM, SSA, MCT, 13385 Marseille, France, snoly@hotmail.fr

* Correspondence: obf@anrb.ru (O.B.K.); bruneljm@yahoo.fr (J.M.B.); Tel.: +7-347-235-6066 (O.B.K.); +33-689-271-645 (J.M.B.)

Abstract: Synthesis of A-ring-modified lupane, oleanane and ursane type triterpenoid conjugates with spermidine through an aldimine linkage or diethylentriamine via an amide bond is described. These derivatives were evaluated for their in vitro antimicrobial properties against human pathogens. Except for derivatives **1** and **7**, all compounds have moderate to weak minimum inhibitory concentrations (MICs) against Gram-positive *Staphylococcus aureus* bacteria, with MICs varying from 3.125 to 200 μ M. Compound **11** is efficient against *Escherichia coli* and *Pseudomonas aeruginosa*, with MICs of 25 and 50 μ M, respectively, while all other derivatives do not possess important antimicrobial activities against these Gram-negative bacteria.

Keywords: triterpenoids; lupane; oleanane; ursane; spermidine; spermine; squalamine; antimicrobial activity; Gram-positive bacteria; Gram-negative bacteria

1. Introduction

Pentacyclic triterpenoids are widely represented in natural products and are useful substrates for the synthesis of various important bioactive compounds [1–3]. Conjugation of a triterpenoid scaffold with amines, amino acids, and polyamines resulted in a series of anticancer, antimicrobial and antiviral agents [4–8]. Chemical antibiotics were one of the great health successes of the 20th century leading to a huge decrease in both morbidity and mortality from bacterial infections. However, this has led to high levels of inappropriate prescribing, which has contributed to a recent rise in the number of antibiotic-resistant bacteria. As a result, multidrug-resistant bacteria such as vancomycin-resistant *Enterococcus* (VRE), methicillin-resistant *Staphylococcus aureus* (MRSA), and multidrug-resistant *Pseudomonas aeruginosa* (MRPA) have appeared [9]. In this context, new types of antibiotics such as antibiotic peptides, lipids, and alkaloids have been isolated as host defense agents from diverse animal species [10–12]. Among these substances, two water-soluble cationic steroids, squalamine, and trodusquamine, have been identified from the dogfish shark *Squalus acanthias*, exhibiting excellent antimicrobial activities (Figure 1) [13–15].

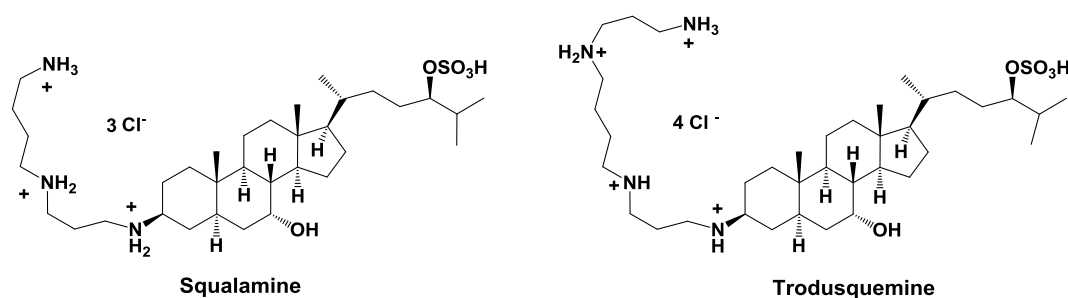


Figure 1. Structure of squalamine and trodusquamine.

Since that time, the synthesis of the triterpenoid analog of squalamine has been accomplished [16], and novel steroidal polyamines, such as Claramine, that exhibit a broad-spectrum antimicrobial bimodal activity, have been discovered [17,18]. Polyamines (putrescine, spermidine, spermine, etc.) play an important role in the physiology of plants and live organisms. They are polycations that interact with negatively charged molecules such as DNA, RNA, and proteins. Betulinic acid-based amides with putrescine and spermine demonstrate a high level of cytotoxicity and antimicrobial activity [19]. At the same time, there are no reports on the synthesis of spermidine conjugates connected to the triterpenoid core via the aldimine group.

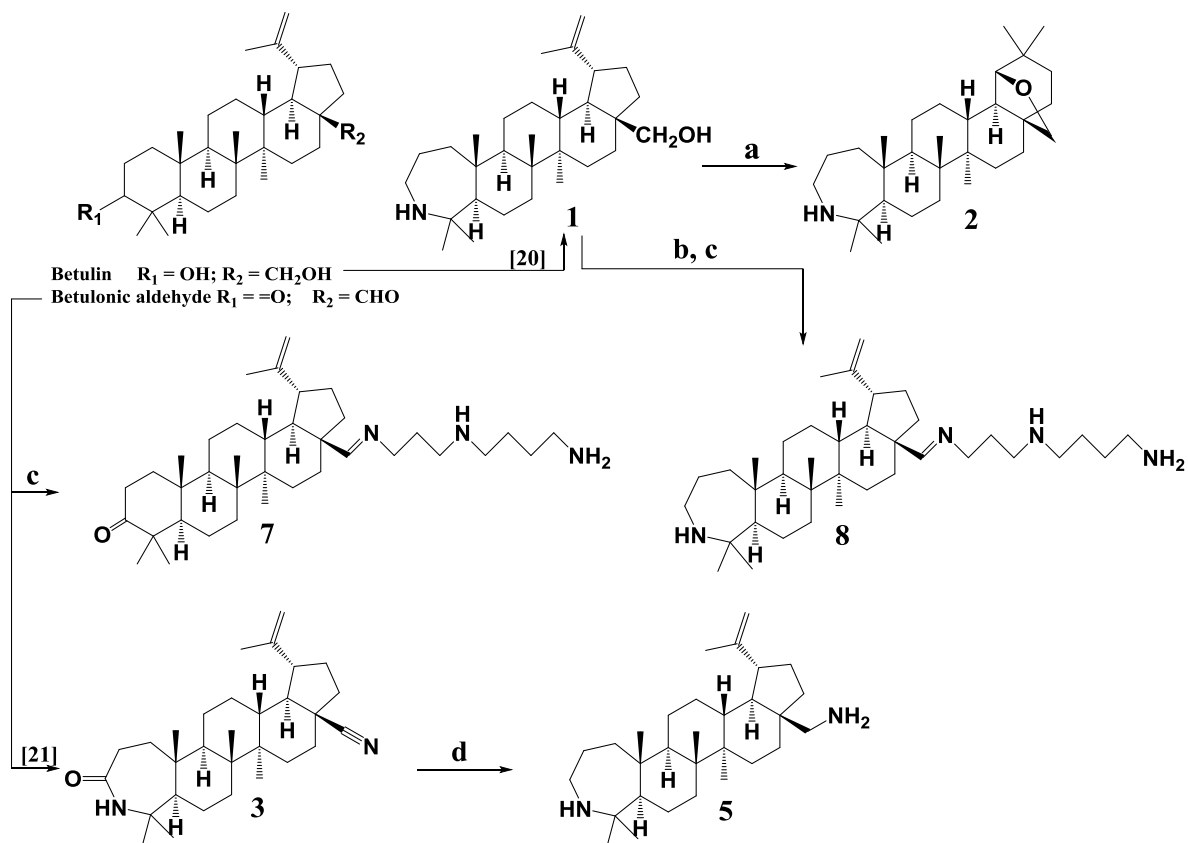
In this work, the first synthesis of triterpenoid spermidino-aldimines, as well as some derivatives with an A-azepano-ring and C28-(diethylenetriamino)-amides is described and the study of their antimicrobial activity is presented.

2. Results and Discussion

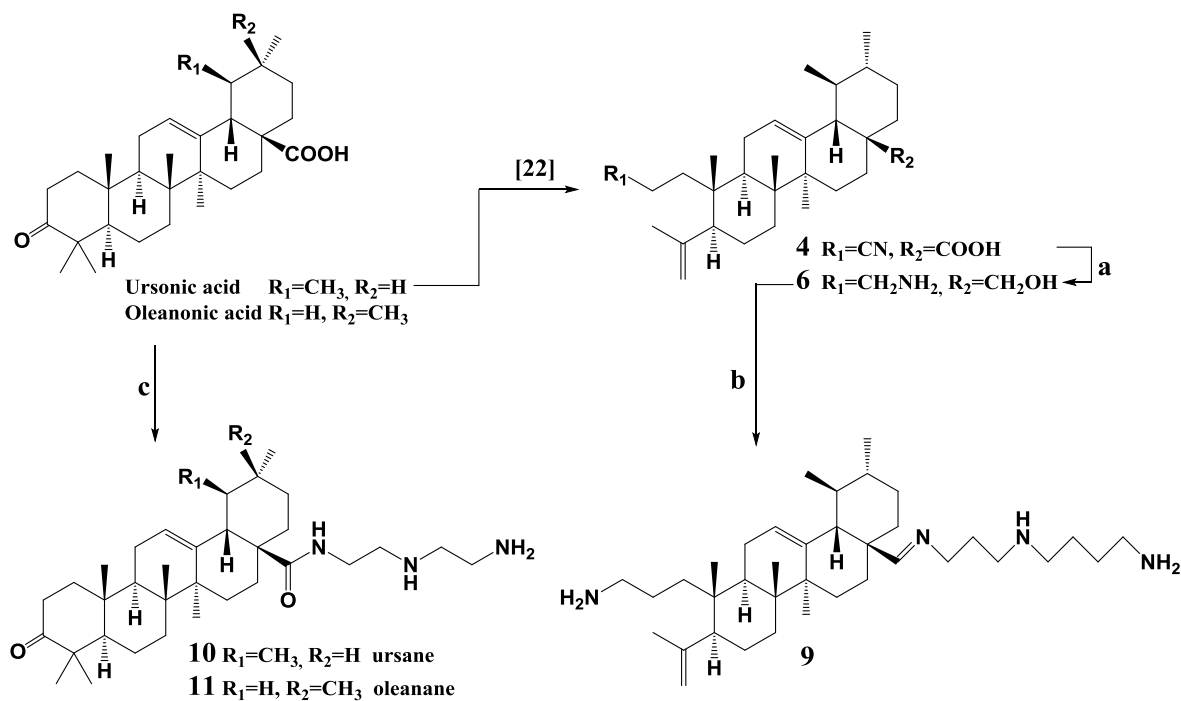
2.1. Chemistry

The synthesis of a series of nitrogen-containing triterpenoids is presented in Schemes 1 and 2. Azepanobetulin **1** was obtained by several stages from naturally-occurring betulin [20]. The following p-TsOH catalyzed rearrangement of compound **1** resulted in azepanoallobetulin **2** with a yield of 86%. Reduction of lupane A-azepanono-C28-nitril **3** [21] and 2-cyano-3,4-seco-ursolic acid **4** [22] with LiAlH_4 under reflux in tetrahydrofuran led to methylenamino-derivatives **5** and **6**. Betulin and C28-hydroxy-triterpenoids **1** or **6** were converted to corresponding aldehydes and were reacted with spermidine in MeOH under reflux to obtain aldimines **7–9** in yields of 65–72%. The reaction of ursonic or oleanonic acid chlorides with diethylenetriamine led to amides **10** and **11** in yields of 85 and 73%, respectively. The data for the latter amide have been previously presented [23].

The structure of the compounds was confirmed by NMR spectroscopy. The ^{13}C -NMR spectra of azepanes **2**, **5** and **8** showed signals of C-3 at δ 62–63 ppm. For aldimines **7–9**, the signals of the $-\text{CH}=\text{N}-$ function were detected at δ 169–173 ppm (^{13}C -NMR) and at δ 7.3–7.8 ppm (^1H -NMR). In the spectra of compound **10**, the signal of amide function $\text{C}(\text{O})\text{NH}$ was detected at δ 178.9 ppm (^{13}C -NMR) and at δ 6.61 ppm (^1H -NMR). Finally, the presence of polyamine moieties at position C-28 of the triterpene core in the structure of compounds **7–11** was confirmed by the characteristic signals of the aminomethylene groups $-\text{CH}_2\text{NH}-$ at δ 2.58–3.70 ppm (^1H -NMR).



Scheme 1. Reagents and conditions: a. TsOH, CHCl_3 , reflux, 4 h; b. PCC, CH_2Cl_2 , rt, 0.5 h; c. $\text{NH}_2(\text{CH}_2)_3\text{NH}(\text{CH}_2)_4\text{NH}_2$, MeOH, reflux, 8 h; d. LiAlH_4 , THF, reflux, 3 h.



Scheme 2. Reagents and conditions: a. LiAlH_4 , THF, reflux, 3 h; b. i PCC, CH_2Cl_2 , rt, 0.5 h, ii $\text{NH}_2(\text{CH}_2)_3\text{NH}(\text{CH}_2)_4\text{NH}_2$, MeOH, reflux, 8 h; c. i $(\text{COCl})_2$, Et_3N , CH_2Cl_2 , 20 °C, 2 h, ii diethylenetriamine, CH_2Cl_2 , Et_3N , 20 °C, 3 h.

2.2. Biology

All the synthesized compounds were screened for antimicrobial activity against both Gram-positive and Gram-negative bacterial strains and found to possess activities (Table 1). Thus, except for derivatives **1** and **7**, all compounds have moderate to weak minimum inhibitory concentrations against Gram-positive *S. aureus* bacteria with MICs varying from 3.125 to 200 μ M. Nevertheless, compound **11** is efficient against *E. coli* and *P. aeruginosa*, with MICs of 25 and 50 μ M, respectively, while all other derivatives do not possess important antimicrobial activities against these Gram-negative bacteria. It is noteworthy that the minimum bactericidal activity remains of interest for all products active against *S. aureus*. These data clearly suggest that the nature of the triterpenoid involved plays an important role in the potential activities of our compounds.

Table 1. Minimum inhibitory concentrations and minimum bactericidal concentration of compounds **1**, **2**, **5**, and **7–11**.

Compound	MIC (μ M)/MBC (μ M)		
	<i>S. aureus</i> DSM789	<i>E. coli</i> ATCC25299	<i>P. aeruginosa</i> PA01
1	>200/>200	>200/>200	>200/>200
2	6.25/25	>200/>200	>200/>200
5	12.5/25	>200/>200	>200/>200
7	>200/>200	>200/>200	>200/>200
8	3.125/6.25	>200/>200	>200/>200
9	12.5/25	200/>200	200/>200
10	12.5/25	200/>200	200/>200
11	12.5/25	25/25	50/>200
Ciprofloxacin	0.20	0.20	0.20
Vancomycin	0.80	>200	>200

3. Materials and Methods

3.1. General Methods and Physical Measurements

The spectra were recorded at the Center for the Collective Use ‘Chemistry’ of the Ufa Institute of Chemistry of the Ufa Federal Research Centre of the Russian Academy of Sciences. ^1H and ^{13}C -NMR spectra were recorded on a “Bruker AM-500” (Bruker, Billerica, MA, USA, 500 and 125.5 MHz respectively, δ , ppm, Hz) in CDCl_3 , internal standard tetramethylsilane. Mass spectra were obtained on a liquid chromatograph–mass spectrometer LCMS-2010 EV (Shimadzu, Kyoto, Japan). Melting points were detected on a micro table “Rapido PHMK05” (Nagema, Dresden, Germany). Optical rotations were measured on a polarimeter “Perkin-Elmer 241 MC” (PerkinElmer, Waltham, MA, USA) in a tube length of 1 dm. Elemental analysis was performed on a Euro EA-3000 CHNS analyzer (Eurovector, Milan, Italy); the main standard is acetanilide. Thin-layer chromatography analyses were performed on Sorbfil plates (Sorbpolimer, Krasnodar, Russian Federation), using the solvent system chloroform–ethyl acetate, 40:1. Substances were detected by 10% H_2SO_4 with subsequent heating to 100–120 $^\circ\text{C}$ for 2–3 min. Betulonic aldehyde [24], compounds **1** [20], **3** [21], ursonic acid and **4** [22], **11** [23] were obtained according to the methods described previously.

3.2. Synthesis of 3-Deoxy-3a-homo-3a-aza-19 β ,28-epoxy-18 α -oleanane (**2**)

A mixture of compound **1** (220 mg, 0.5 mmol) and a catalytic amount of *p*-TsOH (8.6 mg, 0.05 mmol) in dry CHCl_3 (25 mL) was heated under reflux for 4 h, and then the solvent was evaporated under reduced pressure. The product was purified by Al_2O_3 column chromatography using CHCl_3 and a mixture of CHCl_3 –EtOH (100:1) as eluents to afford compound **2** as a beige powder (190 mg, 86%); $[\alpha]_{\text{D}}^{20} +54$ (c 0.05, CHCl_3), m.p. 261 $^\circ\text{C}$. ^1H -NMR (δ , ppm, CDCl_3 , 125.5 MHz): 3.62 and 3.45 (2H, both d, H-28, $J = 7.7$ Hz), 3.41 (1H, s, H-19), 3.21–3.16 (1H, m, H3b), 2.91–2.82 (1H, m, H3a), 2.02–1.16 (26H, m, CH, CH_2), 1.42, 1.29, 1.01, 0.92, 0.81, 0.74, 0.69 (21H, all s, 7 CH_3). ^{13}C -NMR (δ , ppm, CDCl_3 ,

500 MHz): 90.0 (C-19), 72.4 (C-28), 63.0 (C-3), 54.5 (C-9), 48.2 (C-18), 46.6 (C-4), 41.6 (C-5), 41.2, 40.9, 39.9, 36.5, 36.1, 34.4, 33.0, 32.5, 31.5, 28.6, 27.7, 26.6, 26.2, 25.8, 24.4, 22.9, 22.8, 22.0, 21.6, 19.9, 16.2, 15.9, 13.1. Anal. Calcd for C₃₀H₅₁NO: C, 81.57; H, 11.64; N, 3.17. Found: C, 81.60; H, 11.54; N, 3.16. MS (APCI): *m/z* [M + H]⁺ 442.75, calcd for C₃₀H₅₁NO: 441.74.

3.3. Synthesis of 5 and 6

A mixture of compound 3 [21] (225 mg, 0.5 mmol) or compound 4 [22] (226 mg, 0.5 mmol) and LiAlH₄ (230 mg, 0.65 mmol) in dry THF (20 mL) was refluxed for 3 h and then poured into a 5% HCl solution (100 mL). The crude product was extracted with CHCl₃ (3 × 40 mL), and then the organic layer was washed with H₂O, dried over CaCl₂ and evaporated under reduced pressure. The product was purified by Al₂O₃ column chromatography using CHCl₃ and a mixture of CHCl₃–EtOH (100:1; 50:1, 25:1, 10:1) as eluents to afford compound 5 (187 mg, 85%) or compound 6 (174 mg, 79%) as white powders.

3-Deoxy-3a-homo-3a-aza-lup-20(29)-en-28-methylenamine (5): [α]_D²⁰ +96 (c 0.05, CHCl₃), m.p. 246 °C. ¹H-NMR (δ, ppm, CDCl₃, 125.5 MHz): 4.68 and 4.58 (2H, both s, *J* = 2.0 Hz, H-29), 3.80–3.78 (1H, m, H3b), 3.35–3.29 (1H, m, H3a), 2.90–2.78 (2H, m, H-28), 2.40–1.65 (28H, m, CH, CH₂), 1.65 (3H, s, H-30), 1.41, 1.17, 1.05, 0.99, 0.85 (15H, all s, 5CH₃). ¹³C-NMR (δ, ppm, CDCl₃, 500 MHz): 151.0 (C-20), 109.2 (C-29), 63.2 (C-3), 60.5, 54.4, 48.5, 47.8, 47.6, 47.4 (C-28), 43.0, 41.2, 40.9, 40.7, 37.7, 34.0, 29.7, 29.0, 27.9, 26.9, 25.8, 23.0, 22.9, 22.7, 22.0, 21.6, 21.3, 19.1, 16.6, 16.4, 14.5. Anal. Calcd for C₃₀H₅₂N₂: C, 81.75; H, 11.89; N, 6.36. Found: C, 81.69; H, 11.84; N, 6.30. MS (APCI): *m/z* [M + H]⁺ 441.73, calcd for C₃₀H₅₂N₂: 440.75.

3-Amino-3,4-seco-28-hydroxy-urs-12(13)-en (6): [α]_D²⁰ +54 (c 0.05, CHCl₃), m.p. 195 °C. ¹H-NMR (δ, ppm, CDCl₃+C₅D₅N, 125.5 MHz): 5.10 (1H, s, H-12), 4.82 and 4.78 (2H, both d, H-23, *J* = 2.8 Hz), 3.20 and 3.40 (2H, both d, H-28), 3.10 (2H, m, CH₂NH₂), 2.15–1.00 (25H, m, CH and CH₂), 1.65 (3H, s, CH₃, H-24), 1.22, 1.00, 0.92, 0.78, 0.66 (15H, all s, 5CH₃). ¹³C-NMR (δ, ppm, CDCl₃+C₅D₅N, 500 MHz): 148.7 (C-4), 139.2 (C-13), 123.3 (C-12), 115.4 (C-23), 69.9 (C-28), 51.8, 48.4, 43.9, 43.6, 41.8, 40.9, 40.6, 39.4, 38.7, 37.7, 35.8, 34.8, 32.9, 32.6, 32.5, 27.4, 27.1, 25.8, 25.2, 25.1, 24.6, 23.8, 23.0, 21.2, 18.1. Anal. Calcd for C₃₀H₅₁NO: C, 81.57; H, 11.64; N, 3.17. Found: C, 81.49; H, 11.70; N, 3.09. MS (APCI): *m/z* [M + H]⁺ 442.73, calcd for C₃₀H₅₁NO: 441.74.

3.4. Synthesis of Compounds 7–9

To a solution of compound 1 (221 mg, 0.5 mmol) or compound 6 (220 mg, 0.5 mmol) in CH₂Cl₂ (25 mL) pyridinium chlorochromate (PCC) (8.6 mg, 1.2 mmol) was added and the mixture was stirred at room temperature until the starting material disappeared (monitored by TLC). After completion of the reaction, the mixture was passed through Al₂O₃, the organic phase was washed with H₂O (2 × 30 mL), dried over CaCl₂ and evaporated under reduced pressure to yield crude aldehydes. Then, to a solution of betulonic aldehyde (110 mg, 0.25 mmol) [24] or freshly prepared above aldehydes in anhydrous MeOH (25 mL), spermidine (0.05 mL, 0.25 mmol) was added and the reaction mixture was heated under reflux for 8 h, then poured into H₂O (50 mL) and the precipitate was filtered off, washed with water, and dried in air. The products were chromatographed over Al₂O₃ using CHCl₃ and a mixture of CHCl₃–EtOH (100:1; 50:1) as eluents to afford compounds 7 (100 mg, 65%) as a yellow powder, 8 (102 mg, 72%) as a beige powder or 9 (109 mg, 70%) as an orange powder.

3-Oxo-lup-20(29)-en-28-N-(4-aminobutyl)-N-propylamin-28-imine (7): [α]_D²⁰ +3 (c 0.25, CHCl₃), m.p. 201 °C. ¹H-NMR (δ, ppm, CDCl₃, 125.5 MHz): 7.82 (1H, s, CH=N), 4.62 and 4.73 (2H, both d, *J* = 2.0 Hz, H-29), 3.50–2.68 (8H, m, 4CH₂N), 2.10–1.73 (34H, m, CH, CH₂), 1.72 (3H, s, H-30), 1.09, 1.08, 1.01, 0.98, 0.89 (15H, all s, 5CH₃). ¹³C-NMR (δ, ppm, CDCl₃, 500 MHz): 219.8 (C-3), 170.9 (C-28), 150.0 (C-20), 110.1 (C-29), 57.3, 55.1, 49.8, 49.5, 49.3, 48.9, 48.0, 47.9, 47.4, 41.4, 41.2, 39.9, 39.6, 39.0, 37.7, 37.5, 34.3, 33.6, 32.2, 31.9, 30.8, 30.7, 29.0, 28.1, 27.5, 26.7, 25.5, 21.5, 21.0, 19.2, 16.6, 15.7, 14.9. Anal. Calcd for C₃₇H₆₃N₃O: C, 78.53; H, 11.22; N, 7.43. Found: C, 78.49; H, 11.25; N, 7.40. MS (APCI): *m/z* [M + H]⁺ 566.80, calcd for C₃₇H₆₃N₃O: 565.92.

3-Deoxy-3a-homo-3a-aza-lup-20(29)-en-28-N-(4-aminobutyl)-N-propylamin-28-imine (**8**): $[\alpha]_{\text{D}}^{20} +85$ (c 0.1, CHCl_3), m.p. 185 °C. $^1\text{H-NMR}$ (δ , ppm, CDCl_3 , 500 MHz): 7.78 (1H, s, $\text{CH}=\text{N}$), 4.60 and 4.72 (2H, both d, $J = 2.0$ Hz, H-29), 3.48–3.32 (2H, m, H3), 2.80–2.58 (8H, m, $4\text{CH}_2\text{N}$), 2.10–1.42 (35H, m, CH, CH_2), 1.72 (3H, s, H-30), 1.09, 1.08, 1.01, 0.98, 0.89 (15H, all s, 5CH_3). $^{13}\text{C-NMR}$ (δ , ppm, CDCl_3 , 125.5 MHz): 169.1 (C-28), 150.2 (C-20), 109.8 (C-29), 62.3 (C-3), 58.0, 55.0, 52.2, 50.0, 48.7, 48.1, 47.8, 47.7, 42.9, 42.5, 41.4, 41.3, 39.2, 39.1, 37.4, 34.3, 34.0, 32.0, 31.5, 31.0, 30.1, 29.7, 29.4, 29.1, 28.0, 27.3, 26.9, 26.3, 23.2, 19.2, 16.8, 16.6, 14.3. Anal. Calcd for $\text{C}_{37}\text{H}_{66}\text{N}_4$: C, 78.38; H, 11.73; N, 9.88. Found: C, 78.32; H, 11.70; N, 9.85. MS (APCI): m/z $[\text{M} + \text{H}]^+$ 567.93, calcd for $\text{C}_{37}\text{H}_{66}\text{N}_4$: 566.95.

3-Amino-3,4-seco-urs-12(13)-en-28-N-(3-aminopropyl)-N-(butane-1,4-diamine)-N-propylamin-28-imine (**9**): $[\alpha]_{\text{D}}^{20} +91$ (c 0.1, CHCl_3), m.p. 145 °C. $^1\text{H-NMR}$ (δ , ppm, CDCl_3 , 125.5 MHz): 7.31 (1H, s, $\text{CH}=\text{N}$, H-28), 5.10 (1H, s, H-12), 4.79 and 4.59 (2H, both d, H-23, $J = 2.8$ Hz), 3.68–2.58 (8H, m, $4\text{CH}_2\text{N}$), 2.00–1.63 (36H, m, CH and CH_2), 1.62 (3H, s, CH_3 , H-24), 1.18, 0.99, 0.90, 0.74, 0.65 (15H, all s, 5CH_3). $^{13}\text{C-NMR}$ (δ , ppm, CDCl_3 , 500 MHz): 173.6 (C-28), 147.9 (C-4), 139.5 (C-13), 125.2 (C-12), 113.2 (C-23), 54.2, 52.2, 50.4, 42.6, 41.6, 41.4, 40.6, 39.7, 39.1, 38.3, 37.7, 36.5, 35.3, 33.9, 32.3, 31.8, 31.6, 30.6, 29.7, 29.3, 28.0, 26.0, 25.6, 24.4, 23.6, 23.3, 23.0, 21.9, 21.3, 20.7, 20.1, 19.9. Anal. Calcd for $\text{C}_{37}\text{H}_{66}\text{N}_4$: Found: C, 78.38; H, 11.73; N, 9.88. Found: C, 78.29; H, 11.84; N, 9.79. MS (APCI): m/z $[\text{M} + \text{H}]^+$ 567.94, calcd for $\text{C}_{37}\text{H}_{66}\text{N}_4$: 566.95.

3.5. Synthesis of 3-oxo-urs-12(13)-en-28-N-(2-((2-aminoethyl)amino)ethyl)-2-ethylamide (**10**)

A solution of ursonic acid [**22**] (225 mg, 0.5 mmol) in dry CH_2Cl_2 (20 mL) and $(\text{COCl})_2$ (1.5 mmol, 0.13 mL) was stirred at room temperature for 2 h and then was concentrated to dryness under reduced pressure. A resulting ursonic acid chloride (235 mg, 0.5 mmol) was dissolved in dry CH_2Cl_2 (30 mL) and treated with diethylenetriamine (0.05 mL, 0.5 mmol) and Et_3N (0.07 mL, 0.5 mmol). It was then stirred at room temperature for 3 h, washed with 5% HCl solution (2×50 mL) and H_2O (50 mL), dried over CaCl_2 , and the solvent was removed under reduced pressure. The product was chromatographed over a column of Al_2O_3 using CHCl_3 and a mixture of CHCl_3 – EtOH (100:1; 70:1, 40:1) as eluents to afford compound **10** (229 mg, 85%) as a yellow powder. $[\alpha]_{\text{D}}^{20} +35$ (c 0.1, CHCl_3), m.p. 178 °C. $^1\text{H-NMR}$ (δ , ppm, CDCl_3 , 125.5 MHz): 6.61 (1H, s, CONH), 5.30 (1H, s, H-12), 4.50–4.25 (3H, m, NH, NH_2), 3.70–2.70 (8H, m, 4CH_2), 2.55–1.36 (23H, m, CH and CH_2), 1.35, 1.19, 1.10, 0.99, 0.89, 0.75, 0.65 (21H, all s, 7CH_3). $^{13}\text{C-NMR}$ (δ , ppm, CDCl_3 , 500 MHz): 217.6 (C-3), 178.9 (C-28), 139.4 (C-13), 126.8 (C-12), 58.1, 55.2, 54.9, 53.4, 52.5, 48.3, 47.4, 46.7, 42.2, 39.7, 39.5, 38.9, 37.3, 36.6, 34.1, 32.6, 30.9, 28.1, 27.8, 26.6, 24.8, 24.6, 23.5, 23.3, 21.2, 19.6, 19.1, 18.8, 18.4, 17.2. Anal. Calcd for $\text{C}_{34}\text{H}_{57}\text{N}_3\text{O}_2$: C, 75.65; H, 10.64; N, 7.78. Found: C, 75.59; H, 10.58; N, 7.70. MS (APCI): m/z $[\text{M} + \text{H}]^+$ 540.82, calcd for $\text{C}_{34}\text{H}_{57}\text{N}_3\text{O}_2$: 539.84.

4. Biology Methods

4.1. Determination of Minimal Inhibitory Concentrations

The antimicrobial activity of the compounds **1**, **2**, **5**, and **7–11** was studied by determination of minimal inhibitory concentrations (MICs) according to the NCCLS guidelines M7-A2 using the microbroth dilution methods. All of the strains were issued from the Institute Pasteur collection (Paris, France). The bacteria strains were grown on trypticase soy agar (Becton Dickinson) at 37 °C for 24 h (*E. coli* ATCC25299, *S. aureus* DSM789, *P. aeruginosa* (PA01)) in MHII broth for *P. aeruginosa*, *E. coli* and *S. aureus*. Inocula were prepared in MHII by adjusting the turbidity at 623 nm to obtain 10^6 CFU/mL.

Antimicrobial activities of the compounds were determined by using a broth microdilution method performed in sterile 96-well microplates. All compounds were solubilized in methanol at a concentration of 5 mg/mL and were transferred to each microplate well (in all cases concentrations of the desired molecules in methanol did not exceed 2% of the total proportion). In order to obtain a two-fold serial dilution, 100 μL of broth and 100 μL of inocula containing 5×10^5 – 10^6 CFU of each bacteria were added to each well. Several wells were reserved for positive controls, inoculum viability, and solvent effect. After 24 h incubation, growth was assayed by absorbance measurement

at 623 nm with an IEMS Labsystem automatic plate reader. MIC was defined for each agent from triplicate observations as the lowest concentration of compound allowing no visible growth.

5. Conclusions

Finally, a series of triterpenoid C-28 polyamine conjugates or A-azepanes was synthesized and evaluated for their antimicrobial activity. Among them, oleanonic acid amide with diethylentriamine appears to be an interesting antimicrobial candidate against both Gram-positive and Gram-negative bacteria. Current studies are underway to evaluate the potentiality of such derivatives *in vivo*, by determining the cytotoxicity of these compounds, establishing the mechanism of action of this new class of antimicrobial agents and designing more active derivatives by varying the nature of the polyamine chain involved.

Supplementary Materials: ¹H and ¹³C spectra for compounds are available online.

Author Contributions: O.K. brought the idea, managed the research, conducted some synthetic experiments and prepared the manuscript; J.M.B. brought the idea, conducted biological experiments and prepared the manuscript; S.N. conducted some biological experiments, E.K. managed the research and prepared the manuscript; G.G. and T.L. conducted some synthetic experiments; A.P. made structure elucidation and prepared the manuscript.

Funding: The synthesis of compounds 1–3, 5 and 8 was supported by the Russian Foundation for Basic Research (project no. 18-33-00364 mol_a to Tatyana V. Lopatina).

Acknowledgments: This work was supported by the Ministry of Education of the Russian Federation, State task projects no. AAAA-A17-117011910023-2 and AAAA-A19-119020890014-7. The study of antimicrobial activity was supported by Aix-Marseille Université.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Salvador, J.A.R.; Leal, A.S.; Valdeira, A.S.; Gonçalves, B.M.F.; Alho, D.P.S.; Figueiredo, S.A.C.; Silvestre, S.M.; Mendes, V.I.S. Oleanane-, ursane-, and quinone methide friedelane-type triterpenoid derivatives: Recent advances in cancer treatment. *Eur. J. Med. Chem.* **2017**, *142*, 95–130, doi:10.1016/j.ejmech.2017.07.013.
2. Sousa, J.L.C.; Freire, C.S.R.; Silvestre, A.J.D.; Silva, A.M.S. Recent developments in the functionalization of betulinic acid and its natural analogues: A route to new bioactive compounds. *Molecules* **2019**, *24*, E355, doi:10.3390/molecules24020355.
3. Kvasnica, M.; Urban, M.; Dickinson, N.J.; Sarek, J. Pentacyclic triterpenoids with nitrogen- and sulfur-containing heterocycles: Synthesis and medicinal significance. *Nat. Prod. Rep.* **2015**, *32*, 1303–1330, doi:10.1039/c5np00015g.
4. Smirnova, I.E.; Kazakova, O.B. Structure – Anti-influenza Type a Activity Relationship among a Series of Nitrogen Lupane Triterpenoids. *Nat. Prod. Commun.* **2018**, *13*, 1267–1270.
5. Bildziukevich, U.; Rarova, L.; Janovska, L.; Saman, D.; Wimmer, Z. Enhancing effect of cystamine in its amides with betulinic acid as antimicrobial and antitumor agent in vitro. *Steroids* **2019**, *148*, 91–98.
6. Khusnutdinova, E.F.; Apryshko, G.N.; Petrova, A.V.; Kukovinets, O.S.; Kazakova, O.B. The synthesis and selective cytotoxicity of new Mannich bases derivatives of 19- and 28-alkynyltriterpenoids. *Russ. J. Org. Chem.* **2018**, *1*, 123–127, doi:10.1134/S1068162018010090.
7. Bildziukevich, U.; Kaletova, E.; Saman, D.; Sievanen, E.; Kolehmainen, E.T.; Slouf, M.; Wimmer, Z. Spectral and microscopic study of self-assembly of novel cationic spermine amides of betulinic acid. *Steroids* **2017**, *117*, 90–96, doi:10.1016/j.steroids.2016.07.007.
8. Kahnt, M.; Hoenke, S.; Fischer, L.; Al-Harrasi, A.; Csuk, R. Synthesis and cytotoxicity evaluation of DOTA-conjugates of ursolic acid. *Molecules* **2019**, *24*, E2254, doi:10.3390/molecules24122254.
9. Foster, T.J. The *Staphylococcus aureus* “superbug”. *J. Clin. Invest.* **2004**, *114*, 1693–1696, doi:10.1172/JCI23825.
10. Boman, H.G. Antibacterial peptides: Key components needed in immunity. *Cell* **1991**, *65*, 205–207, doi:10.1016/0092-8674(91)90154-q.

11. Zasloff, M. Antibacterial molecules from frogs, sharks and man. In *Phylogenetic Perspective in Immunity: The Insect-Host Defense*. Hoffmann, J., Natori, S., Janeway, C., Eds.; RG Landes Biomedical Publisher, Austin, Texas, USA, **1994**; pp. 31–41.
12. Cho, J.; Kim, Y. Sharks: A potential source of antiangiogenic factors and tumor treatments. *Marine Biotechnology* **2002**, *4*, 521–525, doi:10.1007/s10126-002-0064-3.
13. Moore, K.S.; Wehrli, S.; Roder, H.; Rogers, M.; Forrest, J.N., Jr.; McCrimmon, D.; Zasloff, M. Squalamine: An aminosterol antibiotic from the shark. *Proc. Natl. Acad. Sci.* **1993**, *90*, 1354–1358, doi:10.1073/pnas.90.4.1354.
14. Brunel, J.M.; Letourneux, Y. Recent advances in the synthesis of spermine and spermidine analogs of the shark aminosterol squalamine. *Eur. J. Org. Chem.* **2003**, 3897–3907, doi:10.1002/ejoc.200300167.
15. Alhanout, K.; Rolain, J.M.; Brunel, J.M. Squalamine as an example of a new potent antimicrobial agent class: A critical review. *Curr. Med. Chem.* **2010**, *17*, 3909–3917, doi:10.2174/092986710793205417.
16. Kazakova, O.B.; Giniyatullina, G.V.; Medvedeva, N.I.; Tolstikov, G.A. Synthesis of a triterpene-spermidine conjugate. *Russ. J. Org. Chem.* **2012**, *45*, 1366–1369, doi:10.1134/S107042801.
17. Djouhri-Bouktab, L.; Vidal, N.; Rolain, J.M.; Brunel, J.M. Synthesis of new 3,20-bispolyaminosteroid squalamine analogues and evaluation of their antimicrobial activities. *J. Med. Chem.* **2011**, *54*, 7417–7421, doi:10.1021/jm200506x.
18. Blanchet, M.; Borselli, D.; Rodallec, A.; Peiretti, F.; Vidal, N.; Bolla, J.M.; Digiorgio, C.; Morrison, K.R.; Wuest, W.M.; Brunel, J.M. Claramines: A new class of broad-spectrum antimicrobial agents with bimodal activity. *Chem. Med. Chem.* **2018**, *13*, 1018–1027, doi:10.1002/cmdc.201800073.
19. Bildziukevich, U.; Vida, N.; Rarova, L.; Kolar, M.; Saman, D.; Havlicek, L.; Drasar, P.; Wimmer, Z. Polyamine derivatives of betulinic acid and β -sitosterol: A comparative investigation. *Steroids* **2015**, *100*, 27–35, doi:10.1016/j.steroids.2015.04.005.
20. Medvedeva, N.I.; Kazakova, O.B.; Lopatina, T.V.; Smirnova, I.E.; Giniyatullina, G.V.; Baikova, I.P.; Kataev, V.E. Synthesis and antimycobacterial activity of triterpenic A-ring azepanes. *Eur. J. Med. Chem.* **2018**, *143*, 464–472, doi:10.1016/j.ejmech.2017.11.035.
21. Khusnutdinova, E.F.; Petrova, A.V.; Nguyen, Thi Thu Ha; Le, Thi Tu Anh; Nguyen, Thanh Tra; Ba, Thi Cham; Babkov, D.A.; Kazakova, O.B. Structural modifications of 2,3-indolobetulinic acid: Design and synthesis of highly potent α -glucosidase inhibitors. *Bioorg. Chem.* **2019**, *88*, article 102957, doi:10.1016/j.bioorg.2019.102957.
22. Dalla-Vechia, L.; Dassonville-Klimpt, A.; Grellier, P.; Sonnet, P.; Gosmann G.; Gnoatto, S.C.B. The Beckmann rearrangement applied to ursolic acid with antimalarial activity in medicinal chemistry studies. *Lett. Org. Chem.* **2012**, *9*, 92–95, doi:10.1002/chin.201228213.
23. Giniyatullina, G.V.; Kazakova, O.B.; Salimova, E.V.; Tolstikov, G.A. Synthesis of new betulonic and oleanolic acid amides. *Chem. Nat. Compd.* **2011**, *47*, 68–72, doi:10.1007/s10600-011-9832-5.
24. Flekhter, O.B.; Ashavina, O.Y.; Boreko, E.I.; Karachurina, L.T.; Pavlova, N.I.; Kabal'nova, N.N.; Savinova, O.V.; Galin, F.Z.; Nikolaeva, S.N.; Zarudii, F.S.; et al. Synthesis of 3-acetylbetulinic acid and betulonic aldehydes according to Svern and the pharmacological activity of related oximes. *Pharm. Chem. J.* **2002**, *36*, 303–306, doi:10.1023/A:1020824506140.