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Prognostic significance of *NAB2-STAT6* fusion variants and *TERT* promotor mutations in solitary fibrous tumors/hemangiopericytomas of the CNS: Not (yet) clear.

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Department of Pathology, Radboud University Medical Center, Nijmegen, The Netherlands Rob.Vogels@radboudumc.nl Grading of meningeal solitary fibrous tumors/hemangiopericytomas (SFTs/HPCs) of the central nervous system (CNS) is nowadays based on histologic criteria as described in the revised fourth edition of the WHO Classification of CNS tumors [10] or the more recently published, updated version of the Marseille Grading System (MGS) [11]. Histology based grading of CNS SFTs/HPCs allows for discriminating subgroups with significant differences in prognosis. However, the often-piecemeal resection of these tumors may hamper adequate evaluation of mitotic activity and necrosis, and thereby assessment of malignancy grade. NAB2-STAT6 fusion is the molecular hallmark of both soft tissue SFTs and CNS SFTs/HPCs, and the resulting fusion protein accumulates in the nucleus and acts as a transcriptional activator of early growth response mediated pathways with STAT6 immunohistochemistry being a very sensitive and specific tool for their diagnosis [5, 8, 12, 14]. For soft tissue SFTs, particular NAB2–STAT6 fusion variants as well as telomerase reverse transcriptase (TERT) promoter mutations leading to telomerase activity and tumor cell immortalization have been reported to have prognostic value. Some studies have included CNS SFTs/HPCs in their cohort, but because of small numbers and lack of (long term) follow-up data the prognostic value of these markers for CNS SFTs/HPCs is still unclear [1-4, 6, 7, 9, 13, 15, 16].

To evaluate the prognostic value of *NAB2–STAT6* fusion variants and *TERT* promoter mutations for CNS SFTs/HPCs, we retrospectively analyzed these markers in a cohort of 136 patients with STAT6 nucleopositive CNS tumors. All tumors were graded according to the most recent WHO classification [10] and the updated MGS [11] and were analyzed for type of *NAB2–STAT6* fusion and the presence of *TERT* promoter mutation. For *NAB2–STAT6* fusion analysis, we performed reverse transcriptase polymerase chain reaction (RT-PCR) after RNA extraction and cDNA synthesis using multiple primer sets finding the most common fusion variants which were subsequently grouped based on their hypothesized functional effects [3, 15].

After DNA isolation, amplicons of 163 base pairs spanning *TERT* promoter hot-spot mutations at positions 1,295,228 and 1,295,250 on chromosome 5 were amplified by PCR using the primers hTERT-F CAG CGC TGC CTGAAA CTC and hTERT-R GTC CTG CCC CTT CAC CTT and subsequent sequencing of the products was performed. More detailed information on Materials and Methods is given in the Supplementary Information. For survival analyses, patients who died due to complications of initial therapy (e.g., surgery or radiotherapy) were excluded. Results of

	NAB2-STAT6						TERTp			
	All	Ex4-Ex2	Ex6-Ex16/17	Neg	NA	P value	WT	Mutation	NA	P value
MGS										
1	71	24	24	7	16		43	21	7	
2	54	4	29	10	11		34	15	5	
3	11	0	6	2	3		5	3	3	
Total	136					0.006				0.519
WHO										
1	24	13	1	1	9		15	8	1	
11	47	11	23	6	7		28	13	6	
111	65	4	35	12	14		39	18	8	
Total	136					< 0.0001				0.854
Extradu	ral met	astasis								
Yes	17	0	11	3	3		15	2	0	
No	57	13	33	4	7		33	16	8	
Total	74					0.121				0.09

Table 1 Results of NAB2– STAT6 fusion analysis and TERT promoter mutation analysis: correlation with histological grading and extradural metastatic disease

Significant if p equal or below 0.05

NA not assessable, WT TERTp-wildtype, MGS updated Marseille Grading System

histological grading, analysis of *NAB2–STAT6*f usion and *TERT* promoter mutations and survival analysis are listed in Tables 1 and 2.

Table 2 Association of NAB2–STAT6 fusion analysis and TERT promoter mutation analysis with survival

	No. of patients	PFS (m	onths)		OS (months)			
		Mean	95% CI	Log-rank	Mean	95% CI	Log-rank	
NAB2-STAT6								
All	128ª	124	93-155		346	255-436		
Ex4-Ex2	26	140	94-187		306	265-348		
Ex6-Ex16/17	55	131	83-178		307	186-428		
Neg	17	88	55-122		133	96-171		
NA	30	79	50-107		189	164-215		
				0.280			0.172	
TERTp								
All	128ª	124	93-155		346	255-436		
WT	77	126	85-166		266	151-382		
Mutation	36	111	71-151		237	198-275		
NA	15	90	93-155		272	190-356		
				0.635			0.107	

PFS progression-free survival, OS overall survival, NA not assessable, WT TERTp-wildtype

*8 patients died due to complications of initial therapy (e.g., surgery or radiotherapy) and were excluded for survival analysis

Eighty-seven percent of the tumors could be analyzed for *NAB2–STAT6* fusion variants using RT-PCR (n = 106), in the remaining 30 cases RNA quality was insufficient. Of the 106 tumors, 26% had an exon4–exon2 fusion (n = 28) and 56% (n = 59) an exon6–exon16/17 fusion. In 18% of the tumors (n = 19), no fusion was detected, which may partly be due to our RT-PCR approach which did not cover all possible, rarely detected *NAB2–STAT6* fusion variants.

In our cohort, the *NAB2–STAT6* fusion variants are correlated to both WHO and MGS malignancy grade: The exon6–exon16/17 fusion is significantly more frequent in WHO grade II and III and MGS grade 2 and 3 tumors(p < 0.0001 and p = 0.006, respectively).

Information regarding extradural metastatic disease was available for 74 cases, and 17 of these patients had metastases during their follow-up. In this group of 17 patients, 11(64%) had a tumor with exon6–exon16/17 gene fusion and none an exon4–exon2 fusion. In three cases, no fusion could be detected with our RT-PCR approach and in three cases, RNA quality was insufficient. The results were not statistically significant (p = 0.121), possibly due to the limited number of patients with extradural metastatic disease (n = 17).

In addition, a non-significant trend (p = 0.172) towards shorter overall survival was noted for the exon6–exon16/17 subgroup: of the 19 patients who died from the disease, 12had a tumor with exon6–exon16/17 fusion and only 2 withexon4–exon2 fusion. In the remaining five cases, our fusion analysis approach did not allow for identification of the exact type of *NAB2–STAT6* fusion (n = 3) or could not be assessed due to poor RNA quality (n = 2). *NAB2–STAT6* gene fusion was not correlated to progression-free survival (p = 0.280). *TERT* promoter mutation status could successfully be analyzed in 89% of the tumors (n = 121). Tumors with hot-spot mutations in this promoter region were grouped as '*TERT*p-mutant' and those without such mutations as '*TERT*p-wildtype'. In 68% of the cases, the tumor was found to be *TERT*p-wildtype (n = 82), and in 32% *TERT*p-mutant (n = 39). *TERT* promoter mutation was not significantly correlated with *NAB2–*

STAT6 fusion type (p = 0.164), WHO grade (p = 0.854) or MGS grade (p = 0.519). In addition, *TERT* promoter mutation was not significantly correlated with metastatic disease (p = 0.090), progression-free survival(p = 0.635) or overall survival (p = 0.107), albeit a trend was noted for less aggressive clinical course in *TERT*p-mutant tumors, which is contradictory to published results in soft tissue SFT [2, 7]. Survival curves for *NAB2–STAT6* fusion groups and the presence of *TERT* promoter mutation are provided in the Supplementary Information.

In summary, although in our relatively large cohort of CNS SFTs/HPCs the type of *NAB2–STAT6* fusion lacks significant prognostic value, tumors with an exon6–exon16/17 fusion do show a tendency towards more malignant behavior compared to tumors with exon4–exon2 fusion. This difference in aggressiveness may be explained by the presence of different functional domains within the chimeric *NAB2–STAT6* protein in different fusion groups, with, e.g., the CID domain of *NAB2* only integrated in the exon4–exon2 variant resulting in EGR1 activation and fibrosis as seen in the low-grade groups as hypothesized by Barthelme β et al. [3]. Furthermore, in contrast to studies on mainly extradural SFTs in which *TERT* promoter mutation was shown to indicate poor disease-free survival, in our cohort clear prognostic value of *TERT* promoter mutation status was lacking. Based on our study we conclude that so far, histology-based grading incorporating mitotic activity and necrosis remains the best indicator of prognosis in SFTs/HPCs of the CNS. However, given the trend towards more malignant behavior in the exon6–exon16/17 fusion group future and larger studies are needed to sort out if esp. *NAB2–STAT6* fusion analysis may be of additional value in histology-based prognostic models after all.

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Supplementary Information Text

Materials and Methods

Patient cohort and sample selection

Patients with a diagnosis of meningeal SFT or HPC were retrospectively selected from the (referral) files of the authors. For each case, one representative tumor tissue block of the resection specimen was retrieved from the pathology archives of the participating hospitals and included for histomorphological assessment and gene fusion analysis.

Clinical data including age at first histological diagnosis and follow-up data were extracted from the patient files. Extradural metastatic disease was detected by imaging techniques (with no other malignancies known) and/or proven by histological assessment. Progression-free survival is counted in months from date of first histological diagnosis to date of diagnosis of the (first) local recurrence, growth of residual disease for which therapy was required or detection of metastasis. Overall survival is counted in months from date of first histological diagnosis to date of last known follow-up or date of death. Distinction was made between patients who died due to disease-related factors (i.e. disease-specific survival; DSS) and patients who died due to other causes.

This study was approved by the local scientific review board and was performed in accordance with the Code of Conduct of the Federation of Medical Scientific Societies in the Netherlands. The French Samples used in this study were stored and retrieved from the APHM Tumor Bank (authorization number: AC2018-31053; CRB BB-0033-00097), which respects the ethical charter of the French National Cancer Institute to store and deliver samples for scientific research according to the French Public Health Code (articles L. 1243-4 and R. 1243-61). All samples were obtained after informed consent from patients or their parents when the patients were under the age of 18. As this research was not interventional, it did not require an approval by an ethics committee. The French committee for the treatment of biomedical research information approved the data management of this study (C.C.T.I.R.S. - 09.084Ter).

Assessment of histomorphological features

Four µm thick sections were stained with hematoxylin and eosin and all cases were reviewed by three of the authors (UF, BK, RV). Diagnosis and WHO grade were assigned according to the 2016 WHO

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Classification of Tumours of the Central Nervous System [WHO]. The updated Marseille Grading System [Macagno 2018] was applied on all cases by two authors (NM, CB).

All cases were confirmed by STAT6 nuclear staining of tumor cells using an already published protocol [1].

NAB2-STAT6 gene fusion analysis and TERT promoter mutation analysis

NAB2-STAT6 gene fusion analysis and *TERT* promoter mutation analysis was performed on all cases. Briefly, RNA was extracted from formalin-fixed, paraffin-embedded tissue using RNA-Bee-RNA isolation reagent (Bio-Connect BV, Huissen, the Netherlands). RNA quantity and quality were determined by a NanoDrop measurement (Fisher Scientific, Landsmeer, the Netherlands) and subsequently, cDNA synthesis was performed using Superscript II (Invitrogen Life Technologies Europe, Bleiswijk, the Netherlands) and random hexamers (Promega Nederland, Leiden, the Netherlands). The cDNA was tested by the reverse transcription-polymerase chain reaction (RT-PCR) for the HMBS (hydroxymethylbilase synthase) housekeeping gene using the primers forw150 5'-TGCCAGAGAAGAGTGTGGTG-3' and rev150 5'-ATGATGGCACTGAACTCCTG-3'. Samples without cDNA template were amplified in parallel as negative controls.

For the detection of the most common *NAB2-STAT6* gene fusions, three primers in *NAB2* (NM_005967.3): forw 5'- CAAGTAGCCCGAGAGAGCAC-3' (exon 3), forw 5'- CTCCACTGAAGAAGCTGAAAC-3' (exon 4) and forw 5'-CTGTGTGCCTGCGAAGCC-3' (exon 6) were used in combination with three primers in *STAT6* (NM_001178078.1): rev 5'-GGGAAAGTCGACATAGAGCC-3' (exon 2), rev 5'-

GAGCTGAGCAAGATCCCGG-3' (exon 16) and rev 5'-TTCCACGGTCATCTTGATGG-3' (exon 17). The PCR products were analyzed by agarose gel electrophoresis.

PCR was applied to amplify the TERT promoter region using the following primers: hTERT-F:

CAGCGCTGCCTGAAACTC and hTERT-R GTCCTGCCCCTTCACCTT, amplifying a 163-bp fragment spanning hot-spot mutations at positions 1,295,228 and 1,295,250 on chromosome 5. Sequencing was done on an Illumina MiSeq next generation sequencer. Adapter ligation and barcoding was done using the NEBNext Ultra DNA Library Prep Mastermix Set and NEBNext Multiplex Oligos from New England Biolabs. CLC Cancer Research Workbench software from QIAGEN[®] was performed for sequence analysis. Described briefly, the analysis workflow included adapter trimming and read pair merging before mapping to the human reference genome (hg19). Insertions and deletions as well as single nucleotide variants were detected. Additional information regarding potential mutation type, known single nucleotide polymorphisms and conservation scores was obtained by cross-referencing various databases (COSMIC, ClinVar, dbSNP, 1000 Genomes Project, HAPMAP and PhastCons-Conservation_scores_hg19).

Statistical analysis

Statistical analysis was performed using IBM SPSS Statistics v22.0.

Comparison of different NAB2-STAT6 fusion variants with various clinical and histomorphological

parameters were evaluated by using the χ^2 - or Fisher's exact test as appropriate.

Progression-free- and disease-specific overall survival were used as end points as defined above. Kaplan-

Meier curves were plotted for visualizing survival in different NAB2-STAT6 fusion variant groups and the

log-rank test was used for comparing the difference between the groups. For survival analysis, patients

who died due to complications of initial therapy (e.g. surgery or radiotherapy) were excluded.

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Figure 1a: Overall survival in different *NAB2-STAT6* fusion groups of solitary fibrous tumors/hemangiopericytomas of the central nervous system



Log-rank p = 0.172

Figure 1b: Overall survival in *TERT* promoter-mutant versus *TERT* promoter-wildtype solitary fibrous tumors/hemangiopericytomas of the central nervous system



Log-rank p = 0.107

Figure 2a: Progression-free survival in different *NAB2-STAT6* fusion groups of solitary fibrous tumors/hemangiopericytomas of the central nervous system



Log-rank p = 0.280

Figure 2b: Progression-free survival in *TERT* promoter-mutant versus *TERT* promoter-wildtype solitary fibrous tumors/hemangiopericytomas of the central nervous system



Log-rank p = 0.635