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## **Molecular characterization of fast-growing melanomas**

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**Keywords:** Melanoma, fast growth melanoma, mutations of poor prognosis, FGFR2 mutations

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#### **Abstract**

Background: The rate of growth of primary melanoma is a robust predictor of aggressiveness, but the mutational profile of fast-growing melanomas (FGMM), and its potential to stratify patients at high risk of death, has not been comprehensively studied.

Objective: To investigate the epidemiological, clinical and mutational profile of primary 77 cutaneous melanomas with a thickness  $\geq 1$ mm, stratified by rate of growth (ROG).

Methods: Observational prospective study. Deep-targeted sequencing of 40 melanoma driver genes on formalin fixed, paraffin embedded primary melanoma samples. Comparison of FGMM (ROG >0.5mm/month) and non-FGMM (ROG≤0.5 mm/month).

Results*:* Two hundred patients were enrolled among which 70 were FGMM. The 83 relapse free survival was lower in the FGMM group (p=0.014). FGMM had a higher number of predicted deleterious mutations within the 40 genes than non-FGMM 85 ( $p=0.033$ ). Ulceration ( $p=0.032$ ), thickness ( $p=0.006$ ), lower sun exposure ( $p=0.049$ ), and FGFR2 mutations (p=0.037) were significantly associated with fast growth.

87 Limitations: Single-center study, cohort size, potential memory bias, number of investigated genes.

Conclusion: Fast growth is linked to specific tumor biology and environmental factors. Ulceration, thickness and FGFR2 mutations associate with fast growth. Screening for FGFR2 mutations might provide an additional tool to better identify FGMM which are probably good candidates for adjuvant therapies.

## **Capsule summary**

- Fast growing melanomas are aggressive and linked to early death. Rapid growth is associated to thicker, ulcerated tumors with FGFR2 mutations; and is more frequent in patients with less accumulated sun exposure.
- 98 Ulceration, thickness and FGFR2 mutations are biomarkers for aggressive disease and can stratify patients for adjuvant therapy.
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## **Graphical abstract**



## **Introduction**

106 The incidence of cutaneous melanoma continues to rise worldwide<sup>1-4</sup>. Prevention campaigns promoting early diagnosis underpin an epidemiological shift 108 towards earlier stage disease<sup>5-10</sup> and the majority of new melanoma cases are 109 diagnosed at localized stages $8,11,12$ . Stage II tumors are considered low risk<sup>13</sup>, however, the prediction is that, due to the large prevalence of stage II, they will 111 account for most deaths in the future $8,11,12$ .

Targeted and immunotherapies have transformed the care for metastatic melanomas, and adjuvant trials have demonstrated a high efficacy. New trials are testing anti-PD1 in AJCC IIB/IIC melanomas, expanding the pool of potential candidates for adjuvant immunotherapy. Identification of new biomarkers of aggressiveness is paramount to optimize risk-ratio toxicity and ensure optimal resource allocation.

Primary melanoma growth, defined as the ratio of tumor thickness to patient-119 reported time of melanoma growth<sup>14</sup>, is a validated robust, reproducible and 120 independent prognostic factor of outcome<sup> $14-17$ </sup>. Patients with fast-growing melanoma 121 (FGMM) have aggressive disease that spreads early, leading to shorter survival<sup>14-17</sup>. Fast growth is associated to nodular subtype, trunk location, male sex, previous non-123 melanoma skin cancer, and few childhood sunburns<sup>15-17</sup>. Furthermore, FGMM display 124 high mitotic rates<sup>15-17</sup>, and frequent NRAS<sup>18</sup> and TERT promoter mutations<sup>19</sup>, highlighting growth kinetics is a relevant feature that accurately represents melanoma aggressiveness. The mutational profile of FGMM has not been comprehensively studied, so we compared the epidemiological, clinical and mutational profile of a cohort of FGMM and non-FGMM primary cutaneous melanomas.

#### **Methods**

#### **Clinical data**

We enrolled prospectively consecutive primary melanoma patients with a thickness ≥1mm referred to the dermato-oncology unit of La Timone Hospital, Marseilles, France, between February 2012 and October 2014. We collected epidemiological and clinical characteristics (Mendeley supplemental methods). We calculated rate of growth (ROG) as the ratio between thickness and time taken to 137 melanoma development<sup>14</sup>. FGMM were defined by ROG>0.5mm/month and non-138 FGMM by ROG≤0.5 mm/month<sup>15-17</sup>. Patients filled out a standardized questionnaire to estimate sun exposure (Mendeley supplementary methods). We derived two sun exposure scores to estimate lifelong sun exposure and lifelong sunburn.

#### **Molecular analyses**

Tumor DNA was extracted from hand-macrodissected FFPE melanoma specimens, and normal DNA from patient-matched peritumoural normal skin or wide local excision. DNA was extracted using the GeneRead DNA FFPE kits (Qiagen, Hilden, Germany) and DNA integrity assessed using the NGS FFPE QC Kit (Agilent Technologies). Histopathological variables were extracted from routine reports.

We performed deep-targeted sequencing of 40 melanoma genes, selected by frequency of mutation in the TCGA database (>7%) and/or specific cancer genes of interest (Mendeley supplementary Table 1). TERT promoter was not analyzed in our panel. Three different pipelines were used for variant filtering validation and manual inspection of the "non-consensual" variants was performed on IGV to avoid false positive calls (Mendeley supplementary methods). Only SNVs predicted as 154 pathogenic or likely pathogenic by the COSMIC<sup>22</sup> or Varsome<sup>23</sup> database were considered in the final analysis.

#### **Gene expression analysis**

Cutaneous primary and metastatic melanomas (SKCM) from The Cancer Genome Atlas<sup>24</sup> were divided into *FGFR2* wildtype (WT) and *FGFR2* mutants based on the presence of a deleterious mutation (missense, frame shift insertion/deletion, nonsense mutation) in *FGFR2*. Thirty five cases with a missense mutation were present with matched clinical and gene expression data, and 437 WT cases. For each sample, the G1/S and G2/M gene expression signature scores were determined by calculating the geometric mean of all genes in each signature. G1/S and G2/M specific genes were those used in Tirosh *et al.*<sup>25</sup> to measure proliferating cells.

Somatic mutation and RNA-seq (log2 transformed RSEM) data were downloaded from UCSC Xena TCGA database (https://xenabrowser.net/datapages/) and statistical analysis was performed in GraphPad Prism v8.2.0. Mann-Whitney test was used to compare expression and scores between *FGFR2* mutants and WT samples.

#### **Statistical analysis**

Continuous variables were expressed as means +/- SD or as median with range and categorical variables as count and percentages. Means were compared by student t-test, and percentages by Chi-Square test (or Fisher's exact test, as appropriate). Univariate and stepwise forward multivariate logistic regression models were used to identify factors associated with FGMM. Variables with a p<.05 in univariate analysis were included in multivariate analyses. Recurrence free survival (RFS), distant metastasis free survival (DMFS) and melanoma specific survival (MSS) were calculated from the melanoma diagnosis. RFS, DMFS and MSS curves were estimated by the Kaplan–Meier method and the log-rank test. All tests were two-sided, and statistical significance was defined as *p*<.05. The false discovery rate 181 was controlled with a Benjamini-Hochberg procedure<sup>26</sup>. Statistical analyses were performed using IBM SPSS Statistics version 20 (IBM SPSS Inc., Chicago, IL, USA). Random forest algorithm, bootstrap and multilayer neural perceptron analyses were used to estimate the robustness of the results by using the randomForest and caret packages of R.

## **Results**

#### **Clinical and epidemiological variables associated with fast growth**

Three hundred and fifty-three patients were referred to our institution during the study period. ROG calculation, DNA extraction and molecular analyses were successfully performed for 200 patients (Mendeley Supplementary Figure 1). We compared the clinical, epidemiological and genetic mutations of FGMM (n=70, ROG >0.5 194 mm/month) to the features of non-FGMM (n=130, ROG  $\leq$  0.5/month). The cohort included 112 men, and the median age at diagnosis was 62 years. Most melanomas were located on the trunk (84%) and lower limbs (30.5%), and the most common histological subtype was superficial spreading melanoma (SSM) (63.5%) followed by nodular melanoma (NM) (29.5%). The median Breslow thickness was 2.25 mm. Approximately one-third of melanomas were ulcerated and 54% had a mitotic 200 rate≥1/mm<sup>2</sup>. Regression was present in 18 samples (9%). A sentinel node biopsy was performed in 170 patients (75%). Twelve patients had clinical node involvement 202 at diagnosis (n=12). The AJCC<sup>27</sup> distribution was 35.5% stage I, 39% stage II, 23.5% stage III and 2% stage IV. The median ROG was 0.26 (IQR 0.09-0.77; Mendeley Supplementary Table 2).

Univariate analysis showed that FGMM were more frequently thick (p<0.001), 206 ulcerated  $(p<0.001)$ , nodular  $(p=0.009)$ , with positive sentinel node  $(p=0.01)$ . In 207 addition, they were less frequently located on the upper limb  $(p=0.047)$ , and more frequent in patients medicated with betablockers (p=0.02). Furthermore, we found an association of less lifetime sun exposure (p=0.04) and sunburns (p=0.034) with FGMM, (Table 1 and Mendeley supplementary Table 3). After multivariate analysis,

only ulceration (p=0.016), thickness (p=0.007) and less lifelong sun exposure (p=0.043) were significantly associated to FGMM (Table 2).

We studied the association between survival and melanoma ROG, and found 61 patients relapsed after a median follow-up period of 59.67 months (28 in the FGMM and 33 in the non-FGMM group). Although the median RFS and median DMFS were not reached, the RFS was significantly lower in the FGMM group (5-year FGMM RFS 58.4%, non-FGMM 73.7%, p=0.014 HR 1.9 (1.1-3.1), Figure 1. We assessed distant recurrences, and documented 56 patients (FGMM=27, non-FGMM= 29). The DMFS was significantly lower in the FGMM cohort (5-year DMFS FGMM 61%, non-FGMM 77.2%, p=0.010 HR 1.9 (1.2-3.3), Mendeley Supplementary Figure 2). Finally, 39 patients died during the follow-up period (FGMM=18, non-FGMM=21), and we found a trend for lower MSS in the FGMM cohort (5-year FGMM MSS 74.1%, non-FGMM 83.5%, p=0.092).

#### **Pathogenic mutations associated with FGMM**

We compared the proportion of samples driven by the common oncogenic melanoma mutations BRAF, RAS or NF1, or WT for BRAF, NRAS, NF1 (triple WT), 229 and confirmed a similar distribution to previous cohorts<sup>24,28</sup>. The most commonly mutated genes were BRAF (49%), NRAS (23.5%) and NF1 (17.5%), together with TP53 (12%), which appeared in similar proportions in FGMM and non-FGMM samples. We explored the association between FGMM and mutation burden in the 40 gene panel. FGMM had a higher number of pathogenic mutations than non-FGMM (mean SNVs FGMM=3.17 +-3.58, vs. non-FGMM=2.13 +- 1.931; p=0.033). We next investigated the association between pathogenic mutations in each gene and FGMM and found at least one mutation in one of the 40 genes in 179 patients. We performed univariate analyses and found a higher proportion of pathogenic mutations in FGFR2, ALK, ERBB4, IDH1, PDGFRA, PREX2 and RB1 in FGMM. We corrected for multiple comparison, confirming FGFR2 and IDH1 mutations were associated with fast growth, and noted 15.7% of FGMM presented FGFR2 mutations, in contrast to 2.3% in the non-FGMM group (p=0.0049 HR 7.81 (1.96-45.25; Table 3, Mendeley Supplementary Tables 4, 5). IDH1 mutations were exclusively found in FGMM at a rate of 5.7% (p=0.049).

We reasoned that if FGFR2 mutations are associated to fast growth, melanomas with FGFR2 mutations should present a transcriptional profile of 246 increased cell proliferation<sup>25</sup>. We studied the transcriptional profile of melanomas with FGFR2 mutations using data from TCGA. As the majority of point mutations in 248 melanoma are acquired early and preserved at the metastatic stage<sup>29</sup>, we opted to include both primary and metastatic samples of the SKM cohort in our analysis. We compared the differentially enriched pathways in FGFR2-mutated and FGFR2-wild 251 type samples and performed unbiased gene set enrichment analysis. Remarkably, the expression of cell cycle genes indicating increased melanoma cell proliferation was significantly higher in FGFR2-mutated melanomas (Mendeley Supplementary Figure 3).

#### **Alterations in genetic pathways associated with fast growth**

We analysed the percentage of samples with protein-affecting aberrations in candidate driver genes, grouped by pathway in our 200 sample cohort (Mendeley Supplementary Figure 4). FGMM more frequently presented mutations in RTKs

(ERBB4/PDGFRA/ROS/RET/ALK/KIT/FGFR2; FGMM=32.9%/non-FGMM=18.5%, p=0.033), cell cycle pathway (CDKNA/CDK4/BCLAF1; FGMM=10%/non-FGMM=3.1%, p=0.053) and the methylation pathway gene IDH1 (FGMM=5.7%/non-FGMM=0, p=0.042) after Benjamini-Hochberg adjustment (Mendeley Supplementary Figure 4).

#### **Integrating clinical and molecular biomarkers of FGMM**

To validate the robustness of our biomarkers, we performed stepwise forward logistic regression analysis including variables that were statistically significantly associated with FGMM (tumor location, histological subtype, ulceration, mitotic rate, thickness, sentinel node biopsy, betablocker consumption, lifelong sun exposure, lifelong sunburn and mutations). The final model, which included 114 patients, confirmed ulceration (p=0.032), thickness (p=0.006), less sun exposure (p=0.049), and FGFR2 mutations (p=0.037) are independent features associated with FGMM (Table 4). To further assess how much FGFR2 mutations contributed to fast growth relative to ulceration and thickness we performed a recursive partitioning analysis (Mendeley Supplementary Figure 5) as well as bootstrapping and a multilayer perception network analysis (Mendeley supplementary Figures 6 and 7). All these analyses demonstrated that FGFR2 mutations allowed additional detection of FGMM beyond the other factors.

## **Discussion**

The rapid growth of primary melanoma is recognized as a marker of poor prognosis, frequently described in tumors with additional hallmarks of aggressive 285 disease<sup>14-17</sup>. We studied the clinical and genetic characteristics of FGMM, and we confirm that FGMM metastasize more rapidly, are frequently ulcerated and thicker

287 and inversely associated to lifetime sun exposure<sup>14-17</sup>. Additionally, we observed that FGMM carry an increased number of pathogenic mutations in melanoma driver genes and in a novel finding, we show a strong association with FGFR2 mutations. These results suggest specific molecular changes and environmental factors affect primary ROG and consequently the disease outcome.

292 Our study confirms previous data $15-17$  and sheds light on possible mechanistic drivers of rapid growth. Extensive studies in cancer research show additive 294 oncogenic mutations increase the severity of cancer  $31,32$  and we show that a higher number of pathogenic mutations in 40 melanoma driver genes is linked to fast growth. This suggests that additive genetic damage to key genes will move melanoma forward at a faster pace. Additionally, we show environmental factors influence the ROG, as patient-reported high levels of sun exposure protects from rapid growth. In keeping with this finding, previous work revealed a higher burden of sun-induced mutations across the genome, affecting primarily non-driver genes, is coupled to better outcome<sup>33</sup>. Taken together, these studies validate that additive oncogenic drivers accelerate melanoma, but high levels of sun damage protect from aggressive disease.

Genomic aberrations in melanoma frequently affect key signaling pathways to tumorigenesis. The most affected pathways are the MAP kinase, PI3 kinase and upstream RTKs. In keeping with a faster proliferation, our study revealed FGMM accumulated more mutations in RTKs and showed a trend for more mutations in genes controlling cell cycle. Significantly, we found a robust association between rapid growth and FGFR2 mutations. FGFR2 is involved in tumor cell proliferation, 310 angiogenesis, migration and survival in multiple tissues $34-36$ , and selective FGFR2 inhibitors show a decrease in tumour cell proliferation and promising results in early

312 phase trials for multiple cancer types with activating FGFR2 mutations $36-39$ . Cutaneous melanoma, however, can present both gain-of-function, oncogenic mutations<sup>34</sup> as well as loss-of-receptor function mutations<sup>40</sup> through multiple mechanisms including lower ligand binding affinity, impaired dimerization and 316 reduced kinase activity $34,36$ . These studies highlight FGFR2 signalling can exert opposing functions, either promoting growth or driving senescence, so it is likely the contribution of FGFR2 varies depending on cellular context and tumour type. The signalling consequences of the majority of FGFR2 mutations documented in 320 melanoma are unknown<sup>41,42</sup>, and further work should address if rapidly growing melanomas with FGFR2 mutations are candidates for targeted inhibitor FGFR2 therapies. Although the number of samples in our cohort is small, we found IDH1 mutations might associate with fast growth. We identified hotspot oncogenic IDH1R132C mutations in 4 patients, exclusively in the FGMM group. IDH1 mutations 325 drive a variety of human cancers in addition to melanoma<sup>43,44</sup>. In vitro studies show that mutant IDH1 confers growth and metabolic advantage to melanoma and cancer cells<sup>43,45,46</sup>, and in glioma models, IDH1/2 mutations may shape the immunological 328 landscape of the tumor microenvironment<sup>47,48</sup>. These findings support that IDH1 mutations might drive more aggressive melanomas.

We acknowledge our study has limitations. Twenty percent of patients were not able to provide information required for ROG calculation and were excluded from the study. The size of the population was relatively small and missing data reduced the number of cases included in the multivariate models. Since ROG is calculated prospectively, and FGFR2 is not routinely analysed, we could not corroborate our data in a validation cohort. Finally, TERT promoter sequencing was not included in 336 our panel and is linked to rapid growth<sup>19</sup>. We additionally focused on targeted

mutational analysis, omitting overall tumour mutation burden, mutational signatures, gene fusions and expression.

The multivariate analysis reveals FGFR2 mutations, thickness and ulceration remain robust independent predictor of rapid melanoma growth, a strong indicator of poor outcome. Only patients with stage III or IV resected melanomas are currently eligible for adjuvant therapies. Given the ongoing trials in stage II melanomas, one of the current challenges is to find biomarkers to identify individuals at highest risk of death, who are most likely to benefit from therapies, and to avoid overtreatment and drug toxicity. Screening for FGFR2 mutations might provide an additional tool to better identify the fast-growing tumors which, given their aggressiveness, should be undoubtedly regarded as strong candidates for adjuvant therapies.

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## **Author contributions**

CG and JJG conceived the study. CG and AV carried out experiments and managed the project. CG, AV, NM, AL, EP, LO, TB, PM, GG, LL, MC, RM, RK, RM, EN contributed to method development and analyses. VA contributed resources and RM funding. CG, AV, JJG and EN interpreted the data. CG and AV wrote the manuscript, EN critically reviewed and edited the manuscript.

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# **Figure legends**

# **Figure 1.** Relapse-free survival by rate of growth in the 200-patient cohort.

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## 513 **Tables**



#### 514 **Table 1. Clinical and epidemiological variables associated with fast growth**

515 Univariate analysis, p value <0.05). \* ALM (acral lentiginous melanoma n=6), LM (lentigo

516 maligna n=2), Desmoplastic (n=1), spitzoïd (n=1), malignant blue (n=1), non-assessable

517 (n=3), SSM: superficial spreading melanoma, NM: nodular melanoma.



519

## 520 **Table 2. Clinical and epidemiological variables associated with fast growth**

521 Multivariate stepwise forward model. \*only variables with a p value <0.05 after univariate

522 analysis were included in the model\*\*ALM, LM, Desmoplastic, spitzoïd, malignant blue, non-

523 assessable

## 525 **Table 3**



526 NE: non estimable

527

## 528 **Table 3. Association of mutations in genes with fast growing and non-fast growing**

## 529 **melanoma**

530 Univariate analysis.

### 532 **Table 4**



533 **Table 4. Variables associated with fast growth**

534 Multivariate stepwise forward model. \*only variables with a p value <0.05 after univariate 535 analysis were included in the model. \*\*ALM, LM, Desmoplastic, spitzoïd, malignant blue, 536 non-assessable.

537 Data missing for the following variables: histological subtype (n=1); ulceration (n=4); SN 538 biopsy results (n=30); mitotic rate (n=43); sun exposure score (lifelong) (n=12); sunburn 539 score (lifelong) (n=18).

