

Molecular characterization of fast-growing melanomas

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1 Molecular characterization of fast-growing melanomas

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62 **IRB statement:** This non-interventional study followed French regulations. All 63 patients gave written informed consent.

64

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 mutations
- 67
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71 **Abstract**

<u>Background</u>: The rate of growth of primary melanoma is a robust predictor of
 aggressiveness, but the mutational profile of fast-growing melanomas (FGMM), and
 its potential to stratify patients at high risk of death, has not been comprehensively
 studied.

⁷⁶ <u>Objective</u>: To investigate the epidemiological, clinical and mutational profile of primary ⁷⁷ cutaneous melanomas with a thickness \ge 1mm, stratified by rate of growth (ROG).

<u>Methods</u>: Observational prospective study. Deep-targeted sequencing of 40
 melanoma driver genes on formalin fixed, paraffin embedded primary melanoma
 samples. Comparison of FGMM (ROG >0.5mm/month) and non-FGMM (ROG≤0.5
 mm/month).

<u>Results</u>: Two hundred patients were enrolled among which 70 were FGMM. The relapse free survival was lower in the FGMM group (p=0.014). FGMM had a higher number of predicted deleterious mutations within the 40 genes than non-FGMM (p=0.033). Ulceration (p=0.032), thickness (p=0.006), lower sun exposure (p=0.049), and FGFR2 mutations (p=0.037) were significantly associated with fast growth.

<u>Limitations</u>: Single-center study, cohort size, potential memory bias, number of
 investigated genes.

<u>Conclusion</u>: Fast growth is linked to specific tumor biology and environmental factors.
 Ulceration, thickness and FGFR2 mutations associate with fast growth. Screening for
 FGFR2 mutations might provide an additional tool to better identify FGMM which are
 probably good candidates for adjuvant therapies.

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94 Capsule summary

- Fast growing melanomas are aggressive and linked to early death. Rapid growth is associated to thicker, ulcerated tumors with FGFR2 mutations; and is more frequent in patients with less accumulated sun exposure.
- Ulceration, thickness and FGFR2 mutations are biomarkers for aggressive
 disease and can stratify patients for adjuvant therapy.
- 100

101 Graphical abstract

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105 Introduction

The incidence of cutaneous melanoma continues to rise worldwide¹⁻⁴. Prevention campaigns promoting early diagnosis underpin an epidemiological shift towards earlier stage disease⁵⁻¹⁰ and the majority of new melanoma cases are diagnosed at localized stages^{8,11,12}. Stage II tumors are considered low risk¹³, however, the prediction is that, due to the large prevalence of stage II, they will account for most deaths in the future^{8,11,12}.

Targeted and immunotherapies have transformed the care for metastatic melanomas, and adjuvant trials have demonstrated a high efficacy. New trials are testing anti-PD1 in AJCC IIB/IIC melanomas, expanding the pool of potential candidates for adjuvant immunotherapy. Identification of new biomarkers of aggressiveness is paramount to optimize risk-ratio toxicity and ensure optimal resource allocation.

Primary melanoma growth, defined as the ratio of tumor thickness to patient-118 reported time of melanoma growth¹⁴, is a validated robust, reproducible and 119 independent prognostic factor of outcome¹⁴⁻¹⁷. Patients with fast-growing melanoma 120 (FGMM) have aggressive disease that spreads early, leading to shorter survival¹⁴⁻¹⁷. 121 Fast growth is associated to nodular subtype, trunk location, male sex, previous non-122 melanoma skin cancer, and few childhood sunburns¹⁵⁻¹⁷. Furthermore, FGMM display 123 high mitotic rates¹⁵⁻¹⁷, and frequent NRAS¹⁸ and TERT promoter mutations¹⁹, 124 highlighting growth kinetics is a relevant feature that accurately represents melanoma 125 aggressiveness. The mutational profile of FGMM has not been comprehensively 126 studied, so we compared the epidemiological, clinical and mutational profile of a 127 128 cohort of FGMM and non-FGMM primary cutaneous melanomas.

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130 Methods

131 Clinical data

We enrolled prospectively consecutive primary melanoma patients with a 132 thickness ≥1mm referred to the dermato-oncology unit of La Timone Hospital, 133 Marseilles, France, between February 2012 and October 2014. We collected 134 epidemiological and clinical characteristics (Mendeley supplemental methods). We 135 calculated rate of growth (ROG) as the ratio between thickness and time taken to 136 melanoma development¹⁴. FGMM were defined by ROG>0.5mm/month and non-137 FGMM by ROG≤0.5 mm/month¹⁵⁻¹⁷. Patients filled out a standardized questionnaire 138 to estimate sun exposure (Mendeley supplementary methods). We derived two sun 139 exposure scores to estimate lifelong sun exposure and lifelong sunburn. 140

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142 Molecular analyses

Tumor DNA was extracted from hand-macrodissected FFPE melanoma specimens, and normal DNA from patient-matched peritumoural normal skin or wide local excision. DNA was extracted using the GeneRead DNA FFPE kits (Qiagen, Hilden, Germany) and DNA integrity assessed using the NGS FFPE QC Kit (Agilent Technologies). Histopathological variables were extracted from routine reports.

We performed deep-targeted sequencing of 40 melanoma genes, selected by frequency of mutation in the TCGA database (>7%) and/or specific cancer genes of interest (Mendeley supplementary Table 1). TERT promoter was not analyzed in our panel. Three different pipelines were used for variant filtering validation and manual inspection of the "non-consensual" variants was performed on IGV to avoid false positive calls (Mendeley supplementary methods). Only SNVs predicted as pathogenic or likely pathogenic by the COSMIC²² or Varsome²³ database were 155 considered in the final analysis.

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157 Gene expression analysis

158 Cutaneous primary and metastatic melanomas (SKCM) from The Cancer Genome Atlas²⁴ were divided into *FGFR2* wildtype (WT) and *FGFR2* mutants based on the 159 presence of a deleterious mutation (missense, frame shift insertion/deletion, 160 nonsense mutation) in FGFR2. Thirty five cases with a missense mutation were 161 present with matched clinical and gene expression data, and 437 WT cases. For 162 each sample, the G1/S and G2/M gene expression signature scores were determined 163 by calculating the geometric mean of all genes in each signature. G1/S and G2/M 164 specific genes were those used in Tirosh *et al.*²⁵ to measure proliferating cells. 165

Somatic mutation and RNA-seq (log₂ transformed RSEM) data were downloaded from UCSC Xena TCGA database (https://xenabrowser.net/datapages/) and statistical analysis was performed in GraphPad Prism v8.2.0. Mann-Whitney test was used to compare expression and scores between *FGFR2* mutants and WT samples.

170 Statistical analysis

171 Continuous variables were expressed as means +/- SD or as median with range and categorical variables as count and percentages. Means were compared by 172 student t-test, and percentages by Chi-Square test (or Fisher's exact test, as 173 appropriate). Univariate and stepwise forward multivariate logistic regression models 174 were used to identify factors associated with FGMM. Variables with a p<.05 in 175 univariate analysis were included in multivariate analyses. Recurrence free survival 176 (RFS), distant metastasis free survival (DMFS) and melanoma specific survival 177 (MSS) were calculated from the melanoma diagnosis. RFS, DMFS and MSS curves 178 were estimated by the Kaplan-Meier method and the log-rank test. All tests were 179

two-sided, and statistical significance was defined as p<.05. The false discovery rate was controlled with a Benjamini-Hochberg procedure²⁶. Statistical analyses were performed using IBM SPSS Statistics version 20 (IBM SPSS Inc., Chicago, IL, USA). Random forest algorithm, bootstrap and multilayer neural perceptron analyses were used to estimate the robustness of the results by using the randomForest and caret packages of R.

187 **Results**

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189 Clinical and epidemiological variables associated with fast growth

Three hundred and fifty-three patients were referred to our institution during the study 190 period. ROG calculation, DNA extraction and molecular analyses were successfully 191 performed for 200 patients (Mendeley Supplementary Figure 1). We compared the 192 clinical, epidemiological and genetic mutations of FGMM (n=70, ROG >0.5 193 mm/month) to the features of non-FGMM (n=130, ROG \leq 0.5/month). The cohort 194 included 112 men, and the median age at diagnosis was 62 years. Most melanomas 195 were located on the trunk (84%) and lower limbs (30.5%), and the most common 196 histological subtype was superficial spreading melanoma (SSM) (63.5%) followed by 197 nodular melanoma (NM) (29.5%). The median Breslow thickness was 2.25 mm. 198 Approximately one-third of melanomas were ulcerated and 54% had a mitotic 199 rate≥1/mm². Regression was present in 18 samples (9%). A sentinel node biopsy 200 was performed in 170 patients (75%). Twelve patients had clinical node involvement 201 at diagnosis (n=12). The AJCC²⁷ distribution was 35.5% stage I, 39% stage II, 23.5% 202 stage III and 2% stage IV. The median ROG was 0.26 (IQR 0.09-0.77; Mendeley 203 Supplementary Table 2). 204

Univariate analysis showed that FGMM were more frequently thick (p<0.001), ulcerated (p<0.001), nodular (p=0.009), with positive sentinel node (p=0.01). In addition, they were less frequently located on the upper limb (p=0.047), and more frequent in patients medicated with betablockers (p=0.02). Furthermore, we found an association of less lifetime sun exposure (p=0.04) and sunburns (p=0.034) with FGMM, (Table 1 and Mendeley supplementary Table 3). After multivariate analysis,

only ulceration (p=0.016), thickness (p=0.007) and less lifelong sun exposure (p=0.043) were significantly associated to FGMM (Table 2).

We studied the association between survival and melanoma ROG, and found 213 61 patients relapsed after a median follow-up period of 59.67 months (28 in the 214 FGMM and 33 in the non-FGMM group). Although the median RFS and median 215 DMFS were not reached, the RFS was significantly lower in the FGMM group (5-year 216 FGMM RFS 58.4%, non-FGMM 73.7%, p=0.014 HR 1.9 (1.1-3.1), Figure 1. We 217 assessed distant recurrences, and documented 56 patients (FGMM=27, non-FGMM= 218 29). The DMFS was significantly lower in the FGMM cohort (5-year DMFS FGMM 219 61%, non-FGMM 77.2%, p=0.010 HR 1.9 (1.2-3.3), Mendeley Supplementary Figure 220 2). Finally, 39 patients died during the follow-up period (FGMM=18, non-FGMM=21), 221 and we found a trend for lower MSS in the FGMM cohort (5-year FGMM MSS 74.1%, 222 non-FGMM 83.5%, p=0.092). 223

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225 Pathogenic mutations associated with FGMM

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We compared the proportion of samples driven by the common oncogenic 227 melanoma mutations BRAF, RAS or NF1, or WT for BRAF, NRAS, NF1 (triple WT), 228 and confirmed a similar distribution to previous cohorts^{24,28}. The most commonly 229 mutated genes were BRAF (49%), NRAS (23.5%) and NF1 (17.5%), together with 230 TP53 (12%), which appeared in similar proportions in FGMM and non-FGMM 231 samples. We explored the association between FGMM and mutation burden in the 40 232 gene panel. FGMM had a higher number of pathogenic mutations than non-FGMM 233 (mean SNVs FGMM=3.17 +-3.58, vs. non-FGMM=2.13 +- 1.931; p=0.033). We next 234 investigated the association between pathogenic mutations in each gene and FGMM 235

and found at least one mutation in one of the 40 genes in 179 patients. We 236 performed univariate analyses and found a higher proportion of pathogenic mutations 237 in FGFR2, ALK, ERBB4, IDH1, PDGFRA, PREX2 and RB1 in FGMM. We corrected 238 for multiple comparison, confirming FGFR2 and IDH1 mutations were associated with 239 fast growth, and noted 15.7% of FGMM presented FGFR2 mutations, in contrast to 240 2.3% in the non-FGMM group (p=0.0049 HR 7.81 (1.96-45.25; Table 3, Mendeley 241 Supplementary Tables 4, 5). IDH1 mutations were exclusively found in FGMM at a 242 rate of 5.7% (p=0.049). 243

We reasoned that if FGFR2 mutations are associated to fast growth, 244 melanomas with FGFR2 mutations should present a transcriptional profile of 245 increased cell proliferation²⁵. We studied the transcriptional profile of melanomas with 246 FGFR2 mutations using data from TCGA. As the majority of point mutations in 247 248 melanoma are acquired early and preserved at the metastatic stage²⁹, we opted to include both primary and metastatic samples of the SKM cohort in our analysis. We 249 250 compared the differentially enriched pathways in FGFR2-mutated and FGFR2-wild samples and performed unbiased gene set enrichment analysis³⁰. 251 type Remarkably, the expression of cell cycle genes indicating increased melanoma cell 252 proliferation was significantly higher in FGFR2-mutated melanomas (Mendeley 253 Supplementary Figure 3). 254

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Alterations in genetic pathways associated with fast growth

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We analysed the percentage of samples with protein-affecting aberrations in candidate driver genes, grouped by pathway in our 200 sample cohort (Mendeley Supplementary Figure 4). FGMM more frequently presented mutations in RTKs

(ERBB4/PDGFRA/ROS/RET/ALK/KIT/FGFR2; FGMM=32.9%/non-FGMM=18.5%, 261 (CDKNA/CDK4/BCLAF1; p=0.033), cell cycle pathway FGMM=10%/non-262 FGMM=3.1%, p=0.053) and the methylation pathway gene IDH1 (FGMM=5.7%/non-263 FGMM=0, p=0.042) after Benjamini-Hochberg adjustment (Mendeley Supplementary 264 Figure 4). 265

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Integrating clinical and molecular biomarkers of FGMM

To validate the robustness of our biomarkers, we performed stepwise forward 268 logistic regression analysis including variables that were statistically significantly 269 associated with FGMM (tumor location, histological subtype, ulceration, mitotic rate, 270 thickness, sentinel node biopsy, betablocker consumption, lifelong sun exposure, 271 lifelong sunburn and mutations). The final model, which included 114 patients, 272 confirmed ulceration (p=0.032), thickness (p=0.006), less sun exposure (p=0.049), 273 and FGFR2 mutations (p=0.037) are independent features associated with FGMM 274 (Table 4). To further assess how much FGFR2 mutations contributed to fast growth 275 relative to ulceration and thickness we performed a recursive partitioning analysis 276 (Mendeley Supplementary Figure 5) as well as bootstrapping and a multilayer 277 278 perception network analysis (Mendeley supplementary Figures 6 and 7). All these analyses demonstrated that FGFR2 mutations allowed additional detection of FGMM 279 beyond the other factors. 280

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Discussion 282

The rapid growth of primary melanoma is recognized as a marker of poor 283 prognosis, frequently described in tumors with additional hallmarks of aggressive 284 disease¹⁴⁻¹⁷. We studied the clinical and genetic characteristics of FGMM, and we 285 confirm that FGMM metastasize more rapidly, are frequently ulcerated and thicker 286

and inversely associated to lifetime sun exposure¹⁴⁻¹⁷. Additionally, we observed that FGMM carry an increased number of pathogenic mutations in melanoma driver genes and in a novel finding, we show a strong association with FGFR2 mutations. These results suggest specific molecular changes and environmental factors affect primary ROG and consequently the disease outcome.

Our study confirms previous data¹⁵⁻¹⁷ and sheds light on possible mechanistic 292 drivers of rapid growth. Extensive studies in cancer research show additive 293 oncogenic mutations increase the severity of cancer^{31,32} and we show that a higher 294 number of pathogenic mutations in 40 melanoma driver genes is linked to fast 295 growth. This suggests that additive genetic damage to key genes will move 296 melanoma forward at a faster pace. Additionally, we show environmental factors 297 influence the ROG, as patient-reported high levels of sun exposure protects from 298 rapid growth. In keeping with this finding, previous work revealed a higher burden of 299 sun-induced mutations across the genome, affecting primarily non-driver genes, is 300 coupled to better outcome³³. Taken together, these studies validate that additive 301 oncogenic drivers accelerate melanoma, but high levels of sun damage protect from 302 aggressive disease. 303

Genomic aberrations in melanoma frequently affect key signaling pathways to 304 tumorigenesis. The most affected pathways are the MAP kinase, PI3 kinase and 305 upstream RTKs. In keeping with a faster proliferation, our study revealed FGMM 306 accumulated more mutations in RTKs and showed a trend for more mutations in 307 genes controlling cell cycle. Significantly, we found a robust association between 308 309 rapid growth and FGFR2 mutations. FGFR2 is involved in tumor cell proliferation, angiogenesis, migration and survival in multiple tissues³⁴⁻³⁶, and selective FGFR2 310 inhibitors show a decrease in tumour cell proliferation and promising results in early 311

phase trials for multiple cancer types with activating FGFR2 mutations³⁶⁻³⁹. 312 Cutaneous melanoma, however, can present both gain-of-function, oncogenic 313 mutations³⁴ as well as loss-of-receptor function mutations⁴⁰ through multiple 314 mechanisms including lower ligand binding affinity, impaired dimerization and 315 reduced kinase activity^{34,36}. These studies highlight FGFR2 signalling can exert 316 opposing functions, either promoting growth or driving senescence, so it is likely the 317 contribution of FGFR2 varies depending on cellular context and tumour type. The 318 signalling consequences of the majority of FGFR2 mutations documented in 319 melanoma are unknown^{41,42}, and further work should address if rapidly growing 320 melanomas with FGFR2 mutations are candidates for targeted inhibitor FGFR2 321 therapies. Although the number of samples in our cohort is small, we found IDH1 322 mutations might associate with fast growth. We identified hotspot oncogenic 323 324 IDH1R132C mutations in 4 patients, exclusively in the FGMM group. IDH1 mutations drive a variety of human cancers in addition to melanoma^{43,44}. In vitro studies show 325 that mutant IDH1 confers growth and metabolic advantage to melanoma and cancer 326 cells^{43,45,46}, and in glioma models, IDH1/2 mutations may shape the immunological 327 landscape of the tumor microenvironment^{47,48}. These findings support that IDH1 328 mutations might drive more aggressive melanomas. 329

We acknowledge our study has limitations. Twenty percent of patients were not able to provide information required for ROG calculation and were excluded from the study. The size of the population was relatively small and missing data reduced the number of cases included in the multivariate models. Since ROG is calculated prospectively, and FGFR2 is not routinely analysed, we could not corroborate our data in a validation cohort. Finally, TERT promoter sequencing was not included in our panel and is linked to rapid growth¹⁹. We additionally focused on targeted

mutational analysis, omitting overall tumour mutation burden, mutational signatures,gene fusions and expression.

The multivariate analysis reveals FGFR2 mutations, thickness and ulceration remain 339 robust independent predictor of rapid melanoma growth, a strong indicator of poor 340 outcome. Only patients with stage III or IV resected melanomas are currently eligible 341 for adjuvant therapies. Given the ongoing trials in stage II melanomas, one of the 342 current challenges is to find biomarkers to identify individuals at highest risk of death, 343 who are most likely to benefit from therapies, and to avoid overtreatment and drug 344 toxicity. Screening for FGFR2 mutations might provide an additional tool to better 345 identify the fast-growing tumors which, given their aggressiveness, should be 346 undoubtedly regarded as strong candidates for adjuvant therapies. 347

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354

Author contributions

356 CG and JJG conceived the study. CG and AV carried out experiments and managed 357 the project. CG, AV, NM, AL, EP, LO, TB, PM, GG, LL, MC, RM, RK, RM, EN 358 contributed to method development and analyses. VA contributed resources and RM 359 funding. CG, AV, JJG and EN interpreted the data. CG and AV wrote the manuscript, 360 EN critically reviewed and edited the manuscript.

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502

504 Figure legends

Figure 1. Relapse-free survival by rate of growth in the 200-patient cohort.

513 Tables

Variable	Non-FGMM	FGMM	OR 95%	р
	(n=130)	(n=70)		-
Melanoma location				
Head and Neck	11 (8.5%)	9 (12.9%)	1.26 (0.47-3.38)	0.640
Trunk	51 (39.2%)	33 (47.1%)	1	
Upper Limb	20 (15.4%)	4 (5.7%)	0.31 (0.10-0.99)	0.047
Lower Limb	41 (31.5%)	20 (28.6%)	0.75 (0.38-1.51)	0.423
Hands/Feet/Palm/Nail	7 (5.4%)	4 (5.7%)	0.88 (0.24-3.25)	0.852
Histological subtype				
SSM	90 (69.2%)	37 (52.9%)	1	
NM	30 (23.1%)	29 (41.4%)	2.35 (1.24-4.45)	0.009
Other *	10 (7.7%)	4 (5.7%)	1.08 (0.31-3.73)	0.902
Ulceration				
No	92 (73%)	32 (45.7%)	1	
Yes	34 (27%)	38 (54.3%)	3.21 (1.74-5.93)	<0.001
Missing	4	0		
Thickness (mm)				
Median	1.8 (1.4-3)	4 (2.5-6)	1.61 (1.34-1.93)	<0.001
Mean	2.4 ±1.58	5.70 ±6.68		
1.00-2.00	80 (61.5%)	10 (14.3%)	1	
2.01-4.00	33 (25.4%)	30 (42.9%)	7.27 (3.20-16.56)	<0.001
>4.00	17 (13.1%)	30 (42.9%)	14.12 (5.82-34.26)	<0.001
Sentinel node biopsy				
Positive	19 (16.5%)	19 (34.5%)	2.67 (1.27-5.60)	0.010
Negative	96 (83.5%)	36 (65.5%)	1	
Missing	16	14		
AJCC (7 th classification)				
1	63 (48.5%)	8 (11.4%)	1	
П	43 (33.1%)	35 (50%)	6.41 (2.71-15.11)	<0.001
111	22 (16.9%)	25 (35.7%)	8.95 (3.52-22.74)	<0.001
IV	2 (1.5%)	2 (2.9%)	7.88 (0.97-63.89)	0.053
Beta blokers				
No	123 (94.6%)	59 (84.3%)	1	
Yes	7 (5.4%)	11 (15.7%)	3.28 (1.21-8.88)	0.020
Mean sun exposure score				
during childhood	5.41 ±1.66	4.71 ±1.34	0.75 (0.61-0.92)	0.005
during adulthood	6.88 ±2.11	6.34 ±1.84	0.87 (0.75-1.02)	0.008
all life long	12.3 ±3.42	10.88 ±2.5	0.86 (0.78-0.96)	0.004
Mean sunburn score			. ,	
during childhood	1.55 ±0.9	1.25 ±0.8	0.68 (0.47-0.97)	0.033
during adulthood	1.55 ±0.9	1.33 ±0.9	0.77 (0.55-1.08)	0.124
lifelong	3.10 ±1.6	2.57 ±1.5	0.80 (0.65-0.98)	0.034

514 Table 1. Clinical and epidemiological variables associated with fast growth

515 Univariate analysis, p value <0.05). * ALM (acral lentiginous melanoma n=6), LM (lentigo

516 maligna n=2), Desmoplastic (n=1), spitzoïd (n=1), malignant blue (n=1), non-assessable

517 (n=3), SSM: superficial spreading melanoma, NM: nodular melanoma.

518

	non-FGMM	FGMM	Univariate		Multivariate	
Variables*	(n=130)	(n=70)	OR 95%	р	OR 95%	р
Melanoma location						
Head and Neck	11 (8.5%)	9 (12.9%)	1.26 (0.47-3.38)	0.64		
Trunk	51 (39.2%)	33 (47.1%)	1		1	
Upper Limb	20 (15.4%)	4 (5.7%)	0.31 (0.10-0.99)	0.047		
Lower Limb	41 (31.5%)	20 (28.6%)	0.75 (0.38-1.51)	0.423		
Hands/Feet/palm/nail	7 (5.4%)	4 (5.7%)	0.88 (0.24-3.25)	0.852		
Histological subtype						
SSM	90 (69.2%)	37 (52.9%)	1		1	
NM	30 (23.1%)	29 (41.4%)	2.35 (1.24-4.45)	0.009		
Other**	10 (7.7%)	4 (5.7%)	1.08 (0.31-3.73)	0.902		
Ulceration						
No	92 (73%)	32 (45.7%)	1		1	
Yes	34 (27%)	38 (54.3%)	3.21 (1.74-5.93)	<0.001	3.18 (1.2-8.2)	0.016
Missing	4	0				
Mitotic rate						
0/mm ²	37 (37.4%)	13 (22.4%)	0.48 (0.23-1.01)	0.054		
≥1/mm²	62 (62.6%)	45 (77.6%)	1		1	
Missing	31	12				
Thickness (mm)						
1.00-2.00	80 (61.5%)	10 (14.3%)	1		1	
2.01-4.00	33 (25.4%)	30 (42.9%)	7.27 (3.20-16.56)	<0.001	4.73 (1.55-14. 7)	0.007
>4.00	17 (13.1%)	30 (42.9%)	14.12 (5.82-34.26)	<0.001	7.64 (2.2-27.0)	0.002
Sentinel node biopsy						
Positive	19 (16.1%)	19 (33.3%)	2.67 (1.27-5.60)	0.01		
Negative	96 (81.4%)	36 (63.2%)	1		1	
Missing	16	14				
Beta blokers						
No	123 (94.6%)	59 (84.3%)	1		1	
Yes	7 (5.4%)	11 (15.7%)	3.28 (1.21-8.88)	0.02		
Mean sun exposure score (lifelong)	12.3 (±3.42)	10.88 (±2.5)	0.86 (0.78-0.96)	0.004	0.84 (0.7-0.99)	0.043
Mean sunburn score (lifelong)	3.10 (±1.6)	2.57 (±1.5)	0.80 (0.65-0.98)	0.034		

519

520 Table 2. Clinical and epidemiological variables associated with fast growth

521 Multivariate stepwise forward model. *only variables with a p value <0.05 after univariate

522 analysis were included in the model**ALM, LM, Desmoplastic, spitzoïd, malignant blue, non-

523 assessable

525 **Table 3**

	Non-FGMM	FGMM	OR 95%	Raw p	Adjusted p
	(n=130)	(n=70)			Benjamini- Hochberg
ALK mutation					-
No	124 (95.4%)	62 (88.6%)	1	0.085	0.085
Yes	6 (4.6%)	8 (11.4%)	2.67 (0.89-8.02)		
ERBB4 mutation					
No	121 (93.1%)	59 (84.3%)	1	0.048	0.112
Yes	9 (6.9%)	11 (15.7%)	2.49 (0.88-7.22)		
FGFR2 mutation					
No	127 (97.7%)	59 (84.3%)	1	0.0007	0.0049
Yes	3 (2.3%)	11 (15.7%)	7.81 (1.96-45.25)		
IDH1 mutation					
No	130 (100%)	66 (94.3%)	1	0.014	0.049
Yes	0	4 (5.7%)	NE		
PDGFRA mutation					
No	128 (98.5%)	65 (92.9%)	1	0.052	0.073
Yes	2 (1.5%)	5 (7.1%)	4.92 (0.93-26.07)		
PREX2 mutation					
No	121 (93.1%)	59 (84.3%)	1	0.048	0.084
Yes	9 (6.9%)	11 (15.7%)	2.51 (0.99-6.38)		
RB1 mutation					
No	127 (97.7%)	64 (91.4%)	1	0.068	0.079
Yes	3 (2.3%)	6 (8.6%)	3.97 (0.96-16.39)		

526 NE: non estimable

527

528 Table 3. Association of mutations in genes with fast growing and non-fast growing

529 melanoma

530 Univariate analysis.

532 Table 4

Variables*	Non-FGMM	FGMM	Univariate		Multivariate (Stepwise)	
	(n=130)	(n=70)	OR 95%	р	OR 95%	р
Melanoma location						
Head and Neck	11 (8.5%)	9 (12.9%)	1.26 (0.47-3.38	0.64		
Trunk	51 (39.2%)	33 (47.1%)	1			
Upper Limb	20 (15.4%)	4 (5.7%)	0.31 (0.10-0.99)	0.047		
Lower Limb	41 (31.5%)	20 (28.6%)	0.75 (0.38-1.51)	0.423		
Hands/Feet/palm/nail	7 (5.4%)	4 (5.7%)	0.88 (0.24-3.25)	0.852		
Histological subtype						
SSM	90 (69.2%)	37 (52.9%)	1			
NM	30 (23.1%)	29 (41.4%)	2.35 (1.24-4.45)	0.009		
Other **	10 (7.7%)	4 (5.7%)	1.08 (0.31-3.73)	0.902		
Ulceration						
No	92 (73%)	32 (45.7%)	1		1	
Yes	34 (27%)	38 (54.3%)	3.21 (1.74-5.93)	<0.001	2.90 (1.10-7.63)	0.032
Missing	4	0				
Mitotic rate						
0/mm ²	37(37.4%)	13(22.4%)	0.48 (0.23-1.01)	0.054		
≥1/mm ²	62(62.6%)	45(77.6%)	1			
Missing	31	12				
Thickness (mm)						
1.00-2.00	80 (61.5%)	10 (14.3%)	1		1	
2.01-4.00	33 (25.4%)	30 (42.9%)	7.27 (3.20-16.56)	<0.001	5.41 (1.64-17.82)	0.006
>4.00	17 (13.1%)	30 (42.9%)	14.12 (5.82-34.26)	<0.001	8.88 (2.37-33.26)	0.001
Sentinel node biopsy						
Positive	19 (16.1%)	19 (33.3%)	2.67 (1.27-5.60)	0.01		
Negative	96 (81.4%)	36 (63.2%)	1			
Missing	16	14				
Beta blokers						
No	123 (94.6%)	59 (84.3%)	1			
Yes	7 (5.4%)	11 (15.7%)	3.28 (1.21-8.88)	0.02		
Mean sun exposure	12.3 (±3,42)	10.88 (±2,5)	0.86 (0.78-0.96)	0.004	0.85 (0.71-0.99)	0.049
score (lifelong)						
Mean sunburn score	3.10 (±1.6)	2.57 (±1.5)	0.80 (0.65-0.98)	0.034		
(IITEIONG)						
	107 (07 70/)	50 (94 20/)	1		1	
	127(37.7%)	09 (04.3%)		0.005		0.027
res	J (2.3%)	11 (15.7%)	7.01 (1.90-45.25)	0.005	0.04 (1.14-05.43)	0.037

533 Table 4. Variables associated with fast growth

534 Multivariate stepwise forward model. *only variables with a p value <0.05 after univariate 535 analysis were included in the model. **ALM, LM, Desmoplastic, spitzoïd, malignant blue, 536 non-assessable.

537 Data missing for the following variables: histological subtype (n=1); ulceration (n=4); SN 538 biopsy results (n=30); mitotic rate (n=43); sun exposure score (lifelong) (n=12); sunburn 539 score (lifelong) (n=18).



Months from primary melanoma resection