



HAL
open science

Multi-trophic markers illuminate the understanding of the functioning of a remote, low coral cover Marquesan coral reef food web

Pauline Fey, Valeriano Parravicini, Daniela Bănaru, Jan Dierking, René Galzin, Benoit Lebreton, Tarik Meziane, Nicholas Polunin, Mayalen Zubia, Yves Letourneur

► To cite this version:

Pauline Fey, Valeriano Parravicini, Daniela Bănaru, Jan Dierking, René Galzin, et al.. Multi-trophic markers illuminate the understanding of the functioning of a remote, low coral cover Marquesan coral reef food web. *Scientific Reports*, 2021, 11 (1), 10.1038/s41598-021-00348-w . hal-03404219

HAL Id: hal-03404219

<https://amu.hal.science/hal-03404219v1>

Submitted on 9 Dec 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



OPEN

Multi-trophic markers illuminate the understanding of the functioning of a remote, low coral cover Marquesan coral reef food web

Pauline Fey¹, Valeriano Parravicini², Daniela Bănaru³, Jan Dierking⁴, René Galzin², Benoit Lebreton⁵, Tarik Meziane⁶, Nicholas V. C. Polunin⁷, Mayalen Zubia⁸ & Yves Letourneur¹✉

We studied the food web structure and functioning of a coral reef ecosystem in the Marquesas Islands, French Polynesia, characterized by low coral cover, high sea surface temperature and meso- to eutrophic waters. The Marquesas constitute a relevant ecosystem to understand the functioning of low diversity reefs that are also subject to global change. A multi-tracer assessment of organic matter pathways was run to delineate ecosystem functioning, using analysis of fatty acids, bulk and compound specific stable isotope analysis and stable isotopes mixing models. Macroalgae and phytoplankton were the two major food sources fueling this food web with, however, some marked seasonal variations. Specifically, zooplankton relied on phytoplankton-derived organic matter and herbivorous fishes on macroalgae-derived organic matter to a much higher extent in summer than in winter (~75% vs. ~15%, and ~70 to 75% vs. ~5 to 15%, respectively). Despite remarkably high $\delta^{15}\text{N}$ values for all trophic compartments, likely due to local dynamics in the nitrogen stock, trophic levels of consumers were similar to those of other coral reef ecosystems. These findings shed light on the functioning of low coral cover systems, which are expected to expand worldwide under global change.

Increasing anthropogenic and climatic stressors on Earth ecosystems have motivated a widespread interest in understanding the contribution of species to ecosystem functioning and energy flows between ecological compartments such as trophic guilds. Among marine environments, coral reefs are by far the most diverse ecosystems, hosting thousands of species forming vast, complex and very poorly resolved interaction networks that influence the structure, the functioning and the resilience of the ecosystems¹. However, understanding the processes that drive ecosystem functioning and productivity requires determination of the major sources of organic matter fueling these systems², and the stocks of nutrients³, and quantification of the numerous consumer compartments that rely on sources of organic matter^{4,5}. Elucidating the major energetic pathways that fuel ecosystems across the complexity of the interaction network is crucial not only to assess food web functioning and the resilience of coral reefs but also to anticipate unexpected cascading effects and undesirable ecological surprises related to global changes⁶⁻⁸.

¹UMR ENTROPIE (UR-IRD-CNRS-IFREMER-UNC), LabEx « Corail », Université de La Nouvelle-Calédonie, BP R4, 98851 Nouméa Cedex, New Caledonia. ²CRIOBE, PSL Research University, USR 3278 EPHE-CNRS-UPVD, LabEx « Corail », Université de Perpignan, Avenue Paul Alduy, 66860 Perpignan Cedex, France. ³Mediterranean Institute of Oceanography, UM 110 (AMU-UTV-CNRS-IRD), Campus de Luminy, Case 901, 13288 Marseille Cedex 9, France. ⁴GEOMAR Helmholtz Centre for Ocean Research, Research Division Marine Ecology, Düsternbrooker Weg 20, 24105 Kiel, Germany. ⁵UMR LIENSs 7266 (CNRS-ULR), Institut du Littoral Et de L'environnement, 2 rue Olympe de Gouges, 17000 La Rochelle, France. ⁶Laboratoire BOREA, Muséum National d'Histoire Naturelle, CNRS 7208, IRD 207-SU-UCN-UA, Muséum National d'Histoire Naturelle, 61 rue Buffon, 5 CP 53, 75231 Paris Cedex, France. ⁷Newcastle University, School of Natural and Environmental Sciences, Newcastle-upon-Tyne NE1 7RU, UK. ⁸UMR EIO (UPF-IRD-ILM-IFREMER), Université de La Polynésie Française, LabEx « Corail », BP 6570, 98702 Faa'a, Tahiti, French Polynesia. ✉email: yves.letourneur@unc.nc

Several studies have improved our understanding about flows of energy within coral reefs^{4, 9–13} but the processes involved, the species that support them and the ways these species interact are scarcely resolved. For instance, the high diversity of sources of organic matter available to primary consumers complicates the characterization of energy flows^{2, 3, 14}. Carbon sources may have several origins, continental or marine, and can come from coastal, pelagic or deep waters, and also from living organisms or detrital matter^{15, 16}. Oceanic phyto- and zoo-plankton may also be imported into reefs by currents, and then consumed by phyto- and zoo-planktivorous fish^{17, 18}. While benthic sources represent an important carbon source to reef species^{4, 12, 19}, numerous species rely principally or exclusively on planktonic sources²⁰ and even non-planktivorous species may rely on oceanic production¹². However, in several ecosystems, benthic production and/or terrestrial inputs can also be major contributors to the energy flow in food webs^{2, 3, 21}.

The Marquesas Islands, one of the most isolated archipelagos on Earth, may provide important insights into the functioning of coral reefs characterized by low diversity and low coral cover. Decreasing coral cover and diversity are expected in many reef systems in the world facing increasing anthropogenic pressures^{22–26}. Marquesas Islands coral reefs could not track sea level rise during the last post-glacial period²⁷, and paleo-reefs can now be found at ~60–130 m depth, while modern shallow reefs are essentially rocky reefs with low coral cover. Other distinctive features of the Marquesas include the absence of *Acropora* spp. corals, which are common across other Polynesian coral reefs²⁸. On one of the main islands of the Marquesas, Nuku Hiva, mean coral cover is only ~5%²⁹. Other environmental conditions that make the Marquesas a highly pertinent system for the study of the functioning of their food web are their warm waters and high nutrient levels. The sea surface temperature is usually relatively high all year round (average 28–30°C^{3, 28}), and sometimes higher during ENSO events, and thus substantially warmer than on several other reef systems like those located at higher latitudes and experiencing wider seasonal differences. Surface waters have high loads of nitrate (NO₃⁻) and phosphate (PO₄⁻), with concentrations higher than 1 μM and 0.3 μM, respectively, which exceed ~100-fold those measured in the South Pacific subtropical gyre³⁰. This nutrient richness promotes the development of a high phytoplankton biomass around the Marquesan coasts over the year³⁰.

In coral reef systems worldwide, the questions of ecosystem functioning, food web structure and function and organic matter sources fueling production are gaining attention^{4, 11, 13, 31}. Here we characterize the major energetic pathways that fuel the unusual, low coral cover ecosystem of the Marquesas and assess the extent to which this pelagic production penetrates the coral reef food web. We analysed the trophic relationships among the food web components (sources of organic matter, primary consumers and higher trophic level consumers up to mesopredators) using multiple and complementary trophic markers. We combined information from carbon (δ¹³C) and nitrogen (δ¹⁵N) bulk stable isotope compositions, amino acid compound-specific isotopes compositions (δ¹⁵N) (AA CSIA) and fatty acids data. These three trophic have evidently yet to be combined to analyse any coral reef ecosystems, although they are capable of providing highly complementary information. Fatty acids are used to assess how the various sources of organic matter are integrated by primary consumers (herbivores) and then by secondary consumers, AA-CSIA provide information on both the assimilated baseline and the length of food chains, while bulk stable isotope analyses allow to describe the global structure of the food web. The combination of these methods allowed us to address five major questions: (i) What are the main sources of organic matter that fuel the food web of the Marquesas coral reefs, and (ii) how is this organic matter integrated by primary consumers? What is (iii) the trophic structure of this system, and (iv) the ‘baseline’ allowing us to assess the length of the food chain? (v) Is the uptake of the major sources of organic matter consistent across seasons? We then interpreted the observed patterns in the light of previous observations on other reef systems to draw inferences about possible coral reef functioning in future warming and possibly more eutrophic tropical oceans.

Material and methods

Study site and sampling. Field work was carried out in August 2016 (austral winter) and in March 2017 (austral summer) in southeastern Nuku Hiva (8°54'S, 140°02'W; Marquesas Islands, French Polynesia). The abiotic (rainfall, winds, temperature, hydrodynamics, etc.) and biotic (benthic and pelagic communities) conditions around the islands are well known^{3, 28, 32}.

We sampled the major sources of organic matter (OM), primary consumers (invertebrates and fish) and several higher trophic-level consumers (invertebrates and fish), up to mesopredators. The sampling of OM sources, already described in detail², comprise the particulate OM-hereafter POM mainly consisting of pico-nano phytoplankton (n = 49), sedimentary OM-hereafter SOM (n = 47), micro-phytoplankton-hereafter phytoplankton (n = 49), 11 different locally abundant algae (algal turf and macroalgae, n = 73) and detrital terrestrial material-hereafter DTP-derived from tree leaves transported by rivers (n = 16).

The sampled consumers were 14 invertebrate species (sponges, ascidians, echinoderms, gastropods, bivalves and crustaceans-214 individuals in total), zooplankton (n = 49), and 29 fish species (n = 523). Invertebrates were collected by handpicking during scuba diving, in order to obtain 5–10 individuals per species and per season. For zooplankton, a 125 μm mesh-size WP2 net was used for a vertical tow in the water column (from ~40–50 m depth to the surface). Fish were collected by spearfishing or using an anesthetic (i.e. eugenol diluted at 10% in alcohol), in both seasons. All samples were identified to the lowest possible taxonomic level and were kept in ice chests during sampling and (except for POM, phyto- and zoo-plankton) immediately stored at –20 °C until analysis.

First, we restricted our analysis to species representative of well-known trophic groups across the trophic-level gradient in order (i) to determine how the organic matter is assimilated by primary consumers, and (ii) to assess the length of the food chain. We selected nine primary consumers to study the integration of OM sources. Among these species, three were filter-feeders (the oyster *Pinctada margaritifera*, an unidentified ascidian, the sponge *Sphaciospongia* sp.), one was a phytoplankton browser (zooplankton either 300–500 μm or 1000–2000 μm in size), and five were herbivore-detritivores (the gastropods *Mauritia* spp., the surgeonfishes *Acanthurus nigricans* and

Ctenochaetus marginatus, and the parrotfishes *Scarus koputea* and *S. rubroviolaceus*). We also selected eight secondary-tertiary consumers expected to be at the top of the benthic food webs. These species were one gastropod, *Conus conco*, and seven fish: the snappers *Lutjanus bohar*, *L. gibbus*, and *L. kasmira*, the moray-eel *Enchelycore pardalis*, the scorpionfish *Scorpaenopsis possi*, and the groupers *Cephalopholis argus* and *Epinephelus fasciatus*.

Then, we extended our analysis to all the species for which samples were available, i.e. the 14 invertebrate and 29 fish species collected. We defined the trophic position of all individuals in order to assess their position in the food web relative to the baseline, i.e. linked to OM sources.

Thus, the first analysis, focused on well-known trophic groups, allowed us to determine how the OM is incorporated and what the length of the food chain is. Then, the inclusion of all species allowed depiction of the food-web structure.

Bulk stable isotope analyses. These analyses were run to get a general picture of the food web structure, the role of the OM sources and of various primary and secondary-tertiary consumers within the food web. The carbon and nitrogen stable isotopes (respectively $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$) were used in combination; $\delta^{13}\text{C}$ give information on the origin(s) of the organic matter source(s) used by consumers³³ and $\delta^{15}\text{N}$ is a proxy of trophic level³⁴, thus allowing a depiction of the food webs in bivariate isotopic space³⁵.

A piece of the thallus was sampled for algae, soft muscle for all mollusks, and dorsal white muscle for fish³⁶, ~2–5 g in each case. For ascidians and sponges, ~5–10 g pieces, excluding external theca for ascidians, were taken from each individual. For zooplankton, several entire individuals were grouped to obtain the 5 mg dry mass required for analysis.

Carbon and nitrogen bulk stable isotope compositions ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$) were determined in all samples. Sediment was dried and reduced to a fine powder using a mortar and pestle. POM was collected on precombusted GF/F filters (porosity 0.7 μm), and dried. Plant (algae, DTP) and animal (zooplankton, invertebrates and fish) samples were freeze-dried and ground to fine powder. Approximately 1 mg of powder was precisely weighed and encapsulated for plant/animal samples, and 15–20 mg for SOM and 15–30 mg for POM and phytoplankton (matter extracted by scrapping the filter). For POM and SOM, two subsamples were analyzed: one was acidified to eliminate inorganic carbonates from the sample and used for $\delta^{13}\text{C}$ analysis³⁷, while the other was not acidified and used for $\delta^{15}\text{N}$ analysis⁴³. The other samples were analyzed without prior treatment. Samples were analyzed through continuous-flow isotope-ratio mass spectrometry with a Flash 2000 elemental analyzer equipped with the Smart EA option (Thermo Scientific, Milan, Italy), coupled with a Delta V Advantage isotope ratio mass spectrometer with a ConFlo IV interface (Thermo Scientific, Bremen, Germany) at the Littoral, Environment and Societies Joint Research Unit stable isotope facility (LIENSs) at the University of La Rochelle (France). Isotope compositions were expressed in the δ notation as parts per mil (‰) as deviations from an international standard (i.e. Vienna Pee Dee Belemnite for carbon and atmospheric N_2 for nitrogen) following the formula:

$$\delta X = \left[\left(R_{\text{sample}} / R_{\text{standard}} \right) - 1 \right] \times 1000$$

where X is ^{13}C or ^{15}N , R is the corresponding ratio ($^{13}\text{C}/^{12}\text{C}$ or $^{15}\text{N}/^{14}\text{N}$). Calibration was done using reference materials (USGS-24, -61, -62, IAEA-CH6, -600 for carbon; USGS-61, -62, IAEA-N2, -NO-3, -600 for nitrogen). The analytical precision of the measurements was 0.1‰ for carbon and <0.15‰ for nitrogen based on analyses of USGS-61 and USGS-62 used as laboratory internal standards.

Compound-specific amino acid stable isotope analyses. These analyses were run to (i) help define the ‘baseline’ of the food web through the use of sources amino acids, and (ii) to assess food chain length through both trophic amino acids and bulk stable isotopes. Sr-AA $\delta^{15}\text{N}$ values capture the real baseline isotopic composition much better than those of bulk $\delta^{15}\text{N}$ because the latter value pool several amino acids, whereas Sr-AA $\delta^{15}\text{N}$ values do not fluctuate over different trophic levels, thus capturing the baseline isotope composition of the food web without biases⁵.

Compound-specific $\delta^{15}\text{N}$ values of amino acids (AA-CSIA) were derived from eight selected species having high bulk $\delta^{15}\text{N}$ values (see above). This included the gastropod and the seven secondary-tertiary consumer fish species (total 44 samples for both seasons). Samples were prepared by acid hydrolysis followed by derivatization to produce trifluoroacetic amino acid esters (TFAA) using standard methods³⁸. The $\delta^{15}\text{N}$ values of the TFAA derivatives of amino acids were analyzed using an isotope ratio mass spectrometer (Delta V Plus, Thermo Scientific, Bremen, Germany) interfaced with a gas chromatograph (Trace GC 1300, Thermo Scientific, Bremen, Germany) through a GC IsoLink combustion furnace, and liquid nitrogen cold trap at the University of Davis (California, USA). Measured isotopic values were corrected relative to known $\delta^{15}\text{N}$ values of norleucine, the internal reference material. All samples were analyzed in triplicate. $\delta^{15}\text{N}$ values of the glycine and phenylalanine were then measured for source amino acids ($\delta^{15}\text{N}_{\text{Sr-AA}}$) in order to assess the baseline $\delta^{15}\text{N}$ values (i.e. the $\delta^{15}\text{N}$ of the primary producers at the base of the food web^{5, 39}). So, knowing characteristics of main organic matter sources by one hand (i.e. the ‘baseline’), and knowing species at the top of the food web on the other hand (i.e. the eight selected species), we expected to be able to assess the food chain length.

Trophic positions. We estimated the trophic position (hereafter TP) of several individuals using a combination of AA-CSIA and bulk SIA. In particular, we used the $\delta^{15}\text{N}$ values from bulk SIA of target individuals and from AA-CSIA to characterize the baseline. Then, TP was calculated as follows⁴⁰:

$$\text{TP}_x = \frac{\delta^{15}\text{N}_x - \delta^{15}\text{N}_{\text{baseline}}}{\text{TEF}} + \text{TP}_{\text{baseline}} \quad (1)$$

where x is the species of interest and TEF is the trophic enrichment factor between trophic levels. In our case, we set the TEF at 3.4 ‰ as commonly done in the marine coastal environment⁴⁰. Because the source amino acids (glycine, phenylalanine) represent the $\delta^{15}\text{N}$ value of the baseline, a $\text{TP}_{\text{baseline}}$ of 1 was used.

Fatty acid analyses. These analyses were run to assess the integration of OM sources by primary producers and possibly the use of these herbivores by secondary-tertiary consumers.

Fatty acid (FA) were analysed in the OM sources (SOM, river and marine POM, DTP, algae and phytoplankton²) and in nine primary consumers: zooplankton ($n=23$), four invertebrate species ($n=54$) and four fish species ($n=56$). Lipids were extracted⁴¹, using 5–20 mg of material for POM, 20–30 mg for DTP, algae, phyto- and zooplankton, sponges and ascidians, 30–40 mg for mollusks and fish and 1 g for SOM. Tricosanoic acid (23:0), was added to each sample as an internal standard to measure the FA concentrations. Fatty acid methyl esters (FAMES) were quantified by gas chromatography (Varian 3800-GC), using a flame ionization detector. FAs were identified by comparing retention times with those of a commercial standard (Supelco) and confirmed using a mass spectrometer coupled to a gas chromatograph (Varian 450-GC; Varian 220-MS). Fatty acid concentrations are expressed as % of total FAs, or as absolute concentrations in $\text{mg}\cdot\text{g}^{-1}$.

Functional groups of fatty acids, classified by degree of unsaturation, were used as indicators of different types of organic matter^{42,43}: the saturated fatty acids (SFAs; e.g. 14:0, 16:0 and 18:0), monounsaturated fatty acids (MUFAs; e.g. 16:1 ω 7 and 18:1 ω 9), polyunsaturated fatty acids (PUFAs; e.g. 20:4 ω 6, 20:5 ω 3 and 22:6 ω 3) and branched fatty acids (BrFAs; e.g. iso-15:0 and anteiso-15:0). Among SFAs, the long-chain SFAs (C24–C28) are widely used as an indicator of detrital terrestrial plant (DTP) material⁴⁴. Bacteria and cyanobacteria are also rich in SFAs⁴⁵. C16 and C18 MUFAs are found in algae⁴⁶, bacteria⁴⁷ and zooplankton⁴⁸. PUFAs with an omega-3 (ω 3) or omega-6 (ω 6) terminal end are named essential fatty acids (EFAs) because animals are considered typically to acquire them solely from diet⁴⁵. PUFAs are abundant in fresh plankton material (major components of phytoplankton), but these FAs are rapidly degraded in the water column and the sediment^{49,50}. Iso- and anteiso-FAs (BrFAs) are solely synthesized by bacteria and therefore are deemed to be a good indicator of these microorganisms^{47,51}.

Assessment of OM integration by primary consumers. Assessment of food source uses by primary consumers was achieved through two complementary methods. The first approach is based on the close relationship between the isotope compositions of a consumer and its food sources⁵². The contributions of the different sources to the diets of primary consumers (i.e. macroalgae, algal turf, phytoplankton, POM and/or SOM) were determined using a Bayesian mixing model and the SIAR package⁵³. OM sources with similar isotope composition were pooled to avoid incorrect determination of their relative contribution. Models were run for 200,000 iterations, the burn-in was set at 50,000 iterations and a one-fifteenth thinning was applied. This model provides a range of solutions regarding the proportions of the different sources (i.e. macroalgae, algal turf, phytoplankton, POM and/or SOM). To reduce potential bias associated with the definition of a single trophic enrichment factor (i.e. 3.4‰), we also used TEF ($\Delta^{13}\text{C}$ and $\Delta^{15}\text{N}$) values adapted to the feeding strategies and trophic positions of the studied taxa (Suppl. Figure S1). The determination of $\Delta^{13}\text{C}$ and $\Delta^{15}\text{N}$ values was based on the isotope compositions of the primary consumers (see results) in combination with previous findings³ on OM sources. The herbivore-detritivores showed TEFs values of $1.60 \pm 1.87\text{‰}$ for $\Delta^{13}\text{C}$ and $4.28 \pm 1.05\text{‰}$ for $\Delta^{15}\text{N}$, while filter-feeders and zooplankton had $1.24 \pm 1.16\text{‰}$ for $\Delta^{13}\text{C}$ and $2.71 \pm 0.85\text{‰}$ for $\Delta^{15}\text{N}$.

The second approach was based on the exploration of the links between FAs and primary consumers through a principal component analysis (PCA), using the 25 individual FAs that had an average proportion greater than 1% within at least one group of samples. The statistics and graphical representations were performed using R version 3.4.4⁵⁴, using ggplot2, gridExtra, ggrepel, vegan, FactoMiner and car packages. Identification of FAs bio-indicating particular OM sources allowed us to assess the importance of the integration of various potential OM sources by primary consumers.

Ethical statement. This research received no specific grant from any commercial or not-profit sectors. No coral habitat was degraded during this research. Sample collection was permitted by the French Polynesian government (authorization number: 681/MCE/ENV), which also approved the experimental protocols. All methods were carried out in accordance with relevant guidelines and regulations. All methods were performed in compliance with the ARRIVE guidelines and regulations for ethical treatment of animals⁵⁵.

Results

OM sources in the Marquesan coastal ecosystem. As the data on the different potential sources of organic matter were already widely presented and discussed in a previous article⁴², only a brief outline is given hereafter. The mean $\delta^{13}\text{C}$ values ranged from $-23.9 \pm 1.7\text{‰}$ for algal turf to $-16.4 \pm 2.0\text{‰}$ for macroalgae⁽³⁾, Suppl. Table 1). The mean $\delta^{15}\text{N}$ values ranged from $11.6 \pm 0.9\text{‰}$ for macroalgae to $15.0 \pm 1.8\text{‰}$ for phytoplankton⁽³⁾, Suppl. Table S1).

Primary consumers: trophic marker characteristics and use of OM sources. Mean $\delta^{13}\text{C}$ values ranged from -19.6 ± 2.7 (*Acanthurus nigricans*) to $-14.0 \pm 1.3\text{‰}$ (*Ctenochaetus marginatus*) in winter, and from -20.2 ± 0.5 (small zooplankton) to $-13.9 \pm 1.2\text{‰}$ (*C. marginatus*) in summer (Table 1). Mean $\delta^{15}\text{N}$ values ranged from 12.6 ± 0.9 (small zooplankton) to $19.1 \pm 1.0\text{‰}$ (*C. marginatus*) in winter, and from 14.2 ± 0.4 (*Pinctada margaritifera*) to $17.4 \pm 1.4\text{‰}$ (*C. marginatus*) in summer. The parrotfish and cypraeid species did not differ significantly from each other in mean $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values (Table 1, Kruskal–Wallis tests, $P > 0.05$) and were hereafter pooled with *Mauritia* spp. and Scarinae, respectively.

		Code	n	$\delta^{13}\text{C}\text{‰}$	$\delta^{15}\text{N}\text{‰}$
Invertebrates					
Asciidiidae	Ascidian	Asc	2	-17.8 ± 1.2	14.6 ± 0.3
			15	-18.7 ± 0.4	15.1 ± 1.1
Clionaidae	<i>Spheciospongia</i> sp.	Spsp	28	-18.1 ± 0.5	15.8 ± 0.6
			16	-17.7 ± 0.3	16.1 ± 0.5
Pteriidae	<i>Pinctada margaritifera</i>	Pima	7	-16.7 ± 0.4	15.2 ± 0.5
			17	-17.0 ± 0.3	14.5 ± 0.4
Cypraeidae	<i>Mauritia</i> spp.	Maspp	5	-15.3 ± 1.1	15.8 ± 0.2
			8	-16.4 ± 0.6	16.6 ± 0.9
Zooplankton	300–500 μm	Zoo-300	9	-19.6 ± 0.3	12.6 ± 0.9
			12	-20.2 ± 0.5	14.9 ± 1.1
	1000–2000 μm	Zoo-1000	12	-19.4 ± 0.5	14.1 ± 0.5
			16	-19.2 ± 0.7	17.4 ± 0.5
Fish					
Acanthuridae	<i>Acanthurus nigricans</i>	Acni	11	-19.6 ± 2.7	17.0 ± 0.8
			14	-17.7 ± 1.9	16.1 ± 0.9
	<i>Ctenochaetus marginatus</i>	Ctma	25	-14.0 ± 1.3	19.1 ± 1.0
			13	-13.9 ± 1.2	17.4 ± 1.5
Scarinae	<i>Scarus koputea</i>	Scar	4	-17.7 ± 0.2	19.0 ± 0.3
			9	-14.9 ± 1.2	16.4 ± 1.2
	<i>S. rubroviolaceus</i>		8	-16.6 ± 1.2	18.8 ± 1.1

Table 1. Bulk $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values (means \pm standard deviation) of nine selected primary consumers. Summer with bold italic, winter without bold italic.

We detected 57 FAs, of which 35 had concentrations greater than 1% (see Suppl. Table S2 for details). The two size groups of zooplankton had similar fatty acid compositions (Kruskal–Wallis test, $P > 0.05$) and were thus pooled hereafter. The sponge *Spheciospongia* sp. represented an outlier with extremely high percentages of several SFAs, MUFAs and mostly PUFAs (Suppl. Table S2), and was therefore removed from further analyses.

Different primary consumers were characterized by different FA profiles (Fig. 1, Suppl. Table S2). For instance, zooplankton was mostly characterized by some SFAs (14:0 and 15:0), MUFAs (16:1 ω 7) and PUFAs (16:2 ω 4 and 20:5 ω 3) (Fig. 1). Scarinae were mostly characterized by 16:0 and 18:0 (SFA), 18:1 ω 9, 20:4 ω 6, 22:5 ω 6 (PUFAs), and 22:6 ω 3, whereas *Acanthurus nigricans* was characterized by 16:0 (SFA) and 18:1 ω 9 (Fig. 1, Suppl. Table S2).

We detected differences in the use of major sources of OM among seasons for zooplankton and herbivore-detritivores (Fig. 2). Small zooplankton mostly relied on POM in both seasons ($\sim 60\%$ on average, credibility interval (CI) $\sim 30\text{--}85\%$), with phytoplankton contributing $\sim 20\text{--}30\%$ to isotope composition (CI $\sim 0\text{--}60\%$). Large zooplankton relied mostly on POM during winter ($\sim 60\%$, CI $\sim 40\text{--}85\%$) and on phytoplankton during summer ($\sim 75\%$, CI $\sim 50\text{--}100\%$) (Fig. 2). Similar seasonal differences occurred in the filter-feeders, although with a much lower amplitude. The major sources of OM for ascidians were phytoplankton, algae and POM in relatively equal proportions ($\sim 25\text{--}40\%$ depending on seasons); the sponge *Spheciospongia* sp. did not use a lot of POM, and *Pinctada margaritifera* relied mainly on macroalgae, and less on POM and phytoplankton (Fig. 2). SOM and macroalgae were important OM sources for herbivore-detritivores but with a strong seasonal contrast in some cases, ranging from $\sim 10\%$ (*Acanthurus nigricans* in summer) to $\sim 90\%$ (*Ctenochaetus marginatus* in winter) and from $\sim 10\%$ (*C. marginatus* in winter) to $\sim 75\%$ (*C. marginatus* in summer), respectively (Fig. 2). Algal turf was a marginal OM source for Scarinae and *C. marginatus*, contributing $\sim 10\text{--}15\%$ to the isotope composition of *Mauritia* spp. and up to $\sim 50\%$ (in winter) to that of *A. nigricans*.

Secondary consumers trophic markers and the general structure of the food web. Based on bulk isotope compositions of OM sources, nine primary consumers and eight secondary-tertiary consumers, we were able to delineate the general structure of the major energetic pathways of the Marquesan coastal food web (Fig. 3). For secondary-tertiary consumers, mean bulk $\delta^{13}\text{C}$ values ranged from -16.5 to -14.7‰ , and those of $\delta^{15}\text{N}$ values ranged from 18.7 to 20.4‰ (Fig. 3). The highest mean $\delta^{15}\text{N}$ values were measured in the endemic gastropod *Conus conco* and the grouper *Cephalopholis argus* ($20.4 \pm 0.9\text{‰}$ and $20.3 \pm 0.4\text{‰}$, respectively). For most secondary-tertiary consumers, differences between seasons were often low and non-significant (Suppl. Table S3). Other sampled consumer species not included in Fig. 3 for clarity fitted well within this network based on their $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values (Suppl. Table S3).

On average, the mean $\delta^{15}\text{N}$ value of Sr-AA (phenylalanine and glycine) was $11.6 \pm 3.4\text{‰}$ with values ranging from 7.1‰ for glycine in *Lutjanus bohar* to 17.0‰ for phenylalanine in *Conus conco* (Table 2). Differences in Sr-AA $\delta^{15}\text{N}$ values among species were more evident for glycine than for phenylalanine.

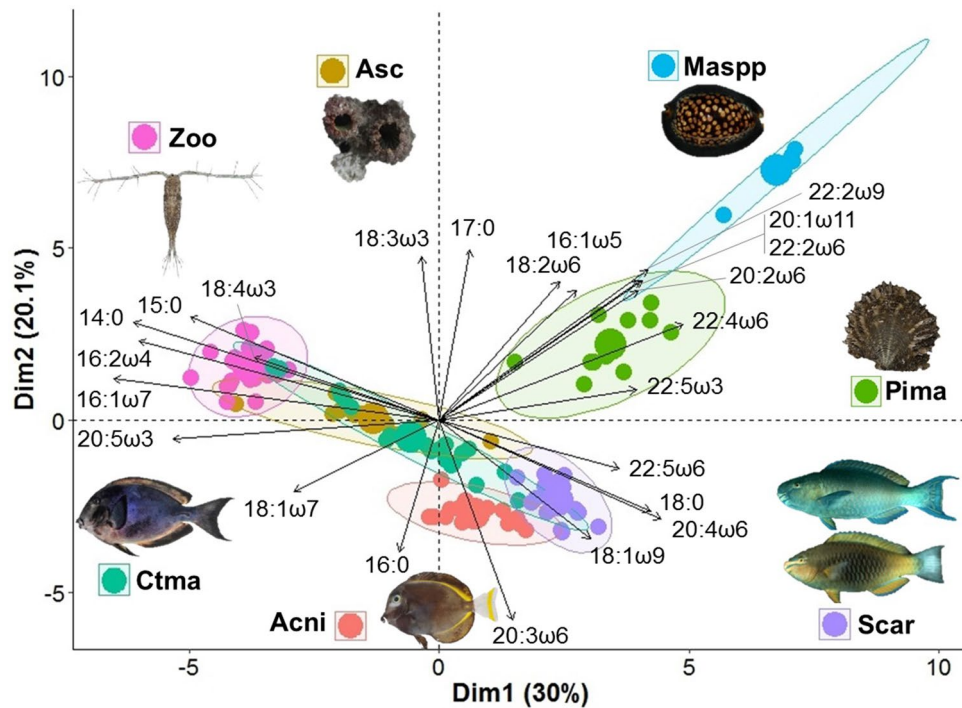


Figure 1. PCA of fatty acid (FA) compositions of the invertebrates (excluding sponges) and fishes. Codes of species: Asc: ascidians, Pima: *Pinctada margaritifera*, Maspp: *Mauritia* spp., Zoo: zooplankton, Acni: *Acanthurus nigricans*, Ctma: *Ctenochaetus marginatus*, Scar: *Scarus koputea* and *S. rubroviolaceus*. Only FAs with a mean percentage > 1% in at least one species were considered (see text).

The eight secondary consumers also displayed seasonal differences in both their mean bulk $\delta^{15}\text{N}$ and Sr-AA $\delta^{15}\text{N}$ values (Fig. 4), although not statistically significant, mean values were higher in winter than summer, in contrast to what was observed for bulk $\delta^{15}\text{N}$ values of phytoplankton and macroalgae (Fig. 4).

Baseline and trophic positions. Although the isotope compositions of primary consumers evidenced the use of several OM sources (Fig. 2), the relatively high concentrations of the fatty acid 20:4 ω 6, a biomarker of macroalgae, in all primary consumers (even for zooplankton although at lower concentrations) allowed us to identify in the macroalgae the main baseline to estimate TP. The Sr-AA $\delta^{15}\text{N}_{\text{phe-gly}}$ values obtained for the eight mesopredators were also used.

The mean bulk $\delta^{15}\text{N}$ values of all individuals ($n = 797$) and species ($n = 43$) belonging to different trophic groups (Suppl. Table S3) provided a detailed picture of their trophic positions in the food web (Fig. 5). Most primary consumers (i.e. filter-feeders, zooplankton and herbivores) displayed a TP of ~ 2 – 2.3 , omnivores and detritivores had a TP of ~ 2.4 – 3 , carnivores were at a TP ~ 3 – 3.2 and piscivores showed a TP of ~ 3.2 – 3.6 , with the highest TP being in *Conus concho* and *Cephalopholis argus* (3.57 ± 0.27 and 3.55 ± 0.23 , respectively; Fig. 5). This global picture was consistent across seasons for all trophic groups, despite higher $\delta^{15}\text{N}$ values in summer than in winter (Suppl. Figure S2).

Discussion

We provide the first combined application of bulk and compound specific stable isotope as well as fatty acid data to a coral reef food web, and elucidate food web functioning including major energetic pathways, OM sources and trophic positions of multiple species on a Marquesan coral reef. Despite the peculiar ecological characteristics of these coral reefs (warm water, high nutrient levels, low coral cover as well as reduced biodiversity compared to many other reefs), fatty acid, C and N stable isotope data evidenced that this system maintains a high productivity fueled by phytoplankton and zooplankton, most likely of pelagic origin. Moreover, despite unusually high $\delta^{15}\text{N}$ values for all groups from OM sources up to mesopredators, trophic positions and food chain lengths were comparable to those documented for other coral reef ecosystems^{56,57}. Since conditions experienced in the Marquesas today are projected for many other reef systems in the future, this study offers valuable insights into the future of changing coral reefs.

The outputs from mixing models highlighted that macroalgae, phytoplankton, POM and SOM are the main sources of OM for the consumers of the Marquesas, while turf algae had a comparatively minor role. Algal turf is often described as a major OM sources in coral reefs, especially for herbivorous fish^{4,10,58,59}. However, at Nuku Hiva, it contributed only $\sim 15\%$ to herbivores, although in *Acanthurus nigricans* its contribution to biomass reached $\sim 50\%$ in winter. In a complex coral reef system in New Caledonia, four major sources of OM used by

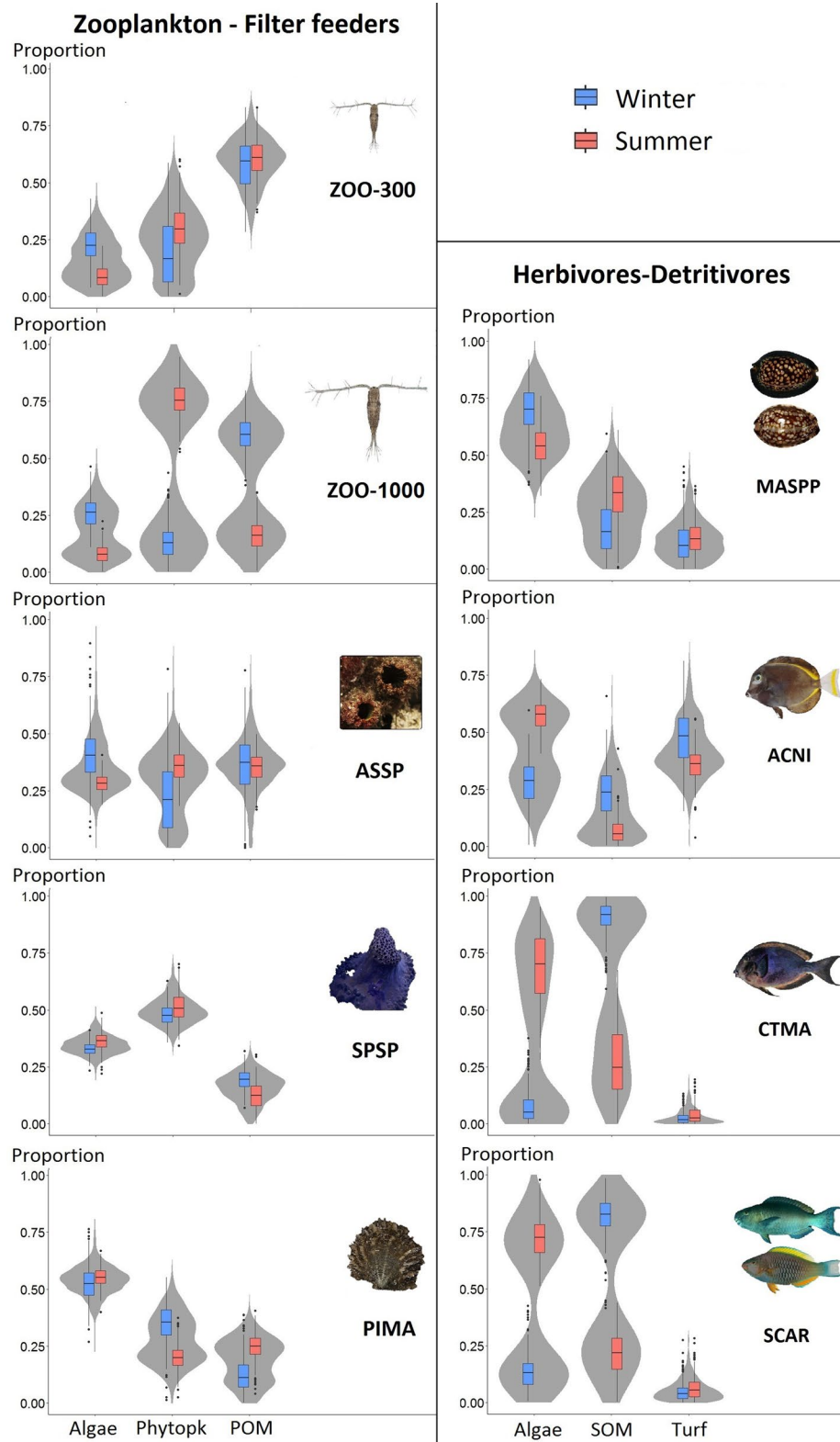


Figure 2. Relative proportions of the different OM sources for filter-feeders and zooplankton (left panel) and for herbivore-detritivores (right panel), based on SIAR mixing model outputs. Phytoplank: Phytoplankton; POM: particulate organic matter; SOM: sedimentary organic matter. Violin plots represent the posterior distribution shape of the data (density estimation) in grey; boxplots represent median, interquartile ranges (red and blue boxes, depending on season), 95% credibility intervals (thin line) and outliers (black dots). ZOO-300: zooplankton 300–500 μm , ZOO-1000: zooplankton 1000–2000 μm , ASSP: ascidians, SPSP: *Spheciospongia* sp., PIMA: *Pinctada margaritifera*, ACNI: *Acanthurus nigricans*, CTMA: *Ctenochaetus marginatus*, SCAR: Scarinae (*Scarus koputea* and *S. rubroviolaceus*), MASPP: *Mauritia* spp.

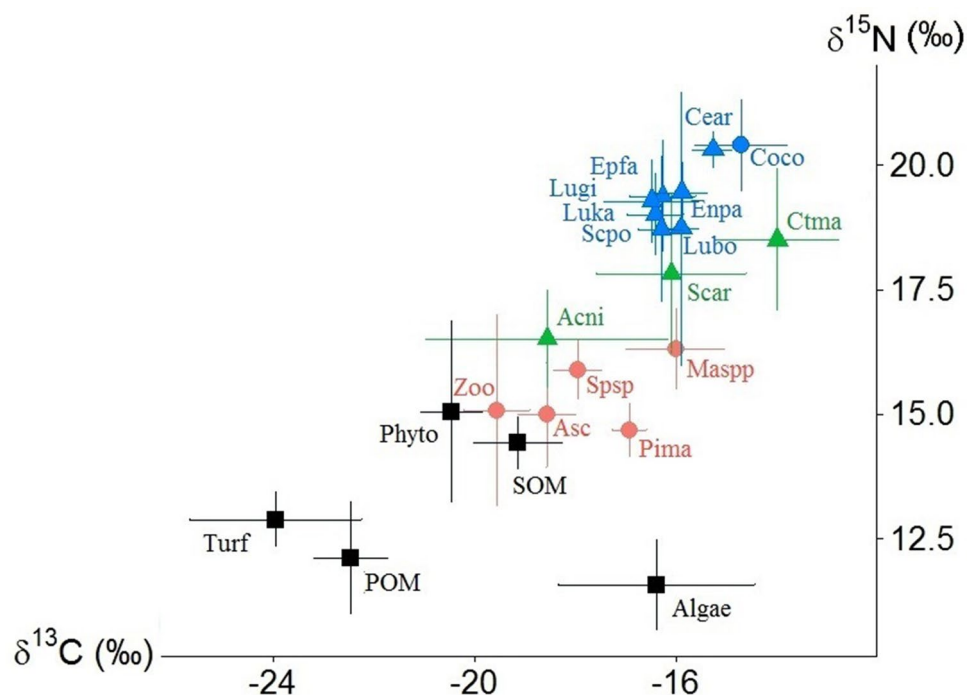


Figure 3. Plot of $\delta^{15}\text{N}$ against $\delta^{13}\text{C}$ values (means \pm standard deviations) of sources of organic matter (black squares, see Fey et al. 2020 for details), nine primary consumers (red circles: invertebrates, green triangles: fish), and eight secondary consumers (blue circle: invertebrate, blue triangles: fish), both seasons pooled. Turf: algal turf, Phyto: phytoplankton, Algae: macroalgae, POM: particulate organic matter, SOM: sedimentary organic matter, Asc: ascidians, Spsp: *Speciospongia* sp., Pima: *Pinctada margaritifera*, Maspp: *Mauritia* spp., Zoo: zooplankton, Acni: *Acanthurus nigricans*, Ctma: *Ctenochaetus marginatus*, Scar: Scarinae, Coco: *Conus conco*, Lubo: *Lutjanus bohar*, Lugi: *L. gibbus*, Luka: *L. kasmira*, Enpa: *Enchelycore pardalis*, Scpo: *Scorpaenopsis possi*, Cear: *Cephalopholis argus*, Epfa: *Epinephelus fasciatus*.

Species	Code	n	Phenylalanine	Glycine	Mean
<i>Conus conco</i>	Coco	6	15.2 \pm 3.9	17.0 \pm 1.6	16.1 \pm 3.0
<i>Lutjanus gibbus</i>	Lugi	6	13.0 \pm 1.7	7.9 \pm 1.6	10.5 \pm 3.1
<i>Lutjanus kasmira</i>	Luka	6	12.3 \pm 3.3	8.8 \pm 1.9	10.5 \pm 3.2
<i>Lutjanus bohar</i>	Lubo	4	12.0 \pm 5.2	7.1 \pm 4.2	9.5 \pm 5.1
<i>Enchelycore pardalis</i>	Enpa	4	11.9 \pm 1.9	13.2 \pm 0.7	12.4 \pm 1.7
<i>Scorpaenopsis possi</i>	Scpo	6	12.2 \pm 3.0	10.5 \pm 1.8	11.4 \pm 2.5
<i>Cephalopholis argus</i>	Cear	6	13.6 \pm 1.9	8.2 \pm 1.2	10.9 \pm 3.2
<i>Epinephelus fasciatus</i>	Epfa	6	11.9 \pm 1.9	10.9 \pm 2.2	11.4 \pm 2.0
Total		44	12.8 \pm 3.0	10.5 \pm 3.6	11.6 \pm 3.4

Table 2. Source amino acids (AA) $\delta^{15}\text{N}$ values (‰) of eight secondary consumers (means \pm standard deviation), both seasons pooled.

consumers were evidenced², i.e. algal turf (the most important one), sedimentary OM mixed with macroalgae, particulate OM, and - to a lower extent - detritus and seagrass. In a Caribbean coral reef, algal turf is also the main OM source for consumers⁵⁷. The reasons for a marginal importance of algal turf in Marquesas Islands remain unclear. Algal turf may lack important nutritive elements⁶⁰ inducing herbivores to shift their diet to other items, such as macroalgae. In contrast to macroalgae, algal turf in Marquesas displayed a high C/N ratio ($\sim 18^3$), much higher than the value of ~ 12 usually considered as characteristic of refractory organic matter⁶¹. This suggests a low nutritional value of this food item and might explain why algal turf was little used in Marquesas, despite its wide distribution. The reason for such a C/N ratio remains unclear and requires further investigations.

The FA compositions of the primary consumers also highlighted the importance of the macroalgae in the food web. Indeed, we detected a high abundance of the FAs 18:3 ω 3 and 20:4 ω 6 in all primary consumers (Fig. 6). Diatom markers (20:5 ω 3, 16:1 ω 7 and 16:2 ω 4) were also present in primary consumers, especially zooplankton, ascidians, and the detritivore-herbivores *Acanthurus nigricans* and *Ctenochaetus marginatus*. Although we did not sample bacteria/cyanobacteria, markers typical of these organisms (FAs 15:0, 17:0, 17:0iso, and 18:1 ω 7) were found in ascidians, *Pinctada margaritifera* and all fish. This suggest a potential role of bacteria in the Marquesan

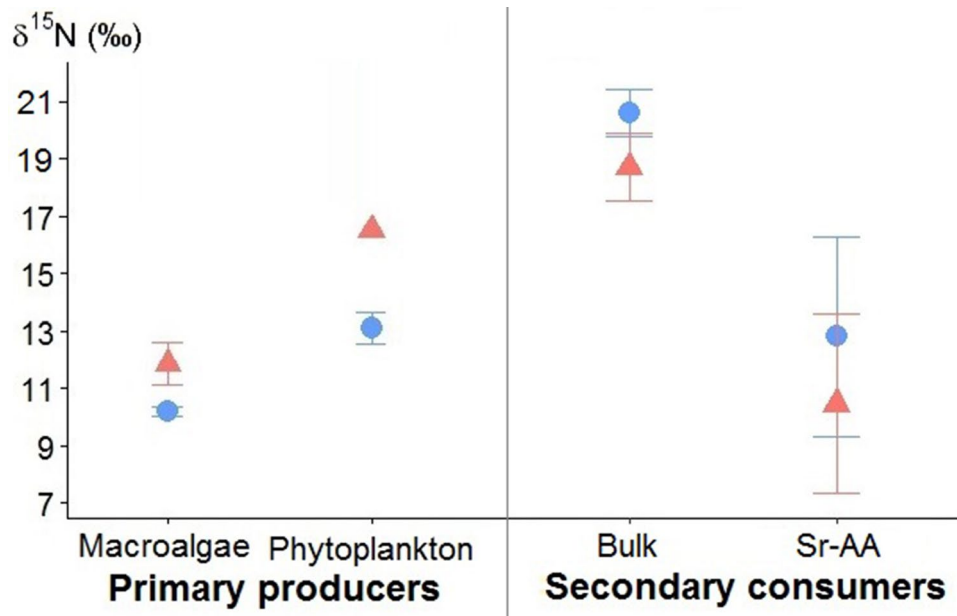


Figure 4. Seasonal variations of bulk $\delta^{15}\text{N}$ values (means \pm standard deviation) for the most integrated primary producers (left panel), and bulk, and Sr-AA (phenylalanine and glycine) for our eight targeted secondary consumers, all species pooled (right panel). Blue dots: winter, red triangles: summer.

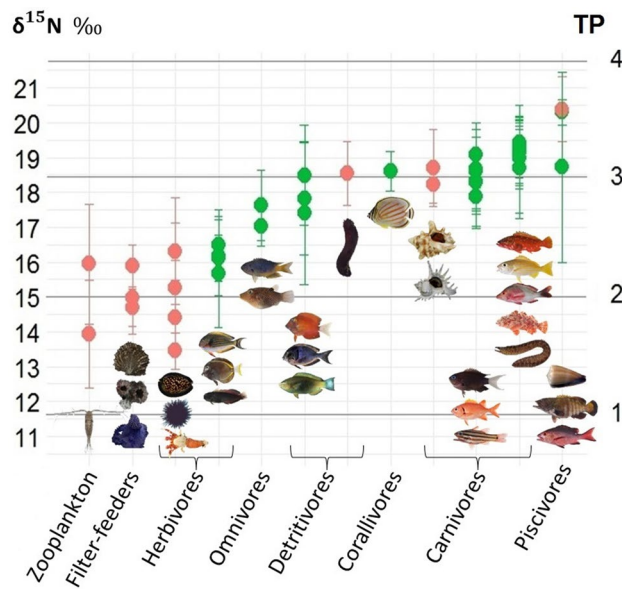


Figure 5. Relationships between mean bulk $\delta^{15}\text{N}$ values and trophic positions (TPs) calculated with Sr-AA $\delta^{15}\text{N}_{\text{phe-gly}}$ values and with macroalgae as the main source of organic matter. Each dot corresponds to the mean TP for a species. Black circles for sources of organic matter, red circles for invertebrates, and green circles for fish.

food web. Bacteria have not been previously shown to contribute substantially to the OM that support reef fishes. Therefore, future studies are needed to clarify their contribution. Cyanobacteria have been recorded on several herbivores fish diets such as in *Scarus* spp.⁶². Our FA data however cannot distinguish between filamentous turf-forming species and non-colonial small species. The strong contribution of the fatty acid 22:6 ω 3, a marker of dinoflagellates found in phytoplankton³, to the total FAs of zooplankton, *P. margaritifera*, *C. marginatus* and *Scarus* spp. indicated that phytoplankton was an important food source not only for zooplankton, but also for filter-feeders and some herbivores. However, we did not identify an important contribution of phytoplankton-derived OM for Acanthuridae and Scarinae, even though these taxa occasionally feed on zooplankton and indirectly assimilate phytoplankton-derived OM⁶³. We suggest that the link between this OM source and these

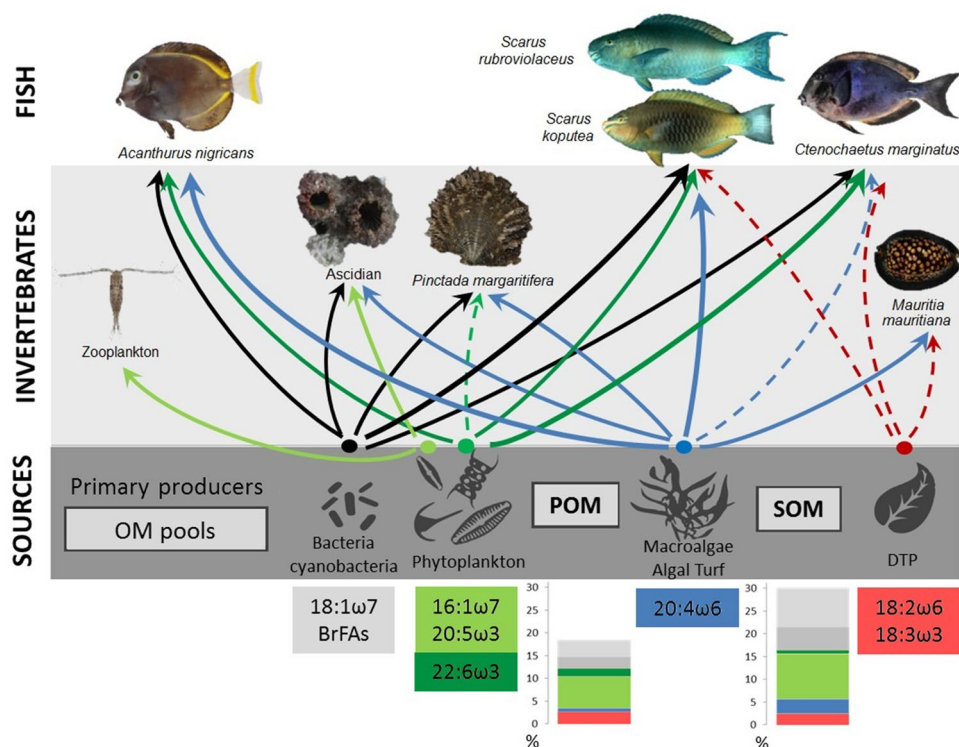


Figure 6. Schematic view of the integration of main OM sources by primary consumers, through fatty acid trophic markers. The relative importance of such integration is indicated by arrows (from dotted lines for minor role, to thick lines for important role). POM: particulate organic matter, SOM: sedimentary organic matter, DTP: detrital terrestrial plant material. For OM pools (POM and SOM), the % indicate the relative importance of primary producers (bacteria, phytoplankton, DTP etc.) assessed by major FAs trophic markers.

primary consumers is probably indirect, through deposition and accumulation of recently dead phytoplankton (or zooplankton) on sediments and macroalgae.

The compound specific stable isotopes (Sr-AA $\delta^{15}\text{N}_{\text{phe-gly}}$) allowed us to define the $\delta^{15}\text{N}$ values of the food web baseline. These values were higher in winter than in summer (Fig. 4), in contrast to the seasonal variability recorded in bulk isotope compositions. This apparent contradiction may be due to a temporal lag. Indeed, the isotope compositions of the source amino acids were measured on consumer, which likely have a longer turnover than primary producers. Also, the estimated $\delta^{15}\text{N}$ values of the food web baseline obtained with bulk stable isotope data reflected relatively recent variations of isotope composition (i.e. isotope composition of the OM sources at the time of collection, typical of the sampled season). In contrast, the amino acid isotope compositions provided a value of the sources referred to a longer time frame corresponding to the renewal time of the tissues analyzed, i.e. ~ 3 months before sampling for fish muscle.

The unusually high bulk $\delta^{15}\text{N}$ values of OM sources up to mesopredators suggest a high $\delta^{15}\text{N}$ value of the food web baseline that then propagates through the system. It is thus essential to understand why this pattern exists and why $\delta^{15}\text{N}$ values remain high across seasons despite winter / summer fluctuations (Fig. 7). The $\delta^{15}\text{N}$ values measured in the primary producers in the Marquesas were $\sim 8\text{--}10\text{‰}$ higher than in other Pacific sites^{2,9,10,64}. These $\delta^{15}\text{N}$ values are probably due to the ^{15}N enrichment of nutrient reservoirs in Marquesan waters, and seasonal changes in the strength of hydrodynamic processes such as eddies and upwelling³². $\delta^{15}\text{N}$ values increase in declining nitrate reservoirs, due to the more rapid assimilation of nitrates containing the light isotope (^{14}N) during photosynthesis⁶⁵. The high phytoplankton biomass throughout the year in the Marquesas Islands³⁰, and its use of nitrate (NO_3^-) helps explain the ^{15}N enrichment of the reservoirs of residual nutrients⁶⁶. In the Marquesas, the high nutrient intake in summer may promote significant fractionation and enhance the $\delta^{15}\text{N}$ values of the residual nitrate^{65,67} (Fig. 7). By assimilating ^{15}N -enriched nitrates, primary producers such as phytoplankton and macroalgae thus increase their $\delta^{15}\text{N}$ values. This is not the case for turf algae that sustain similar $\delta^{15}\text{N}$ values over seasons. Other processes may drive the ^{15}N enrichment of nitrates and primary producers, such as higher bacterial denitrification in summer⁶⁵, a possibly important role of bacteria in the functioning of the Marquesan coral reef ecosystem. Mineralization, nitrogen fixation, assimilation, nitrification and/or denitrification processes influence the ^{15}N composition of both inorganic (e.g. N_2 , NO_3^- , NO_2^-) and organic nitrogen species⁵⁰.

The assessment of trophic positions (TPs) of organisms within food webs is essential to understand ecosystem structure and functioning⁶⁸, although a thorough quantification remains challenging given the large number of factors involved (e.g. ontogeny of species, change of dietary regime). The $\delta^{15}\text{N}$ values of the source amino acids phenylalanine and glycine indicate that the Marquesan coastal food web is mostly supported by macroalgae.

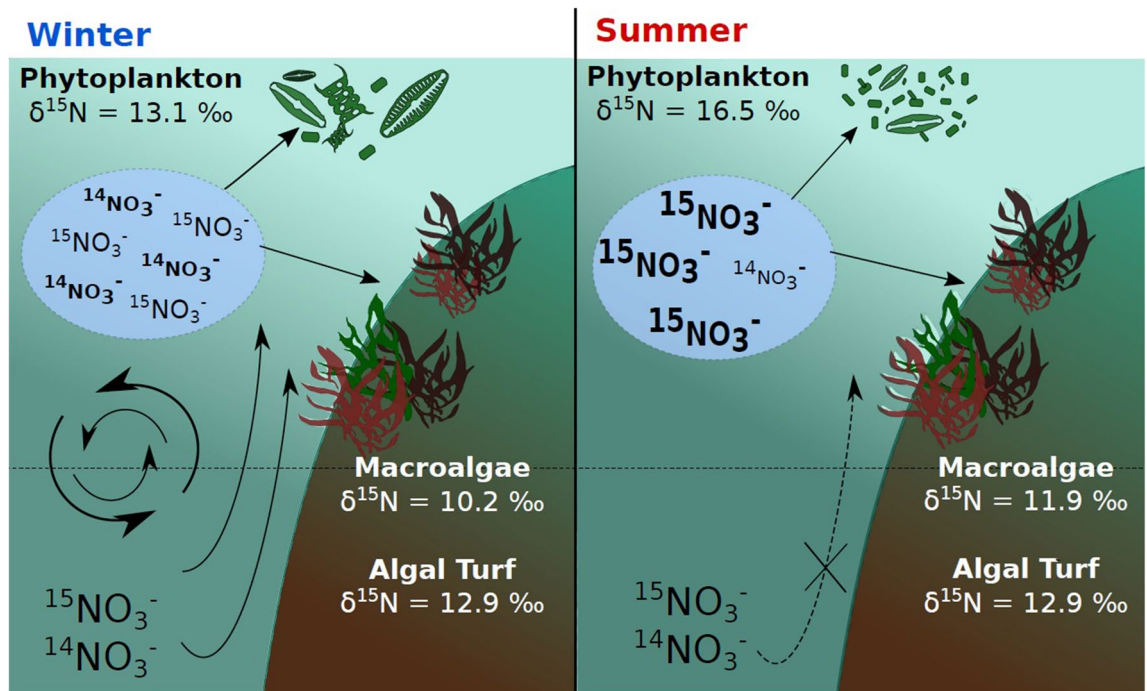


Figure 7. Schematic view of seasonal processes that could explain seasonal differences in nitrogen isotope compositions in primary producers.

However, the high variability around the mean implies that this finding should be considered with some caution. Seasonal fluctuations in phytoplankton isotope composition in the Marquesas make this compartment an important food source for several primary consumers³. This suggests that coupled pelagic-benthic processes likely drive the general characteristics and properties of OM sources over seasons. The TP values found in other studies are close to ours, despite very different bulk $\delta^{15}\text{N}$ values. For instance in a Caribbean coral reef, the five carnivorous fish having the highest TPs reached values of ~ 3.3 , for bulk $\delta^{15}\text{N}$ values of $\sim 9.5\text{‰}$ ⁵⁷. In a Polynesian coral reef, those values reached ~ 3.5 and $\sim 11\text{‰}$, respectively⁵⁶. Overall, the TPs we obtained for the different species or trophic groups are very similar to those from these studies, implying similar food chain lengths.

The overall increase in the TP for all consumers in summer did not alter this general pattern. Under the hypothesis that the Marquesas may be considered a present-day reference for degraded reefs, the Nuku Hiva data suggest that the length of the food chain may remain unchanged in future coral reefs subject to anthropogenic degradations, as also observed in Mexico⁵⁷. However, systems with similar food chain lengths may strongly differ in energy flows. For instance, the seasonality in TP values in the Marquesas, which were mostly apparent for omnivores and some carnivores (Suppl. Figure S2), also reflected a change in the use of OM and in the energy pathway within food webs. Coral losses alter isotope compositions of coral reef fish⁶⁹. Since environmental changes may produce substantial effects on fish abundance and biomass⁷⁰, ongoing coral reefs degradations may drastically affect fluxes of energy and ecosystem functioning⁷. While some coral reefs appeared to be resilient to disturbances, mostly thanks to herbivory⁷¹, a shift from coral towards algal reefs seems likely in the coming decades^{72–74}. On such reefs, the role of macroalgae and plankton driven by nutrient enrichments especially for nitrogen might grow. Although the present Nuku Hiva data were mainly qualitative, a greater potential role of bacterial / cyanobacterial communities as significant sources of OM in future coastal systems needs more consideration than it has had to date^{75, 76}.

Conclusions

Although some characteristics appear to be similar to other tropical reef systems, in many respects the food web functioning of the Marquesas Islands coral reef ecosystems is quite atypical. Fatty acid data highlighted that the roles of several sources of OM, such as bacteria / cyanobacteria which are generally difficult to collect in classical food web studies, may have been underestimated in other reef systems. Pelagic-derived OM may exceed the key-role of macroalgae as the main baseline of this food web. Similar conclusions regarding the importance of pelagic-derived organic matter as a food source for coral reefs were drawn from other studies^{13, 77}. This reinforces the hypothesis of a food web based on pelagic-benthic coupling³, and suggests that such functioning might become an increasingly important characteristic in future coral reefs. Despite the unusually high $\delta^{15}\text{N}$ values in all compartments of the studied food web, the trophic positions of consumers were comparable to other reef systems. Although this suggests that food chain lengths might not be much affected by the expected loss of corals, seasonal and more long-term variations in the use of OM by consumers will likely affect energy flows. Much further research is needed to better assess how coral reef flows of energy and organic matter might change in the next decades. The current Marquesas Islands coral reefs offer one plausible future scenario for functioning

of this ecosystem. The resilience of the Marquesan reef ecosystem may be low and this raises concerns about its capacity to resist future changes.

Received: 10 December 2020; Accepted: 11 October 2021

Published online: 25 October 2021

References

- Folke, C. *et al.* Regime shifts, resilience, and biodiversity in ecosystem management. *Annu. Rev. Ecol. Evol. Syst.* **35**, 557–581. <https://doi.org/10.1146/annurev.ecolsys.35.021103.105711> (2004).
- Briand, M. J., Bonnet, X., Goiran, C., Guillou, G. & Letourneur, Y. Major sources of organic matter in a complex coral reef lagoon: Identification from isotopic signatures ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$). *PLoS ONE* **10**, e0131555. <https://doi.org/10.1371/journal.pone.0131555> (2015).
- Fey, P. *et al.* Sources of organic matter in an atypical phytoplankton rich coral ecosystem, Marquesas Islands: composition and properties. *Mar. Biol.* **167**, 92. <https://doi.org/10.1007/s00227-020-03703-z> (2020).
- Briand, M. J., Bonnet, X., Guillou, G. & Letourneur, Y. Complex food webs in highly diversified coral reefs: insights from $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ stable isotopes. *Food Webs* **8**, 12–22. <https://doi.org/10.1016/j.fooweb.2016.07.002> (2016).
- Bierwagen, S. L., Heupel, M. R., Chin, A. & Simpfendorfer, C. A. Trophodynamics as a tool for understanding coral reef ecosystems. *Front. Mar. Sci.* <https://doi.org/10.3389/fmars.2018.00024> (2018).
- Halpern, B. S. *et al.* A Global map of human impact on marine ecosystems. *Science* **319**, 948–952. <https://doi.org/10.1126/science.1149345> (2008).
- Hughes, T. P. *et al.* Global warming transforms coral reef assemblages. *Nature* **556**, 492–496. <https://doi.org/10.1038/s41586-018-0041-2> (2018).
- Hughes, T. P. *et al.* Global warming impairs stock–recruitment dynamics of corals. *Nature* **568**, 387–390. <https://doi.org/10.1038/s41586-019-1081-y> (2019).
- Wyatt, A. S. J., Waite, A. M. & Humphries, S. Stable isotope analysis reveals community-level variation in fish trophodynamics across a fringing coral reef. *Coral Reefs* **31**, 1029–1044 (2012).
- Letourneur, Y. *et al.* Identifying carbon sources and trophic position of coral reef fishes using diet and stable isotope ($\delta^{15}\text{N}$ and $\delta^{13}\text{C}$) analyses in two contrasted bays in Moorea, French Polynesia. *Coral Reefs* **32**, 1091–1102. <https://doi.org/10.1007/s00338-013-1073-6> (2013).
- Zhu, Y., Newman, S. P., Reid, W. D. K. & Polunin, N. V. C. Fish stable isotope community structure of a Bahamian coral reef. *Mar. Biol.* **166**, 160. <https://doi.org/10.1007/s00227-019-3599-9> (2019).
- McMahon, K. W., Thorrold, S. R., Houghton, L. A. & Berumen, M. L. Tracing carbon flow through coral reef food webs using a compound-specific stable isotope approach. *Oecologia* **180**, 809–821. <https://doi.org/10.1007/s00442-015-3475-3> (2015).
- Skinner, C. *et al.* Offshore pelagic subsidies dominate carbon inputs to coral reef predators. *Sci. Adv.* <https://doi.org/10.1126/sciadv.abf3792> (2021).
- Mann, K. H. Production and use of detritus in various freshwater, estuarine and coastal marine ecosystems. *Limnol. Oceanogr.* **33**, 910–930 (1988).
- Antonio, B., Maria Teresa, A.-O. & Manuel, V. Phytoplankton and macrophyte contributions to littoral food webs in the Galician upwelling estimated from stable isotopes. *Mar. Ecol. Prog. Ser.* **318**, 89–102 (2006).
- Gazeau, F., Smith, S. V., Gentili, B., Frankignoulle, M. & Gattuso, J.-P. The European coastal zone: characterization and first assessment of ecosystem metabolism. *Est. Coast. Shelf Sci.* **60**, 673–694. <https://doi.org/10.1016/j.ecss.2004.03.007> (2004).
- Hamner, W. M., Jones, M. S., Carleton, J. H., Hauri, I. R. & Williams, D. M. Zooplankton, planktivorous fish, and water currents on a windward reef face: great Barrier Reef, Australia. *Bull. Mar. Sci.* **42**, 459–479 (1988).
- Hamner, W. M., Colin, P. L. & Hamner, P. P. Export–import dynamics of zooplankton on a coral reef in Palau. *Mar. Ecol. Prog. Ser.* **334**, 83–92 (2007).
- Carassou, L., Kulbicki, M., Nicola, T. J. R. & Polunin, N. V. C. Assessment of fish trophic status and relationships by stable isotope data in the coral reef lagoon of New Caledonia, southwest Pacific. *Aquat. Living Resour.* **21**, 1–12 (2008).
- Frédérich, B., Fabri, G., Lepoint, G., Vandewalle, P. & Parmentier, E. Trophic niches of thirteen damselfishes (Pomacentridae) at the Grand Récif de Toliara, Madagascar. *Ichthyol. Res.* **56**, 10–17. <https://doi.org/10.1007/s10228-008-0053-2> (2009).
- Riera, P. & Richard, P. Isotopic determination of food sources of *Crassostrea gigas* along a trophic gradient in the estuarine bay of Marennes-Oléron. *Estuar. Coast. Shelf Sci.* **42**, 347–360 (1996).
- Hoegh-Guldberg, O. Climate change, coral bleaching and the future of the world's coral reefs. *Mar. Fresh. Wat. Res.* **50**, 839–866 (1999).
- Bellwood, D. R., Hughes, T. P., Folke, C. & Nyström, M. Confronting the coral reef crisis. *Nature* **429**, 827–833 (2004).
- Bruno, J. F. & Selig, E. R. Regional decline of coral cover in the Indo-Pacific: Timing, extent, and subregional comparisons. *PLoS ONE* **2**, e711. <https://doi.org/10.1371/journal.pone.0000711> (2007).
- Roff, G. *et al.* Porites and the Phoenix effect: unprecedented recovery after a mass coral bleaching event at Rangiroa Atoll, French Polynesia. *Mar. Biol.* **161**, 1385–1393. <https://doi.org/10.1007/s00227-014-2426-6> (2014).
- Hoey, A. *et al.* Recent advances in understanding the effects of climate change on coral reefs. *Diversity* **8**, 12 (2016).
- Cabioch, G. *et al.* Successive reef depositional events along the Marquesas foreslopes (French Polynesia) since 26 ka. *Mar. Geol.* **254**, 18–34. <https://doi.org/10.1016/j.margeo.2008.04.014> (2008).
- Galzin, R., Duron, S. D. & Meyer, J. Y. *Biodiversité terrestre et marine des îles Marquises, Polynésie française*. (Société française d'Ichtyologie, 2016).
- SO CORAIL. *Site d'observation CORAIL*, <https://sextant.ifremer.fr/record/le51de1b-7979-4487-b5d5-329394d166da> (2018).
- Martinez, E., M., R. & Maamaatuaiahutapu, K. in *Biodiversité terrestre et marine des îles Marquises, Polynésie française* (eds Galzin R., Duron S.-D., & Meyer J.-Y) 123–136 (Société Française d'Ichtyologie, 2016).
- Houk, P. & Musburger, C. Trophic interactions and ecological stability across coral reefs in the Marshall Islands. *Mar. Ecol. Prog. Ser.* **488**, 23–34 (2013).
- Raapoto, H., Martinez, E., Petrenko, A., Doglioli, A. M. & Maes, C. Modeling the Wake of the Marquesas Archipelago. *J. Geophys. Res. Oceans* **123**, 1213–1228. <https://doi.org/10.1002/2017jc013285> (2018).
- Vander Zanden, M. J. & Rasmussen, J. B. Variation in $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ trophic fractionation: Implications for aquatic food web studies. *Limnol. Oceanogr.* **46**, 8 (2001).
- De Niro, M. J. & Epstein, S. Influence of diet on the distribution of carbon isotopes in animals. *Geochim. Cosmochim. Acta* **42**, 495–506 (1978).
- Layman, C. A. *et al.* Applying stable isotopes to examine food-web structure: an overview of analytical tools. *Biol. Rev.* **87**, 545–562. <https://doi.org/10.1111/j.1469-185X.2011.00208.x> (2012).
- Pinnegar, J. & Polunin, N. V. C. Differential fractionation of d^{13}C and d^{15}N among fish tissues: implications for the study of trophic interactions. *Funct. Ecol.* **13**, 225–231 (1999).

37. De Niro, M. J. & Epstein, S. Influence of diet on the distribution of carbon isotopes in animals. *Geochim. Cosmochim. Acta* **42**, 495–506 (1978).
38. Hannides, C. C. S., Popp, B. N., Landry, M. R. & Graham, B. S. Quantification of zooplankton trophic position in the North Pacific Subtropical Gyre using stable nitrogen isotopes. *Limnol. Oceanogr.* **54**, 50–61. <https://doi.org/10.4319/lo.2009.54.1.0050> (2009).
39. Hannides, C. C. S., Popp, B. N., Choy, C. A. & Drazen, J. C. Midwater zooplankton and suspended particle dynamics in the North Pacific Subtropical Gyre: a stable isotope perspective. *Limnol. Oceanogr.* **58**, 1931–1946. <https://doi.org/10.4319/lo.2013.58.6.1931> (2013).
40. Post, D. M. Using stable isotopes to estimate trophic position: models, methods, and assumptions. *Ecology* **83**, 703–710 (2002).
41. Meziane, T. *et al.* Inter-specific and geographical variations in the fatty acid composition of mangrove leaves: implications for using fatty acids as a taxonomic tool and tracers of organic matter. *Mar. Biol.* **150**, 1103–1113. <https://doi.org/10.1007/s00227-006-0424-z> (2007).
42. Parrish, C. C. *et al.* in *Marine Chemistry* (ed P. J. Wangersky) 193–223 (Springer Berlin Heidelberg, 2000).
43. Alfaro, A. C., Thomas, F., Sergent, L. & Duxbury, M. Identification of trophic interactions within an estuarine food web (northern New Zealand) using fatty acid biomarkers and stable isotopes. *Est. Coast. Shelf Sci.* **70**, 271–286. <https://doi.org/10.1016/j.ecss.2006.06.017> (2006).
44. Meyers, P. A. Organic geochemical proxies of paleoceanographic, paleolimnologic, and paleoclimatic processes. *Org. Geochem.* **27**, 213–250. [https://doi.org/10.1016/S0146-6380\(97\)00049-1](https://doi.org/10.1016/S0146-6380(97)00049-1) (1997).
45. Dalsgaard, J., St. John, M., Kattner, G., Müller-Navarra, D. & Hagen, W. in *Advances in Marine Biology* Vol. 46 225–340 (Academic Press, 2003).
46. Volkman, J. K., Jeffrey, S. W., Nichols, P. D., Rogers, G. I. & Garland, C. D. Fatty acid and lipid composition of 10 species of microalgae used in mariculture. *J. Exp. Mar. Biol. Ecol.* **128**, 219–240. [https://doi.org/10.1016/0022-0981\(89\)90029-4](https://doi.org/10.1016/0022-0981(89)90029-4) (1989).
47. Volkman, J. K., Johns, R. B., Gillan, F. T., Perry, G. J. & Bavor, H. J. Microbial lipids of an intertidal sediment—I. Fatty acids and hydrocarbons. *Geochimica et Cosmochimica Acta* **44**, 1133–1143. [https://doi.org/10.1016/0016-7037\(80\)90067-8](https://doi.org/10.1016/0016-7037(80)90067-8) (1980).
48. Lee, R. F., Hirota, J. & Barnett, A. M. Distribution and importance of wax esters in marine copepods and other zooplankton. *Deep Sea Res. A* **18**, 1147. [https://doi.org/10.1016/0011-7471\(71\)90023-4](https://doi.org/10.1016/0011-7471(71)90023-4) (1971).
49. Wakeham, S. G., Hedges, J. I., Lee, C., Peterson, M. L. & Hernes, P. J. Compositions and transport of lipid biomarkers through the water column and surficial sediments of the equatorial Pacific Ocean. *Deep Sea Res. Part II* **44**, 2131–2162. [https://doi.org/10.1016/S0967-0645\(97\)00035-0](https://doi.org/10.1016/S0967-0645(97)00035-0) (1997).
50. Budge, S. M. & Parrish, C. C. Lipid biogeochemistry of plankton, settling matter and sediments in Trinity Bay, Newfoundland. II. Fatty acids. *Organic Geochem.* **29**, 1547–1559. [https://doi.org/10.1016/S0146-6380\(98\)00177-6](https://doi.org/10.1016/S0146-6380(98)00177-6) (1998).
51. Meziane, T., Agata, D. F. & Lee, S. Y. Fate of mangrove organic matter along a subtropical estuary: small-scale exportation and contribution to the food of crab communities. *Mar. Ecol. Prog. Ser.* **312**, 15–27 (2006).
52. Phillips, D. L. & Gregg, J. W. Source partitioning using stable isotopes: coping with too many sources. *Oecologia* **136**, 261–269 (2003).
53. Parnell, A. C., Inger, R., Bearhop, S. & Jackson, A. L. Source partitioning using stable isotopes: Coping with too much variation. *PLoS ONE* **5**, e9672. <https://doi.org/10.1371/journal.pone.0009672> (2010).
54. R Core Team. (R Foundation for Statistical Computing, Vienna, Austria, 2018).
55. du Percie, S. *et al.* Reporting animal research: explanation and elaboration for the ARRIVE guidelines 2.0. *PLOS Biol.* **18**, e3000411. <https://doi.org/10.1371/journal.pbio.3000411> (2020).
56. Page, H. M. *et al.* Stable isotopes reveal trophic relationships and diet of consumers in temperate kelp forest and coral reef ecosystems. *Oceanography* **26**, 180–189 (2013).
57. Morillo-Velarde, P. S. *et al.* Habitat degradation alters trophic pathways but not food chain length on shallow Caribbean coral reefs. *Sci. Rep.* **8**, 4109. <https://doi.org/10.1038/s41598-018-22463-x> (2018).
58. Bellwood, D. R. & Choat, J. H. A functional analysis of grazing in parrotfishes (family Scaridae): The ecological implications. *Environ. Biol. Fish.* **28**, 189–214 (1990).
59. Choat, J. H., Clements, K. D. & Robbins, W. D. The trophic status of herbivorous fishes on coral reefs. I: Dietary analyses. *Mar. Biol.* **140**, 613–623 (2002).
60. Dromard, C. R. *et al.* Resource use of two damselfishes, *Stegastes planifrons* and *Stegastes adustus*, on Guadeloupean reefs (Lesser Antilles): Inference from stomach content and stable isotope analysis. *J. Exp. Mar. Biol. Ecol.* **440**, 116–125. <https://doi.org/10.1016/j.jembe.2012.12.011> (2013).
61. Hedges, J. I. *et al.* Compositions and fluxes of particulate organic material in the Amazon River. *Limnol. Oceanogr.* **31**, 717–738. <https://doi.org/10.4319/lo.1986.31.4.0717> (1986).
62. Nicholson, G. M. & Clements, K. D. Resolving resource partitioning in parrotfishes (Scarini) using microhistology of feeding substrata. *Coral Reefs* **39**, 1313–1327. <https://doi.org/10.1007/s00338-020-01964-0> (2020).
63. Clements, K. D., German, D. P., Piché, J., Tribollet, A. & Choat, J. H. Integrating ecological roles and trophic diversification on coral reefs: multiple lines of evidence identify parrotfishes as microphages. *Biol. J. Lin. Soc.* **120**, 729–751. <https://doi.org/10.1111/bij.12914> (2016).
64. Bradley, C. J., Longenecker, K., Pyle, R. L. & Popp, B. N. Compound-specific isotopic analysis of amino acids reveals dietary changes in mesophotic coral-reef fish. *Mar. Ecol. Prog. Ser.* **558**, 65–79 (2016).
65. Raimbault, P., Garcia, N. & Cerutti, F. Distribution of inorganic and organic nutrients in the South Pacific Ocean—evidence for long-term accumulation of organic matter in nitrogen-depleted waters. *Biogeosciences* **5**, 281. <https://doi.org/10.5194/bg-5-281-2008> (2008).
66. Savoye, N. *et al.* Dynamics of particulate organic matter d15N and d13C during spring phytoplankton blooms in a macrotidal ecosystem (Bay of Seine, France). *Mar. Ecol. Prog. Ser.* **255**, 27–41 (2003).
67. Montoya, J. P. & McCarthy, J. J. Isotopic fractionation during nitrate uptake by phytoplankton grown in continuous culture. *J. Plankton Res.* **17**, 439–464. <https://doi.org/10.1093/plankt/17.3.439> (1995).
68. Hussey, N. E. *et al.* Rescaling the trophic structure of marine food webs. *Ecol. Lett.* **17**, 239–250. <https://doi.org/10.1111/ele.12226> (2014).
69. Letourneur, Y., Briand, M. J. & Graham, N. A. J. Coral reef degradation alters the isotopic niche of reef fishes. *Mar. Biol.* **164**, 224. <https://doi.org/10.1007/s00227-017-3272-0> (2017).
70. Graham, N. A. J. *et al.* Extinction vulnerability of coral reef fishes. *Ecol. Lett.* **14**, 341–348 (2011).
71. Viviani, J. *et al.* Synchrony patterns reveal different degrees of trophic guild vulnerability after disturbances in a coral reef fish community. *Divers. Distrib.* **25**, 1210–1221. <https://doi.org/10.1111/ddi.12931> (2019).
72. Diaz-Pulido, G., Gouezo, M., Tilbrook, B., Dove, S. & Anthony, K. R. N. High CO₂ enhances the competitive strength of seaweeds over corals. *Ecol. Lett.* **14**, 156–162. <https://doi.org/10.1111/j.1461-0248.2010.01565.x> (2011).
73. Koch, M., Bowes, G., Ross, C. & Zhang, X.-H. Climate change and ocean acidification effects on seagrasses and marine macroalgae. *Glob. Change Biol.* **19**, 103–132. <https://doi.org/10.1111/j.1365-2486.2012.02791.x> (2013).
74. Ainsworth, T. D. *et al.* Climate change disables coral bleaching protection on the Great Barrier Reef. *Science* **352**, 338–342. <https://doi.org/10.1126/science.aac7125> (2016).
75. Jackson, J. B. C. What is natural in the coastal oceans?. *Proc. Natl. Acad. Sci. USA* **98**, 5411–5418 (2001).

76. Bourne, D. G., Morrow, K. M. & Webster, N. S. Insights into the coral microbiome: underpinning the health and resilience of Reef ecosystems. *Annu. Rev. Microbiol.* **70**, 317–340. <https://doi.org/10.1146/annurev-micro-102215-095440> (2016).
77. Morais, R. A. & Bellwood, D. R. Pelagic subsidies underpin fish productivity on a degraded coral reef. *Curr. Biol.* **29**, 1521–1527. e1526. <https://doi.org/10.1016/j.cub.2019.03.044> (2019).

Acknowledgements

This article is a part of the first author's PhD, funded by the LabEx "Corail" (project RETROMAR). We would like to thank the Centre de Plongée des Marquises (X. Curvat and his crew) for field assistance and J.Y. Meyer from the Délégation à la Recherche de la Polynésie française for its financial support. We are also grateful to C. Berthe, F. Bouilleret, D. Lecchini, R. Sauze, and M. Taquet for their field assistance. We thank the staff of the CRILOBE, especially C. Sidobre and B. Espiau, for laboratory assistance and N. Thiney for her invaluable help for FAs analyses in the Muséum National d'Histoire Naturelle, Paris. G. Guillou and C. Yarnes are acknowledged for running bulk and compound-specific stable isotope analyses, respectively. Compound specific stable isotope analyses were supported by the Cluster of Excellence 80 "The Future Ocean" within the Excellence Initiative by the Deutsche Forschungsgemeinschaft (DFG).

Author contributions

P.F., V.P., R.G. and Y.L. conceived and designed the research. P.F., V.P., R.G., M.Z. and Y.L. conducted the field-work. P.F., D.B., B.L., T.M., M.Z. and Y.L. analyzed the samples. P.F. and Y.L. wrote the manuscript, with significant inputs from V.P., D.B., J.D., R.G., B.L., T.M., N.V.C.P. and M.Z. All authors contributed critically to the drafts and gave final approval for publication.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1038/s41598-021-00348-w>.

Correspondence and requests for materials should be addressed to Y.L.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2021, corrected publication 2021