

Parkinson disease: Protective role and function of neuropeptides

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24 Abstract

Neuropeptides are bioactive molecules, made up of small chains of amino acids, with many 25 neuromodulatory properties. Several lines of evidence suggest that neuropeptides, mainly expressed 26 in the central nervous system (CNS), play an important role in the onset of Parkinson's Disease 27 28 (PD) pathology. The wide spread disruption of neuropeptides has been excessively demonstrated to be related to the pathophysiological symptoms in PD where impairment in motor function per 29 example was correlated with neuropeptides dysregulation in the substantia niagra (SN). Moreover, 30 31 the levels of different neuropeptides have been found modified in the cerebrospinal fluid and blood of PD patients, indicating their potential role in the manifestation of PD symptoms and 32 dysfunctions. In this review, we outlined the neuroprotective effects of neuropeptides on 33 dopaminergic neuronal loss, oxidative stress and neuroinflammation in several models and tissues 34 of PD. Our main focus was to elaborate the role of orexin, pituitary adenylate cyclase activating 35 polypeptide (PACAP), vasoactive intestinal peptide (VIP), opioids, angiotensin, carnosine and 36 37 many others in the protection and/or involvement in the neurodegeneration of striatal dopaminergic cells. Further studies are required to better assess the mode of action and cellular mechanisms of 38 neuropeptides in order to shift the focus from the *in vitro* and *in vivo* testing to applicable clinical 39 40 testing. This review, allows a support for future use of neuropeptides as therapeutic solution for PA 41 pathophysiology.

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43 Keywords:

44 Parkinson disease, neuropeptide, orexin, PACAP, VIP, opioid, angiotensin, carnosine

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52 1. Introduction

53 Parkinson's disease (PD) is one of the most common neurodegenerative diseases, second only to Alzheimer's disease with a late onset of 40-60's years of age [1]. Traditionally, it's characterized by 54 55 dopaminergic degeneration in the substancia niagra pars compacta (SNpc), a midbrain basal ganglia 56 structure implicated mainly in movement, with a deposition of insoluble cytoplasmic proteins in 57 surviving cells called "Lewy bodies". PD leads to an abundance of motor dysfunctions such as bradykinesia (slowness of movements), muscular rigidity, rest tremor, and postural and gait 58 59 impairment [2,3]. Several other brain structures are also involved in PD pathophysiology: lesions in all of 13 nuclei in the hypothalamus were linked to sleep disorders and cognitive impairment. The 60 posterior cingulum, a limbic lobe structure activated during retrieval of episodic memory, is 61 62 affected by the neurodegeneration, astrogliosis and demyelination in PD [4]. Besides the involvement of many brain regions, even peripheral tissues are affected by PD such as the 63 cardiovascular system, the urinary tract, the skin and sweat glands, gastrointestinal tract, and the 64 65 neuroendocrine structures. As a result, PD comes with other non-motor symptoms summarized by affective, cognitive, dizziness, olfactory dysfunctions among many others [5]. This broad range of 66 disabling symptoms causes not only a reduction of quality of life but also an emotional and 67 68 economic burden on the caregivers [6].

⁶⁹ 2. Parkinson's disease pathophysiology

70 2.1. Direct/indirect pathways

In the direct pathway, the substantia niagra activates D1 receptors, via dopaminergic stimulation, 71 72 leading to the activation of GABAergic MSNs (striatal medium spiny neurons, targets of gamma-73 aminobutyric acid) that inhibits the internal pallidum, leading to the disinhibition of the thalamic glutamatergic neurons that have an excitatory glutamatergic action on the cortex. This leads to 74 75 locomotor activation. Conversely, in the indirect pathway, the substantia niagra (SN) activates D2 receptors which inhibits the GABAergic neurons on the external pallidum, leading to the 76 disinhibition of the glutamatergic neurons of the sub thalamic nucleus (STN). These in turn 77 78 activates the internal pallidum's GABAergic neurons that project to the thalamus. This results in the

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- reduction of locomotor activity. PD is a result of dopamine (DA) depletion in the SN pars compacta
- 80 which results in an overstimulation in the indirect pathway and a reduced activation of the direct
- 81 pathway. Thus, causing overall reduced locomotor activity [7] (Figure 1).



83 Figure 1. SN modality role is executed through dopaminergic projection to the striatum with D1 excitatory 84 synapse and D2 inhibitory synapse. The downstream effect of nigrostriatal D1 excitation of the striatum will result in an inhibition of the internal Globus Pallidus by GABAergic projection from the striatum to the 85 86 internal globus palidus through the direct pathway. Its inhibition will allow the activation of motor activity 87 through the thalamus. In the indirect pathway, the SN will inhibit the GABAergic inhibitory projection from 88 the striatum to the external globus pallidus, this will therefore allow the subthalamic nucleus to activate the 89 internal globus pallidus through glutamate projection in order to inhibit the thalamus and result in the 90 inhibition of the motor activity. Therefore, the SN will allow motor activity through the direct pathway and 91 inhibit it through the indirect pathway. In the PD case, SN modulation of the striatum is attenuated by the 92 dopaminergic degeneration. The activity of the direct pathway will be decreased, and the activity of the 93 indirect pathway will be increased, leading to the overinhbition of the thalamus that will decrease the motor 94 activity.

95 2.2. Alpha synuclein

 α -synuclein is a 140 amino acid protein mostly found in the brain, especially in dopaminergic

- 97 neurons of the SN in form of Lewy bodies [8]. α -synuclein's proper structure hasn't been clearly
- 98 identified, which is why its termed as a natively unfolded protein. It can have several

conformations: monomeric, fibrillar, α-helical, β-pleated sheet and an amyloid structure in the 99 fibrils of Lewy bodies, a hallmark of PD pathophysiology [9,10]. Mutations occurring in the 3 100 101 domains of a-synuclein lead to its pathological folding, oligomerization, formation of fibrils that 102 leads to Lewy bodies, are responsible of the toxicity induced by α -synuclein found in familiar PD [11]. α -synuclein is expressed in all brain regions, however, Lewy bodies and neurodegenerations 103 104 are abundant especially in the SNpc. This could be related to the major presence of the DA neurons in the SN. In fact, during the metabolism of DA, reactive oxygen species (ROS) that stimulate 105 106 apoptosis, might interact with the α -synuclein and aggregation them into protofibrils. The latter 107 have been shown to play major role in α -synuclein toxicity [10].

108 2.3. Oxidative stress neural inflammation and mitochondrial dysfunction

The pathophysiological hallmark of PD is indisputably considered as the neurodegeneration of 109 SNpc dopaminergic projection to the striatum. But the true origin of such insult is a subject of 110 discussion. Hence, many theories are trying to decipher the underlying cause of DA neurons death. 111 112 The literature presents oxidative stress, neural inflammation, and mitochondrial dysfunction to 113 converge and intertwine to cause cellular death and degeneration [12]. In fact, oxidative stress is 114 manifested when the balance is disrupted between the ROS being produced and accumulated in 115 cells and tissues, and their detoxification by the biological system [13]. Superoxide, hydroxyl radical, peroxynitrite, and H₂O₂ are ROS that are produced by the biological system through 116 117 enzymatic or non-enzymatic reactions. At low concertation, they can present a physiological cellular role such as in macrophage phagocytosis. But at higher concertation, they can alter and 118 119 damage several structures in the cell such as membrane, organelles, and even DNA leading to many chronic and neurodegenerative diseases [14]. Innately dopamine neurons are highly susceptible to 120 exhibit high ROS levels and oxidative stress: (i) they require a large amount of energy and have 121 long axons of 1-4-meter-long and a large number of synapses which are hard to sustain and require 122 high energy consumption; (ii) they exhibit an increased level of cytosolic dopamine and its 123 metabolite which can cause toxicity ROS formation [12]. 124

125 Neural inflammation is another player of PD neurodegeneration evidenced by the increase in pro-

inflammatory cytokines and factors in PD patients and animal models [15, 16]. One of the major

127 players of neural inflammation are microglial cells. These cells are residing macrophages in the

brain tissue responsible for monitoring brain homeostasis. Microglial cells can be activated into M1

129 phenotypes (proinflammatory) or M2 (anti-inflammatory) phenotype. Misfolded proteins and toxins

lead to the hyperactivation of the M1 phenotypes microglia in PD animal models. This 130

hyperactivation is theorized to cause DA cell death by phagocytosis of the neurons and by the 131

increase of ROS formation through inductible nitric oxide synthase (INOS) and nitric oxide 132

synthase (NOS) Enzymes [17]. 133

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Dysregulated mitochondrial protein, DNA, and function in PD hint at the involvement of 134 mitochondrial dysfunction in PD pathology of sporadic and familial PD [18,19]. Changes in

mitochondrial morphology and dynamic, inhibition of complex 1 of electron-transport chain, and 136

formation of ROS lead to cell energetic failure and toxicity and eventually degeneration [20]. 137

3. Parkinson's disease models 138

Various toxins induce motor deficits occurring in PD. Among these toxins MPTP (1-Methyl-4-139 phenyl-1,2,3,6-tetrahydropyridine), 6-OHDA (6-hydroxydopamine), and rotenone. MPTP, after 140 passing through the blood brain-barrier (BBB), is transformed by monoamine oxidase B into its 141 active metabolite MPP+. The dopamine transporter (DAT) carries MPP+ into dopaminergic 142 neurons of the SNpc, where it stops mitochondrial complex I activity causing the death of these 143 neurons. 6-OHDA, once injected locally (SNpc or medial forebrain bundle (MFB) due to its 144 inability to cross the BBB), builds up in the cytosol of dopaminergic neurons and rapidly auto-145 146 oxidizes leading to the formation of high levels of hydrogen peroxide. Thus, causing the degeneration of the nigrostriatal pathway marking the highest level of nigral cell loss and striatal 147 148 DA depletion obtainable in PD animal models. Rotenone, being highly lipophilic easily crosses the 149 BBB, once in the cell, it stops complex I activity, leading to massive formation of ROS, and 150 inhibition of the proteasome activity, thus causing proteolytic stress [21].

4. Neuropeptides 151

One of the main hypotheses proposed for PD pathology is the impairment in main protein's 152

degradation systems, such as ubiquitin proteasome system (UPS) and autophagy-lysosomal 153

pathways (ALP), that leads to accumulation of proteins, followed by cell toxicity and eventually 154

cell death [22]. A proteomic analysis done in PD patients, showed that 221 proteins were 155

156 dysregulated in the SN [23]. As well as in other biological samples, such as cerebrospinal fluid 157 (CSF), saliva and plasma, and other brain regions such as the caudate, the putamen and the globus

- pallidus where multiple dysregulations were found in several neuropeptides: Neuropeptide Y,
- Ghrelin, Substance P and Neurotensin [24]. Hence, the importance in studying the variation of thedifferent neuropeptides in PD.

161 First and foremost, a peptide is a chain of 2 to 50 amino acids, joined by covalent binds, of 50 kD mass [25]. When it's secreted by the neurons in the brain it's called a "neuropeptide". By definition 162 "a neuropeptide is a small proteinaceous substance produced and released by neurons through the 163 regulated secretory route and acting on neural substrates". According to H. Burbach, there are three 164 165 main pillars that identify neuropeptides: (i) their genes and proteins are, simultaneously, expressed 166 and released by neurons; (ii) they are stored in dense vesicles and released when needed and (iii) they have the ability to regulate, via neuroreceptors, the functioning of neurons. Neuropeptides' 167 168 release and activation is a multi-step progressive process from the endoplasmic reticulum, through the Golgi apparatus into secretory vesicles called dense core vesicles. All neuropeptides descend 169 170 from a precursor protein called pro-peptide obtained from the cleavage of a signal peptide, a proprepeptide modified by C-terminal amidation, in the endoplasmic reticulum. The pro-peptide then 171 172 undergoes multiple modifications, such as glycosylation and proteolytic cleavage, when passing the secretory route through the Golgi apparatus. The peptide is then transported through the dense core 173 174 vesicles where, in addition to proteolytic cleavage, C-terminal cleavage and C-terminal amidation occur for some peptides. The vesicles, now full of accumulated peptides, are transported from the 175 cell body down the axon. Upon stimulation, depolarization or other signals, the neuropeptides are 176 then released by calcium-dependent exocytosis [26, 27]. 177

Once released by neurons, neuropeptides can act on different cell and tissue types inside and outside 178 179 the brain, such as endocrine system, the blood, digestive system. Hence, their pleiotropic 180 capabilities. They show multi-level effects on genes, cells, organism and even on behaviors. 181 Interestingly, neuropeptides use volume transmission, a dispersion method independent from synaptic communication that allows them to diffuse from the neurons to large distances in the 182 extracellular fluid and CSF. Volume transmission, as well as lack of neuropeptides reuptake, eases 183 their travel across the brain into different organs, where they have several different effects. It's of 184 185 great interest to note the difference between neuropeptides and peptides. Both are involved in the

paracrine, autocrine and endocrine signaling. They are both synthesized and released by the sameenzymes but neuropeptides are discreetly secreted by neurons.

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Neuropeptides can have both neuroprotective and neurodegenerative effects. It's of great interest to

study their neuroprotective effect and their further therapeutic properties in neurodegenerative diseases. It's also important to investigate the neurodegenerative effects of some neuropeptides, in hopes of using them as biomarkers in the diagnosis of early-stage neurodegeneration, which will help in applying all neuroprotective agents as early as possible to ameliorate the disease prognosis.
In this article, we overviewed the neuroprotective effects of many neuropeptides in PD like: orexin, opioids, angiotensin, vasoactive intestinal peptide (VIP), pituitary adenylate cyclase activating polypeptide (PACAP), Neuropeptide Y (NPY), somatostatin, galanin, norepinephrine,

196 octadecaneuropeptide (ODN) and nerve growth hormone (VGF), a well as other peptides such as

197 carnosine, selanx, kinin and gastrointestinal peptide (GIP) and pancreatic peptide (PP). We included

198 studies on neuropeptides that are equal or smaller than 100 amino acids, and published between

199 2008 to June 2021. We investigated their effects on DA neuronal loss, inflammation, oxidative stress

and apoptosis on the cellular levels in different animal models of PD and in PD patients from

201 different biological samples. We also assessed the current state of neuropeptide' application on

these PD models which endeavor to find their effects on behavioral, cognitive and motor functions.

203 We aimed to present a solid basis that supports the use of neuropeptides as future therapies for PD.

A summary table classifying studies by location of neuropeptide's effect in PD and/or

205 neuropeptide's level in different locations is shown below. The major findings regarding cellular

206 effect (oxidative stress, inflammation, astrogliosis), motor dysfunctions, sleep disturbances and

207 cognitive/memory impairment are presented (+ signified increased; – signified decreased).

Study	Peptide	Location	Levels in PD	Cellular effect	Motor impairment	Sleep deficits	Cognitive/memory impairment
Huang <i>et al.</i> 2021	Orexin-A	Plasma	+ in plasma Orexin-A levels (early-stage and medium-stage PD)			+ in REM sleep	- in cognitive dysfunction
			- (advanced-stage PD)				
Stanojlovic, Pallais and Kotz 2021	Orexin	Lateral hypothalamus			+ exploratory locomotion & spontaneous physical activity	-	
Stanojlovic, Pallais, Lee <i>et al.</i> 2019	Orexin-A	Hippocampus		No effect on inflammation and astrogliosis			 hippocampus-dependent memory impairment
Stanojlovic, Pallais, Yllescas <i>et al.</i> 2019	Orexin-A	Medial prefrontal cortex					- social cognition impairment
Hadadianpour <i>et al.</i> 2017)	Orexin-A	CSF			+ locomotor activity+ ability to hold wire		
Wienecke <i>et al.</i> 2012	Hypocretin (Orexin)	CSF	 over time in advanced stage PD 			No association	
Xue et al. 2016	Orexin-A	Globus pallidus			+ body swing test		
Drouot <i>et al.</i> 2011	Orexin- A/hypocretin-1	Ventricular CSF	 over time in late-stage PD 			No association	
Compta <i>et al.</i> 2009	Orexin- A/hypocretin-2	CSF	No difference				No association
Bridoux <i>et al.</i> 2013	Orexin-A	CSF				+ correlation between Orexin-A and REM epochs without atonia	
Stanojlovic, Pallais, and Kotz 2019	Orexin-A	Hippocampus and motor cortex			+ distance covered in locomotion test		
Giancola <i>et al.</i> 2017	VIP	Submucosal neurons	 of submucosal neurons containing VIP due to α-syn overexpression 				
Yelkenli <i>et al.</i> 2016	VIP	CSF		- GAD astrocytes	+ motor activity		

Yu et al. 2020	VIP and	Striatum		+ oxidative stress	+ suspension test		
	PACAP			- inflammation	score		
				- neuronal death			
Olson et al. 2015	VIP	Substantia niagra		- inflammation			
				+ surviving dopaminergic neurons			
				- microgliosis			
Tunçel et al.	VIP	Striatum		 oxidative stress 			
2012				- apoptosis			
				- NO levels			
Korkmaz et al. 2012	VIP	Striatum and SNpc		- neuronal death			
Mosley et al.	VIP	SN		- inflammation			
2019				- neuronal death			
				VIPR2 agonist protects from α -syn induced inflammation, degeneration			
Korkmaz <i>et al.</i> 2010	VIP	CSF		+ NGF			
Federoff, 2009	NURR 1 and VIP		 due to α-syn overexpression 	 expression of DAT due to NuIP 			
				 VIP mRNA in embryos with loss of Nurr1 function 			
Gao et al. 2017	Angiotensin	SN	+	+ apoptosis			
				+ oxidative stress			
Villar-Cheda et al. 2010	Angiotensin	SN striatum		+ oxidative stress in SN & striatum			
Rocha et al. 2016	Angiotensin	Plasma	-			+ cognitive dysfunction	
Villar-Cheda et	Angiotensin II		+	+ oxidative stress			
<i>u</i> . 2012				+ inflammation			
				+ neuronal death			
Rodriguez-	Angiotensin II			+ inflammation			
2012				+ neuronal death			

Kubota <i>et al.</i> 2020	Carnosine	Hypothalamus		 neuronal death inflammatory response ROS production 				
Bermúdez, Seroogy, and Genter, 2019	Carnosine	In vivo and cells			 progression of motor deficits 			
Bermúdez, Skelton, and Genter, 2018	Carnosine			 oxidative stress inflammation 				
Maasz <i>et al</i> . 2017	РАСАР	CNS of snail SN of rats		 DA loss alpha-syn and Lewy bodies (through PARK7) 	-			
Yu <i>et al.</i> 2020	PACAP	Striatum			+ oxidative stress - inflammation - neuronal death	+ suspension test score		
Shivers et al. 2014	PACAP	Striatum		- neuronal loss	- motor deficits			
Brown et al. 2014	PACAP	Culture in toxic THP-1 cells medium		 inflammation neuronal death 				
Jouve <i>et al.</i> 2010	Pronkephaline	Striatum	+ in PD, – after treatment					
Sun <i>et al.</i> 2011	Enkephalin			 – oxidative stress – neuronal loss 				
Vinuela <i>et al.</i> 2008	Enkephalin	Striatum	+ in PD, - after treatment					
Ljungdahl <i>et al.</i> 2011	Dynorphin	SN	+		+ dyskinesia			
Hanrieder <i>et al.</i> 2011	Dynorphin	Striatum	+					
Elson <i>et al.</i> 2018	Galanin	Pedunculopontine nucleus	+	+ neuronal loss				
Niewiarowska- Sendo <i>et al.</i> 2016	Bradykinin			+ apoptotic processes + ROS release + NO release				
Lazarova <i>et al.</i> 2018	Neurotensin	Striatum		+ DA levels	 – fall time 		+ time to enter dark roo	m
Umehara <i>et al.</i> 2018	Norepinephrine	Plasma					+ cognition	
Pain <i>et al.</i> 2019	Neuropeptide Y	Striatum and SN		- inflammation				
Decressac <i>et al.</i> 2012	Neuropeptide Y	SNpc, striatum		 cell death -nigral dopamine neurons 				
Escamilla-Sevilla et al. 2011	Neuropeptide Y	Plasma	+ over time					
Bahdoudi <i>et al.</i> 2018	Octadecane- uropeptide	Striatum and SN nigral tissues and ventral tegmental area		 neuronal degeneration inflammation apoptosis 				
Knudsen <i>et al.</i> 2017	Pancreatic Polypeptide	Plasma	- in PD over time					
Slominsky <i>et al.</i> 2017	Semax	SNpc	+		No effect		+ anxiety	
Shiraishi <i>et al.</i> 2010	Somatostatin	Plasma	+					

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213 **4.1.** Orexin

Orexin was discovered in 1998 by Sakurai *et al.*, gets its name from orexis, Greek word for

²¹⁵ "appetite" [28]. It's also known as hypocretin, due to its expression in the posterior hypothalamus

[29]. Orexin is neuropeptide mostly abundant in the neurons of the perifornical area of the lateral

217 hypothalamus (LH). Orexinergic neurons send excitatory projections to arousal promoting areas in

the brain stem such as the locus coeruleus (LC) and the dorsal raphe (DR) nuclei involved in
increasing wakefulness and vigilance state [30]. Alongside its important role in the circadian
rhythm, orexin is also involved in promoting feeding via stimulation of the arcuate nucleus (Arc)
and ventromedial hypothalamic nucleus (VMH), promoting energy expenditure via interaction with
the brown adipose tissue and involved in neuroendocrine responses via projections to the pituitary
gland. Indirectly, orexin activates the reward system by its projections to the ventral tegmental area

224 (VTA) [8],[31].

Orexin has two homologous molecular forms: orexin-A and orexin-B, both derived from the same precursor peptide (prepro-orexin) [32]. Orexin A and B exert their various functions by binding and activating two G-protein-coupled receptors (GPCRs) called Orexin receptor 1 (OX1R) and Orexin receptor 2 (OX2R). OX1R is selective to orexin-A, binds to it with great affinity, however, OXR2 is selective and binds to both orexins [33].

230 Recent studies are showing the involvement of the orexin/receptor system in important

physiological processes of the brain such as sleep-wake cycle, as well as in pathophysiological

processes of several neurological disorders [33]. Orexin or hypocretin deficiency is the main cause

of human narcolepsy [34]. It has also been found reduced in neuromuscular disorders like myotonic

dystrophy and in immune-mediated diseases like Multiple Sclerosis (MS). Most interestingly,

orexin has been involved in several neurodegenerative disorders such as Alzheimer's disease and

PD, which is of great interest to this review [35].

237 Significant differences in the levels of orexin-A have been found in the CSF of late-stage PD

patients. Both CSF and plasma studies in PD showed a decreasing trend in orexin levels with the

increasing severity of the PD, as measured by the Unified Parkinson's Disease Rating Scale Part III

240 (UPDRS III) [36,37]. Drourot *et al.* also found further decrease in CSF orexins levels in a

longitudinal study for 4 and 5 years in 2 PD patients [37]. This negative correlation, shows the

progressive neurodegeneration of orexinergic neurons over time in PD patients [38]. However,

another study reported unchanged levels of orexin in CSF [39]. The decreased CSF levels of

orexin/hypocretin don't necessarily reflect direct correlation with the loss of orexin cells in the

245 hypothalamus, as some orexinergic neurons might compensate for the neuronal loss by increasing

the levels of orexin in the CSF [40]. Additionally, in PD the BBB becomes more permeable,

allowing leakage of orexin/hypocretin in the blood [36]. As lumbar CSF and Plasma levels don't

reflect accurately the orexinergic loss in the LH in PD's brain, other tools could be used to assessthe levels of orexin in PD and its reflection on degeneration.

Orexin plays a major role in the sleep/wake cycle. It has been shown to contribute to excessive day 250 time sleep (EDS) in advanced PD [37],[41]. Dopaminergic neuronal loss in advanced PD did not 251 252 correlate with EDS, and early-stage PD didn't majorly affect sleep. Interestingly, in an 253 electrophysiologically controlled prospective study, sleep measures of nocturnal sleep, deep sleep, sleep architecture and efficiency, as well as EDS are all most significantly disrupted in advanced 254 255 PD, where the orexin/hypocretin neuronal loss is most abundant [41]. Therefore, orexin 256 neurodegeneration promotes sleep in advanced PD. PD treatment with dopamine agonist 257 replacement therapy in 4 PD patients with sleep attacks showed decreased orexin levels in correlation with increased sleep attacks, suggesting the implication of orexin in reward circuit [42]. 258 259 On the contrary, Bridoux *et al.* found an increase in CSF orexin-A levels that correlated positively with epochs of rapid eye movement (REM) sleep without atonia, in patients during deep brain 260 261 stimulation who underwent full night polysomnography pre surgery. In fact, orexin is therefore involved in the loss of REM atonia, implicating the involvement of orexin in REM behavioral 262 263 disorder. Moreover, this study had a small sample size of 8 patients, where only 4 showed the significant correlation. It also lacked a control group, alongside the limitations electromyography 264 265 technique imposes [43].

266 An increasing body of evidence is recently studying the relationship between orexin and motor 267 activity in PD. The administration of orexin in the intraventricular CSF and globus pallidus (GP) in 6-OHDA mice models of PD has shown its promising effects in improving locomotor activity and 268 muscles weakness [44,45]. Hanging and rotarod tests following the administration of orexin, 269 270 showed an increased time on rod and latency to fall [44]. Also, elevated body swing test post orexin 271 administration resulted in rotation in the direction contralateral to the lesioned side [45]. On the 272 other hand, orexin increased the firing rate of the GP neurons via ORX1 and ORX2 receptors in both ODHA and MPTP mice models of PD [45,46]. Orexin-A and B increased significantly the 273 firing rate as recorded by the *in vivo* single unit electrophysiological recording, probably via their 274 action on the Ca^{2+} influx through the L-type Ca^{2+} channel [46]. Xue *et al.* showed that orexin-A 275 increases the firing rate of the GP via its action on the OX1R and OX2R [45]. Therefore, orexin 276 achieves its therapeutic effects on motor symptoms via the activation of the GP, leading to the 277

278 inhibition the STN activity and therefore disinhibiting the thalamus, allowing the motor signal to pass from the motor cortex to the muscles, improving the posture and motor behaviors [46]. 279 280 However, the activation of the external and the internal GP, although both GABAergic neurons, 281 have opposing effects on motor activity. The external one is implicated in the activation of motor neurons by inhibition of the STN. The internal GP inhibits the thalamus and therefore the voluntary 282 movement [47]. Other studied reported the increased firing rate of the internal GP neurons and the 283 STN in health rats [48,49]. The present finding proves the implication of orexin activation in 284 pathophysiology of PD. However, its precise action on both the external and internal GP and STN 285 need to be further studied. 286

287 Orexin alleviate motor deficits by its projections from the LH to other motor structures like the 288 cerebellum and vestibular nucleus [50]. Inhibition of the orexinergic neurons, using designer 289 receptors exclusively activated by designer drugs (DREADD) approach, has shown decreased the locomotor functions, energy expenditures and body composition as shown by the open field test, 290 291 calorimetry assessment of energy expenditures and spontaneous physical activity in A53-T mice models of PD [51,52]. In fact, orexinergic neurons recruit glutamic acid decarboxylase 65kD 292 293 (GAD65) cells in the LH implicated in voluntary motor control. Hence, its inhibition, reduces the overactivation of GAD64 and the locomotor activity as quantified using treadmill rotarod testing 294 295 [53].

296 It was also displayed, that orexin activation via DREADDS improves hippocampal memory 297 impairment and social memory [54,55]. Activation of orexin by using DREADDS or by injecting it in the hippocampus, ameliorated the hippocampal dependent memory impairment as seen by the 298 299 increased the latency to escape the Barnes maze and increased the total exploration time in the contextual object recognition test in 3, 5 and 7 months of A52T mice of models of PD [54]. The 300 301 administration of orexin/hypocretin significantly restored social memory in PD model, during the 302 automated two-enclosure homecage social test, that testes for sociability and social novelty (56). It's possible that orexin/hypocretin consolidates the social memory via its direct projection to the 303 304 hippocampus or indirectly thought the medial septum, thought enhancement of synaptic plasticity and the modulation of the cyclic adenosine monophosphate (cAMP) response element-binding 305 306 protein phosphorylation [54], [56]. Orexin is also a critical molecule in the execution of alertness and arousal. Stanojlovic et al. reported increased anxiety and social cognition after activation of 307

orexin neurons in the LH and the medial prefrontal cortex (mPFC) reciprocally [36],[52],[55]. The
LH, the hippocampus and the mPFC are all part of the reward system in the brain. Orexin could
have restored anxiety and social cognition by modulating the firing rate in any of these structures
[52].

312 4.2. Opioids

313 Endogenous opioids peptides are a class of molecules that are synthesized in the brain and can act locally or circulate for more widespread systemic action. They are termed opioids for their 314 315 presumed or confirmed binding to an opioid receptor in brain tissue [57]. Opioids peptides are a product of larger opioids precursors cleavage categorized into 3 families of molecules: enkephalin, 316 317 dynorphin and endorphin. Met-enkephalins and leu-enkephalins are end-products of preproenkephalin (PPE or PPE-A) cleavage the precursor belonging to the enkephalins family. 318 319 Similarly, prodynorphin (Pdyn or PPE-B) of the dynorphins family is cleaved to yield dynorphin-A, dynorphin-B and α or β neodynorphin. Lastly, pro-opiomelanocortin belonging to the endorphins 320 family is cleaved to yield melanocyte-stimulating hormone (MSH), adrenocorticotropic hormone 321

322 (ACTH) β -lipoproteins and β -endorphins [58,59].

323 After their secretion by a neuron, opioids peptides undergo enzymatic inactivation or transformation 324 to stop or regulate their action on opioid receptors. Most dynorphins are later cleaved to Leu-enkarg which is the end product of dynorphins transformation while the removal of tyrosine of 325 326 dynorphins peptides usually deactivates their action on opioids receptors [59]. Classical opioids peptides receptors are a family of three receptors termed Kappa opioids receptor (KOPR), mu 327 opioids receptors (MOPR) and delta opioids receptors (DOPR)[60]. Opioids have different affinities 328 329 to opioids receptors depending on their type: dynorphin-B has a higher affinity to KOPR and α -330 neodynorphin has no preferred affinity. Besides, opioid dynamics play a major role in receptor 331 affinity as such, dynorphin-B, has higher KOPR affinity but leu-enk-arg has higher DOPR affinity 332 [59]. Understanding opioids' nature and dynamics are very important to unravel their role and action 333 in the healthy and the pathological brain.

Opioids in general are extremely important in brain activity and homeostasis through their action as neurotransmitters and neuromodulators to regulate different behavior and functions such as pain, memory, addiction, and many more. Consequently, a disruption in this system will likely cause many physiological responses and malfunctions [61]. Disruption in opioid functions is non-evitable

in PD as opioid modulators are involved in the basal ganglia circuitry the center of PD

339 pathophysiology: dynorphin is involved in the direct pathway while enkephalin is specialized in the

indirect pathway making changes in their expression a likely outcome in PD [62]. As such, early

341 studies regarding opioids in PD revealed an increase of enkephalin expression in the striatum with

an increase in the GP in MPTP mice and marmosets reinforcing opioids evolvement in PD

343 pathophysiology [63]. Nowadays, there's an increasing line of evidence suggesting the involvement

of opioids in L-dopa induced dyskinesia (LID) an involuntary movement of the face, limbs, or trunk
caused by L-dopa administration [64].

Imaging mass spectrometry revealed dynorphin-B, α -neodynorphin leu-enk-arg (resulting from dynorphin-B degradation and non-other) were increased in SN of highly dyskinetic 6-OHDA rats in comparison to their no or low dyskinetic counterpart. The degree of dyskinesia was assessed by blind observation of limb dyskinesia, mouth and tongue movement, axial dystonia, and rotational behavior. This increase in peptide levels was correlated with higher dyskinetic scores [59]. In a similar study assessing opioids levels in the striatum of unilaterally 6-OHDA lesioned rats, dynorphin-B and α -neodynorphin were also increased showing similar correlative trends with

353 dyskinetic scores [65].

Multipliexing ion masss spectrometry images of dynorphin-B and leu-enk-arg in the SN of higly diskenetic mice revealed that the latter can only results of biodegeredation of dynorphin-B. This enzymatic conversion of dynorphin-B to Leu-enk-arg is of great interest, as it shifts opioids receptor affinity from KOPR to DOTR/MOTR in the SN. This receptor affinity switch could have a major implication in dyskinesia pathology in PD and it's fortified by the concomitant increase in α neodynorphin that can bind all receptors [59]. Indeed nalbuphine, a KOPR agonist and MOPR antagonist, was found to be a good anti-LID agent in the primate model of PD [66].

361 Dyskinesia can be explained as long-term potentiation (LTP) of striatal N-methyl-D-aspartate
362 (NMDA) receptors after L-dopa administration. Therefore, there is a link between over-excitation
363 of the striatum by L-dopa with NMDA receptor and DA receptors. This LTP does not depotentiate
364 causing the motor complications of LID [67]. Interestingly, Des-dynorphin can bind to NMDA
365 receptors possibly exhibiting an inhibitory effect on the striatum. As such, the Des-tyrosin-

dynorphin increase in the striatum could be implicated in a compensatory mechanism to alleviate
 striatal overstimulation of D1 receptor by unregulated L-dopa activity of L-dopa drugs [65].

Lucia Frau et al. found that the first movement after denervation of 6-OHDA rats induced by 368 dopamine agonism could be responsible for dyskinetic features and symptoms in these PD rat 369 370 models. Consequently, prohibiting rats from executing movement after apomorphine stimulation by 371 placing them in a restrainer significantly reduced dyskinesia vs rats placed in hemispherical bowls where the movement was possible as measured by contralateral movement behavior. Using *in situ* 372 373 hybridization, mRNA levels of the transcription factor zif-268 were differentially increased in 374 dynorphin + striatal cells of freely moving mice after stimulation with SKF 38393, a D1 agonist, 375 hinting at a change in molecular gene expression of dynorphin neurons following L-dopa agonism in this "priming model" of PD dyskinesia [68]. 376

377 Following dynorphin's trends enkephalins peptides were increased in the striatum of 6-OHDA dyskinetic rats in L-dopa treated side vs saline treated side as measured by Nilsson et al. using 378 quantitative mass spectrometry [69]. Besides, in a study done by Jouve et al. 2010, in situ 379 hybridization revealed that PPE mRNA levels were elevated in the striatum of 6-OHDA rat escorted 380 by worsening of sensiomotor and motor functions measured by corridor task and cylinder test 381 382 respectively as well as increased forelimb dyskinesia. Consequently, sub-chronic high-frequency stimulation of the thalamic centromedian-parafasicular complex was able to reduce the PPE mRNA 383 384 level elevation, thus normalizing motor dysfunctions and partially reducing dyskinesia [70]. 385 Paralleled, PPE RNA leveled normalization was found in an attempt of grafting ventral mesencephalic cells from wild-type or DAT kappa opioids (KO) in 6-OHDA mice model with 386 387 dyskinesia assessed by blinded observation of abnormal involuntary movements. Wild-type transplantation was found to be better at alleviating dyskinesia and showed less diffuse extra and 388 389 intracellular DA levels assessed with high-pressure liquid chromatography [67]. Taken together, 390 these findings reinforce enkephalin implication in PD and overall opioids role in LID with a close association with L-dopa regulation and administration. An interesting study on restless leg 391 392 syndrome found that delta-opioid agonist D-Ala2, D-Leu5-enkephalin (DADLE) was able to protect DA neurons in vitro against iron deficiency which is related to ROS formation and DA neuron 393 394 degeneration. Although, opioid implication in PD is complex making the need for in vivo testing to assess DADLE's significance in PD [70,71]. 395

17

Lastly, we should mention the possible implication of a rather newly discovered opioid peptide called orpahin FQ or nociceptin (OFQ/N). OFQ/N is a rat neuropeptide involved in the indirect striatum/thalamus pathway. When activated in PD, it increases the secretion of OFQ from the subthalamic nucleus's glutamatergic terminal nerves, or by the reduction GABAergic activation from the SN [79]. Hence, the increased OFQ expression in the SNC pars reticulata, SNpc and the subthalamic nucleus, as well as in the CSF of PD patients [80].

402 **4.3.** Vasoactive Intestinal Neuropeptide

403 VIP is neuropeptide, made of 28 amino acids, secreted in the central and peripheral nervous system [73]. As its name suggests, VIP was initially discovered in the intestine of the hog [74]. It also plays 404 405 an important role as an immunoregulatory neuropeptide, produced by lymphocytes T and B cells, also by mast cells and eosinophils stimulated by cytokines like interleukin (IL)-6, and IL-1 β [75]. 406 407 VIP has numerous and diverse biological functions such as respiratory stimulation, hypotension, vasodilation and hyperglycemia [74]. It's involved in the digestive system as a regulator of smooth 408 muscle contraction and epithelial secretion and blood flow. VIP downregulation is associated with 409 chronic constipation in PD patients [76]. Due to its various functions, VIP is produced in the 410 central and peripheral system by neurons, in the endocrine system by glands and in the immune 411 system by immune cells and nodes, as well as in the heart, the lungs, the kidneys, the thyroid 412 glands, the gastrointestinal (GI) tracts [73]. VIP is a member of the secretin (a peptide hormone 413 composed of 27 amino acids) receptor family with related structure that includes growth hormone 414 releasing hormone (GHRH), glucan, glucagon like peptides and PACAP. Due to their homologous 415 416 amino acids sequences, both VIP and PACAP share 3 GPCR: PAC1-receptor (PAC1-R), the 417 VPAC1-receptor (VPAC1-R) and VPAC2-receptor (VPAC2-R) [77].

The hallmark of PD must be the neurodegeneration of dopaminergic neurons in the SNpc, that disrupts the direct and indirect pathway from the striatum to the motor cortex, inducing impaired motor skills [47]. VIP has proved its neuroprotective function in PD, by increasing neuronal cells viability, decreasing dopaminergic neuronal loss and promoting the activity of DAT [77,78]. In 6-OHDA parkinsonian rats, VIP was capable of protecting not only the neurons but also the myelin sheath and reverse the motor defects [80]. Tunçel *et al.* reported direct evidence on VIP antiapoptotic and anti-oxidant properties, as shown by the decreased DNA fragmentation and lipid

425 peroxidation respectively. It also regulated the levels of the first line antioxidant defense system:

426 superoxide dismutase (SOD) and catalase (CAT) that were found disrupted in PD mice model [78].

427 Golgi-Cox staining and stereological methods were used, respectively, to determine the

428 morphological densitretic changes occurring in the neurons of the striatum and their number in the

429 SNpc in 6-OHDA PD rat model. The intra peritoneal administration of VIP showed its effects on

430 restoring the density of the striatal dendritic spines and the number of dopaminergic neurons

431 reduction in PD [79, 80].

VIP once released in the body, can cross the BBB into the CNS. However, due its short half-life, its 432 433 rapidly metabolized and cleared out. Therefore, limiting its potential use as therapeutic candidate 434 for PD [80, 81]. Hence why several studies are tagging TAT, a cationic Arginine-rich peptide 435 (CARP), to VIP in hopes of increasing its potency and traversing capabilities. Only VIP-TAT enhanced the motor deficits, increased the levels of DA and its metabolites (levodopa (L-dopa) and 436 437 homovanilic acid (HVAC)) and SOD levels in MPTP mice model of PD when compared to VIP, using the pole-climbing and Paws suspension test, the liquid chromatography mass spectrometry 438 439 analysis system, and the antioxidative activity SOD evaluation kit in the striatum reciprocally. The improved neuroprotective effects are mostly caused by the enhanced potency of VIP-TAT and 440 441 affinity to activate PAC1-R. The electrostatic interaction between the catatonic TAT domain and the anionic PAM (positive allosteric modulation) domain of the extracellular domain of PAC1-R 442 443 (PAC1-EC1) induces conformational changes in PAC1-EC1 that results in the interaction between the PAC1-R and VIP-TAT necessary to its activation [79]. The activation of PAC1-R contributes to 444 the activation of the CARP mechanisms and the extracellular signal-regulated kinases (ERK) 445 pathway, that promote neuroprotection against PD pathophysiology [78],[82]. 446

447 In another approach to use VIP as a drug to target PD, VIP-receptor 1 and VIP-receptor 2

448 antagonists, LB 3393 and LBT-3627, neuroprotective properties have been investigated. Treatment

449 with LBT-3627 gives rise to a link between the neuroprotection and the adaptive immune response

450 [82]. This VIP-receptor 2 antagonist is an effective modulator of PD pathophysiology, as it

451 decreased in a dose dependent manner the dopaminergic loss, reduced the reactive microglia's

452 levels and decreased the expression of genes associated with pro-inflammation and oxidative stress

453 in CD11+ cells (microglia). Interestingly, in a MPTP models of PD, LBT-3627 treatment elicits

454 immunosuppressive effects on adaptive immunity where it increased inhibition of responder T cells

455 by regulatory T cells, improved the activity of Tregs, as well as inhibited the production of

456 proinflammatory cytokines by helper T cells. Therefore, LBT-3627 modulation of the adaptive

- 457 response association with inflammation by creating a shift from pro-inflammation towards anti-
- 458 inflammation and ultimately neuroprotection. A cross-talk between the CD4+ T cells and the CD11
- 459 microglia cells could have taken place in creating the link between immuno-suppression and
- 460 neuroprotection [81],[83].
- 461 Moreover, the motor deficits in PD have been linked the GABA neurotransmitter. VIP recovered
- the motor deficits by increasing GABA levels in the ventral anterior nucleus of the thalamus
- 463 (VATH), as well as in the neurons of the striatum without modulating the dopamine turnover
- 464 [84,85]. In 6-OHDA brain slices of the VATH from rat model of PD, immunohistological staining
- showed that the administration of VIP at 25 ng/KG is capable of restoring the levels of GABA and
- the number of nerve growth factor (NGF) and the rat mast cell protease II (RMCPII) -
- immunoreactive mast cells in the thalamus of PD models [84].
- 468 Interesting, Federoff *et al.* describes PD as a developmental disorder. NuIP, the Nurr1 (nuclear
- receptor 1) interacting protein, a modulator of DA phenotype has a crucial role in the survival of
- 470 DA neurons during the brain development. It stimulated the transcription of Nurr1 which in turn
- induces the expression of DA proteins (DAT and TH) and enhances VIP production. VIP in turn
- 472 activates neurotrophic factors necessary for DA survival and microglia as immune-suppressor in the
- 473 CNS. Therefore, insuring DA survival and function [87]. However, environmental modulators and
- 474 increase in α-synuclein the CNS might downregulate NuIP during development resulting in loss of
- 475 DA neurons and PD pathology [86, 87].

476 **4.4.** Angiotensin

Two independent renin-angiotensin-systems (RAS) exist in the human organism: the circulating 477 478 cardiovascular hormonal system and the brain RAS [89]. Hormonal RAS regulates the blood pressure, the sodium homeostasis and the volume of body fluids, by enhancing the reabsorption of 479 480 water and sodium. Renin is secreted in the afferent arteria of the kidney. Once released, renin cleaves angiotensinogen, found circulating in the plasma and secreted by the liver, into the inactive 481 precursor angiotensin I. Angiotensin-converting enzyme (ACE) converts the latter into the leading 482 effector of the RAS: angiotensin II (AII) [90]. In the brain, AII is mostly secreted by glial cells (i.e. 483 microglia, astrocytes, and oligodendrocytes) in regions responsible of cardiovascular functions and 484

other brain regions (hypothalamus, etc...) where it plays an important role in memory, anxiety,
bipolar disorder and especially in PD [88],[90].

487 Angiotensin receptors type 1 and type 2, AT1R and AT2R, are located in different regions in the brain and along the spinal cord [91]. AT1R is expressed in the CNS by astrocytes, microglia and 488 489 even by neurons in particularly DA neurons. AT1 isn't only expressed in the membrane of the cells but also intracellularly in the nuclei membrane, inside the nuclei and in the membranes of the 490 endoplasmic reticulum [92]. The AII/AT1R axis induce a pro-inflammatory and pro-oxidative states 491 492 by production of ROS and activation of nicotinamide adenine dinucleotide phosphate (NADPH) 493 complex. Its overactivation induces harmful effects upon the brain: neurotoxicity, 494 neuroinflammation, as well as facilitated release of DA in the CNS. Interestingly, it exerts inhibitory effects on GABA and excitatory effects on glutamate, therefore decreasing GABA and 495 496 increasing glutamate release, which suggest an important role of AT1R activation in the indirect pathway of the SN, especially in motor deficits in PD. As opposed to AT1R functions, AII/AT2R 497 498 activation yields neuroprotective effects in the brain. AT2R activates nitric oxid (NO) production, enhances neurite outgrowth and improves memory learning and cognition [93]. Decreased levels of 499 500 AII, measured using the ELISA test, is associated with increased severity of depression symptoms 501 in thirty PD patients [94]. Hence why the study of RAS in PD is of major interest.

The AII/AT1R axis triggers dopaminergic neuronal loss, especially in the SN as seen by the 502 503 reduction in 70% of tyrosine hydroxylase neurons due to AII administration and restored with 504 candesartan (AII inhibitor), in rotenone, MPTP and 6-OHDA PD mice models [94-96]. A 505 multifactorial mechanism seems to be at the base of the pro-apoptotic activity of AT1R in dopaminergic cells: via NADPH mediated oxidative stress or via mitochondrial apoptotic signaling 506 507 [95]. In different PD models, in the DA neurons of the SN, AII enhanced the expression mRNA and 508 protein AT1 receptor and induced its overactivation. AT1R activates mitogen-activated protein 509 kinase (MAPK) cascade initiated by the protein kinase C (PKC), the critical actor of AT1R effects. PKC binds to and catalysis p47phbox, therefore activating NADPH complex, the main source of 510 511 ROS production in the cell [98]. In a more direct evidence, especially the activation of the intracellular NADPH-oxidase 4 (NOX4), induces the activation of the superoxide cascade, the 512 513 overexpression of oxidized guanosine 8-OH guanosine, and activation of caspase-3, giving rise to DNA damage in DA neurons of PD model, and resulting in apoptosis of DA neurons [91],[94]. 514

515 AII also stimulates ROS generation by activation of the intracellular mitochondria. A cross talk

- 516 between cytosolic NADPH complex and mitochondrial ATP-sensitive potassium channels
- 517 (mito(KATP)) exists where NADPH activates mito(KATP) oxidase production and the latter
- 518 modulates the extramitochondrial oxidative stress. In DA neurons of PD mice models, the inhibition
- of mito(KATP) inhibits the DA neuronal loss, blocks the superoxide induced ROS and reduces the
- 520 inner mitochondrial membrane potential induced by AII administration. All of which confirms the
- enhancing role of mito(KATP) on AII induced dopaminergic neurodegeneration [97].

522 Moreover, AT2R exerts its neuroprotective effects by decreasing the NADPH oxidative stress

activation, therefore restoring the DA neurons in the SN [95],[97]. The opposing relationship

between A1TR and A2TR has been extensively studied and shows a counterbalance between the

action of the two receptors. AT1R induction of ROS is counteracted by activation of AT2R that

526 decreases NADPH expression. Therefore, the unbalance between the activation of the two receptors

- 527 towards an overly activate AT1R pathway may increase the risk of PD via enhanced oxidative
- stress production and increased DA neurodegeneration [97,98].

529 DA depletion exerts a counter regulatory effect on AT1R expression. In 6-OHDA mice model of
530 PD, depleted DA neurons show increased AT1R expression [95-98]. However, in L-dopa presence

its expression is reduced [98]. Adversely, in aging rats AT1R expression was found upregulated and

- 532 AT2R expression is downregulated, which may lead to a pro-oxidative and pro-inflammatory state
- that increases dopaminergic neuron's vulnerability [99]. Hence, why it's important to question the

following: does the neurodegeneration of DA neurons in PD lead to the dysfunction in AII/AT1R

axis, in a manner where the AII/AT1R/NOX4 seeks to protect DA neurons and promote neuronal

regeneration and resilience, but when the neuronal stress is extended over time the same mechanism

537 induces oxidative stress and apoptosis [92]. Or, does the increase in AII/AT1R axis upregulation,

due to normal aging, increase DA neuron's vulnerability and impose a risk factor for PD?

539 Labandeira-Garcia *et al.* suggested a double mechanism of action of AII on DA neurons and on

540 microglia. In DA neurons, AII activates AT1R that induces NADPH NOX activation and produces

- 541 low levels of ROS in the nuclei. In microglia cells, AII induces the release of high levels of ROS
- and superoxide-activated ROS, that damages of DA neurons. Microglial polarization is the shift
- from the M1, proinflammatory and death inducer, phenotype to the M2, immunoregulatory and
- regenerative, phenotype in an inflammatory response [100]. It has been reported that microglia play

an important role in enhancing the effects of AII on ROS production in PD models, and inducing

neurodegeneration [97]. As well as, interacting with the neuroprotective estrogen in the SN, that

regulates AT1R post-transcriptional expression to reduce DA neuronal loss [96]. This suggests that

the proinflammatory M1 microglia is activated by the AII/AT1R axis and the M2

549 immunoregulatory microglia is activated by the AII/AT2R axis.

550 *4.5. Carnosine*

551 Carnosine or (β -alanyl-L-histidine) is a dipeptide found in vertebrate's skeletalmuscles and brains 552 making animal dietary consumption of fish and meat one of its best sources [101]. Taken as a supplement for exercise with the aim of better physical performance, many other roles have been 553 554 attributed to this peptide since its discovery in Russia in 1990. Indeed, this dipeptide is proven to have anti-oxidant and antiglycation properties as well as having a role in Cu and Zn chelation and 555 556 wound healing promotion [100, 101]. Carnosine, as well as its precursors but to a lesser extent, is 557 capable of crossing the blood-brain barrier making the use of exogenous carnosine in 558 neuroprotection more applicable. Carnosine also exist in endogenous form. It is synthesized by muscle cells, oligodendrocytes and olfactory neurons and latter metabolized by carnosinase in the 559 560 kidneys, liver, and serum [103].

Interest regarding carnosine neuroprotective effect is on the rise, carnosine additive administration 561 562 has been found to alleviate disease-related symptoms better than classic therapy in PD patients in a 563 controlled clinical trial done by Boldyrev et al. 2008 [104] as measured my UPDRS rating scale. 564 Also, this dipeptide caused less behavioral and motor dysfunctions than DA alone in rat model when administered as a pharmacological compound with DA. These rats showed less locomotion 565 566 depression and more entries and time spent in open zones [105]. Similarly, when carnosine was 567 introduced intranasally in th1-asyn mice models, the latter showed less motor deficit compared to 568 no treatment group as assessed using the challenging beam traversal test [106].

569 Carnosine can exhibit its protective effect by inhibiting ROS formation [103], [105, 106] through an

570 enhancement of glutathione peroxidase (GPX) in a-syn mice model measured by RTPCR [108] and

571 through SOD in MPTP and 6-OHDA rat models [104], [108] as revealed using superoxide

dismutase (SOD) measuring assay. SOD and GPX are two enzymes involved in cellular oxidative

573 stress resistance, thus an increase in SOD and GPX activity reinforces carnosine's role as an anti-

574 oxidant. Although this stress-related response seems to have a threshold for its activity as carnosine
575 alone [105, 106] or in presence of low oxidative states [105] doesn't show this same effect.
576 Consequently, western blot analysis showed that similar ROS inhibition by carnosine pre-treatment
577 led to a down-regulation of stress-activated protein kinases (SAPK)/ Jun amino-terminal kinases

578 (JNK) pathway of the MAPK family involved majorly in cell death and inflammation in 6-OHDA

immortalized hypothalamic neuronal GT1-7 cells. In this case, carnosine successfully suppressed 6-

580 OHDA induced upregulation of JNK. In the same cell line, carnosine also showed inhibition of

581 integrated stress response genes such *Chop*, *Gadd34*, and *Atf4* further enhancing cell survivability

as revealed by RT-PCR [107]. These 2 pathways, among others, could work hand in hand to exhibit

neuroprotective effects by slowing down disease progression and onset through reduction of cellular

584 oxidative state.

585 Carnosine plays a major role in protecting against neural inflammation. It has been found to reduce

the activation of many pro-inflammatory cytokines induced by different PD models, such as IL-

587 1β [106-108], IL-6 [107], [108] and MCP-1 [108]. As well as, tumor necrosis factor-alpha

588 (*Tnfa*), cyclooxygenase 2 (*Cox2*), and toll-like receptor 4 (*Tlr4*) [107] as revealed using

589 ELISA[106], [108] or RT-PCR [107] methods. Interestingly, carnosine administration has an

inhibitory effect on INOS and NOS in MPTP mice two pro-inflammatory enzymes known to

promote inflammatory interleukins production under *Tnfa* activation [108]. Using in silico method

of upstream regulator analysis Bermudez *et al.* 2008 predicted and normalized Trem1 (trigerring

receptor expressed on myeloid cells 1) activation in a-syn mouse models when treated with

carnosine in drinkable water to be similar level of healthy controls [109]. Trem1 is an activator of

tumour necrosis factor alpha (TNF- α). Therefore Trem-1 activation of TNF- α leads to increased

596 INOS and NOS enzymes activity through NF-kappa-B pathway and thus increasing anti-

inflammatory cytokines [106],[108]. It would be interesting to further test carnosine's effect on this

pathway to better understand the mode of action of carnosine in neural inflammation. Lastly,

599 carnosine's effect is not limited to neural inflammation and ROS control as it was able to re-

600 increase the expression of the mitochondria genes complexes I, IV and V after it was attenuated by

 α -synuclein overexpression in mice, therefore, increasing mitochondrial function and respiration

602 [109].

603

4.6. Pituitary adenylate cyclase-activating polypeptide

606 PACAP is a 27- or 38-amino acid neuropeptide, belonging to the intestinal polypeptide/glucagon/secretin family [110]. This peptide exists in two forms termed and PACAP27 607 608 (27 amino acids) and PACAP28 (28 amino acids) which is an N-terminal cleaved form of the latter. 609 This peptide was discovered in 1989 in the ovine hypothalamus but since then, PACAP was also 610 found in other brain regions such as the amygdala, the septum, the GP, the midbrain, and many 611 others. PACAP can also be found in non-CNS tissues such as the digestive and respiratory systems [110, 111]. Studies regarding this peptide found that it has many roles like regulating insulin 612 613 secretion and immunity. It also exhibits an anti-apoptotic effect and can promote cell proliferation 614 [113]. Neural inflammation and apoptosis are well-documented pathophysiological hallmarks of PD 615 [12]. Taking this into consideration many studies were interested in unraveling its possible 616 implication in PD by protecting the dopaminergic cells from degeneration.

617 Indeed, PACAP seems to be effective at alleviating Parkinson-like symptoms and increasing

618 survivability in animals. As such, it was able to increase the survivability of the rotenone snail

model of PD concomitantly improving motor functions and feeding [114]. Similarly, PACAP

620 reverted parkinsonian-like symptoms induced by prostaglandin injection in mice, MPTP in mice,

and 6-OHDA in rats [113-115]. MPTP mice treated with PACAP showed higher pole climbing and

suspension scores thus better coordination and motor function than untreated counterparts [115]. As

to mice infused unilaterally with prostaglandins in the SN, they showed worse forelimb asymmetry

and postural stability measured with cylinder test and curling test respectively. Co-injection of

625 PACAP with prostaglandins normalized motor dysfunctions score to be comparable with healthy

626 controls [116].

604

627 High pressure liquid chromatography analysis of neurotransmitters levels of PD animal models

628 found that, although PACAP seems to be also capable of restoring DA striatal levels in 6-OHDA

629 mice all while rescuing dopaminergic degeneration it was unable to restore dopamine levels in

630 MTTP regardless of its positive effect on motor symptoms [115]. PACAP did not affect 5HT

restoration in the striatum of 6-OHDA mice and in the Retonon snail model. Lastly, PACAP failed

to inhibit microglial activation in PG-J2 (prostaglandin J2) mice. Co-administration of PACAP with

prostaglandins did not reduce the amount of IBA1+ cells in brain tissues of treated mice as assessed
by immunostaining [113],[115].

PACAP shares 68% structural similarities with VIP. Therefore, VIP and PACAP share two GPCR, 635 VPACR-1 and VPACR-2. PACAP can also specifically bind to its specific PAC-1 receptor found 636 637 distributed in brain tissues but also in the SN [111]. There's growing evidence suggesting that 638 PACAP exhibits its protective effect in PD through this receptor by inhibiting apoptosis. The downstream effect of PACAP activation of PAC-1 is the induction of adenyl cyclase and later 639 protein kinase A (PKA). PKA can have many effectors including the activation of cyclic AMP 640 response element binding protein (CREB) which promotes brain-derived neurotrophic factor 641 642 (BDNF) and inhibits caspase 3, therefore, promoting cell survival [113],[116]. Indeed, PACAP was successfully able to prevent neuroblastoma-derived cell line (SH-SY5Y) death expressing high 643 644 levels of DA when introduced in the toxic medium of stimulated pro-inflammatory THP-1 cells. Western blot analysis of the surviving cells showed low lower levels of caspase 3 and a higher level 645 646 of CREB and BDNF when co-administered in medium with PACAP but not when in presence of

647 PAC-1 antagonist [117].

This pathway also explains why PACAP couldn't deactivate microglial activation in PG-j2 mice.

649 PG-j2 acts through GPCR coupled with Gi protein to inhibit cAMP which can be countered by

650 PACAP through PAC-1. Although PG-j2 can also bind protein on the sulfhydryl ramification

651 causing chronic inflammation and proteins aggregation which may not be altered by using PACAP

[117]. Another pathway of PACAP's action can be through PARK7 (protein coding gene associated

with PD7) which was rescued by PACAP in 6-OHDA rats. PARK7 is a chaperon protein with anti-

oxidant and anti-apoptotic properties. PARK 7 can also stop apodosis by inhibiting the proapoptotic

protein-associated splicing factor (PSF). Besides, it could be involved in inhibiting ROS formation

by stabilizing NRF2 (nuclear factor erythroid 2 related factor 2) protein, a regulator of anti-

oxidative stress. PACAP's possible implication in oxidative stress is further supported by an

elevated SOD level in MTTP mice after its administration. Lastly, PARK7 can stop mutant α -

659 synuclein aggregation which is another important pathological hallmark of PD [113,114].

660 Briefly, PACAP seems to exhibit some neuroprotective effects through inhibition of apoptosis and

oxidative stress *in vivo* and in vitro but the extent of its protection differs depending on animal

models. More *in vivo* studies should explore the molecular aspect of such discrepancies beforestudying its application in humans.

664 **4.7.** Other neuropeptides

PD's most pathological characteristic is the presence of Lewy bodies, their fibrils are mostly made 665 of α -synuclein. Lewy bodies are widely distributed across the central and peripheral autonomic 666 667 nervous systems including the enteric system, cardiac plexuses, adrenal gland, etc... [118]. It's especially found in the dorsal motor nucleus of the vagus nerves in the medulla oblongata. The 668 669 secretion of both somatostatin and pancreatic polypeptide (PP) from the pancreas is controlled by 670 the vagus nerve, which is why they're used as markers for vagal nerve integrity [119]. Somatostatin 671 is known as a growth hormones inhibitor, and has various inhibitory effects on GI tract, endocrine 672 and exocrine systems, as well as modulates neurotransmission and formation of memory in the 673 CNS and most importantly has anti-proliferative effects [120]. On the other hand, PP is secreted by 674 islet and acinar cells in the pancreas. Its major role is to regulate the pancreatic secretion, as well as 675 modulate the contraction of the GI tract and gall bladder, hence its importance in regulating the digestive system that's disrupted in PD [121]. In fact, in the plasma of PD patients, somatostatin is 676 677 upregulated and PP is downregulated. These results show the contribution of somatostatin in PD pathophysiology in the digestive system and PP involvement in the disrupted function of the GI 678 tract found in PD [119], [122]. NPY has also been found increased in the plasma of PD patients 679 680 undergoing deep brain stimulation [123]. The neuroprotective role of NPY in PD has been widely studied in both 6-OHDA PD mice models and in vitro with neuroblastoma cells. NPY has been 681 682 shown to protect DA neurons and their terminals from cell loss, in both cells and mice PD models, in the SN and the striatum, via activation of their Y2 receptor specifically, elucidating its 683 neuroprotective effects. It also showed significant results in restoring the expression of DAT and 684 the concentration of DA and its metabolites. This neuroprotection might be mediated by the 685 activation of ERK ¹/₂ and protein kinase B (AKT) pathways, or by the inhibition of the 686 neuroinflammatory process elicited by the excessive activation of microglia. Blocking microglial 687 688 activation, reduces apoptosis of DA neurons therefore alleviates the behavioral and motor impairments found in PD [124]. Interestingly, Apelin, a novel neuropeptide, is showing its 689 regulating and neuroprotective effects on brain functions. In a 6-OHDA model of PD, motor 690 691 assessment tests like accelerating rotarod, balance beam and beam traversal tests and the bar tests

692 showed significant motor improvement when lesioned rats were treated with increasing concentrations of Apelin-13. Not only Apelin decreased the motor impairment in PD rat models but 693 694 also, increased the dopaminergic receptor's levels, attenuated the DA loss in the substantia nigra. 695 The destruction of the dopaminergic pathway in the SN can affect the functioning of the striatum and therefore lead to a loss of its synaptic plasticity, which is represented by the loss of the 696 697 membrane associated proteins such postsynaptic density protein 95 (PSD-95), neuroligin, neurexin1, metabotropic glutamate receptor (mGlu R1). The administration of Apelin-13 698 699 significantly increased the expression of these proteins in the PD lesioned model, therefore 700 protecting the synaptic plasticity in the striatum [124]. Octadecaneuropeptide (ODN) is a 701 neuropeptide secreted extensively by astrocytes in the CNS. It also exerts neuroprotective action on dopaminergic neurons and astrocytes through preventing astrogliosis, oxidative stress and 702 703 inflammation in the SNpc, the striatum and in the ventral tegmental area [125]. Emerging evidence is showing that the motor and non-motor symptoms occurring in PD are in fact, induced by 704 705 degeneration in the cholinergic neurons rather than in the dopaminergic neurons alone [126]. The 706 pedunculopontine nucleus (PPN), is extremely rich in cholinergic neurons and plays a major role in 707 the regulation of movement execution, maintenance of arousal and awareness in posture and balance [127]. 708

709 Galanin, is a neuropeptide distinguished from other peptides by its co-existence with neurotransmitters such as dopamine and acetylcholine. In a Lactacystin mice model of PD, galanin 710 711 was increased in cholinergic neurons in PPN, especially near the side of the toxin injection. This upregulation marks the pro-survival, anti-inflammatory and anti-oxidative stress action of galanin 712 713 towards the toxin. It might show the compensatory role of galanin, it compensates for the damage 714 induces by the neurovegetative toxin to maintain movement and arousal as long as possible. Further 715 studies need to demonstrate if the rise in galanin has significant change on motor impairment in PD and investigate its therapeutic properties [126]. Locomotor impairment has been reported reduced 716 717 by analogues GIP in chronic PD mice models [128]. GIP or glucose-dependent insulinotropic polypeptide, secretion is essential to stimulate insulin release. It also has anti-apoptotic effects and 718 induces growth [129]. Through the reduction of oxidative stress, astrogliosis, microglial activation, 719 inhibition of the α -synuclein expression as well as protection of dopamine synapses and 720 mitochondrial activity, GIP exerts its anti-parkinsonian action on chronic PD, along with the 721 722 increase in the BNDF expression to induce neuronal growth and fight neurodegeneration [128],

723 [130]. Several other neuropeptides have been found modified in PD patient's plasma, such as decreased VGF cleaved peptides and increased norepinephrine, and in 6-OHAD mice model of PD 724 725 where selanx was found increased as a marker of anxiety [129-131]. Moreover, neuropeptides can 726 also have neurodegenerative properties and induce parkinsonian motor and non-motor symptoms. A great example of toxicity enhancer is the kinin peptide: bradykinin. In a cellular model of PD, 727 bradykinin not only activates pro-apoptotic pathways but also activates inflammatory response 728 enhancing the release of cytokines by neurons: IL-1 β , IL-6 and TNF- α , as well as activates the 729 oxidative stress leading to casapase-3 activation. All of which inhibits cell survival, enhances DA 730 neurons loss. However, if the activation of bradykinin is a consequence or a cause to PD 731 pathophysiology is a matter that needs to be studied in further experiments [134]. The formation 732 and release of neuropeptide vesicles are also impaired due to the endoplasmic reticulum lipids that 733 734 create contact surfaces with mitochondrial membranes and induces morning anticipation deficits and increases awakening at night in drosophila model of PD. These results have been evolutionary 735 736 conserved from drosophila to hypothalamic neuron in PD patients [135]. Deep brain stimulation is a 737 therapeutic procedure used in PD to help alleviate the motor disorders patients experience. The 738 connection between the brain-gut axis has led to investigating the effects of deep brain stimulation (DBS) on the secretion and concentration levels of several neuropeptides such as ghrelin, especially 739 740 the acetylated ghrelin peptide produced in the stomach. Before the DBS in PD patients, a decrease in level of plasma acetyl ghrelin during fasting and postprandial. However, after the DBS procedure 741 742 the PD patients showed an increase in this concentration. These results suggest an impairment in the normal functioning and secretion of ghrelin in the stomach. These results are of high importance 743 because they shed a light on the less studied brain-gut axis in PD [135]. 744

745 5. Conclusion

This review shows indisputable implications of several neuropeptides in the neuroprotection or symptoms in PD. Orexin alleviates the motor impairment, regulates the sleep disorders and improves hypothalamic memory in PD pathology. AII activates the pro-inflammatory pathways via its AT1 receptor and increases DA neuronal cell death, but protects the latter via activation of its AR2 receptor. VIP also activates its anti-apoptotic and anti-oxidative stress activity to insure the protection for the DA neurons. Carnosine was found to be able to alleviate motor symptoms by protecting against oxidative stress neural inflammation and mitochondrial dysfunction in PD. 753 Opioids disruption in the basal ganglia and striatum is highly linked with LID but it's unclear 754 whether it's a compensatory mechanism or the cause of the symptom itself. Lastly PACAP seem to 755 exhibit a neuroprotective of effect on striatal neurons in vivo and in vitro through regulation of apoptosis and inflammation. The presented findings urge the need for a shift towards a more 756 advanced understanding of neuropeptides mechanism and involvement in clinical trials. Further 757 studies need to take in consideration the major limitations found in the reviewed studies such as the 758 sex difference between group subjects, the small sample size used in a study, and the lack of results 759 verification through the use of multiple techniques. The difference in animal models used in studies 760 sometimes yielded difference in the results even when same measurement techniques were applied 761 making a deeper understanding of the molecular aspect of each animal model very important for a 762 more unified assessment of Parkinson pathophysiology and symptoms. It's also important to take 763 into consideration that the basal ganglia have many subdivisions and roles, that's why a more 764 detailed and specific approach needs to be applied. This specific approach firstly, help to unravel a 765 more zoomed in effect of these peptides on the basal ganglia's more specific regions to help in 766 future development of a targeted therapeutic approach. And secondly, aid in the construction of a 767 768 more global view to understand the multi-level effect of these peptides on brain tissues.

769

770 6. Conflict of interest

771 None.

772 7. Acknowledgements

773 None.

774 8. Abreviations

775 CNS: central nervous system ; PD : Parkinson's disease; SN: substantia niagra; PACAP: pituitary adenylate cyclase

activating polypeptide ; VIP: vasoactive intestinal peptide ; SNpc: substantia niagra pars compacta; MSNs: striatal

777 medium spiny neurons; GABA: gamma-aminobutyric acid; STN: subthalamic nucleus; DA: dopamine; ROS: oxygen

reactive species; iNOS: inductible nitric oxide synthase; NOS: nitric oxide synthase; MPTP: 1-methyl-4-phenyl-1,2,3,6-

tetrahydropyridine; 6-OHDA: 6-hydroxydopamine; BBB: blood brain barrier; DAT: dopamine transporter; MFB:

780 medial forebrain bundle; UPS: ubiquitin proteasome system; ALP: autophagy-lysosomal pathways; CSF: cerebrospinal

fluid; NPY: neuropeptide Y; ODN: octadecaneuropeptide; VGF: nerve growth hormone; GIP: gastrointestinal peptide;

782 PP: pancreatic peptide; LH: lateral hypothalamus; LC: locus coeruleus; DR: dorsal raphe; Arc: arcuate neucleus; VMH: 783 ventromedial hypothalamic nucleus; VTA: ventral tegmental area; GPCRs : protein-coupled receptors; OX1R: orexin 784 receptor 1; OX2R: orexin receptor 2; MS: multiple sclerosis; UPDRS III: unified parkinson's disease rating scale part 785 III; EDS: excessive day time sleep; REM: rapid eye movement; GP: globus pallidus; DREADD: designer receptors 786 exclusively activated by designer drugs; GAD65: glutamic acid decarboxylase 65kD; cAMP: cyclic adenosine 787 monophosphate; mPFC: medial prefrontal cortex; PPE or PPE-A: preproenkephalin; Pdyn or PPE-B: prodynorphin; 788 MSH: melanocyte-stimulating hormone; ACTH: adrenocorticotropic hormone; KOPR: kappa opioids receptor; MOPR: 789 mu opioids receptors; DOPR: delta opioids receptors; LID: L-dopa induced dyskinesia; LTP: long-term potentiation; 790 NMDA: N-methyl-D-aspartate; KO: kappa opioids; DADLE: D-Ala2, D-Leu5-enkephalin; OFO/N: orpahin FQ or 791 nocicpetin; IL: interleukin; GHRH: growth hormone releasing hormone; PAC1-R: PAC1-receptor; VPAC1-R: VPAC1-792 receptor; VPAC2-R: VPAC2-receptor; SOD: superoxide dismutase; CAT: catalase; CARP : cationic Arginine-rich 793 peptide; L-dopa: levodopa; HVAC: Homovanillic acid; PAM: positive allosteric modulation; ERK: Extracellular 794 signal-regulated kinases; VATH: ventral anterior nucleus of the thalamus; NGF: nerve growth factor; RMCPII : rat 795 mast cell protease II; NuIP or Nurr1: nuclear receptor 1; RAS: renin-angiotensin-systems; ACE: angiotensin-converting 796 enzyme; AII: angiotensin II; AT1R: angiotensin receptors type 1; AT2R: angiotensin receptors type 2; NADPH: 797 nicotinamide adenine dinucleotide phosphate; NO: nitric oxid; MAPK: mitogen-activated protein kinase; PKC: protein 798 kinase C; NOX4: NADPH oxidase 4 ; mito(KATP): mitochondrial ATP-sensitive potassium channel; GPX: glutathione 799 peroxidase; SOD: superoxide dismutase; SAPK: stress-activated protein kinases; JNK: jun amino-terminal kinases; 800 Tnfa: tumor necrosis factor-alpha; Cox2: cyclooxygenase 2; Tlr4: toll-like receptor 4; Trem1: triggering receptor 801 expressed on myeloid cells; TNF- α : tumour necrosis factor alpha; PKA: protein kinase A; CREB: cAMP response 802 element binding protein; BDNF: brain-derived neurotrophic factor; PG-j2: prostaglandin j2; PARK7 : protein coding 803 gene associated with PD7; PSF: proapoptotic protein-associated splicing factor; PP: pancreactic polypeptide; 804 AKT: protein kinase B; PSD-95: postsynaptic density protein 95; mGluR1: metabotropic glutamate receptor;

805 ODN: octadecaneuropeptide; PPN: pedunculopontine nucleus.

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807 **References**

808 [1] S. D. Kim, N. E. Allen, C. G. Canning, and V. S. C. Fung, "Parkinson disease," in *Handbook of Clinical Neurology*, vol. 159, Elsevier, 2018, pp. 173–193. doi: 10.1016/B978-0-444-63916-5.00011-2.

[2] A. Berardelli, J. C. Rothwell, P. D. Thompson, and M. Hallett, "Pathophysiology of bradykinesia in
Parkinson's disease," *Brain J. Neurol.*, vol. 124, no. Pt 11, pp. 2131–2146, Nov. 2001, doi:
10.1093/brain/124.11.2131.

[3] L. V. Kalia and A. E. Lang, "Parkinson's disease," *The Lancet*, vol. 386, no. 9996, pp. 896–912,
Aug. 2015, doi: 10.1016/S0140-6736(14)61393-3.

- K. M. Prakash, N. V. Nadkarni, W.-K. Lye, M.-H. Yong, and E.-K. Tan, "The impact of non-motor symptoms on the quality of life of Parkinson's disease patients: a longitudinal study," *Eur. J. Neurol.*, vol. 23, no. 5, pp. 854–860, May 2016, doi: 10.1111/ene.12950.
- 818 [5] S. Jain and D. S. Goldstein, "What ARE Parkinson disease? Non-motor features transform
- conception of the shaking palsy," *Neurobiol. Dis.*, vol. 46, no. 3, pp. 505–507, Jun. 2012, doi:
 10.1016/j.nbd.2012.04.013.
- W. Muangpaisan, A. Mathews, H. Hori, and D. Seidel, "A systematic review of the worldwide
 prevalence and incidence of Parkinson's disease," *J. Med. Assoc. Thail. Chotmaihet Thangphaet*, vol. 94, no.
 6, pp. 749–755, Jun. 2011.
- P. Calabresi, B. Picconi, A. Tozzi, V. Ghiglieri, and M. Di Filippo, "Direct and indirect pathways of
 basal ganglia: a critical reappraisal," *Nat. Neurosci.*, vol. 17, no. 8, pp. 1022–1030, Aug. 2014, doi:
 10.1038/nn.3743.
- M. Skrzypski, M. Billert, K. W. Nowak, and M. Z. Strowski, "The role of orexin in controlling the activity of the adipo-pancreatic axis," *J. Endocrinol.*, vol. 238, no. 2, pp. R95–R108, Aug. 2018, doi: 10.1530/JOE-18-0122.
- [9] L. Stefanis, "α-Synuclein in Parkinson's Disease," *Cold Spring Harb. Perspect. Med.*, vol. 2, no. 2,
 p. a009399, Feb. 2012, doi: 10.1101/cshperspect.a009399.
- E. Maries, B. Dass, T. J. Collier, J. H. Kordower, and K. Steece-Collier, "The role of α-synuclein in
 Parkinson's disease: insights from animal models," *Nat. Rev. Neurosci.*, vol. 4, no. 9, pp. 727–738, Sep.
 2003, doi: 10.1038/nrn1199.
- [11] L. Xu and J. Pu, "Alpha-Synuclein in Parkinson's Disease: From Pathogenetic Dysfunction to
 Potential Clinical Application," *Park. Dis.*, vol. 2016, p. e1720621, Aug. 2016, doi: 10.1155/2016/1720621.
- 837 [12] W. Poewe *et al.*, "Parkinson disease," *Nat. Rev. Dis. Primer*, vol. 3, p. 17013, Mar. 2017, doi: 10.1038/nrdp.2017.13.
- [13] L. Puspita, S. Y. Chung, and J.-W. Shim, "Oxidative stress and cellular pathologies in Parkinson's disease," *Mol. Brain*, vol. 10, no. 1, p. 53, Nov. 2017, doi: 10.1186/s13041-017-0340-9.
- 841 [14] "Oxidative Stress: Harms and Benefits for Human Health."
- 842 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5551541/ (accessed Aug. 27, 2021).
- [15] S. J.-P. Haas, X. Zhou, V. Machado, A. Wree, K. Krieglstein, and B. Spittau, "Expression of Tgfβ1 and Inflammatory Markers in the 6-hydroxydopamine Mouse Model of Parkinson's Disease," *Front. Mol. Neurosci.*, vol. 9, p. 7, Feb. 2016, doi: 10.3389/fnmol.2016.00007.
- J. R. Pereira *et al.*, "IL-6 serum levels are elevated in Parkinson's disease patients with fatigue
 compared to patients without fatigue," *J. Neurol. Sci.*, vol. 370, pp. 153–156, Nov. 2016, doi:
 10.1016/j.jns.2016.09.030.
- [17] Q. Wang, Y. Liu, and J. Zhou, "Neuroinflammation in Parkinson's disease and its potential as
 therapeutic target," *Transl. Neurodegener.*, vol. 4, no. 1, p. 19, Dec. 2015, doi: 10.1186/s40035-015-0042-0.
- [18] B. J. Ryan, S. Hoek, E. A. Fon, and R. Wade-Martins, "Mitochondrial dysfunction and mitophagy in
 Parkinson's: from familial to sporadic disease," *Trends Biochem. Sci.*, vol. 40, no. 4, pp. 200–210, Apr.
- 853 2015, doi: 10.1016/j.tibs.2015.02.003.

- 854 [19] M. Rango, G. Dossi, L. Squarcina, and C. Bonifati, "Brain mitochondrial impairment in early-onset
- Parkinson's disease with or without PINK1 mutation," *Mov. Disord. Off. J. Mov. Disord. Soc.*, vol. 35, no. 3,
 pp. 504–507, Mar. 2020, doi: 10.1002/mds.27946.
- A. Bose and M. F. Beal, "Mitochondrial dysfunction in Parkinson's disease," *J. Neurochem.*, vol.
 139 Suppl 1, pp. 216–231, Oct. 2016, doi: 10.1111/jnc.13731.
- [21] F. Blandini and M.-T. Armentero, "Animal models of Parkinson's disease," *FEBS J.*, vol. 279, no. 7,
 pp. 1156–1166, Apr. 2012, doi: 10.1111/j.1742-4658.2012.08491.x.
- V. Licker and P. R. Burkhard, "Proteomics as a new paradigm to tackle Parkinson's disease research
 challenges," *Transl. Proteomics*, vol. 4–5, pp. 1–17, Dec. 2014, doi: 10.1016/j.trprot.2014.08.001.
- 863 [23] C. J. Werner, R. Heyny-von Haussen, G. Mall, and S. Wolf, "Proteome analysis of human substantia
 864 nigra in Parkinson's disease," *Proteome Sci.*, vol. 6, p. 8, Feb. 2008, doi: 10.1186/1477-5956-6-8.
- Y. Zheng, L. Zhang, J. Xie, and L. Shi, "The Emerging Role of Neuropeptides in Parkinson's
 Disease," *Front. Aging Neurosci.*, vol. 13, 2021, doi: 10.3389/fnagi.2021.646726.
- 867 [25] J. Forbes and K. Krishnamurthy, "Biochemistry, Peptide," in *StatPearls*, Treasure Island (FL):
- 868 StatPearls Publishing, 2021. Accessed: Aug. 25, 2021. [Online]. Available:
- 869 http://www.ncbi.nlm.nih.gov/books/NBK562260/
- 870 [26] B. Jp, "What are neuropeptides?," *Methods Mol. Biol. Clifton NJ*, vol. 789, 2011, doi: 10.1007/978871 1-61779-310-3_1.
- 872 [27] A. F. Russo, "Overview of Neuropeptides: Awakening the Senses?," *Headache*, vol. 57 Suppl 2, pp.
 873 37–46, May 2017, doi: 10.1111/head.13084.
- [28] T. Sakurai *et al.*, "Orexins and orexin receptors: a family of hypothalamic neuropeptides and G
 protein-coupled receptors that regulate feeding behavior," *Cell*, vol. 92, no. 4, pp. 573–585, Feb. 1998, doi:
 10.1016/s0092-8674(00)80949-6.
- [29] L. de Lecea *et al.*, "The hypocretins: Hypothalamus-specific peptides with neuroexcitatory activity," *Proc. Natl. Acad. Sci. U. S. A.*, vol. 95, no. 1, pp. 322–327, Jan. 1998.
- [30] A. L. Gotter, A. L. Webber, P. J. Coleman, J. J. Renger, and C. J. Winrow, "International Union of
 Basic and Clinical Pharmacology. LXXXVI. Orexin receptor function, nomenclature and pharmacology," *Pharmacol. Rev.*, vol. 64, no. 3, pp. 389–420, Jul. 2012, doi: 10.1124/pr.111.005546.
- [31] T. E. Scammell, "Wakefulness: An eye-opening perspective on orexin neurons," *Curr. Biol.*, vol. 11, no. 19, pp. R769–R771, Oct. 2001, doi: 10.1016/S0960-9822(01)00466-3.
- [32] T. Voisin, P. Rouet-Benzineb, N. Reuter, and M. Laburthe, "Orexins and their receptors: structural aspects and role in peripheral tissues," *Cell. Mol. Life Sci. CMLS*, vol. 60, no. 1, pp. 72–87, Jan. 2003, doi: 10.1007/s000180300005.
- [33] C. Wang *et al.*, "The Orexin/Receptor System: Molecular Mechanism and Therapeutic Potential for
 Neurological Diseases," *Front. Mol. Neurosci.*, vol. 11, p. 220, Jun. 2018, doi: 10.3389/fnmol.2018.00220.
- [34] J. M. Siegel, R. Moore, T. Thannickal, and R. Nienhuis, "A brief history of hypocretin/orexin and
- narcolepsy," *Neuropsychopharmacol. Off. Publ. Am. Coll. Neuropsychopharmacol.*, vol. 25, no. 5 Suppl, pp.
 S14-20, Nov. 2001, doi: 10.1016/S0893-133X(01)00317-7.

- [35] R. Fronczek, C. R. Baumann, G. J. Lammers, C. L. Bassetti, and S. Overeem, "Hypocretin/orexin disturbances in neurological disorders," *Sleep Med. Rev.*, vol. 13, no. 1, pp. 9–22, Feb. 2009, doi: 10.1016/j.smrv.2008.05.002.
- 895 [36] S. Huang *et al.*, "Increased plasma orexin-A concentrations are associated with the non-motor
 896 symptoms in Parkinson's disease patients," *Neurosci. Lett.*, vol. 741, p. 135480, Jan. 2021, doi:
- 897 10.1016/j.neulet.2020.135480.
- [37] X. Drouot *et al.*, "Low level of ventricular CSF orexin-A is not associated with objective sleepiness
 in PD," *Sleep Med.*, vol. 12, no. 9, pp. 936–937, Oct. 2011, doi: 10.1016/j.sleep.2011.08.002.
- [38] R. Fronczek *et al.*, "Hypocretin (orexin) loss in Parkinson's disease," *Brain J. Neurol.*, vol. 130, no.
 Pt 6, pp. 1577–1585, Jun. 2007, doi: 10.1093/brain/awm090.
- 902 [39] Y. Compta *et al.*, "Cerebrospinal hypocretin, daytime sleepiness and sleep architecture in
 903 Parkinson's disease dementia," *Brain J. Neurol.*, vol. 132, no. Pt 12, pp. 3308–3317, Dec. 2009, doi:
 904 10.1093/brain/awp263.
- 905 [40] D. Gerashchenko *et al.*, "Relationship between CSF hypocretin levels and hypocretin neuronal loss,"
 906 *Exp. Neurol.*, vol. 184, no. 2, pp. 1010–1016, Dec. 2003, doi: 10.1016/S0014-4886(03)00388-1.
- M. Wienecke *et al.*, "Progressive dopamine and hypocretin deficiencies in Parkinson's disease: is
 there an impact on sleep and wakefulness?," *J. Sleep Res.*, vol. 21, no. 6, pp. 710–717, Dec. 2012, doi:
 10.1111/j.1365-2869.2012.01027.x.
- 910 [42] H. Asai *et al.*, "Cerebrospinal fluid-orexin levels and sleep attacks in four patients with Parkinson's
 911 disease," *Clin. Neurol. Neurosurg.*, vol. 111, no. 4, pp. 341–344, May 2009, doi:
 912 10.1016/j.clineuro.2008.11.007.
- 913 [43] A. Bridoux *et al.*, "Ventricular orexin-A (hypocretin-1) levels correlate with rapid-eye-movement
 914 sleep without atonia in Parkinson's disease," *Nat. Sci. Sleep*, vol. 5, pp. 87–91, 2013, doi:
 915 10.2147/NSS.S41245.
- [44] Z. Hadadianpour, F. Fatehi, F. Ayoobi, A. Kaeidi, A. Shamsizadeh, and I. Fatemi, "The effect of
 orexin-A on motor and cognitive functions in a rat model of Parkinson's disease," *Neurol. Res.*, vol. 39, no.
 9, pp. 845–851, Sep. 2017, doi: 10.1080/01616412.2017.1352185.
- 919 [45] Y. Xue *et al.*, "Orexin-A increases the activity of globus pallidus neurons in both normal and
 920 parkinsonian rats," *Eur. J. Neurosci.*, vol. 44, no. 5, pp. 2247–2257, Sep. 2016, doi: 10.1111/ejn.13323.
- [46] Y. Wang *et al.*, "Orexins alleviate motor deficits via increasing firing activity of pallidal neurons in a
 mouse model of Parkinson's disease," *Am. J. Physiol. Cell Physiol.*, vol. 317, no. 4, pp. C800–C812, Oct.
 2019, doi: 10.1152/ajpcell.00125.2019.
- [47] W. Poewe *et al.*, "Parkinson disease," *Nat. Rev. Dis. Primer*, vol. 3, p. 17013, Mar. 2017, doi: 10.1038/nrdp.2017.13.
- [48] H.-R. Gao *et al.*, "Orexin Directly Enhances the Excitability of Globus Pallidus Internus Neurons in
 Rat by Co-activating OX1 and OX2 Receptors," *Neurosci. Bull.*, vol. 33, no. 4, pp. 365–372, Apr. 2017, doi:
 10.1007/s12264-017-0127-0.
- [49] Q. Sheng *et al.*, "The Subthalamic Neurons are Activated by Both Orexin-A and Orexin-B," *Neuroscience*, vol. 369, pp. 97–108, Jan. 2018, doi: 10.1016/j.neuroscience.2017.11.008.

- [50] X.-Y. Zhang, L. Yu, Q.-X. Zhuang, J.-N. Zhu, and J.-J. Wang, "Central functions of the orexinergic
 system," *Neurosci. Bull.*, vol. 29, no. 3, pp. 355–365, Jun. 2013, doi: 10.1007/s12264-012-1297-4.
- [51] M. Stanojlovic, J. P. Pallais, and C. M. Kotz, "Inhibition of Orexin/Hypocretin Neurons Ameliorates
 Elevated Physical Activity and Energy Expenditure in the A53T Mouse Model of Parkinson's Disease," *Int. L. Mal. Sci.*, vol. 22, p. 20, p. 705, Jap. 2021, doi: 10.3200/jime22020705
- 935 *J. Mol. Sci.*, vol. 22, no. 2, p. 795, Jan. 2021, doi: 10.3390/ijms22020795.
- 936 [52] M. Stanojlovic, J. P. Pallais, and C. M. Kotz, "Chemogenetic Modulation of Orexin Neurons
- Reverses Changes in Anxiety and Locomotor Activity in the A53T Mouse Model of Parkinson's Disease," *Front. Neurosci.*, vol. 13, p. 702, 2019, doi: 10.3389/fnins.2019.00702.
- [53] C. Kosse, C. Schöne, E. Bracey, and D. Burdakov, "Orexin-driven GAD65 network of the lateral
 hypothalamus sets physical activity in mice," *Proc. Natl. Acad. Sci. U. S. A.*, vol. 114, no. 17, pp. 4525–
 4530, Apr. 2017, doi: 10.1073/pnas.1619700114.
- 942 [54] M. Stanojlovic, J. P. Pallais, M. K. Lee, and C. M. Kotz, "Pharmacological and chemogenetic
 943 orexin/hypocretin intervention ameliorates Hipp-dependent memory impairment in the A53T mice model of
 944 Parkinson's disease," *Mol. Brain*, vol. 12, no. 1, p. 87, Oct. 2019, doi: 10.1186/s13041-019-0514-8.
- [55] S. M, P. Y. Jp, V. A, and K. C, "Early Sociability and Social Memory Impairment in the A53T
 Mouse Model of Parkinson's Disease Are Ameliorated by Chemogenetic Modulation of Orexin Neuron
- 947 Activity," *Mol. Neurobiol.*, vol. 56, no. 12, Dec. 2019, doi: 10.1007/s12035-019-01682-x.
- L. Yang *et al.*, "Hypocretin/orexin neurons contribute to hippocampus-dependent social memory and
 synaptic plasticity in mice," *J. Neurosci. Off. J. Soc. Neurosci.*, vol. 33, no. 12, pp. 5275–5284, Mar. 2013,
 doi: 10.1523/JNEUROSCI.3200-12.2013.
- 951 [57] I. S. Zagon and P. J. McLaughlin, "Endogenous Opioids in the Etiology and Treatment of Multiple
- 952 Sclerosis," in *Multiple Sclerosis: Perspectives in Treatment and Pathogenesis*, I. S. Zagon and P. J.
- McLaughlin, Eds. Brisbane (AU): Codon Publications, 2017. Accessed: Aug. 26, 2021. [Online]. Available:
 http://www.ncbi.nlm.nih.gov/books/NBK470156/
- E. E. Benarroch, "Endogenous opioid systems: Current concepts and clinical correlations," *Neurology*, vol. 79, no. 8, pp. 807–814, Aug. 2012, doi: 10.1212/WNL.0b013e3182662098.
- [59] A. Ljungdahl, J. Hanrieder, M. Fälth, J. Bergquist, and M. Andersson, "Imaging Mass Spectrometry
 Reveals Elevated Nigral Levels of Dynorphin Neuropeptides in L-DOPA-Induced Dyskinesia in Rat Model
 of Parkinson's Disease," *PLoS ONE*, vol. 6, no. 9, p. e25653, Sep. 2011, doi: 10.1371/journal.pone.0025653.
- 960 [60] R. Al-Hasani and M. R. Bruchas, "Molecular Mechanisms of Opioid Receptor-Dependent Signaling
 961 and Behavior," *Anesthesiology*, vol. 115, no. 6, pp. 1363–1381, Dec. 2011, doi:
 10 1007 (ALN 0) 012, 21022011 (
- 962 10.1097/ALN.0b013e318238bba6.
- [61] R. J. Bodnar, "Endogenous opiates and behavior: 2019," *Peptides*, vol. 141, p. 170547, Jul. 2021, doi: 10.1016/j.peptides.2021.170547.
- 965 [62] H. Steiner and C. R. Gerfen, "Role of dynorphin and enkephalin in the regulation of striatal output
 966 pathways and behavior," *Exp. Brain Res.*, vol. 123, no. 1–2, pp. 60–76, Nov. 1998, doi:
 967 10.1007/s002210050545.

- [63] X.-Y. Chen, Y. Xue, H. Chen, and L. Chen, "The globus pallidus as a target for neuropeptides and
 endocannabinoids participating in central activities," *Peptides*, vol. 124, p. 170210, Feb. 2020, doi:
 10.1016/j.peptides.2019.170210.
- [64] J. Pan and H. Cai, "Opioid system in L-DOPA-induced dyskinesia," *Transl. Neurodegener.*, vol. 6,
 p. 1, 2017, doi: 10.1186/s40035-017-0071-y.
- J. Hanrieder, A. Ljungdahl, M. Fälth, S. E. Mammo, J. Bergquist, and M. Andersson, "L-DOPAinduced dyskinesia is associated with regional increase of striatal dynorphin peptides as elucidated by
 imaging mass spectrometry," *Mol. Cell. Proteomics MCP*, vol. 10, no. 10, p. M111.009308, Oct. 2011, doi:
 10.1074/mcp.M111.009308.
- 977 [66] L. F. Potts *et al.*, "Dual κ-agonist / μ-antagonist opioid receptor modulation reduces L-dopa induced
 978 dyskinesia and corrects dysregulated striatal changes in the non-human primate model of Parkinson's
 979 disease," *Ann. Neurol.*, vol. 77, no. 6, pp. 930–941, Jun. 2015, doi: 10.1002/ana.24375.
- [67] A. Vinuela *et al.*, "Implanted reuptake-deficient or wild-type dopaminergic neurons improve ON ldopa dyskinesias without OFF-dyskinesias in a rat model of Parkinson's disease," *Brain*, vol. 131, no. 12,
 pp. 3361–3379, Dec. 2008, doi: 10.1093/brain/awn192.
- [68] L. Frau, M. Morelli, and N. Simola, "Performance of movement in hemiparkinsonian rats influences
 the modifications induced by dopamine agonists in striatal efferent dynorphinergic neurons," *Exp. Neurol.*,
 vol. 247, pp. 663–672, Sep. 2013, doi: 10.1016/j.expneurol.2013.03.002.
- [69] A. Nilsson *et al.*, "Striatal Alterations of Secretogranin-1, Somatostatin, Prodynorphin, and
 Cholecystokinin Peptides in an Experimental Mouse Model of Parkinson Disease," *Mol. Cell. Proteomics MCP*, vol. 8, no. 5, pp. 1094–1104, May 2009, doi: 10.1074/mcp.M800454-MCP200.
- [70] L. Jouve, P. Salin, C. Melon, and L. Kerkerian-Le Goff, "Deep brain stimulation of the center
 median-parafascicular complex of the thalamus has efficient anti-parkinsonian action associated with
 widespread cellular responses in the basal ganglia network in a rat model of Parkinson's disease," *J. Neurosci. Off. J. Soc. Neurosci.*, vol. 30, no. 29, pp. 9919–9928, Jul. 2010, doi: 10.1523/JNEUROSCI.140410.2010.
- Y. Y. Yien and B. H. Paw, "A role for iron deficiency in dopaminergic neurodegeneration," *Proc. Natl. Acad. Sci. U. S. A.*, vol. 113, no. 13, p. 3417, Mar. 2016, doi: 10.1073/pnas.1601976113.
- Y.-M. J. Sun, T. Hoang, J. A. Neubauer, and A. S. Walters, "Opioids protect against substantia nigra
 cell degeneration under conditions of iron deprivation: a mechanism of possible relevance to the Restless
 Legs Syndrome (RLS) and Parkinson's disease," *J. Neurol. Sci.*, vol. 304, no. 1–2, pp. 93–101, May 2011,
 doi: 10.1016/j.jns.2011.02.003.
- 1000 [73] A. J. Harmar *et al.*, "Pharmacology and functions of receptors for vasoactive intestinal peptide and
 1001 pituitary adenylate cyclase-activating polypeptide: IUPHAR Review 1," *Br. J. Pharmacol.*, vol. 166, no. 1,
 1002 pp. 4–17, May 2012, doi: 10.1111/j.1476-5381.2012.01871.x.
- 1003 [74] S. I. Said and V. Mutt, "Polypeptide with broad biological activity: isolation from small intestine,"
 1004 *Science*, vol. 169, no. 3951, pp. 1217–1218, Sep. 1970, doi: 10.1126/science.169.3951.1217.
- 1005 [75] M. Iwasaki, Y. Akiba, and J. D. Kaunitz, "Recent advances in vasoactive intestinal peptide
 1006 physiology and pathophysiology: focus on the gastrointestinal system," *F1000Research*, vol. 8, p. F1000
 1007 Faculty Rev-1629, Sep. 2019, doi: 10.12688/f1000research.18039.1.

1008 [76] F. Giancola *et al.*, "Downregulation of neuronal vasoactive intestinal polypeptide in Parkinson's
1009 disease and chronic constipation," *Neurogastroenterol. Motil. Off. J. Eur. Gastrointest. Motil. Soc.*, vol. 29,
1010 no. 5, May 2017, doi: 10.1111/nmo.12995.

1011 [77] T. Hirabayashi, T. Nakamachi, and S. Shioda, "Discovery of PACAP and its receptors in the brain,"
1012 *J. Headache Pain*, vol. 19, no. 1, p. 28, Apr. 2018, doi: 10.1186/s10194-018-0855-1.

- 1013 [78] N. Tunçel, O. T. Korkmaz, N. Tekin, E. Şener, F. Akyüz, and M. Inal, "Antioxidant and anti-apoptotic activity of vasoactive intestinal peptide (VIP) against 6-hydroxy dopamine toxicity in the rat
 1015 corpus striatum," *J. Mol. Neurosci. MN*, vol. 46, no. 1, pp. 51–57, Jan. 2012, doi: 10.1007/s12031-011-96181016 z.
- 1017 [79] R. Yu *et al.*, "TAT-tagging of VIP exerts positive allosteric modulation of the PAC1 receptor and
 1018 enhances VIP neuroprotective effect in the MPTP mouse model of Parkinson's disease," *Biochim. Biophys.*1019 *Acta Gen. Subj.*, vol. 1864, no. 8, p. 129626, Aug. 2020, doi: 10.1016/j.bbagen.2020.129626.
- 1020 [80] O. Korkmaz, H. Ay, E. Ulupinar, and N. Tunçel, "Vasoactive intestinal peptide enhances striatal
 1021 plasticity and prevents dopaminergic cell loss in Parkinsonian rats," *J. Mol. Neurosci. MN*, vol. 48, no. 3, pp.
 1022 565–573, Nov. 2012, doi: 10.1007/s12031-012-9781-x.
- 1023 [81] C. M. White, S. Ji, H. Cai, S. Maudsley, and B. Martin, "Therapeutic potential of vasoactive
 1024 intestinal peptide and its receptors in neurological disorders," *CNS Neurol. Disord. Drug Targets*, vol. 9, no.
 1025 5, pp. 661–666, Nov. 2010.
- 1026 [82] K. E. Olson *et al.*, "Selective VIP Receptor Agonists Facilitate Immune Transformation for
 1027 Dopaminergic Neuroprotection in MPTP-Intoxicated Mice," *J. Neurosci.*, vol. 35, no. 50, pp. 16463–16478,
 1028 Dec. 2015, doi: 10.1523/JNEUROSCI.2131-15.2015.
- 1029 [83] A. Bohush, G. Niewiadomska, and A. Filipek, "Role of Mitogen Activated Protein Kinase Signaling
 1030 in Parkinson's Disease," *Int. J. Mol. Sci.*, vol. 19, no. 10, p. 2973, Sep. 2018, doi: 10.3390/ijms19102973.
- 1031 [84] R. L. Mosley *et al.*, "A Synthetic Agonist to Vasoactive Intestinal Peptide Receptor-2 Induces
 1032 Regulatory T Cell Neuroprotective Activities in Models of Parkinson's Disease," *Front. Cell. Neurosci.*, vol.
 1033 13, p. 421, 2019, doi: 10.3389/fncel.2019.00421.
- 1034 [85] O. T. Korkmaz, N. Tunçel, M. Tunçel, E. M. Oncü, V. Sahintürk, and M. Celik, "Vasoactive
 1035 intestinal peptide (VIP) treatment of Parkinsonian rats increases thalamic gamma-aminobutyric acid (GABA)
 1036 levels and alters the release of nerve growth factor (NGF) by mast cells," *J. Mol. Neurosci. MN*, vol. 41, no.
 1037 2, pp. 278–287, Jun. 2010, doi: 10.1007/s12031-009-9307-3.
- 1038 [86] İ. H. Yelkenli *et al.*, "Modulation of Corpus Striatal Neurochemistry by Astrocytes and Vasoactive
 1039 Intestinal Peptide (VIP) in Parkinsonian Rats," *J. Mol. Neurosci. MN*, vol. 59, no. 2, pp. 280–289, Jun. 2016,
 1040 doi: 10.1007/s12031-016-0757-0.
- 1041 [87] H. J. Federoff, "Nur(R1)turing a notion on the etiopathogenesis of Parkinson's disease," *Neurotox.*1042 *Res.*, vol. 16, no. 3, pp. 261–270, Oct. 2009, doi: 10.1007/s12640-009-9056-7.
- 1043 [88] C. Jia *et al.*, "α-Synuclein Negatively Regulates Nurr1 Expression Through NF-κB-Related
 1044 Mechanism," *Front. Mol. Neurosci.*, vol. 0, 2020, doi: 10.3389/fnmol.2020.00064.

1045 [89] A. Mascolo *et al.*, "New and old roles of the peripheral and brain renin-angiotensin-aldosterone
1046 system (RAAS): Focus on cardiovascular and neurological diseases," *Int. J. Cardiol.*, vol. 227, pp. 734–742,
1047 Jan. 2017, doi: 10.1016/j.ijcard.2016.10.069.

E. Farag *et al.*, "The renin angiotensin system and the brain: New developments," *J. Clin. Neurosci. Off. J. Neurosurg. Soc. Australas.*, vol. 46, pp. 1–8, Dec. 2017, doi: 10.1016/j.jocn.2017.08.055.

1050 [91] M. J. McKinley *et al.*, "The brain renin-angiotensin system: location and physiological roles," *Int. J.*1051 *Biochem. Cell Biol.*, vol. 35, no. 6, pp. 901–918, Jun. 2003, doi: 10.1016/s1357-2725(02)00306-0.

W. M. Zawada *et al.*, "Loss of angiotensin II receptor expression in dopamine neurons in
Parkinson's disease correlates with pathological progression and is accompanied by increases in Nox4- and
8-OH guanosine-related nucleic acid oxidation and caspase-3 activation," *Acta Neuropathol. Commun.*, vol.
3, p. 9, Feb. 2015, doi: 10.1186/s40478-015-0189-z.

1056 [93] C. Cosarderelioglu *et al.*, "Brain Renin–Angiotensin System at the Intersect of Physical and
1057 Cognitive Frailty," *Front. Neurosci.*, vol. 14, p. 981, 2020, doi: 10.3389/fnins.2020.586314.

1058 [94] N. P. Rocha *et al.*, "Peripheral levels of angiotensins are associated with depressive symptoms in
1059 Parkinson's disease," *J. Neurol. Sci.*, vol. 368, pp. 235–239, Sep. 2016, doi: 10.1016/j.jns.2016.07.031.

1060 [95] Q. Gao *et al.*, "Azilsartan ameliorates apoptosis of dopaminergic neurons and rescues characteristic
1061 parkinsonian behaviors in a rat model of Parkinson's disease," *Oncotarget*, vol. 8, no. 15, pp. 24099–24109,
1062 Apr. 2017, doi: 10.18632/oncotarget.15732.

1063 [96] A. I. Rodriguez-Perez, R. Valenzuela, B. Villar-Cheda, M. J. Guerra, J. L. Lanciego, and J. L.
1064 Labandeira-Garcia, "Estrogen and angiotensin interaction in the substantia nigra. Relevance to
1065 postmenopausal Parkinson's disease," *Exp. Neurol.*, vol. 224, no. 2, pp. 517–526, Aug. 2010, doi:
1066 10.1016/j.expneurol.2010.05.015.

1067 [97] J. Rodriguez-Pallares, J. A. Parga, B. Joglar, M. J. Guerra, and J. L. Labandeira-Garcia,
1068 "Mitochondrial ATP-sensitive potassium channels enhance angiotensin-induced oxidative damage and
1069 dopaminergic neuron degeneration. Relevance for aging-associated susceptibility to Parkinson's disease,"
1070 *Age*, vol. 34, no. 4, pp. 863–880, Aug. 2012, doi: 10.1007/s11357-011-9284-7.

1071 [98] B. Villar-Cheda *et al.*, "Nigral and striatal regulation of angiotensin receptor expression by
1072 dopamine and angiotensin in rodents: implications for progression of Parkinson's disease," *Eur. J. Neurosci.*,
1073 vol. 32, no. 10, pp. 1695–1706, Nov. 2010, doi: 10.1111/j.1460-9568.2010.07448.x.

1074 [99] B. Villar-Cheda, R. Valenzuela, A. I. Rodriguez-Perez, M. J. Guerra, and J. L. Labandeira-Garcia,
1075 "Aging-related changes in the nigral angiotensin system enhances proinflammatory and pro-oxidative
1076 markers and 6-OHDA-induced dopaminergic degeneration," *Neurobiol. Aging*, vol. 33, no. 1, p. 204.e1–11,
1077 Jan. 2012, doi: 10.1016/j.neurobiolaging.2010.08.006.

1078 [100] J. L. Labandeira-Garcia, A. I. Rodríguez-Perez, P. Garrido-Gil, J. Rodriguez-Pallares, J. L.
1079 Lanciego, and M. J. Guerra, "Brain Renin-Angiotensin System and Microglial Polarization: Implications for
1080 Aging and Neurodegeneration," *Front. Aging Neurosci.*, vol. 9, p. 129, 2017, doi: 10.3389/fnagi.2017.00129.

[101] A. A. Boldyrev, G. Aldini, and W. Derave, "Physiology and pathophysiology of carnosine," *Physiol. Rev.*, vol. 93, no. 4, pp. 1803–1845, Oct. 2013, doi: 10.1152/physrev.00039.2012.

- 1083 [102] M. Kawahara, K.-I. Tanaka, and M. Kato-Negishi, "Zinc, Carnosine, and Neurodegenerative
 1084 Diseases," *Nutrients*, vol. 10, no. 2, p. E147, Jan. 2018, doi: 10.3390/nu10020147.
- 1085 [103] M. Schön *et al.*, "The Potential of Carnosine in Brain-Related Disorders: A Comprehensive Review 1086 of Current Evidence," *Nutrients*, vol. 11, no. 6, p. 1196, May 2019, doi: 10.3390/nu11061196.
- 1087 [104] A. Boldyrev et al., "Carnosine [corrected] increases efficiency of DOPA therapy of Parkinson's
- 1088 disease: a pilot study," *Rejuvenation Res.*, vol. 11, no. 4, pp. 821–827, Aug. 2008, doi:
- 1089 10.1089/rej.2008.0716.
- 1090 [105] P. Sozio *et al.*, "New L-dopa codrugs as potential antiparkinson agents," *Arch. Pharm. (Weinheim)*,
 1091 vol. 341, no. 7, pp. 412–417, Jul. 2008, doi: 10.1002/ardp.200700228.
- 1092 [106] M.-L. Bermúdez, K. B. Seroogy, and M. B. Genter, "Evaluation of Carnosine Intervention in the
 1093 Thy1-aSyn Mouse Model of Parkinson's Disease," *Neuroscience*, vol. 411, pp. 270–278, Jul. 2019, doi:
 1094 10.1016/j.neuroscience.2019.05.026.
- 1095 [107] M. Kubota, N. Kobayashi, T. Sugizaki, M. Shimoda, M. Kawahara, and K.-I. Tanaka, "Carnosine 1096 suppresses neuronal cell death and inflammation induced by 6-hydroxydopamine in an in vitro model of 1097 Parkinson's disease," *PloS One*, vol. 15, no. 10, p. e0240448, 2020, doi: 10.1371/journal.pone.0240448.
- 1098 [108] S.-J. Tsai, W.-W. Kuo, W.-H. Liu, and M.-C. Yin, "Antioxidative and anti-inflammatory protection
 1099 from carnosine in the striatum of MPTP-treated mice," *J. Agric. Food Chem.*, vol. 58, no. 21, pp. 11510–
 1100 11516, Nov. 2010, doi: 10.1021/jf103258p.
- [109] M.-L. Bermúdez, M. R. Skelton, and M. B. Genter, "Intranasal carnosine attenuates transcriptomic alterations and improves mitochondrial function in the Thy1-aSyn mouse model of Parkinson's disease," *Mol. Genet. Metab.*, vol. 125, no. 3, pp. 305–313, Nov. 2018, doi: 10.1016/j.ymgme.2018.08.002.
- 1104 [110] S. Shioda, F. Takenoya, N. Wada, T. Hirabayashi, T. Seki, and T. Nakamachi, "Pleiotropic and
 1105 retinoprotective functions of PACAP," *Anat. Sci. Int.*, vol. 91, no. 4, pp. 313–324, Sep. 2016, doi:
 1106 10.1007/s12565-016-0351-0.
- 1107 [111] T. Hirabayashi, T. Nakamachi, and S. Shioda, "Discovery of PACAP and its receptors in the brain,"
 1108 *J. Headache Pain*, vol. 19, no. 1, p. 28, Apr. 2018, doi: 10.1186/s10194-018-0855-1.
- 1109 [112] A. Arimura, "Perspectives on pituitary adenylate cyclase activating polypeptide (PACAP) in the
 1110 neuroendocrine, endocrine, and nervous systems," *Jpn. J. Physiol.*, vol. 48, no. 5, pp. 301–331, Oct. 1998,
 1111 doi: 10.2170/jjphysiol.48.301.
- 1112 [113] N. M. Sherwood, S. L. Krueckl, and J. E. McRory, "The Origin and Function of the Pituitary
- Adenylate Cyclase-Activating Polypeptide (PACAP)/Glucagon Superfamily*," *Endocr. Rev.*, vol. 21, no. 6,
 pp. 619–670, Dec. 2000, doi: 10.1210/edrv.21.6.0414.
- 1115 [114] G. Maasz et al., "Pituitary adenylate cyclase-activating polypeptide (PACAP) has a neuroprotective
- function in dopamine-based neurodegeneration in rat and snail parkinsonian models," *Dis. Model. Mech.*,
 vol. 10, no. 2, pp. 127–139, Feb. 2017, doi: 10.1242/dmm.027185.
- 1118 [115] R. Yu *et al.*, "TAT-tagging of VIP exerts positive allosteric modulation of the PAC1 receptor and
- 1119 enhances VIP neuroprotective effect in the MPTP mouse model of Parkinson's disease," *Biochim. Biophys.* 1120 Acta Can Subi yol 1864 no 8 p 120626 Aug 2020 doi: 10.1016/j.bbogon.2020.120626
- 1120 Acta Gen. Subj., vol. 1864, no. 8, p. 129626, Aug. 2020, doi: 10.1016/j.bbagen.2020.129626.

- 1121 [116] K.-Y. Shivers, A. Nikolopoulou, S. I. Machlovi, S. Vallabhajosula, and M. E. Figueiredo-Pereira,
- 1122 "PACAP27 prevents Parkinson-like neuronal loss and motor deficits but not microglia activation induced by
- 1123 prostaglandin J2," Biochim. Biophys. Acta, vol. 1842, no. 9, pp. 1707–1719, Sep. 2014, doi:
- 1124 10.1016/j.bbadis.2014.06.020.
- 1125 [117] D. Brown, A. Tamas, D. Reglodi, and Y. Tizabi, "PACAP protects against inflammatory-mediated
- toxicity in dopaminergic SH-SY5Y cells: implication for Parkinson's disease," *Neurotox. Res.*, vol. 26, no. 3,
 pp. 230–239, Oct. 2014, doi: 10.1007/s12640-014-9468-x.
- 1128 [118] K. Wakabayashi, K. Tanji, F. Mori, and H. Takahashi, "The Lewy body in Parkinson's disease:
- 1129 molecules implicated in the formation and degradation of alpha-synuclein aggregates," Neuropathol. Off. J.
- 1130 Jpn. Soc. Neuropathol., vol. 27, no. 5, pp. 494–506, Oct. 2007, doi: 10.1111/j.1440-1789.2007.00803.x.
- [113] [119] K. Knudsen *et al.*, "Pancreatic Polypeptide in Parkinson's Disease: A Potential Marker of
 Parasympathetic Denervation," *J. Park. Dis.*, vol. 7, no. 4, pp. 645–652, 2017, doi: 10.3233/JPD-171189.
- 1133 [120] T. J. O'Toole and S. Sharma, "Physiology, Somatostatin," in *StatPearls*, Treasure Island (FL):
- 1134 StatPearls Publishing, 2021. Accessed: Aug. 20, 2021. [Online]. Available:
- 1135 http://www.ncbi.nlm.nih.gov/books/NBK538327/
- 1136 [121] J. Lonovics, P. Devitt, L. C. Watson, P. L. Rayford, and J. C. Thompson, "Pancreatic polypeptide. A
- 1137 review," Arch. Surg. Chic. Ill 1960, vol. 116, no. 10, pp. 1256–1264, Oct. 1981, doi:
- 1138 10.1001/archsurg.1981.01380220010002.
- 1139 [122] S. M, K. T, W. H, K. T, and H. Y, "Serum somatostatin in early-stage Parkinson's disease," *Acta Neurol. Scand.*, vol. 121, no. 4, Apr. 2010, doi: 10.1111/j.1600-0404.2009.01178.x.
- 1141 [123] F. Escamilla-Sevilla *et al.*, "Change of the melanocortin system caused by bilateral subthalamic
 1142 nucleus stimulation in Parkinson's disease," *Acta Neurol. Scand.*, vol. 124, no. 4, pp. 275–281, Oct. 2011,
 1143 doi: 10.1111/j.1600-0404.2010.01469.x.
- 1144 [124] "Neuroprotection by neuropeptide Y in cell and animal models of Parkinson's disease PubMed."
 1145 https://pubmed.ncbi.nlm.nih.gov/21816512/ (accessed Aug. 20, 2021).
- 1146 [125] S. Bahdoudi et al., "Neuroprotective effects of the gliopeptide ODN in an in vivo model of
- Parkinson's disease," *Cell. Mol. Life Sci. CMLS*, vol. 75, no. 11, pp. 2075–2091, Jun. 2018, doi:
 10.1007/s00018-017-2727-2.
- 1149 [126] J. L. Elson, R. Kochaj, R. Reynolds, and I. S. Pienaar, "Temporal-Spatial Profiling of
- Pedunculopontine Galanin-Cholinergic Neurons in the Lactacystin Rat Model of Parkinson's Disease,"
 Neurotox. Res., vol. 34, no. 1, pp. 16–31, Jul. 2018, doi: 10.1007/s12640-017-9846-2.
- I. T. French and K. A. Muthusamy, "A Review of the Pedunculopontine Nucleus in Parkinson's Disease," *Front. Aging Neurosci.*, vol. 10, p. 99, 2018, doi: 10.3389/fnagi.2018.00099.
- 1154 [128] Y. Li, W. Liu, L. Li, and C. Hölscher, "D-Ala2-GIP-glu-PAL is neuroprotective in a chronic
- 1155 Parkinson's disease mouse model and increases BNDF expression while reducing neuroinflammation and
- lipid peroxidation," *Eur. J. Pharmacol.*, vol. 797, pp. 162–172, Feb. 2017, doi:
- 1157 10.1016/j.ejphar.2016.11.050.

- 1158 [129] C. H. S. McIntosh, S. Widenmaier, and S.-J. Kim, "Glucose-dependent insulinotropic polypeptide 1159 (Gastric Inhibitory Polypeptide; GIP)," *Vitam. Horm.*, vol. 80, pp. 409–471, 2009, doi: 10.1016/S0083-
- 1159
 (Gastric Infibitory Polypeptide, GIP), Vitam. Horm., vol. 80, pp. 409–471, 2009, doi: 10.1010/S0

 1160
 6729(08)00615-8.

[130] Z. Q. Zhang and C. Hölscher, "GIP has neuroprotective effects in Alzheimer and Parkinson's disease
models," *Peptides*, vol. 125, p. 170184, Mar. 2020, doi: 10.1016/j.peptides.2019.170184.

- 1163 [131] C. Cocco et al., "Distribution of VGF peptides in the human cortex and their selective changes in
- Parkinson's and Alzheimer's diseases," J. Anat., vol. 217, no. 6, pp. 683–693, Dec. 2010, doi:
- 1165 10.1111/j.1469-7580.2010.01309.x.
- 1166 [132] T. Umehara, H. Oka, A. Nakahara, H. Matsuno, and C. Toyoda, "High norepinephrinergic
 1167 orthostatic hypotension in early Parkinson's disease," *Parkinsonism Relat. Disord.*, vol. 55, pp. 97–102, Oct.
 1168 2018, doi: 10.1016/j.parkreldis.2018.05.025.
- 1169 [133] "Peptides semax and selank affect the behavior of rats with 6-OHDA induced PD-like parkinsonism
 1170 PubMed." https://pubmed.ncbi.nlm.nih.gov/28702721/ (accessed Aug. 21, 2021).
- 1171 [134] A. Niewiarowska-Sendo, A. Kozik, and I. Guevara-Lora, "Kinin Peptides Enhance Inflammatory
- and Oxidative Responses Promoting Apoptosis in a Parkinson's Disease Cellular Model," *Mediators*
- 1173 Inflamm., vol. 2016, p. 4567343, 2016, doi: 10.1155/2016/4567343.
- 1174 [135] "ER Lipid Defects in Neuropeptidergic Neurons Impair Sleep Patterns in Parkinson's Disease -
- 1175 PubMed." https://pubmed.ncbi.nlm.nih.gov/29887339/ (accessed Aug. 21, 2021).
- 1176