

# Peptostreptococcus faecalis sp. nov., new bacterial species isolated from healthy indigenous congolese volunteer

Fatima Mekhalif, Rita Zgheib, Jean Akiana, Melhem Bilen, Geor Mongo Ndombe, Florence Fenollar, Pierre-Edouard Fournier, Didier Raoult, Stéphane Alibar, Oleg Mediannikov, et al.

### ▶ To cite this version:

Fatima Mekhalif, Rita Zgheib, Jean Akiana, Melhem Bilen, Geor Mongo Ndombe, et al.. Peptostreptococcus faecalis sp. nov., new bacterial species isolated from healthy indigenous congolese volunteer. Heliyon, 2022, 8 (3), pp.e09102. 10.1016/j.heliyon.2022.e09102. hal-03678949

## HAL Id: hal-03678949 https://amu.hal.science/hal-03678949

Submitted on 22 Jul 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



## Peptostreptococcus faecalis sp. nov., new bacterial species isolated from healthy 1 2 indigenous Congolese volunteer 3 Fatima Mekhalif<sup>1,2</sup>, Rita Zgheib<sup>2,3</sup>, Jean Akiana<sup>4</sup>, Melhem Bilen<sup>1,3</sup>, Geor Mongo Ndombe<sup>4†</sup>, 4 Florence Fenollar<sup>2,5</sup>, Pierre-Edouard Fournier<sup>2,5</sup>, Didier Raoult<sup>1,2</sup>, Stéphane Alibar<sup>2</sup>, Oleg 5 Mediannikov<sup>1,2</sup> and Cheikh Ibrahima Lo<sup>2,5\*</sup> 6 7 <sup>1</sup>Aix Marseille Univ, IRD, AP-HM, MEPHI, Marseille, France 8 9 <sup>2</sup>IHU-Méditerranée Infection, Marseille, France 10 <sup>3</sup>University of Balamand, Faculty of medicine and medical sciences, Amioun, Lebanon 11 <sup>4</sup>Laboratoire National de Santé Publique, Brazzaville, Congo <sup>5</sup>Aix Marseille Univ, IRD, AP-HM, SSA, VITROME, Marseille, France 12 13 \*Corresponding author: Cheikh Ibrahima Lo, Institut Hospitalo-Universitaire Méditerranée-14 15 Infection, 19-21 Boulevard Jean Moulin, 13385, Marseille cedex 05, France; tel +33 413 732 401; fax: +33 413 732 402; e-mail: cibrahimalo@gmail.com. 16 17 **Keywords:** Peptostreptococcus faecalis; gut microbiote; indigenous Congolese; culturomics; 18 19 taxonogenomics 20 † - Geor Mongo Ndombe passed away. 21

22

23

Word abstract count: 149

Word text count: 1,056

#### **ABSTRACT**

24

The Microbial Culturomics Project aiming to discover several bacterial species made it 25 possible to isolate the strain Marseille-P4308<sup>T</sup> from a stool sample of a healthy indigenous 26 Congolese volunteer. Strain Marseille-P4308<sup>T</sup> is a Gram-positive coccus shaped bacterium 27 that optimally grows at 37°C. The 16S rRNA gene sequence of the strain has a 96.2% 28 sequence similarity to *Peptostreptococcus anaerobius* strain NCTC 11460<sup>T</sup> (GenBank 29 accession number: NR 042847.1). In addition, the average nucleotide identity of strain 30 Marseille-P4308<sup>T</sup> with its closest related species was 71.1%, which was far below the 31 recommended threshold (>95-96%). The genome of the strain Marseille-P4308<sup>T</sup> has a length 32 of 2.14 Mbp with G+C content of 30.4 mol%. Based on phenotypic, biochemical, genomic 33 and phylogenetic analysis, strain Marseille-P4308<sup>T</sup> (= CSUR P4308 = CECT 9960) clearly 34 appears to be a new species for which the name *Peptostreptococcus faecalis* sp. nov., is 35 36 proposed.

#### INTRODUCTION

37

38 Different genera of anaerobic cocci bacteria are involved in a broad range of infections, occurring in all parts of the human body [1]. In various environments, members of the 39 Peptostreptococcaceae family belonging to the order Clostridiales, phylum Firmicutes can be 40 found in the human body, manure, soil and sediment [2]. The closest phylogenetic neighbours 41 42 to this family belong to the genera Alkaliphilus, Natronincola, and Tindallia [2]. The species 43 of the genus *Peptostreptococcus* are commensal species that colonise almost every mucosal human tissue, forming a part of the intestinal, urinary, vaginal, oral tract microbiotas and also 44 the skin. They are pathogenic under certain circumstances and can cause bacteraemia and 45 46 abscesses in different organs [3]. The genus *Peptostreptococcus* is a group of Gram-positive, mostly anaerobic cocci species 47 that are very diverse phenotypically and phylogenetically [4]. Cells measure between 0.3 and 48 49 2.0 µm and are arranged in chains, pairs, tetrads or masses. Some are aero-tolerant but do not form spores. Their ability to use carbohydrates varies considerably. The main source of 50 51 energy would be the products of protein metabolism [1]. However, the pathogenic character 52 of some members of the genus *Peptostreptococcus* is complex to determine, because some members are part of the normal microflora, while species such as Peptostreptococcus 53 54 anaerobius and Peptostreptococcus stomatis have been frequently identified from clinical samples of diseased individuals [5–7]. Indeed, *Peptostreptococcus russellii* was isolated in 55 environment as a storage pit [8]. 56 Numerous previously uncultured members of the human microbiome have been isolated by 57 the culturomics method [9]. Isolated species are then described by multiphasic approach 58 including a MALDI-TOF MS identification, 16S rRNA gene sequencing and the phenotypic 59 and biochemical analysis of the strain [10]. We report here a taxonogenomic description of 60

- 61 Peptostreptococcus faecalis sp. nov., strain Marseille-P4308<sup>T</sup> isolated from human gut
- 62 microbiota.

#### 63 MATERIAL AND METHODS

- 64 Ethics and sample collection
- The stool sample was collected from a healthy indigenous Congolese volunteer living in Bene
- 66 Gain (1°59'24.7"S 15°52'18.1"E), in the Republic of the Congo, in August 2015
- 67 (supplementary Figure S1) [11]. Approval for this study was obtained from the Ministry for
- Health of the Republic of Congo (000208/MSP/CAB.15 du Ministère de la Santé et de la
- 69 Population, 20 August 2015). The study was also approved by the ethics committee of the
- 70 Institut Fédératif de Recherche IFR48, Faculty of Medicine, Marseille, France, under
- reference number 09-022. Prior to sampling, an informed consent form was obtained from
- each individual. In the presence of representatives from a local health centre and village
- elders, full information was given orally in the local language (Lingala) to ensure that the
- 74 project was fully understood, as the participants were illiterate.
- 75 *Strain isolation and growth conditions*
- One gram of the stool sample was pre-incubated in a blood culture bottle containing rumen
- and sheep blood, as described above for culturing human samples [12]. The culture medium
- was serially diluted and inoculated on Columbia agar with 5% sheep blood (BioMérieux,
- 79 Marcy-L'Etoile, France). All pure colonies obtained by culture were identified by MALDI-
- TOF MS [13]. If, despite the good quality of the bacterial spectrum, the bacteria could not be
- 81 identified, sequencing of the 16S rRNA gene of the bacterium was performed for
- 82 identification.
- 83 *16S rRNA sequencing and phylogenetic analysis*
- The DNA of the bacterial strain was extracted using the EZ1 (Qiagen, Venlo, The
- Netherlands) DNA tissue kit on the EZ1 (Qiagen) automate. The universal primers fD1 and

rP2 (Eurogentec, Angers, France) were used to amplify the 16S rRNA gene sequence. 86 87 Sequencing was performed using the BigDye® Terminator v1.1 Cycle Sequencing Kit and ABI Prism 3130xl Genetic Analyzer capillary sequencer (Thermo Fisher, Saint-Aubin, 88 France), as previously described [14]. Using Codon Code Aligner software 89 (http://www.codoncode.com), the 16S rRNA nucleotide sequences were assembled and 90 91 corrected. Consensus sequence from 16S rRNA gene sequencing was compared by BLASTn 92 within the NCBI 16S rRNA database (https://blast.ncbi.nlm.nih.gov/). In order to create a robust phylogenetic tree, the 16S rRNA sequences of species with a validly published name 93 were downloaded from the LPSN website (https://lpsn.dsmz.de/). Using MEGA X software 94 95 [15], the sequences were aligned and a phylogenetic tree was constructed with 1,000 bootstrap replicates. 96 Genome annotation and comparison 97 Genomes of closely related species were downloaded from the GenBank database and 98 annotated with Prokka software v1.14.6 [16]. Coding sequences were predicted using 99 100 Prodigal software 2.6 [17], then the predicted bacterial protein sequences were searched 101 against the GenBank (https://www.ncbi.nlm.nih.gov/genbank/) database using BLASTp. ARAGORN software 1.2 [18] was used to find tRNA and mRNA genes, whereas rRNA 102 103 genes were predicted with Barrnap software 0.4. 104 The genome of strain Marseille-P4308<sup>T</sup> was compared to the genomes of the following closely related species, including *Peptostreptococcus anaerobius* NCTC11460<sup>T</sup> (GenBank 105 accession: UGTB00000000), Peptostreptococcus russellii RT-10B<sup>T</sup> (JYGE00000000), 106 Peptostreptococcus stomatis DSM 17678<sup>T</sup> (ADGQ0000000), Peptostreptococcus canis 107 DSM 27025<sup>T</sup> (JABGBW00000000), Asaccharospora irregularis DSM 2635<sup>T</sup> 108

(FQWX0000000), Clostridioides difficile ATCC 9689<sup>T</sup> (AUOX0000000), Clostridioides

mangenotii LM2 (JIAA0000000) and Clostridium hiranonis DSM 13275<sup>T</sup> (CP036523). The

109

DNA-DNA hybridisation (dDDH) was calculated to assess similarity between studied genome sequences using the online server of Genome-to-Genome Distance Calculator (GGDC 2.1) (http://ggdc.dsmz.de/), taking into account the method and formula two, as suggested [19,20]. Furthermore, the genomic average nucleotide identity (ANI) was determined using OrthoANI software v0.93.1 [21]. Clusters of Orthologous genes (COGs) were detected by performing BLASTp of genomes against the COG database [22]. Conditions of growth The strain was grown under different conditions. In terms of temperature, the growth of the strain was evaluated at room temperature, 28°C, 37°C, 45°C and 55°C and incubated under aerobic, anaerobic and microaerophilic conditions on Colombia agar enriched with 5% sheep blood (BioMérieux), using the GENbag anaer and GENbag microaer systems (ThermoFisher Scientific, Basingstoke), respectively. To determine the tolerated salt concentration and the optimum pH for growth, the strain was cultured in different media with varied pH (6, 6.5, 7 and 8.5) and at different NaCl concentrations (5, 10, 50, 75 and 100 g/L NaCl) at 37°C under anaerobic conditions. Phenotypic characteristics Motility test and Gram staining of the strain were verified using a DM1000 photomicroscope (Leica Microsystems, Nanterre, France) under a 100× objective. In addition, a bacterial suspension was fixed with a 2.5% glutaraldehyde solution at 0.1 mol/L with the aim of observing the morphology of cells using a Hitachi TM4000 electron microscope (Hitachi Group, Krefeld, Germany). Spore-forming was investigated by exposing a bacterial suspension for 10 minutes under thermal shock at 80°C. Multiple biochemical criteria from strain Marseille-P4308<sup>T</sup> were revealed using API tests (50CH, ZYM and 20A; bioMérieux). Oxidase and catalase reactions were determined using a BD BBL<sup>TM</sup> DrySlide (Becton Dickinson, Le Pont-de-Claix, France).

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

Antibiotic susceptibility 136 The E test method was used to obtain an antimicrobial sensitivity profile and minimum 137 inhibitory concentration (µg/mL) of the strain Marseille-P4308<sup>T</sup> [23]. Antimicrobial discs 138 placed on a blood agar Petri dish were amikacin, amoxicillin, benzylpenicillin, ceftazidime, 139 ceftriaxone, ciprofloxacin, clindamycin, daptomycin, erythromycin, imipenem, linezolid, 140 metronidazole, minocycline, rifampicin, teicoplanin, tigecycline, 141 142 trimethoprim/sulfamethoxazole, tobramycin and vancomycin. **RESULTS** 143 Identification and classification 144 Analysis of the 16S rRNA gene sequence on NCBI Blast showed that the Marseille-P4308<sup>T</sup> 145 strain has the highest similarity score of 96.2% to Peptostreptococcus anaerobius strain 146 NCTC 11460<sup>T</sup> (GenBank accession number: NR 042847.1). This value obtained is below the 147 148 threshold value recommended for delimiting a new species of prokaryote [24]. Therefore, the strain Marseille-P4308<sup>T</sup> is presumed to be a potential new bacterial species belonging to the 149 150 genus Peptostreptococcus within the Peptostreptococcaceae family and phylum Firmicutes. 151 The phylogenetic tree constructed on the basis of the sequences of the 16S rRNA gene shows the strain Marseille-P4308<sup>T</sup> anchored in the *Peptostreptococcus* sp group with closely related 152 153 species (Figure 1). The position is confirmed with the phylogenomic tree basing on the genomic sequences of the closely related species (Figure 2). 154 Phenotypic characteristics 155 Strain Marseille-P4308<sup>T</sup> grows under anaerobic and micro-aerobic conditions between 25°C 156 and 42°C, with an optimal growth at 37°C. It is a Gram-positive coccus-shaped bacterium, 157 non-motile, non-spore-forming, oxidase-negative and catalase-negative. Using a Hitachi 158 159 electron microscope, the strain's morphology is highlighted; cells had a mean length of 0.84 μm and a mean diameter of 0.7 μm (supplementary Figure S2). Furthermore, this strain was 160

able to grow on media at a pH between 6 and 8.5 and tolerated NaCl concentration up to 50 161 162 g/L. Colonies were smooth, convex and regular in appearance, with a mean diameter of 0.5 mm on Columbia Agar with 5% Sheep Blood (bioMérieux). 163 Using API 50CH, the following carbohydrates are fermented: glycerol, erythritol, D-164 arabinose, D-ribose, L-xylose, D-adonitol, methyl β-D-xylopyranoside, D-galactose, D-165 fructose, D-mannose, L-sorbose, L-rhamnose, dulcitol, inositol, D-sorbitol, methyl α-D-166 167 mannopyranoside, methyl α-D-glucopyranoside, N-acetylglucosamine, amygdalin, arbutin, esculin ferric citrate, D-cellobiose, D-melibiose, D-saccharose, D-trehalose, inulin, D-168 melezitose, D-raffinose, starch, glycogen, xylitol, gentiobiose, D-lyxose, D-tagatose, L-169 170 fucose, D-arabitol, L-arabitol, potassium gluconate and potassium 5-ketogluconate. Negative reactions were observed for D-glucose, D-lactose, D-maltose, D-mannitol, D-turanose, D-171 xylose, salicin and sucrose. Indeed, the use of API ZYM strips revealed positive reactions for 172 173 esterase, leucine arylamidase, cystine arylamidase,  $\alpha$ -glucosidase and  $\alpha$ -galactosidase, while the following tests, such as alkaline phosphatase, esterase lipase, lipase (C14), valine 174 175 arylamidase, trypsin,  $\alpha$ -chymotrypsin, acid phosphatase,  $\beta$ -galactosidase,  $\beta$ -glucosidase, N-176 acetyl- $\beta$ -glucosaminidase,  $\alpha$ -mannosidase and  $\alpha$ -fucosidase were negative. Finally, the use of API 20A strips shows that strain Marseille-P4308<sup>T</sup> was positive for gelatin and esculin ferric 177 citrate, while it was negative for tryptophan, urea, D-glucose, D-mannitol, D-lactose, D-178 179 sucrose, D-maltose, salicin, and D-xylose. The comparison of the main phenotypic and chemical differences between strain Marseille-P4308<sup>T</sup> and its phylogenetically related species 180 is given in Table 1. 181 Antimicrobial susceptibility testing of strain Marseille-P4308<sup>T</sup> showed the following minimal 182 inhibitory concentrations (in parenthesis) for the following antibiotics: ceftazidime (0.75 183 184 μg/mL), trimethoprim (0.25 μg/mL), daptomycin (0.016 μg/mL), tobramycin (8 μg/mL), benzylpenicillin (5 µg/mL), ciprofloxacin (1 µg/mL), ceftriaxone (0.5 µg/mL), linezolid 185

(0.094 μg/mL), ciprofloxacin (0.75 μg/mL), clindamycin (0.064 μg/mL) and amikacin (16 186 187  $\mu g/mL$ ). Genome properties and comparison 188 Strain Marseille-P4308<sup>T</sup> has a genome size of 2,145,294 bp, assembled into 29 contigs with 189 30.4 mol% G+C content. Its genome possesses 2,095 predicted genes, of which 2,031 are 190 protein-coding genes and 64 code for RNAs (9 rRNA genes, 53 tRNA genes and 2 tmRNA 191 gene). Of the 2,031 protein-coding genes, 1,442 were assigned to COG functional categories. 192 193 The genomic structure of the strain is represented by a circular map showing the coding regions of the genome (supplementary Figure S3). The genome of strain Marseille-P4308<sup>T</sup> 194 was compared with eight other genomes belonging to closely-related species in the 195 Peptostreptococcaceae family (Table 2). Among the Peptostreptococcus species, strain 196 Marseille-P4308<sup>T</sup> has the second largest genome (2.14 Mbp), shorter only than the genome of 197 Peptostreptococcus anaerobius ATCC 27337<sup>T</sup> (2.25 Mbp). The distribution of genes among 198 the different COG categories is almost the same proportion in all the genomes of the 199 200 compared *Peptostreptococcus* species (supplementary Figure S4). The DDH values obtained after genomic analysis vary for strain Marseille-P4308<sup>T</sup> from 201 18.9% with Asaccharospora irregularis DSM 2635<sup>T</sup> to 23.7% with Clostridioides mangenotii 202 DSM 1289<sup>T</sup>. Among species within *Peptostreptococcus* genus, DDH values ranged from 203 19.5%, between P. faecalis Marseille-P4308<sup>T</sup> and P. canis CCUG 57081<sup>T</sup>, to 27.9% between 204 P. stomatis W2278<sup>T</sup> and P. anaerobius ATCC 27337<sup>T</sup> (Table 3). These values are therefore 205 lower than the recommended threshold value of 70% for predicting a new prokaryote species 206 [19,24]. However, on this basis, strain Marseille-P4308<sup>T</sup> is considered to be a new species in 207 the genus *Peptostreptococcus*. In addition, among the *Peptostreptococcus* species studied, the 208 lowest OrthoANI value was 71.16% (between P. faecalis Marseille-P4308<sup>T</sup> and P. anaerobius 209 ATCC 27337<sup>T</sup> or P. stomatis W2278<sup>T</sup>) while 73.25% (between P. canis CCUG 57081<sup>T</sup> and P. 210

russellii RT-10B<sup>T</sup>) was the highest OrthoANI value obtained in this analysis (Table 3). The 211 mean nucleotide identity (ANI) analysis based on the genomes of species close to the 212 Marseille-P4308<sup>T</sup> strain revealed genomic sequence similarities of less than 80%. Indeed, the 213 cut-off value of 95 ~ 96% was recommended to delineate the species barrier in 214 prokaryotes[25,26]. Therefore, our strain Marseille-P4308<sup>T</sup> sharing with its all other related 215 species is considered as a new bacterial species. 216 217 **Conclusion** Based on the results of phylogenetic, phenotypic and biochemical analyses, it appeared that 218 Peptostreptococcus faecalis sp. nov., strain Marseille-P4308<sup>T</sup>, is formally considered as a new 219 species within the genus *Peptostreptococcus*. The type strain of this new species is Marseille-220 P4803<sup>T</sup>. 221 Description of Peptostreptococcus faecalis sp. nov. 222 Peptostreptococcus faecalis (fae.ca'lis. N.L. fem. adj. faecalis of faeces). Bacterial colonies 223 of strain Marseille-P4308<sup>T</sup> are convex, smooth and regular with a mean diameter of 0.7 mm 224 on blood agar. Cells are non-motile and non-spore-forming. They are Gram-positive cocci 225 bacteria and are oxidase and catalase negative. Glycerol, erythritol, D-arabinose, potassium 226 gluconate, D-ribose, L-xylose, D-adonitol, methyl \( \beta - \D-xylopyranoside, \) D-galactose, D-227 fructose, D-mannose, L-sorbose, L-rhamnose, dulcitol, inositol, D-sorbitol, methyl α-D-228 229 mannopyranoside, methyl α-D-glucopyranoside, N-acetylglucosamine, amygdalin, arbutin, 230 esculin ferric citrate, D-cellobiose, D-melibiose, D-saccharose, D-trehalose, inulin, Dmelezitose, D-raffinose, starch, glycogen, xylitol, gentiobiose, D-lyxose, D-tagatose, L-231 232 fucose, D-arabitol, L-arabitol, potassium 5-ketogluconate, esterase, leucine arylamidase, cystine arylamidase,  $\alpha$ -galactosidase,  $\alpha$ -glucosidase and gelatin are fermented. The genome 233 size of strain Marseille-P4308<sup>T</sup> is about 2.14 Mbp with a 30.4 mol% of G+C content. The 16S 234 rRNA and genome sequences are deposited in the GenBank database under Accession 235 numbers LT960583 and OERU00000000, respectively. 236

237	The type strain Marseille-P4308 <sup>T</sup> (=CSUR P4308 = CECT 9960) was isolated from stool
238	sample from a healthy indigenous Congolese volunteer.
239	Author Contributions:
240	Conceptualization and supervision, C.I.L., P.E.F., F.F., and D.R. Methodology, C.I.L., J.A.,
241	O.M., and F.F.; Validation, C.I.L. and F.F; Formal analysis, F.M., C.I.L., R.Z., and G.M.N.;
242	Strain culture and collection, S.A.; Investigation, M.B. and F.M.; Writing-original draft
243	preparation, F.M., and R.Z.; Writing-review and editing, C.I.L.; P.E.F., J.A., O.M., F.F., and
244	D.R; Funding acquisition, F.F. All authors have read and accepted this version of the
245	manuscript.
246	Acknowledgements:
247	The authors thank Professor Henri-Joseph Parra, Doctor Bernard Davoust, and Doctor Cheikh
248	Sokhna for their help in setting up the fieldwork, Ludivine Brechard for sequencing the
249	genome and Aurelia Caputo for submitting the genomic sequence to GenBank. The authors
250	would like to thank the Hitachi Company for supplying the TM4000 Plus tabletop
251	microscope.
252	
253	Funding sources:
254	This study was supported by the Institut Hospitalo-Universitaire (IHU) Méditerranée
255	Infection, the National Research Agency under the "Investissements d'avenir" programme,
256	reference ANR-10-IAHU-03, the Region Provence-Alpes-Côte d'Azur and European ERDF
257	PRIMI funding.
258	
259	Conflicts of interest:

None to declare.

#### 261 **REFERENCES:**

- [1] D.A. Murdoch, Gram-Positive Anaerobic Cocci, Clin Microbiol Rev. 11 (1998) 81–120. 262
- A. Slobodkin, The Family *Peptostreptococcaceae*, in: E. Rosenberg, E.F. DeLong, S. 263 Lory, E. Stackebrandt, F. Thompson (Eds.), The Prokaryotes: Firmicutes and 264

265 Tenericutes, Springer, Berlin, Heidelberg, 2014: pp. 291–302. https://doi.org/10.1007/978-3-642-30120-9 217. 266

- J.J. Berman, The Magnitude and Diversity of Infectious Diseases, in: Taxonomic Guide 267 to Infectious Diseases, Elsevier, 2012: pp. 3–5. https://doi.org/10.1016/B978-0-12-268 415895-5.00001-5. 269
- D.A. Murdoch, K.A. Young, J.T. Magee, Description of Three New Species of the 270 Genus Peptostreptococcus from Human Clinical Specimens: Peptostreptococcus harei 271 272 sp. nov., Peptostreptococcus ivoni sp. nov., and Peptostreptococcus octavius sp. nov., (1997) 7.273
- J. Downes, W.G. Wade, *Peptostreptococcus stomatis* sp. nov., isolated from the human 274 oral cavity, Int J Syst Evol Microbiol. 56 (2006) 751–754. 275 https://doi.org/10.1099/ijs.0.64041-0. 276
- Y.-L. Chen, S.-H. Tsai, K.-C. Hsu, C.-S. Chen, C.-W. Hsu, Primary sternal osteomyelitis 277 278 due to Peptostreptococcus anaerobius, Infection. 40 (2012) 195–197. https://doi.org/10.1007/s15010-011-0171-z. 279
- I. Bolivar, K. Whiteson, B. Stadelmann, D. Baratti-Mayer, Y. Gizard, A. Mombelli, D. 280 281 Pittet, J. Schrenzel, Geneva Study Group on Noma (GESNOMA), Bacterial diversity in oral samples of children in niger with acute noma, acute necrotizing gingivitis, and 282 healthy controls, PLoS Negl Trop Dis. 6 (2012) e1556. 283 284 https://doi.org/10.1371/journal.pntd.0001556.
- T.R. Whitehead, M.A. Cotta, E. Falsen, E. Moore, P.A. Lawson, *Peptostreptococcus* 285 russellii sp. nov., isolated from a swine-manure storage pit, Int J Syst Evol Microbiol. 61 286 (2011) 1875–1879. https://doi.org/10.1099/ijs.0.023762-0. 287
- [9] J.-C. Lagier, S. Khelaifia, M.T. Alou, S. Ndongo, N. Dione, P. Hugon, A. Caputo, F. 288 Cadoret, S.I. Traore, E.H. Seck, G. Dubourg, G. Durand, G. Mourembou, E. Guilhot, A. 289 Togo, S. Bellali, D. Bachar, N. Cassir, F. Bittar, J. Delerce, M. Mailhe, D. Ricaboni, M. 290 Bilen, N.P.M. Dangui Nieko, N.M. Dia Badiane, C. Valles, D. Mouelhi, K. Diop, M. 291 Million, D. Musso, J. Abrahão, E.I. Azhar, F. Bibi, M. Yasir, A. Diallo, C. Sokhna, F. 292 Diossou, V. Vitton, C. Robert, J.M. Rolain, B. La Scola, P.-E. Fournier, A. Levasseur, 293
- 294 D. Raoult, Culture of previously uncultured members of the human gut microbiota by culturomics, Nat Microbiol. 1 (2016) 16203. 295
- https://doi.org/10.1038/nmicrobiol.2016.203. 296
- [10] I. Pagnier, O. Croce, C. Robert, D. Raoult, B. La Scola, Non-contiguous finished 297 genome sequence and description of Anaerococcus provenciensis sp. nov., Stand 298 299 Genomic Sci. 9 (2014) 1198–1210. https://doi.org/10.4056/sigs.5501035.
- [11] E. Angelakis, D. Bachar, M. Yasir, D. Musso, F. Djossou, B. Gaborit, S. Brah, A. Diallo, 300 G.M. Ndombe, O. Mediannikov, C. Robert, E.I. Azhar, F. Bibi, N.S. Nsana, H.-J. Parra, 301 J. Akiana, C. Sokhna, B. Davoust, A. Dutour, D. Raoult, *Treponema* species enrich the 302 303 gut microbiota of traditional rural populations but are absent from urban individuals,

New Microbes New Infect. 27 (2018) 14-21. 304

- https://doi.org/10.1016/j.nmni.2018.10.009. 305
- [12] J.-C. Lagier, P. Hugon, S. Khelaifia, P.-E. Fournier, B. La Scola, D. Raoult, The Rebirth 306 307 of Culture in Microbiology through the Example of Culturomics To Study Human Gut Microbiota, Clin Microbiol Rev. 28 (2015) 237-264. 308
- https://doi.org/10.1128/CMR.00014-14. 309

- 131 P. Seng, M. Drancourt, F. Gouriet, B. La Scola, P.-E. Fournier, J.M. Rolain, D. Raoult, Ongoing revolution in bacteriology: routine identification of bacteria by matrix-assisted laser desorption ionization time-of-flight mass spectrometry, Clin Infect Dis. 49 (2009) 543–551. https://doi.org/10.1086/600885.
- 314 [14] M. Drancourt, C. Bollet, A. Carlioz, R. Martelin, J.P. Gayral, D. Raoult, 16S ribosomal 315 DNA sequence analysis of a large collection of environmental and clinical unidentifiable 316 bacterial isolates, J. Clin. Microbiol. 38 (2000) 3623–3630.
- 317 [15] S. Kumar, G. Stecher, M. Li, C. Knyaz, K. Tamura, MEGA X: Molecular Evolutionary 318 Genetics Analysis across Computing Platforms, Mol. Biol. Evol. 35 (2018) 1547–1549. 319 https://doi.org/10.1093/molbev/msy096.
- 320 [16] T. Seemann, Prokka: rapid prokaryotic genome annotation, Bioinformatics. 30 (2014) 2068–2069. https://doi.org/10.1093/bioinformatics/btu153.
- [17] D. Hyatt, G.-L. Chen, P.F. LoCascio, M.L. Land, F.W. Larimer, L.J. Hauser, Prodigal:
  prokaryotic gene recognition and translation initiation site identification, BMC
  Bioinformatics. 11 (2010) 119. https://doi.org/10.1186/1471-2105-11-119.
- [18] D. Laslett, ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences, Nucleic Acids Research. 32 (2004) 11–16.
  https://doi.org/10.1093/nar/gkh152.
- [19] A.F. Auch, M. von Jan, H.-P. Klenk, M. Göker, Digital DNA-DNA hybridization for microbial species delineation by means of genome-to-genome sequence comparison,
  Standards in Genomic Sciences. 2 (2010) 117–134. https://doi.org/10.4056/sigs.531120.
- [20] J.P. Meier-Kolthoff, A.F. Auch, H.-P. Klenk, M. Göker, Genome sequence-based
  species delimitation with confidence intervals and improved distance functions, BMC
  Bioinformatics. 14 (2013) 60. https://doi.org/10.1186/1471-2105-14-60.
- [21] I. Lee, Y. Ouk Kim, S.-C. Park, J. Chun, OrthoANI: An improved algorithm and software for calculating average nucleotide identity, International Journal of Systematic and Evolutionary Microbiology. 66 (2016) 1100–1103.
  https://doi.org/10.1099/ijsem.0.000760.
- 338 [22] R.L. Tatusov, M.Y. Galperin, D.A. Natale, E.V. Koonin, The COG database: a tool for genome-scale analysis of protein functions and evolution, Nucleic Acids Res. 28 (2000) 33–36. https://doi.org/10.1093/nar/28.1.33.
- 341 [23] H.S. Sader, A.C. Pignatari, E test: a novel technique for antimicrobial susceptibility testing, Sao Paulo Med J. 112 (1994) 635–638. https://doi.org/10.1590/s1516-31801994000400003.
- [24] M. Kim, H.-S. Oh, S.-C. Park, J. Chun, Towards a taxonomic coherence between
  average nucleotide identity and 16S rRNA gene sequence similarity for species
  demarcation of prokaryotes, Int J Syst Evol Microbiol. 64 (2014) 346–351.
  https://doi.org/10.1099/ijs.0.059774-0.
- 348 [25] M. Richter, R. Rosselló-Móra, Shifting the genomic gold standard for the prokaryotic 349 species definition, PNAS. 106 (2009) 19126–19131. 350 https://doi.org/10.1073/pnas.0906412106.
- [26] J. Chun, A. Oren, A. Ventosa, H. Christensen, D.R. Arahal, M.S. da Costa, A.P. Rooney,
  H. Yi, X.-W. Xu, S. De Meyer, M.E. Trujillo, Proposed minimal standards for the use of
  genome data for the taxonomy of prokaryotes, Int J Syst Evol Microbiol. 68 (2018) 461–
  466. https://doi.org/10.1099/ijsem.0.002516.
- [27] P.A. Lawson, C.N. Johnson, L. Bengtsson, G. Charalampakis, G. Dahlén, E. Moore, E.
  Falsen, *Peptostreptococcus canis* sp. nov., isolated from subgingival plaque from canine oral cavity, Anaerobe. 18 (2012) 597–601.
- 358 https://doi.org/10.1016/j.anaerobe.2012.10.008.

- [28] J. Gerritsen, S. Fuentes, W. Grievink, L. van Niftrik, B.J. Tindall, H.M. Timmerman,
  G.T. Rijkers, H. Smidt, Characterization of Romboutsia ilealis gen. nov., sp. nov.,
  isolated from the gastro-intestinal tract of a rat, and proposal for the reclassification of
  five closely related members of the genus *Clostridium* into the genera *Romboutsia* gen.
  nov., *Intestinibacter* gen. nov., *Terrisporobacter* gen. nov. and *Asaccharospora* gen.
  nov., Int J Syst Evol Microbiol. 64 (2014) 1600–1616.
  https://doi.org/10.1099/ijs.0.059543-0.
- [29] N. Saitou, M. Nei, The neighbor-joining method: a new method for reconstructing phylogenetic trees, Mol Biol Evol. 4 (1987) 406–425.
  https://doi.org/10.1093/oxfordjournals.molbev.a040454.
- [30] S.V. Angiuoli, S.L. Salzberg, Mugsy: fast multiple alignment of closely related whole
  genomes, Bioinformatics. 27 (2011) 334–342.
  https://doi.org/10.1093/bioinformatics/btq665.
- [31] I. Letunic, P. Bork, Interactive Tree Of Life (iTOL) v5: an online tool for phylogenetic
  tree display and annotation, Nucleic Acids Research. 49 (2021) W293–W296.
  https://doi.org/10.1093/nar/gkab301.

**Table 1.** Differential characteristics of Marseille-P4308<sup>T</sup>, *Peptostreptococcus stomatis* W2278 <sup>T</sup> [5], *Peptostreptococcus canis* CCUG 57081<sup>T</sup> [27], and *Peptostreptococcus anaerobius* ATCC 27337<sup>T</sup> [28].

Characteristics	Marseille- P4308 <sup>T</sup>	W2278 <sup>T</sup>	CCUG 57081 <sup>T</sup>	ATCC 27337 <sup>T</sup>	
	Strictly	Strictly	Facultative	Strictly	
Oxygen requirement	anaerobic	anaerobic	anaerobic	anaerobic	
Acid production					
from:					
D-Cellobiose	+	_	_	+	
α-Galactosidase	+	+	_	+	
D-Glucose	_	+	_	+	
α-Glucosidase	+	+	NP	+	
D-Lactose	_	_	_	+	
D-Mannose	+	_	+	NP	
D-Raffinose	+	_	_	NP	
Sucrose	_	_	_	+	
G+C content (mol%)	30.4	36	30.8	34-36	
Habitat	Human	Human	Dog	Cat	

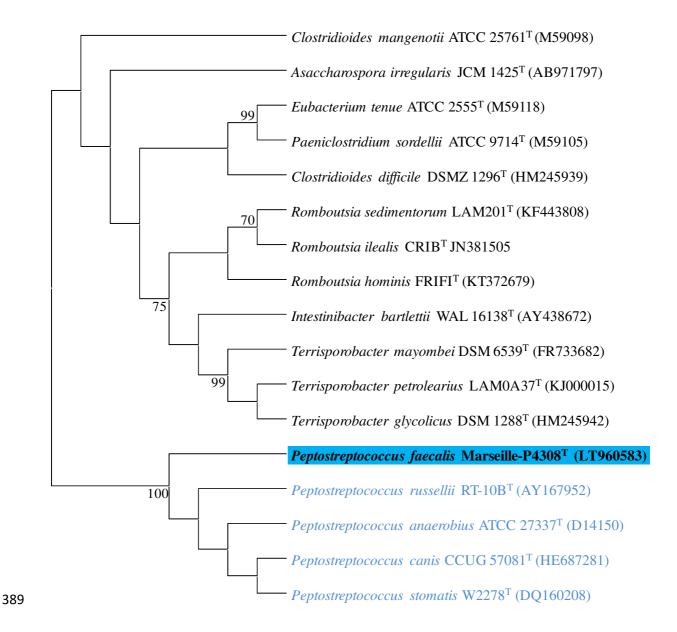
NP, not performed.

**Table 2.** Comparison of the size, the content of G+C mol% and the number of proteins of the genome of *Peptostreptococcus faecalis* sp. nov., strain Marseille-P4308<sup>T</sup> with the other genomes of related species.

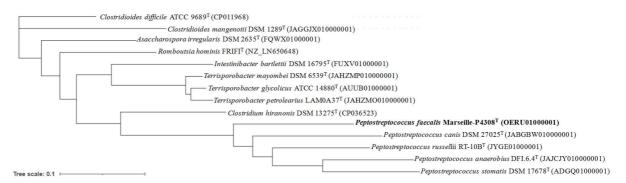
Species	Strain Number	Genbank accession	Size (bp)	G+C mol%	Nb of proteins
Peptostreptococcus faecalis	Marseille-P4308 <sup>T</sup>	OERU00000000	2,145,294	30.4	2,031
Peptostreptococcus anaerobius	ATCC 27337 <sup>T</sup>	UGTB00000000	2,256,756	35.7	2,070
Peptostreptococcus canis	CCUG 57081 <sup>T</sup>	JABGBW000000000	2,064,240	30.2	1,834
Peptostreptococcus russellii	$RT-10B^T$	JYGE00000000	2,082,949	30.9	1,756
Peptostreptococcus stomatis	$W2278^{T}$	ADGQ00000000	1,988,044	36.6	1,799

**Table 3.** Average nucleotide identity (ANI) and dDDH values (%) obtained by pairwise comparison of the nine studied genomes. Compared genomes: 1, *Peptostreptococcus faecalis* Marseille-P4308<sup>T</sup>; 2, *Peptostreptococcus anaerobius* ATCC 27337<sup>T</sup>; 3, *Peptostreptococcus canis* CCUG 57081<sup>T</sup>; 4, *Peptostreptococcus russellii* RT-10B<sup>T</sup>; 5, *Peptostreptococcus stomatis* W2278<sup>T</sup>; 6, *Asaccharospora irregularis* DSM 2635<sup>T</sup>; 7, *Clostridioides difficile* DSM 1296<sup>T</sup>; 8, *Clostridioides mangenotii* DSM 1289<sup>T</sup>; 9, *Clostridium hiranonis* DSM 13275<sup>T</sup>. OrthoANI values are shown on right bottom (in bolt) and dDDH values calculated using GGDC formula 2 software (DDH estimates based on HSP identities / length) shown on upper left. The empty boxes between them are 100%.

Strains	Marseille-P4308 <sup>T</sup>	ATCC 27337 <sup>T</sup>	CCUG 57081 <sup>T</sup>	RT-10B <sup>T</sup>	W2278 <sup>T</sup>	DSM 2635 <sup>T</sup>	DSM 1296 <sup>T</sup>	DSM 1289 <sup>T</sup>	DSM 13275 <sup>T</sup>
Marseille-P4308 <sup>T</sup>		22.6%	19.5%	21.2%	20.7%	18.9%	19.3%	23.7%	23.1%
ATCC 27337 <sup>T</sup>	71.16%		24.0%	22.5%	27.9%	20.4%	21.3%	28.3%	26.0%
CCUG 57081 <sup>T</sup>	71.17%	71.68%		21.3%	25.1%	18.9%	18.8%	25.8%	25.7%
$RT-10B^T$	73.25%	72.26%	73.83%		23.3%	25.7%	19.4%	27.1%	22.6%
$W2278^{T}$	71.16%	73.83%	71.63%	72.29%		19.4%	21.2%	28.4%	27.8%
DSM 2635 <sup>T</sup>	69.14%	68.03%	68.85%	70.06%	67.83%		21.1%	20.3%	20.7%
DSM 1296 <sup>T</sup>	69.16%	68.00%	69.31%	69.40%	67.60%	75.83%		19.8%	21.6%
DSM 1289 <sup>T</sup>	68.82%	68.28%	68.58%	68.80%	67.50%	72.90%	73.53%		26.1%
DSM 13275 <sup>T</sup>	69.71%	69.54%	69.13%	69.66%	68.38%	72.03%	71.95%	71.65%	



**Figure 1.** Phylogenetic tree with the 16S rRNA gene sequences indicating the position of *Peptostreptococcus faecalis* sp. nov. Marseille-P4308<sup>T</sup> among other closely related species. Sequences were aligned using MUSCLE, and phylogenetic inferences were obtained using the Neighbor-Joining method [29] within the MEGA X software [15]. Accession numbers are indicated in parenthesis.



**Figure 2.** Phylogenomic tree basing on genomic sequences of *Peptostreptococcus faecalis* sp. nov., Marseille-P4308<sup>T</sup> and other its related species. Sequences were concatenated with union EMBOSS (https://www.bioinformatics.nl/cgi-bin/emboss/union) and aligned by Mugsy software [30]. The tree was built using iTOL an online bioinformatic tool [31].