

# Genomic features of an isolate of Empedobacter falsenii harbouring a novel variant of metallo- $\beta$ -lactamase, blaEBR-4 gene

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Ahmed Olowo-Okere, Yakubu Kokori Enevene Ibrahim, Busayo Olalekan Olayinka, Yahaya Mohammed, Larbi Zakaria Nabti, et al.. Genomic features of an isolate of Empedobacter falsenii harbouring a novel variant of metallo- $\beta$ -lactamase, blaEBR-4 gene. Infection, Genetics and Evolution, 2022, 98, pp.105234. 10.1016/j.meegid.2022.105234. hal-03684605

# HAL Id: hal-03684605 https://amu.hal.science/hal-03684605v1

Submitted on 22 Jul 2024

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Version of Record: https://www.sciencedirect.com/science/article/pii/S1567134822000314 Manuscript\_7035f76d1db61797ffc9326d0b32482d

## 1 Genomic features of an isolate of *Empedobacter falsenii* harbouring a novel variant of

# 2 metallo- $\beta$ -lactamase, *bla*<sub>EBR-4</sub> gene

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#### 23 Abstract

Empedobacter falsenii is an emerging opportunistic pathogen that has been 24 occasionally implicated in various human infections. In this study, we described the genomic 25 26 features of a multidrug resistant E. falsenii Q1655 obtained from a patient attending a public hospital in Sokoto, northwest Nigeria. The isolate, E. falsenii Q1655, was isolated from the 27 stool sample of a patient in Sokoto, Nigeria. The identity of the isolate was confirmed by 28 MALDITOF-MS. The disc diffusion test and modified Carba-NP test were used for 29 phenotypic antibiotic susceptibility test and carbapenemase enzyme production test, 30 31 respectively. The whole genome of the strain was sequenced using the Illumina MiSeq technique. Resistome analysis was done by annotation of the WGS against the ARG-ANNOT 32 database. The isolate was resistant to all  $\beta$ -lactam antibiotics with the exception of cefepime. 33 34 The MICs of imipenem and ertapenem as determined by E-test were 12  $\mu$ g/ml and 2  $\mu$ g/ml, respectively. Modified Carba NP test showed that the strain was carbapenemase producing. 35 Resistome analysis revealed the presence of a novel metallo- $\beta$ -lactamase, a chromosomal 36 *bla*<sub>EBR-4</sub> (Accession No: MN997121), which exhibited 94.92 % and 97.02 % nucleotide and 37 protein sequence identities respectively with *bla*<sub>EBR-3</sub> gene of *E. falsenii* 174820. Seven and 38 39 eight amino-acid substitutions were observed with the  $bla_{EBR-1}$  and  $bla_{EBR-2}$ , respectively. We reported the first isolation and genomic description of an extensively drug resistant isolate of 40 41 Empedobacter falsenii in Nigeria. This report broadens our knowledge of carbapenem 42 resistance in *E. falsenii* and it will serve as a useful guide in the development of antibiotic use policy. 43

44 **Keywords:** *Empedobacter falsenii*, metallo-β-lactamase, *bla*<sub>EBR-4</sub>, *tet*X

#### 45 Introduction

Empedobacter falsenii (formally Wautersiella falsenii) is a member of the family 46 Flavobacteriaceae [1,2]. The bacteria species was first isolated and described in 2006 in 47 Belgium by Kampfe et al. [3]. The bacterium is an emerging opportunistic, non-fermentative 48 Gram-negative pathogen that has been occasionally implicated in various human infections 49 50 [4]. The strain has been previously isolated from the respiratory tract of an immunocompromised patient and also from the urine of an infant with pyelonephritis, among 51 others [5,6]. The increasing identification of this strain from varying human clinical samples 52 is facilitated by improvement in the clinical microbiology laboratory capacity including the 53 use of quick and reliable matrix-assisted laser desorption ionization time-off-light mass 54 spectrometry [7,8]. 55

56 Metallo- $\beta$ -lactamases (MBLs) are bacterial enzymes which catalyse the formation of a noncovalent reactive complex with  $\beta$ -lactam rings through a hydrolytic reaction facilitated by 57 one or two essential divalent cations, typically zinc, at their active site [9,10]. The reaction 58 results in the formation of microbiologically inactive molecule that is incapable of forming an 59 enzymatically productive complex with a penicillin binding protein (PBP), ultimately leading 60 61 to loss of activity of  $\beta$ -lactam antibiotics [10]. Several MBLs have been described but the most commonly encountered are the Verona integron encoded metallo-β-lactamase (VIM), 62 Imipenemase (IMP) and New-Delhi Metallo-β-lactamase (NDM) (Bonomo & Szabo, 2006). 63 Other MBL are confined to a specific bacteria species such as *bla*<sub>EBR</sub> identified in 64 Empedobacter spp. or to a particular geographical location such as SPM-1 (Sao Paulo MBL), 65 GIM-1 (German imipenemase); SIM-1 (Seoul imipenemase); TMB-1 (Tripoli MBL) [11]. 66 This is likely due to the localisation of the corresponding genes on narrow-host- range 67 plasmids or chromosomes, which makes wide diffusion unlikely [12]. 68

Although *E. falsenii* is not a common cause of infection in hospitalized patients, its genome is replete with genes mediating resistance to several antibiotics classes particularly cephalosporin and carbapenems [7,13]. Strain of *E. falsenii* resistant to colistin has also been reported [14]. This makes this bacterium a pathogen of concern as it may serve as reservoir for dissemination of antibiotic resistant to genes to clinically important bacteria. In this study, we described the genomic features of a multidrug resistant isolate of *E. falsenii* obtained from a patient attending a public hospital in Sokoto, northwest Nigeria.

76

#### 77 Methods

78 The approval to conduct this study was granted by the Sokoto State Ministry of 79 Health (Approval number: SMH/1580/VIV) and the Research and Ethics Committee of the hospital (Approval number: UDUTH/HREC/2019/770. In April 2019, a strain of E. falsenii 80 Q1655 was isolated from the stool sample of a 54 years old male patient admitted into the 81 medical ward of a tertiary healthcare facility in Sokoto, Nigeria as part of antibiotics 82 resistance surveillance. The identity of the isolates was confirmed by the matrix assisted laser 83 84 desorption ionisation time of flight mass spectrometry (MALDITOF) following a previously described protocol [8]. The disc diffusion test and modified Carba-NP test were used for 85 phenotypic detection of antibiotic susceptibility and production of carbapenemase enzyme, 86 respectively [15,16]. The result of antibiotic susceptibility test was interpreted using the 87 EUCAST breakpoints for non-Enterobacteriaceae. Detection of genes encoding ESBLs 88 (*bla*<sub>CTX-M</sub>, *bla*<sub>TEM</sub>, *bla*<sub>SHV</sub>) and carbapenemases (*bla*<sub>OXA-48</sub>, *bla*<sub>VIM</sub>, *bla*<sub>IMP</sub>, *bla*<sub>NDM</sub> and *bla*<sub>KPC</sub>) 89 90 was done using the real-time PCR (qPCR) as previously described [17]. The whole genome of the strain was thereafter sequenced using the MiSeq technique (Illumina Inc., San Diego, 91 CA, USA). The reads obtained were assembled using Spades software and annotated by the 92

Prokka bacterial genome annotation pipeline [18,19]. The Plasmid incompatibility (*Inc*) groups and resistome analysis were determined *in-silico* using the PlasmidFinder 2.0 platform and annotation of the whole genome sequence (WGS) against antibiotic resistance gene annotation (ARG-ANNOT) database, respectively [20,21]. Alignment of the nucleotide sequences and amino acid sequences of the  $\beta$ -lactamase gene and its variants was done using the Network Protein Sequence Analysis server [22].

99

#### 100 Results and Discussion

The antibiotic susceptibility assay revealed that the strain was resistant to several 101 antibiotics including penicillins (amoxicillin, amoxicillin-clavulanate), second generation 102 103 cephalosporin (cephalothin), third-generation cephalosporin (ceftriaxone) and carbapenems (imipenem and ertapenem) with the exception of cefepime (Table 1). The MIC of imipenem 104 and ertapenem were respectively 12 µg/ml and 2 µg/ml. Phenotypic modified Carba NP test 105 conducted according to the previously published protocol showed that the strain was 106 carbapenemase producing. The PCR investigation of resistance mechanism revealed the 107 108 absence of common acquire β-lactamase genes (i.e. *bla*<sub>SHV</sub>, *bla*<sub>TEM</sub>, *bla*<sub>CTX-M</sub>, *bla*<sub>NDM</sub>. *bla*<sub>OXA-</sub> 48, *bla*<sub>VIM</sub>, *bla*<sub>KPC</sub>, *bla*<sub>IMP</sub>). This was what prompted the genomic investigation of the strain. 109

110 The genome assembly results to a chromosome size of 3,482,659-bp with a G+C 111 content of 34.4 % (Table 1). Resistome analysis of the WGS of *E. falsenii* Q1655 against 112 ARG-ANNOT database revealed the presence of a novel metallo- $\beta$ -lactamase type 2, a 113 chromosomal *bla*<sub>EBR-4</sub> which exhibited 94.92 % and 97.02 % nucleotide and protein identity 114 with *bla*<sub>EBR-3</sub> gene of *E. falsenii* 174820 (Accession No: NG\_066507). Pairwise alignment of 115 the amino acid sequence revealed that the *bla*<sub>EBR-4</sub> differs from *bla*<sub>EBR-3</sub> by only 6 amino-acids 116 (fig. 1). Seven and eight amino-acid substitutions were observed with the *bla*<sub>EBR-1</sub> of 117 Empedobacter brevis (Accession No: WP\_063860216) and blaEBR-2 of E. falsenii (Accession No: WP\_038333643), respectively. The various mutations were expected to affect the 118 biological activity of the enzyme. Surprisingly, the enzyme retains its carbapenem 119 120 hydrolysing activity as demonstrated by its resistance to carbapenem and positive Carba NP test result. The absence of plasmid in the genome of the strain indicates that the bla<sub>EBR-4</sub> is 121 chromosomal. This is in line with a previous description of the gene [23]. The  $bla_{EBR}$  family 122 of metallo-\beta-lactamase enzyme, efficiently catalyzes the hydrolysis of all penicillin, 123 carbapenems and cephalosporin with a notable exception of cefepime [24]. So far, all 124 125 detected *bla*<sub>EBR</sub> genes have been detected in the genus *Empedobacter*. With the exception of bla<sub>EBR-1</sub> which was detected in *E. brevis*, the rest were all detected in *E. falsenii* [23,24]. The 126 phylogenetic tree showing the relationship of the  $bla_{EBR-4}$  with other  $\beta$ -lactamase genes 127 128 downloaded from the gene bank is presented in Figure 2. In addition, the genome also contains tet(X2) gene, a tetracycline-modifying enzyme involved in enzymatic inactivation of 129 all generations of tetracycline including tigecycline [25]. 130

The resistance of the strain to other antibiotics such as fosfomycin, nitrofurantoin, trimethoprim/sulphamethoxazole may be due to non-permeability of its outer membrane similar to other non-fermentative bacteria [26,27]. Similar to a previously published report, the strain was resistant to colistin [14]. *Empedobacter falsenii* has been suggested to be naturally resistant to colistin [13].

## 136 Conclusion

We reported the first isolation and genomic description of an extensively drug
resistant isolate of *Empedobacter falsenii* in Nigeria. The isolate was resistant to several
medically important antibiotics including third generation cephalosporin, carbapenems,
fosfomycin and colistin and harbours a novel variant of metallo-β-lactamase, *bla*<sub>EBR-4</sub>. This

- 141 report broadens our knowledge of carbapenem resistance in *E. falsenii* and it will serve as a
- 142 useful baseline for understanding resistance mechanism in this strain in the future.

143

#### Sequences accession number:

The Whole Genome Shotgun project of *E. falsenii* Q1655 has been deposited at DDBJ/ENA/GenBank under the accession JAAGKM000000000. The isolate information, sequencing metrics, and genomic data of the strain are listed in Table 1.

# 147 **Funding information:**

This work was supported by the French Government under the Investments for the
Future program managed by the National Agency for Research, Méditerranée Infection, 10IAHU-03.

# 151 Acknowledgement

We sincerely thank Dr Linda Hadjadj for the technical support given to us during thecourse of this study and for deposition of the WGS in NCBI database.

# 154 **Conflict of interest:**

155 We declare no conflicts of interest.

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# **Table 1:** Genetic and phenotypic features of *Empedobacter falsenii* Q1655 strain

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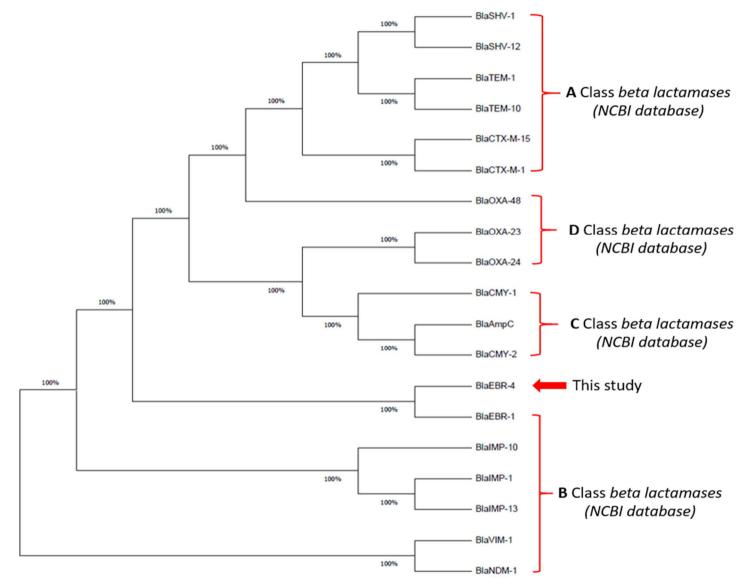
Strain name	Genome size (bp)	% GC content	Antibiotic resistance genes	<b>Resistance phenotype</b>	Susceptible phenotype		
E. falsenii Q1655	3,482,659	34.4	$bla_{\rm EBR-4}$ and $tet({\rm X})$	Amoxicillin, amoxicillin/clavulanate, cefalotin, ceftriaxone, ertapenem, imipenem, fosfomycin, nitrofurantoin, trimethoprim/sulphamethoxazole, colistin	Ciprofloxacin, gentamicin, doxycycline, amikacin and cefepime		

	10	20	30	40	50 I	60	70			
WP_063860216_ <i>E.brevis</i> _EBR_1	I MKKLFSLIALIGSFA	 \FGQIKPIQI	I D <mark>PINNNLFV</mark> Y	QTFNSFNGVE	I YNANGMYLVTN	ا KGIVLFDVP	I IQKSQYQELNDML	.QE		
WP_038333643_E.falsenii_EBR_2	MKKIFSLIALIGSFA	FGQIKPIQI	DPINNNLFVY	QTFNSFNDVE	YNANGMYLVTN	KGIVLFDVP	QKSQYQELNDIL	QE		
WP_150823468_E.falsenii_EBR_3	MKKIFSLIALIGSFA	FGQIKPIQI	D <mark>SINNNLFV</mark> Y	QTFNSFNGVE	YNANGMYLVTN	KGIVLFDVP	QKSQYQELNDML	.QE		
Q1655_01285_E.falsenii_EBR_4	MKKLFSLIALIGSFA	FGQIKPIQI	D <mark>PINNNLFV</mark> Y	QTFNSFNGVE	YNANGMYLVTN	KGIVLFDVP	QKSQYQELNDML	.QE		
	*************	*******	* *******	*********	********	********	*********	**		
	89 I	99 I	109	119	129 I	139	149 I			
WP_063860216_ <i>E.brevis</i> _EBR_1	KYNLPVIAVFATHSH	IDDRAGDLSF	YNELNIPTYA	TSLTNSKLKK	EGKATSKFEI	ELGKTYKFGNE	KFVFEYFGEGHT	SD		
WP_038333643_E.falsenii_EBR_2	KYNLPVIAVFATHSH	IDDRAGDLSF	YNELNIPTYA	TSLTNSKLKK	EGKATSKFEI	ELGKTYKFGNE	KFVVEYFGEGHT	SD		
WP_150823468_ <i>E.falsenii</i> _EBR_3	KYNLPVIAVFATHSH	IDDRAGDLSF	YNDLNIPTYA	TSLTNSKLKK	EGKATSKFEI	ELGKTYKFGNE	KFVVEYFGEGHT	SD		
Q1655_01285_E.falsenii_EBR_4	KYNLPVIAVFATHSH	IDDRAGDLSF	YNDLNIPTYA	TSLTNSKLKK	EGKATSKFEI	LGKTYKFGNE	KFVVEYFGEGHT	SD		
	***************************************									
	168 	178 	188 	198 	208 	218 	228 			
WP_063860216_E.brevis_EBR_1	NVVVWFPKYKVLNGG	GCLIKGADAV	NLGYTGEANV	VEWPKTVHKL	VAKHPTIKQVI	PGHDNWKATO	HIENTFKLLEKK	1		
WP_038333643_E.falsenii_EBR_2	NVVVWFPKYKVLNGGCLIKGADAVNLGYTGEANVVEWPKTVHKLVAKHPTIKQVIPGHDNWKATGHIENTFKLLEKK									
WP_150823468_ <i>E.falsenii</i> _EBR_3	NVVVWFPKYKVLNGGCLIKGADAVNLGYTGEANVAEWPKTVHKLVAKHPTIKQVIPGHDNWKATGHIENTFKLLEKK									

Q1655\_01285\_E.falsenii\_EBR\_4 NVVVWFPKYKVLNGGCLIKGADAVNLGYTGEANVTEWPKTVQKLVTKHPKIKQVIPGHDNWKVTGHIENTFKLLEKK

232

**Figure 1:** Protein sequence alignment of the novel metallo-β-lactamase EBR-4 protein and others ERB sequences.



**Figure 2:** Phylogenetic tree of the novel  $bla_{EBR-4}$  gene with other  $\beta$ -lactamase genes downloaded from the NCBI Genbank database.