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Highlighting the Dystonic Phenotype Related to GNAO1

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ABSTRACT: Background: Most reported patients carrying GNAO1 mutations showed a severe phenotype characterized by early-onset epileptic encephalopathy and/or chorea.

Objective: The aim was to characterize the clinical and genetic features of patients with mild GNAO1-related phenotype with prominent movement disorders.

Methods: We included patients diagnosed with GNAO1-related movement disorders of delayed onset (>2 years). Patients experiencing either severe or profound intellectual disability or early-onset epileptic encephalopathy were excluded.

Results: Twenty-four patients and 1 asymptomatic subject were included. All patients showed dystonia as prominent movement disorder. Dystonia was focal in 1, segmental in 6, multifocal in 4, and generalized in 13. Six patients showed adolescence or adulthoodonset dystonia. Seven patients presented with parkinsonism and 3 with myoclonus. Dysarthria was observed in 19 patients. Mild and moderate ID were present in 10 and 2 patients, respectively.

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Conclusion: We highlighted a mild GNAO1-related phenotype, including adolescent-onset dystonia, broadening the clinical spectrum of this condition. © 2022 The Authors. Movement Disorders published by Wiley Periodicals LLC on behalf of International Parkinson and Movement Disorder Society

Key Words: dystonia; GNAO1; phenotypes; mutation

GNAO1 mutations have been associated with two phenotypes: a severe, early-infantile epileptic encephalopathy with burst-suppression (EIEE17, OMIM 6154731) and a neurodevelopmental disorder with involuntary movements (NEDIM, OMIM 617493^{2-4}), with or without seizures. GNAO1 encodes the α-subunit of a heterotrimeric guanine nucleotide-binding protein (G_{co}) , which is widely expressed in the central nervous system, playing an important role in signal transduction through AMPc metabolism in the striatum.^{2,5,6} As the number of reports increased, it became evident that GNAO1-related encephalopathies encompass a continuous spectrum of neurological syndromes featuring variable association of movement disorders, psychomotor delay, intellectual disability (ID), and different types of epilepsy. 2.7 GNAO1-related movement disorder usually starts in infancy. Choreoathetosis is usually described with spontaneous or trigger-induced exacerbations, potentially leading to status dystonicus, as a hallmark of the disease.² Most patients reported so far showed a severe phenotype, with recurrent exacerbations and significant disability. However, in a few atypical, milder cases, with movement disorder onset in late childhood or adolescence, no acute exacerbation and less-severe disability have been identified using next-generation sequencing techniques. $8-10$ In this study, we characterized the clinical and genetic features of a cohort of patients with mild GNAO1-related phenotype characterized by prominent movement disorders, further expanding the spectrum of this condition.

Patients and Methods

Patients

Patients carrying causative heterozygous variants in GNAO1 and exhibiting mild phenotypes were included from 18 neurology and neuropediatric movement disorders reference centers from the United States, France, Israel, Switzerland, the United Kingdom, and Italy. Mild phenotype was defined by (1) lack of severe or profound ID, (2) lack of early-onset epileptic encephalopathy, (3) late-onset (ie, after age 2 years) appearance of movement disorders, and (4) acquisition of walk. Patients were

recruited through an international collaboration mediated by the online platform Genematcher.¹¹ All patients were assessed by neurologists or neuro-pediatricians with an expertise in movement disorders. Patients' phenotypes from family 6 and family 4, which were previously reported elsewhere, were added in the cohort as further clinical data were obtained.

Genetic Analysis

CGH-array, gene panel, exome, and genome sequencing were performed as previously reported.^{9,10,12-15} Detailed procedures of the sequencing, including library preparation and bioinformatic analysis, are available in Supplementary Data. Variants were considered as causative if they fulfilled the following criteria: (1) known disease mutation reported in ClinVar; (2) loss-offunction variant, including protein truncating variants, frameshift indel, large deletion, and splice site changes predicted to cause aberrant splicing; or (3) missense variant with a CADD score >20, absent in GnomAD and predicted to be deleterious by at least two additional algorithms (Polyphen-2, SIFT and Mutation taster). In addition, variant class of pathogenicity was reported according to the American College of Medical Genetics and Genomics (ACMG) guidelines.¹⁶

Ethics

All patients and relatives provided written informed consent before genetic analysis. Strasbourg University Hospital review board gave approval for the exome sequencing of families 4 and 6 that was performed in a research framework. Genetic analysis for other families was performed for diagnostic purposes.

Results

We included 24 patients (15 women) and 1 asymptomatic carrier from 20 different families. Patients' clinical characteristics are provided in Table 1. Mean age at inclusion was 23.8 years (range: 5–66), mean age at disease onset was 6.6 years (range: 0.25–47), and mean age of dystonia onset was 10.1 years (range: 2–47). Initial manifestations included dystonia in 10 (41%), myoclonus or seizure in 1, developmental delay in 13, language delay in 4, motor delay in 9, and hypotonia in 4 patients. Seven patients were from three unrelated families showing autosomal dominant inheritance, while all others were sporadic cases due to de novo mutations. Pedigrees of these three families and videos of patients are available in Supplementary Data.

Dystonia was the main movement disorder in all patients, prominently affecting multiple segments of the upper body part in 21 patients or being limited to the cervical segment in 1 patient. Dystonia was isolated, namely not associated to any other symptom, in

(Continues)

TABLE 1 Continued TABLE 1 Continued

Abbreviations: htz: heterozygous, NA, not avaliable; BFMDRS: Burke Falm Marsden Dystomia Rating Scale, dystomia score; GPi-DBS: globus pallidus internal deep brain strimulation; MDD, major depressive disorder; ADHD, attent Abbreviations: htz. heterozygous, NA, not available; BFMDRS: Burke Falm Marsden Dystonia Rating Scale, dystonia score; GFi-DBS: globus pallidus internal deep brain stimulation; MDD, major depressive disorder; ADHD, attenti disorder; ASD, autism spectrum disorder; ID, intellectual disability.

7 patients (29%). Dystonia was the only movement disorder in 14 patients and was combined with other movement disorders in 10, namely myoclonus in 3, chorea in 2, and parkinsonism in 7 (with 2 patients combining three movement disorders). Only 3 patients presented an acute exacerbation of dystonia. Dystonia course was nonprogressive for 11 patients. Dystonia topography was generalized in 13 patients (54%), multifocal in 4 patients, segmental in 6 patients, and focal in 1 patient. Dystonia was associated with dysarthria/anarthria in 19 patients. Early-onset hypotonia preceded dystonia in 11 patients. Dystonia was associated with ID in 12 patients (mild for 10 and moderate for 2). Seizures occurred in 3 patients between age 4 and 19 years. Magnetic resonance imaging was unremarkable for all patients.

Movement disorders response to medication, including anticholinergic drugs, levodopa, tetrabenazine, amantadine, clonazepam, or methylphenidate, was variable. Six patients received pallidal deep brain stimulation (DBS), with significant improvement for 5 of them. Detailed treatment outcomes are available in Supplementary Data.

Mutations carried by the patients are presented in Table 1. Details regarding pathogenicity assessment are available in Supplementary Data. Apart from the p.R206Q, all variants were classified as pathogenic (class V) according to the ACMG criteria. In family 4, we identified 3 patients with the R206Q variant, which was classified as a variant of uncertain significance due to the presence of an unaffected carrier, 4-D, son of 4-C, despite meeting other criteria of pathogenicity (absent from GnomAD, unanimously predicted damaging by in silico tools, affecting a highly conserved residue located in a hot spot without benign variation) (criteria PM1, PM2, PP2, and PP3). A recurring splicing variant $(c.724-8G > A)$, previously reported in ClinVar, was identified in 8 patients (33%) showing late-onset and/or segmental dystonia. Previous report showed this variant to damage the natural splice acceptor site and create a stronger cryptic splice acceptor site in intron 6, resulting in the insertion of two amino acids leading to protein mislocation.¹⁷ Two patients were carrying a nonsense variant (p.N256*), and one was carrying a

FIG. 1. Impact of the mutations on the protein. (A) Position of the variant sites on the heterotrimeric complex containing the G α subunit. The heterotrimer is depicted in the resting state (GDP-bound, PDBcode 1GG2). Subunits α, β, and γ are colored in green, cyan, and gray, respectively. Affected residues in this cohort are in red, and their position is indicated both on the human Gαo1 and on rat Gαi1 (UniProtKB ID P10824, between brackets). GDP-binding residues are colored in yellow. Previously reported GNAO1 variants are in blue. On the right, a focused view of the GDP-binding site is shown. (B) Cartoon model of the heterotrimeric-αβγ G-protein coupled-receptor on the synaptic cleft. (C) Schematic representation of the disease-causing variants on GNAO1 transcript (NM_020988.3) and protein (UniprotKB ID P09471-1). The amino acids impacted by the mutations identified in this work are in red, whereas previously reported variants are in black. The blue bar on the transcript indicates the translated region. The blue segments in the protein sequence indicate the G-motifs (containing the nucleotide binding residues)—numbered from 1 to 5. Molecular graphics are realized with UCSF Chimera ([http://www.rbvi.ucsf.edu/chimera\)](http://www.rbvi.ucsf.edu/chimera), developed by the Resource for Biocomputing, Visualization, and Informatics at the University of California, San Francisco, with support from NIH P41-GM103311. The cartoon has been created with [BioRender.com.](http://biorender.com) aa, amino acids; nt, nucleotides; UTR, untranslated region. [Color figure can be viewed at [wileyonlinelibrary.com\]](http://wileyonlinelibrary.com)

large deletion encompassing GNAO1. Other mutations (p.L23P; p.K46R; p.R179G; p.R206Q; p.E208N; p.C215Y; and p.N242T and p.E246V) were all missense variants absent from GnomAD. The previously reported pathogenic p.C215Y variant 8 was found in three unrelated families. All variants were close to known mutational hot spots (Fig. 1).

Discussion

Here, we report a large cohort of patients with mild GNAO1-related phenotypes, experiencing prominent movement disorders without severe chronic encephalopathy. The typical phenotype was a nonprogressive generalized or focal/segmental upper-body dystonia appearing beyond infancy, associated with dysarthria. Acute exacerbation occurred only in 3 patients, and 29% of patients showed isolated dystonia without additional neurological manifestation. Our inclusion criteria were able to identify these phenotypes that were in contrast with most of the previously reported patients with GNAO1-related movement disorders, who showed severe hyperkinetic encephalopathy with recurrent dystonic exacerbations,^{18,19} and profound developmental delay^{3,7} with or without epilepsy in the first year of life.^{1,7}

Dystonia distribution was segmental or focal in 7 patients, and clinical course was nonprogressive in 11 patients, while most of the previously reported patients with GNAO1-related movement disorders had generalized and rapidly progressive dystonia.² Dystonia topography revealed prominent upper-body distribution in most of our patients, reminiscent of the clinical pictures associated with other dystonia-related genes, such as $GNAL^{20,21}$ or $ANO3$. $22,23$ Seven patients also exhibited mild parkinsonism, which is consistent with the role of $G_{\alpha\alpha}$ in the signal transduction within the striatal projection neurons downstream of the dopamine receptors. $6,24$

We identified 3 autosomal dominant families where multiple symptomatic relatives carried heterozygous variants, which was in contrast with all the previously reported patients who showed de novo mutations.² One p.R206Q carrier did not present any clinical sign evocative of GNAO1-related disorders. The similarities between this family's phenotype and the other cases all showing upper-body distribution—argue for the implication of the variant, while no other class IV to V variant in a dystonia-related gene was identified. In addition, a family member carrying this variant had disease onset in his 40s, meaning the 30-year-old asymptomatic carrier could be potentially presymptomatic. Future identification of autosomal dominant family with GNAO1-related dystonia and follow-up of this patient might confirm whether incomplete penetrance is possible in GNAO1-related disorders.

Response to medication was variable in our cohort. No significant response to levodopa was identified in our cohort, but 3 patients had partial response to anticholinergic drugs, which was in accordance with previous findings from the literature.^{2,4,25} Conversely, the outcome</sup> was good in 5 of 6 patients who received DBS, further confirming its efficacy in GNAO1-related dystonia.²⁶

Most of the variants identified in the present work were not reported among previously published cases showing severe phenotype, and two variants recurred in multiple families, suggesting that mild phenotypes could be related to specific mutations. However, the variants we identified were close to previously reported hot spots (Fig. 1), leading to amino-acid substitution in the same functional domains. Further studies are needed to elucidate if these different variants have a milder impact on protein function. In addition, we identified two putative loss-of-function variants (a nonsense variant and a whole-gene deletion), presumably affecting protein expression and possibly causing GNAO1 haploinsufficiency. All 3 carriers were presenting latechildhood/adolescence onset dystonia without ID. Thus far, no report described the phenotype of patients harboring GNAO1-nonsense variants. A few patients with chromosome 16q deletions encompassing GNAO1 have been described, all harboring significantly larger deletions compared to our case and showing variable associations of dysmorphisms, microcephaly, seizures, and developmental delay.²⁷ Although previous research suggested that loss-of-function variants were mainly responsible for epileptic encephalopathy while gain-offunction mutations were mostly associated with a movement disorders prominent phenotype, $\frac{5}{3}$ recent evidence suggests that pathogenic variants exert their effect through a combination of dominant-negative and loss-of-function mechanisms, and each mutation likely produces circuit-selective effects through a peculiar mechanism of signaling disruption.²⁸ The expanding spectrum of associated phenotypes and disease-causing variants provides further evidence that genotype– phenotype correlations are nuanced, and GNAO1 related disorders shape a continuous spectrum of overlapping phenotypes rather than distinct entities.²⁸ Our study carries some limitations, including the retrospective design and the lack of formal assessment in several cases. Here, we highlighted the milder GNAO1-related phenotypes, broadening this condition-clinical spectrum. GNAO1 mutations should be considered as a cause of adolescent or adult-onset nonprogressive dystonia, particularly in the presence of a speech involvement even in the absence of seizures or ID.

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Data Availability Statement

Anonymized data pertaining to the research presented will be made available upon reasonable request from external investigators.

References

- 1. Nakamura K, Kodera H, Akita T, Shiina M, Kato M, Hoshino H, et al. De novo mutations in GNAO1, encoding a Gαo subunit of heterotrimeric G proteins, cause epileptic encephalopathy. Am J Hum Genet 2013;93(3):496-505.
- 2. Schirinzi T, Garone G, Travaglini L, Vasco G, Galosi S, Rios L, et al. Phenomenology and clinical course of movement disorder in GNAO1 variants: results from an analytical review. Parkinsonism Relat Disord 2019;61:19–25.
- 3. Kulkarni N, Tang S, Bhardwaj R, Bernes S, Grebe TA. Progressive movement disorder in brothers carrying a GNAO1 mutation responsive to deep brain stimulation. J Child Neurol 2016;31(2):211–214.
- 4. Danti FR, Galosi S, Romani M, Montomoli M, Carss KJ, Raymond FL, et al. GNAO1 encephalopathy: broadening the phenotype and evaluating treatment and outcome. Neurol Genet 2017;3(2):e143
- 5. Feng H, Khalil S, Neubig RR, Sidiropoulos C. A mechanistic review on GNAO1-associated movement disorder. Neurobiol Dis 2018; 116:131–141.
- 6. Delorme C, Giron C, Bendetowicz D, Méneret A, Mariani L-L, Roze E. Current challenges in the pathophysiology, diagnosis, and treatment of paroxysmal movement disorders. Expert Rev Neurother 2021;21(1):81–97.
- 7. Kelly M, Park M, Mihalek I, Rochtus A, Gramm M, Pérez-Palma E, et al. Spectrum of neurodevelopmental disease associated with the GNAO1 guanosine triphosphate-binding region. Epilepsia 2019; 60(3):406–418.
- 8. Carecchio M, Invernizzi F, Gonzàlez-Latapi P, Panteghini C, Zorzi G, Romito L, et al. Frequency and phenotypic spectrum of KMT2B dystonia in childhood: a single-center cohort study. Mov Disord 2019;34(10):1516–1527.
- 9. Graziola F, Garone G, Stregapede F, Bosco L, Vigevano F, Curatolo P, et al. Diagnostic yield of a targeted next-generation sequencing gene panel for pediatric-onset movement disorders: a 3-year cohort study. Front Genet 2019;10:1026
- 10. Wirth T, Tranchant C, Drouot N, Keren B, Mignot C, Cif L, et al. Increased diagnostic yield in complex dystonia through exome sequencing. Parkinsonism Relat Disord 2020;74:50–56.
- 11. Sobreira N, Schiettecatte F, Valle D, Hamosh A. GeneMatcher: a matching tool for connecting investigators with an interest in the same gene. Hum Mutat 2015;36(10):928-930.
- 12. Guillen Sacoto MJ, Tchasovnikarova IA, Torti E, Forster C, Andrew EH, Anselm I, et al. De novo variants in the ATPase module of MORC2 cause a neurodevelopmental disorder with growth retardation and variable craniofacial Dysmorphism. Am J Hum Genet 2020;107(2):352–363.
- 13. Valence S, Cochet E, Rougeot C, Garel C, Chantot-Bastaraud S, Lainey E, et al. Exome sequencing in congenital ataxia identifies two new candidate genes and highlights a pathophysiological link between some congenital ataxias and early infantile epileptic encephalopathies. Genet Med Off J Am Coll Med Genet 2019;21(3): 553–563.
- 14. Cif L, Demailly D, Lin J-P, Barwick KE, Sa M, Abela L, et al. KMT2B-related disorders: expansion of the phenotypic spectrum and long-term efficacy of deep brain stimulation. Brain J Neurol 2020;143(11):3242–3261.
- 15. Smol T, Thuillier C, Boudry-Labis E, Dieux-Coeslier A, Duban-Bedu B, Caumes R, et al. Neurodevelopmental phenotype associated with CHD8-SUPT16H duplication. Neurogenetics 2020;21(1):67–72.
- 16. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med Off J Am Coll Med Genet 2015;17(5):405–424.
- 17. Miyamoto S, Nakashima M, Fukumura S, Kumada S, Saitsu H. An intronic GNAO1 variant leading to in-frame insertion cause movement disorder controlled by deep brain stimulation. Neurogenetics 2022;23(2):129-35.
- 18. Ananth AL, Robichaux-Viehoever A, Kim Y-M, Hanson-Kahn A, Cox R, Enns GM, et al. Clinical course of six children with GNAO1 mutations causing a severe and distinctive movement disorder. Pediatr Neurol 2016;59:81–84.
- 19. Honey CM, Malhotra AK, Tarailo-Graovac M, van Karnebeek CDM, Horvath G, Sulistyanto A. GNAO1 mutationinduced pediatric dystonic storm rescue with Pallidal deep brain stimulation. J Child Neurol 2018;33(6):413–416.
- 20. Fuchs T, Saunders-Pullman R, Masuho I, Luciano MS, Raymond D, Factor S, et al. Mutations in GNAL cause primary torsion dystonia. Nat Genet 2013;45(1):88–92.
- 21. Geoghegan AR, Al Hussona M, Beauchamp NJ, Hutchinson M, Sean O'Riordan MB, Lynch T, et al. A novel GNAL mutation in familial dystonia presenting with childhood tremor and myoclonus. Mov Disord Off J Mov Disord Soc 2019;34(6):923–924.
- 22. Charlesworth G, Plagnol V, Holmström KM, Bras J, Sheerin U-M, Preza E, et al. Mutations in ANO3 cause dominant Craniocervical dystonia: Ion Channel implicated in pathogenesis. Am J Hum Genet 2012;91(6):1041–1050.
- 23. Lange LM, Junker J, Loens S, Baumann H, Olschewski L, Schaake S, et al. Genotype-phenotype relations for isolated dystonia genes: MDSGene systematic review. Mov Disord Off J Mov Disord Soc 2021;36(5):1086–1103.
- 24. Corvol J-C, Muriel M-P, Valjent E, Féger J, Hanoun N, Girault J-A, et al. Persistent increase in olfactory type G-protein alpha subunit levels may underlie D1 receptor functional hypersensitivity in Parkinson disease. J Neurosci Off J Soc Neurosci 2004;24(31): 7007–7014.
- 25. Yamashita Y, Ogawa T, Ogaki K, Kamo H, Sukigara T, Kitahara E, et al. Neuroimaging evaluation and successful treatment by using directional deep brain stimulation and levodopa in a patient with GNAO1-associated movement disorder: a case report. J Neurol Sci 2020;411:116710
- 26. Koy A, Cirak S, Gonzalez V, Becker K, Roujeau T, Milesi C, et al. Deep brain stimulation is effective in pediatric patients with GNAO1 associated severe hyperkinesia. J Neurol Sci 2018;15(391):31–39.
- 27. Apuzzo D, Cappuccio G, Vaisanen T, Alagia M, Pignataro P, Genesio R, et al. Two cases of 16q12.1q21 deletions and refinement of the critical region. Eur J Med Genet 2020;63(6):103878
- 28. Muntean BS, Masuho I, Dao M, Sutton LP, Zucca S, Iwamoto H, et al. Gαo is a major determinant of cAMP signaling in the pathophysiology of movement disorders. Cell Rep 2021;34(5):108718

Supporting Data

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.