



HAL
open science

Editorial: Venoms, animal and microbial toxins, volume II

Zhijian Cao, Delavar Shahbazzadeh, Hervé O Kovacic, Patrick Michael Mcnutt, Jing-Lin Wang, Heike Wulff, Yuri Utkin, Jean-Marc Sabatier

► To cite this version:

Zhijian Cao, Delavar Shahbazzadeh, Hervé O Kovacic, Patrick Michael Mcnutt, Jing-Lin Wang, et al.. Editorial: Venoms, animal and microbial toxins, volume II. *Frontiers in Pharmacology*, 2022, 13, pp.973628. 10.3389/fphar.2022.973628 . hal-03981943

HAL Id: hal-03981943

<https://amu.hal.science/hal-03981943v1>

Submitted on 26 Oct 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



OPEN ACCESS

EDITED AND REVIEWED BY
Diana Conte Camerino,
University of Bari Aldo Moro, Italy

*CORRESPONDENCE
Jean-Marc Sabatier,
sabatier.jm1@libertysurf.fr

SPECIALTY SECTION
This article was submitted to
Pharmacology of Ion Channels and
Channelopathies,
a section of the journal
Frontiers in Pharmacology

RECEIVED 20 June 2022
ACCEPTED 06 July 2022
PUBLISHED 19 August 2022

CITATION
Cao Z, Shahbazzadeh D, Kovacic H,
McNutt PM, Wang J-L, Wulff H, Utkin Y
and Sabatier J-M (2022), Editorial:
Venoms, animal and microbial toxins,
volume II.
Front. Pharmacol. 13:973628.
doi: 10.3389/fphar.2022.973628

COPYRIGHT
© 2022 Cao, Shahbazzadeh, Kovacic,
McNutt, Wang, Wulff, Utkin and Sabatier.
This is an open-access article
distributed under the terms of the
[Creative Commons Attribution License
\(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or
reproduction in other forums is
permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original
publication in this journal is cited, in
accordance with accepted academic
practice. No use, distribution or
reproduction is permitted which does
not comply with these terms.

Editorial: Venoms, animal and microbial toxins, volume II

Zhijian Cao¹, Delavar Shahbazzadeh², Hervé Kovacic³,
Patrick Michael McNutt⁴, Jing-Lin Wang⁵, Heike Wulff⁶,
Yuri Utkin⁷ and Jean-Marc Sabatier^{3*}

¹College of Life Sciences, Wuhan University, Wuhan, China, ²Pasteur Institute of Iran (PII), Tehran, Iran, ³UMR7051 Institut de NeuroPhysiopathologie, Aix-Marseille Université, Marseille, France, ⁴Wake Forest Institute for Regenerative Medicine, Wake Forest School of Medicine, Winston-Salem, NC, United States, ⁵Beijing Institute of Microbiology and Epidemiology, Beijing, China, ⁶Department of Pharmacology, University of California, Davis, Davis, CA, United States, ⁷Laboratory of Molecular Toxinology, Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry RAS, Moscow, Russia

KEYWORDS

venom, animal toxin, plant toxin, bacterial toxin, viral toxin, structure

Editorial on the Research Topic

Venoms, animal and microbial toxins, volume II

Our Research Topic titled *Venoms, animal and microbial toxins, volume II* is centered on the characteristics of animal, plant, microbial toxins, and their molecular/cellular targets. It also addresses whole animal venoms, which contain a complex mixture of diverse toxins. Venomous animals and microbes are the natural sources of many toxins. These toxins vary in size, nature, and mode of action. They mainly act on ion channels, enzymes, receptors, and neurotransmitter release to produce an acute pathophysiological effect. Animal venom and microbial molecules behave as candidate therapeutics or biological weapons because of their unique potencies, rapid mode of action and wide range of bioactivities.

This Research Topic compiles twenty research and review articles to describe venoms and toxins (or derivatives), through an “in-depth” analysis of their structure, pharmacology, synergistic effects, and structure-function relationship.

Several articles are focused on the structural and/or functional characterization of animal and microbial toxins, as well as other animal venom compounds. A first article by Ullah *et al.* describes the three-dimensional (3-D) structure of the exfoliative toxin D (ETD) from the pathogenic bacterium *Staphylococcus aureus* responsible for skin disorders. The authors predicted the 3-D structure of ETD using optimized molecular modeling techniques, and compared it to those of four other known exfoliative toxins (A, B, C and E). The authors then used the predicted ETD structure for *in silico* docking simulations of natural and synthetic inhibitors, which is important to inform the design of new potent inhibitors to treat staphylococcal scalded skin syndrome. The article by Peng *et al.* reports on the isolation and characterization of δ -theraphotoxin-Gr4b, a novel toxin (37-residue peptide crosslinked by 3 disulfide bridges) from the venom of the spider *Grammostola*

rosea. Interestingly, this toxin inhibits the fast inactivation of the $\text{Na}_v1.9$ channel through a unique mechanism: binding to domains III and IV voltage sensors. Since $\text{Na}_v1.9$ plays a key role in pain perception, these findings may inform a therapeutic development of novel analgesics. In another article, [Dongol et al.](#) screened venoms from Australian theraphosid spider species against the human pain target $\text{hNa}_v1.7$ and isolated Ssp1a, a 33-residue peptide with an “inhibitor cystine knot” motif. Functional characterization of recombinant Ssp1a on neuronal hNav subtypes suggested Ssp1a interacts with the voltage-sensing domain II of $\text{hNa}_v1.7$ to trap the channel in a resting state. [George et al.](#) investigated the effects of synthetic peptide toxins predicted from Arizona bark scorpion venom on the voltage-gated sodium channel $\text{Na}_v1.8$, another channel involved in pain signaling. The authors found the peptide NaTx36 inhibited sodium current recorded from a recombinant grasshopper mouse $\text{Na}_v1.8$ channel ($\text{OtNa}_v1.8$) via interactions with the domain I voltage sensor and domain II pore module. [Ho et al.](#) described the structural and functional properties of an α -conotoxin OmIA isolated from *Conus omaria* venom. OmIA behaves as a potent antagonist of $\alpha 7$ nicotinic acetylcholine receptors (nAChRs). The authors analyzed a co-crystal structure of OmIA with the acetylcholine binding protein of gastropod *Lymnae stagnalis*, and highlighted key amino acid residues responsible for the high potency of OmIA against these receptors. [Huynh et al.](#) characterized α -elapitoxin-Oh3a, a 72-residue post-synaptic neurotoxin derived from venom of the Asian *Ophiophagus hannah* king cobra. This toxin, which causes neuromuscular paralysis, inhibited the contractile responses of tissues to exogenous carbachol and acetylcholine in chick *Biventer cervicis* nerve-muscle samples. King cobra antivenom prevented α -elapitoxin-Oh3a-induced neurotoxicity *in vitro*, suggesting an active role for α -elapitoxin-Oh3a during king cobra intoxication. [Yu et al.](#) focused on *Nemopilema nomurai* jellyfish envenomation, a threat to humans in Asian waterways. Metalloproteinases are reportedly the main toxic components of the venom, causing inflammation and damage. The authors isolated and characterized high proteinase activity fractions in tentacle autolysis using complementary purification techniques. Unexpectedly, these fractions did not show any proteinase activity except when mixed together, highlighting some synergistic effects among components of the fractions. [Elrayess et al.](#) characterized Smp24 and Smp43, two “novel” cationic antimicrobial peptides (AMPs) from the venom of the scorpion *Scorpio maurus palmatus*. The authors examined peptide-induced cytotoxicity on various cell lines, including leukaemia and non-cancer cells. The peptides were found to alter the viability of all cells tested (although HaCaT human skin keratinocytes were less sensitive). This decreased cell viability was accompanied by a selective up-regulation of the caspase-1 gene (except for HaCaT cells), whereas all tested cells showed an increase in downstream interleukin-1 β expression.

These data suggested scorpion venom AMPs activate pyroptosis, a highly inflammatory signaling cascade leading to a lytic programmed cell death.

Other articles of the Research Topic are focused on the structural characteristics, mode of action, targets and fields of application of toxins, venom components or whole venoms. For example, [Wu et al.](#) characterized FM-CATH, a “novel” cathelicidin from the *Fejervarya multistriata* paddy frog skin. FM-CATH has potent antimicrobial properties against both bacteria and fungi. It binds to lipopolysaccharides and lipoteichoic acid, and induces agglutination of bacteria. It also alters enzymatic activities (plasmin, thrombin, tissue plasminogen activator, β -tryptase) thus inhibiting the coagulation process both *in vitro* and *in vivo*. FM-CATH increased survival of septic mice, suggesting it might be of value in the treatment of sepsis. [Wang et al.](#) studied PcActx, a toxic peptide from the zoantharian *Palythoa caribaeorum* with potential inhibitory activity on the transient receptor potential cation channel subfamily V member 1 (TRPV1). TRPV1 conducts Ca^{2+} , is widely expressed in sensory neurons but is also expressed in epileptic brain areas and is thought to be a potential target to prevent epileptic seizures. At non-toxic doses, PcActx peptides (reduced and folded oxidized forms) reversed pentylentetrazol-induced seizure-related behavior in zebrafish larvae by limiting the production of reactive oxygen species (ROS) and modulating the expression of genes involved in Ca^{2+} and GABA-glutamate signaling. The authors conclude PcActx is a potential novel treatment for epilepsy. Another study by [Wu et al.](#) focused on Cath-MH, an AMP from the skin of the frog *Microhyla heymonsivogt*. The authors investigated the antimicrobial potential of Cath-MH on *Propionibacterium acnes*. Cath-MH had bacteriocidal effects through membrane disruption. *In vivo*, Cath-MH showed agglutination activity, and reduced edema and infiltration of inflammatory cells in an acne mouse model. Cath-MH also suppressed bacterial growth and production of pro-inflammatory cytokines *in vivo*, suggesting it might be useful to manage acne vulgaris and related skin disorders. [Gutierrez et al.](#) examined the protective effects of the phospholipase A2 inhibitor Varespladib on the deleterious actions of the venom from the southern American bushmaster *Lachesis muta rhombeata* pit viper snake. The enzymatic, coagulant and hemorrhagic activities of venom were studied in the presence of varespladib (with or without addition of a commercial antivenom). Varespladib potentially antagonized PLA2 activity preventing venom-induced coagulation, whereas it had little or no effects on esterase, caseinolytic, and hemorrhagic activities. [Eisele et al.](#) studied the binary C2 toxin of the highly pathogenic *Clostridium botulinum* bacteria. The binary C2 toxin consists of two proteins: C2I (enzyme) and C2II (binding/transport) subunits. To exert toxic effects on mammal cells, C2II needs to be proteolytically cleaved to the pore-forming subunit C2IIa. The authors

demonstrate that C2IIa reduces the chemotactic translocation of human neutrophils (polymorphonuclear leukocytes), thus have potential to down-modulate the excessive and deleterious recruitment of neutrophils into organs after trauma. Barros et al. reviews bioactive peptides and alkaloids identified in skin secretions of Urodele amphibians, which include antimicrobials, antioxidants, immune system modulators, vasoactive and coagulation-acting compounds and which could serve as “new” scaffolds for drug design.

Another set of articles of the Research Topic are focused on the potential treatment of envenomation using antibodies or other compounds. Johnston et al. introduced the “Australian Snakebite Project” (ASP-24), and detailed the epidemiology and clinical presentation of Australian sea snake envenoming. The efficacies of antivenoms in preventing myotoxicity and neurotoxicity are presented. The morbidity and mortality related to sea snake envenoming are discussed and it is concluded that early antivenom treatment after host envenomation is a key therapeutic intervention to prevent severe myotoxicity and death. Hmaidi et al. described the molecular basis of the interaction between the cardiotoxic α -neurotoxin AahII (scorpion *Androctonus australis hector*) and the (cardiac) voltage-gated $\text{Na}_v1.5$ channel. The authors showed that AahII slows the fast inactivation of $\text{Na}_v1.5$ channels expressed in HEK293 cells. A highly neutralizing anti-AahII nanobody (previously produced Nb10) was shown to fully reverse the effects of AahII on the kinetics of channel inactivation. Computer-aided docking experiments suggest that at AahII molecule, Nb10 might bind to the same binding sites as $\text{Na}_v1.5$. Sachetto et al. studied the neutralization of toxins in venom from the lethal Brazilian pit viper *Bothrops jararaca*. The bioflavonoids rutin and its water-soluble derivative rutin succinate were used to assess their protective potential against the snake venom in *in vitro* and *in vivo* assays (*in vivo*, mice were injected with venom, or venom preincubated with rutin or rutin succinate). The data indicated that both flavonoids prevent venom-induced lethality through multiple mechanisms (e.g., coagulation, metalloproteinases). Interestingly, rutin, and rutin succinate showed different modes of action on homeostasis, which would deserve a more detailed analysis of the structure-activity relationships. A similar approach was followed by Heber et al. using ambroxol to neutralize exotoxins TcdA and TcdB from the enterobacterium *Clostridioides difficile*. TcdA and TcdB are the main virulence factors produced by the bacterium and the cause of *C. difficile* associated diseases (CDAD). To exert their toxic effects, the two exotoxins are internalized into the cells via receptor-mediated endocytosis. Translocation of exotoxins from endosomal vesicles into the cytosol requires the acidification of endosomes; ambroxol prevents such acidification. The authors therefore examined the potential protective effects of ambroxol on TcdA- and TcdB-induced cytotoxicity. Ambroxol was found to inhibit key (exotoxin-induced) events (i.e., endosome acidification, morphological changes, glycosylation of Rac1), whereas it also

unexpectedly decreases the intracellular enzyme activity of exotoxins. Ambroxol thus behaves as a candidate therapeutic against CDAD. Silva et al. discussed translational concerns regarding the use of rodent lethality models to evaluate antivenoms for human envenoming. To illustrate this problem, it was shown that human nicotinic acetylcholine receptors (nAChRs) have an exceptionally low affinity for the short-chain α -neurotoxins compared to long-chain α -neurotoxins, while both types of α -neurotoxins bind to mouse nAChRs with high affinity. The authors pointed out that the effects of purified toxins or animal venoms on natural prey species are likely to be different from the effects on non-prey species, including humans.

A final article by Hirschenberger et al. describes CRISPA, a transient and non-viral technique to deliver Cas9 endonuclease into specific cells. The strategy developed by the authors is based on the translocation machinery of the *Bacillus anthracis* anthrax toxin, PA (protective antigen). The PA transporter, which normally mediates the entry of anthrax lethal factor and edema factor into the cells, might be optimized for a cell-type specific delivery of Cas9. Therefore, CRISPA potentially represents a step forward, in the translation of the CRISP/Cas9 genome editing technology into clinics.

Taken together, Volume II of this Research Topic contributes to a better understanding of venoms, venom compounds and toxins (or derivatives) opening the way to new exciting research and discoveries. We do believe that these articles exploring a particularly complex world, will inspire researchers and clinicians worldwide.

Author contributions

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.