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Review

Montelukast and Acute Coronary Syndrome: The Endowed Drug

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Abstract: Acute coronary syndrome (ACS) is a set of signs and symptoms caused by a reduction of coronary blood flow with subsequent myocardial ischemia. ACS is associated with activation of the leukotriene (LT) pathway with subsequent releases of various LTs, including LTB₄, LTC₄, and LTD₄, which cause inflammatory changes and induction of immunothrombosis. LTs through cysteine leukotriene (CysLT) induce activation of platelets and clotting factors with succeeding coronary thrombosis. CysLT receptor (CysLTR) antagonists such as montelukast (MK) may reduce the risk of the development of ACS and associated complications through suppression of the activation of platelet and clotting factors. Thus, this critical review aimed to elucidate the possible protective role of MK in the management of ACS. The LT pathway is implicated in the pathogenesis of atherosclerosis, cardiac hypertrophy, and heart failure. Inhibition of the LT pathway and CysL1TR by MK might be effective in preventing cardiovascular complications. MK could be an effective novel therapy in the management of ACS through inhibition of pro-inflammatory CysLT1R and modulation of inflammatory signaling pathways. MK can attenuate thrombotic events by inhibiting platelet activation and clotting factors that are activated during the development of ACS. In conclusion, MK could be an effective agent in reducing the severity of ACS and associated complications. Experimental, preclinical, and clinical studies are recommended to confirm the potential therapeutic of MK in the management of ACS.

Keywords: acute coronary syndrome; leukotriene; montelukast; antileukotrienes therapy

1. Introduction

Acute coronary syndrome (ACS) is a set of signs and symptoms caused by a reduction of coronary blood flow with subsequent myocardial ischemia [1]. According to the electrocardiographic (ECG) changes and duration of symptoms, ACS is divided into unstable angina, which represents 38%; ST-elevation myocardial infarction (STEMI) representing 30%, and non-ST-elevation myocardial infarction (NSTEMI) representing 25%. Unstable angina of ACS is differentiated from stable angina, as it occurs suddenly at rest with little response to treatment [2].

The cardinal clinical features of ACS are chest pain, dyspnea, nausea, vomiting, sweating, and tachycardia [3]. ACS is mainly caused by coronary thrombosis, coronary vasospasm, aortic valve stenosis, severe anemia, and pulmonary hypertension [4]. The underlying pathogenic mechanisms of ACS are related to rupture of atheroma in 60% and atheroma erosion in 30%. Most of the atheroma rupture induces the development of STEMI, while the development of atheroma erosion induces propagation of NSTEMI [5].

Plaque rupture provokes *in situ* inflammatory reactions and thrombosis with subsequent ischemic-reperfusion injury (IRI). Thrombolytic therapy is indicated for STEMI, while nitroglycerin, antiplatelet, low-molecular-weight heparin (LMWH) such as enoxaparin, and heparin-like drugs such as fondaparinux are effective in this state [6,7].

It has been reported that leukotriene receptor (LT) receptor antagonists such as montelukast (MK) may have potential beneficial effects in the cardiovascular field [8]. Thus, this critical review aimed to elucidate the possible protective role of MK in the management of ACS.

2. Leukotriene Pathway

The LT name was initially described in 1979 by Swedish Bengt Samuelsson as a slow-reaction, smooth-muscle-stimulating substance that was introduced by Feldberg and Kellaway between 1938 and 1940 [9]. LTs are inflammatory eicosanoid mediators generated by leukocytes through oxidation of arachidonic acid (AA) and eicosapentaenoic acid (EPA) by the action of 5-lipoxygenase (5-LO) [9]. LTs use lipid signaling to express and convey information in autocrine and paracrine patterns to control immune functions. Usually, the production of LTs is correlated with the release of prostaglandins and histamines [10]. By the action of 5LO or 5LO-activating protein (FLAP), the AA is converted to LTA₄, which is converted through LTA₄ hydrolase and LTC₄ synthase to LTB₄ and cysteinyleukotriene (CysLT), respectively. LTB₄ is inhibited by LY-293111 a specific inhibitor of the LTB₄ receptor, while MK, pranlukast, and zafirlukast block CysLT receptor (CysLTR) type 1 [11] (Figure 1).

Thus, there are two types of LTs, namely LTB₄ and CysLT, which include LTC₄, LTE₄, and LTD₄. CysLTs are involved in the development of allergy and anaphylaxis. Specifically, LTB₄ induces recruitment of neutrophils with induction of tissue injury and release of pro-inflammatory cytokines. Therefore, LTB₄ antagonists inhibit the development and propagation of neutrophil-mediated disorders [12,13]. LTA₄ is mainly converted to LTB₄ and has few inflammatory effects. LTC₄ is mainly expressed by eosinophil and mast cells, which are converted outside the cells to form LTD₄ and LTE₄, which have inflammatory effects. Moreover, LTG₄ is regarded as an active metabolite of LTE₄ that has unknown biological effects [14,15].

There are two main types of CysLT receptors, which are CysLTR1 and CysLTR2. CysLTR1 is a G-protein coupled receptor that is mainly activated by LTD₄ and LTC₄, causing airway bronchoconstriction and increasing vascular permeability, the influx of neutrophils and eosinophils, fibrosis and collagen deposition, and membrane hypertrophy of respiratory epithelium [16]. Additionally, activation of CysLTR1 is associated with disruption of the blood–brain barrier (BBB) with induction of neuroinflammation and convulsion. Therefore, CysLTR1 antagonists could be effective against the development of Parkinson's disease, Alzheimer's disease, encephalomyelitis, and multiple sclerosis [17]. CysLTR1 is inhibited by MK, pranlukast, and zafirlukast [17]. CysLTR2 is activated by LTD₄ and LTC₄, and it is involved in the development of hypersensitivity and allergic reactions [18,19]. CysLTR2 is chiefly expressed in the platelets, eosinophils, macrophages, and mast cells. Furthermore, CysLTR2 is highly expressed in the heart, adrenal gland, endothelial cells, smooth muscle cells, and airway epithelial cells [18]. MK and other CysLTs only block CysLTR1; however, gemilukast inhibits both CysLTR1 and CysLTR2, which can be effective in the management of asthma [20]. Other types of LT receptors such as GPR99, which is called oxoglutarate or CysLTR3, are stimulated by LTE₄ and α -ketoglutarate, while

GPR99 is inhibited by MK [21]. Overexpression of GPR99 is linked with the development of hypertension [21].

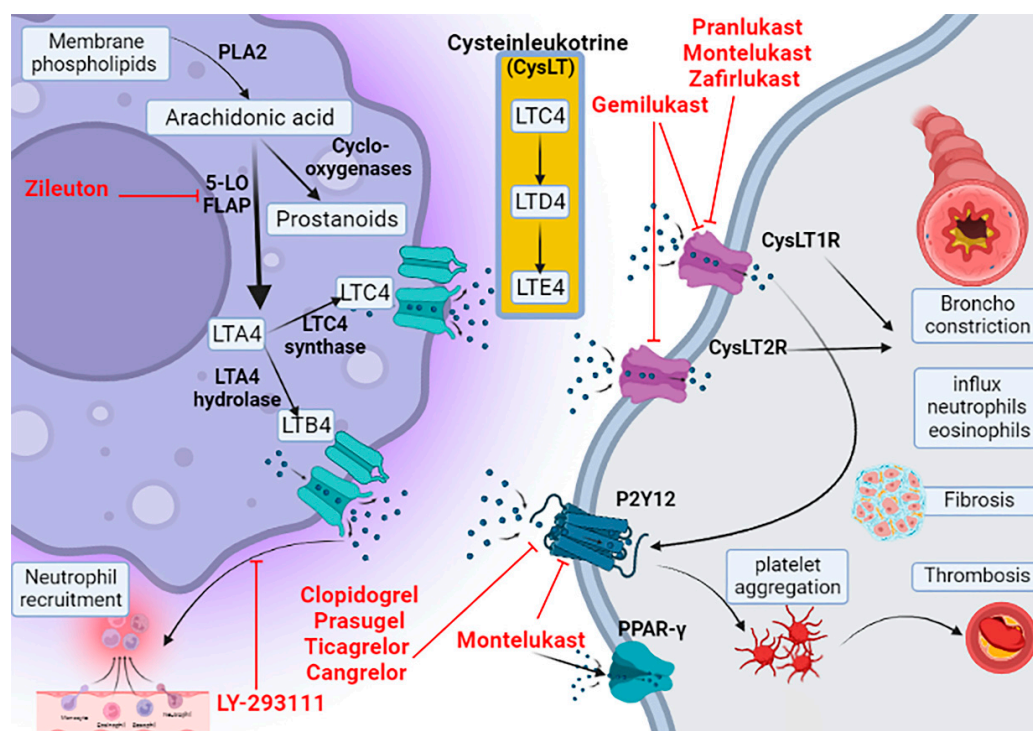


Figure 1. Leukotriene pathway and its inhibitors: Membrane phospholipids hydrolyzed by phospholipase A to arachidonic acid (AA), which, by the action of 5-lipoxygenase (5-LO), convert to leukotriene A4 (LTA4). Further, LTA4 is converted to LTB4 by the action of LTA4 hydrolase and to cysteinleukotriene (CysLT) by the action of LTC4 synthase. CysLTs act on CysLT receptors (CysLT1R and CysLT2R).

GPR17 is a G-protein coupled receptor stimulated by LTC4, LTD4, and uracil nucleotides. GPR17 chemical structure is intermediate between CysLTR1 and CysLTR2. GPR17 is highly distributed in the central nervous system (CNS) mainly in the microglia cells [22]. Cangrelor, which is a purinergic receptor (P2Y) antagonist, blocks GPR17 [23].

Furthermore, P2Y12 is activated following activation of CysLTR1 by LTs; therefore, inhibition of P2Y12 by a specific antagonist attenuates the CysLTR1-dependent effect of LTs. P2Y12 is a chemoreceptor stimulated by adenosine diphosphate (ADP) and belongs to G-protein coupled purinergic receptor group [24]. P2Y12 is involved in platelet aggregation and thrombosis; it is inhibited by antiplatelet drugs such as clopidogrel, prasugrel, ticagrelor, and cangrelor [24]. Notably, P2Y12 is also expressed in the vascular smooth muscle and immune cells and involved in the pathogenesis of inflammation through the promotion of the interaction between platelets and leukocytes with the release of inflammatory cytokines [24]. Kang et al. illustrated that MK attenuates bone loss through inhibition expression of P2Y12 [25]. Moreover, LTs can activate peroxisome proliferator-activated receptors (PPAR). Interestingly, zafirlukast and MK are regarded as a potent modulators of PPAR- γ [26]. LTs are intricate in the pathogenesis of asthma, dementia, and cognitive dysfunction [26].

These findings suggest that LTs through activation of LTRs and other related receptors may be involved in the pathogenesis of respiratory and systemic disorders.

3. Pharmacology of Montelukast

MK is a monocarboxylic acid with aliphatic sulfide from the quinolone family (Figure 2).

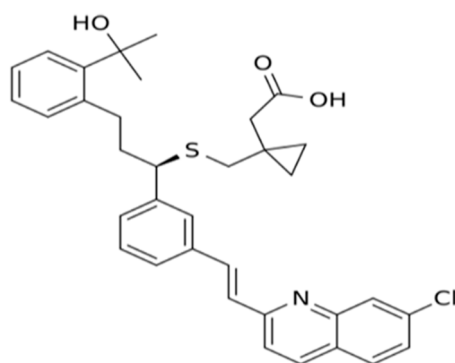


Figure 2. Chemical structure of montelukast (Reprinted from Ref. [27]).

MK is an orally active CysLTR1 antagonist that blocks the action of LTD₄ used in the prophylaxis of asthma and management of various inflammatory disorders [28]. Similarly, MK is indicated in the management of urticarial and allergic rhinitis [28]. MK was approved by the Food and Drug Administration (FDA) in 1998 in the USA. The use of MK is associated with the development of many adverse effects, including vomiting, nausea, skin rash, angioedema, mild fever, malaise, paresthesia, muscle cramps, and seizure [28,29]. Prolonged use of MK is linked with the development of neuropsychiatric disorders, including nightmares, insomnia, depression, anxiety, and aggression [29]. In 2009, the FDA declared that long-term use of MK was associated with increased risk of suicidal behavior [29]. In 2020, the FDA put MK in a boxed warning about the risk of neuropsychiatric disorders and advises to limit the use of MK for simple allergic reactions when safe alternatives are available [30].

MK is rapidly absorbed from the intestine with 64% bioavailability, and its absorption is not affected by food. MK plasma concentration is attained within 2–4 h with high plasma protein binding. MK has few drug–drug interactions though it inhibits liver metabolizing enzyme CYP2C8. MK and its metabolites are highly distributed and excreted by bile [31]. The lethal dose in 50% (LD₅₀) of MK is more than 5000 mg/kg [31].

4. Montelukast and Cardiovascular Complications

The general expression of CysLTR is associated with the propagation of different cardiovascular complications [32]. CysLT₂R is highly expressed in the human atrium, ventricles, apex, septum, and Purkinje cells. In addition, CysLT₂R is expressed on the endothelial cells, monocytes, and smooth muscle cells, while CysLT₁R is chiefly present in the macrophages and monocytes [32]. Surprisingly, only CysLT₂R is expressed in coronary smooth muscles, whereas aortic smooth muscles highly expressed CysLT₁R [32]. Of note, CysLTR exerts negative inotropic effects and reduces coronary blood flow without chronotropic effects [33].

MK and other CysLTR antagonists were originally developed for the management of asthma. In virtue of its anti-inflammatory property, MK could be effective in treating atherosclerosis and metabolic syndrome [34]. It has been shown that asthmatic patients on MK treatments showed a low cardiovascular risk index since MK decreases the risk of stroke, myocardial infarction, and other cardiovascular complications [8]. Of interest, MK inhibits epoxide hydrolase, which hydrolyzes beneficial epoxyeicosatrienoic acid to inactive dihydroxyeicosatrienoic acid [35]. Thus, MK and other CysLTR antagonists through activation of PPAR- γ and inhibition of epoxide hydrolase may reduce cardiovascular complications through the improvement of endothelial function and insulin sensitivity [36]. Animal model studies revealed that MK reduced the risk for the development of myocardial infarction, ischemic reperfusion injury, and atherosclerosis [37,38]. Ge et al. observed that MK had an anti-atherogenic effect by inhibiting the expression of monocyte chemoattractant protein 1 (MCP-1) in rabbits with induced carotid injury [38]. Likewise, Liu's experimental study confirmed that MK reduced the propagation of atherosclerosis by inhibiting the

expression of matrix metalloproteinases (MMPs) (Figure 3) [39]. Activation of PPAR- γ and inhibition of soluble epoxide hydrolase by MK attenuates propagation of oxidative stress-induced endothelial injury through suppression expression of AA metabolites [38]. Wang and colleagues observed that over-expression of soluble epoxide hydrolase was engaged with the development of atherosclerosis by reducing the anti-inflammatory anti-thrombotic effects of epoxyeicosatrienoic acid [40].

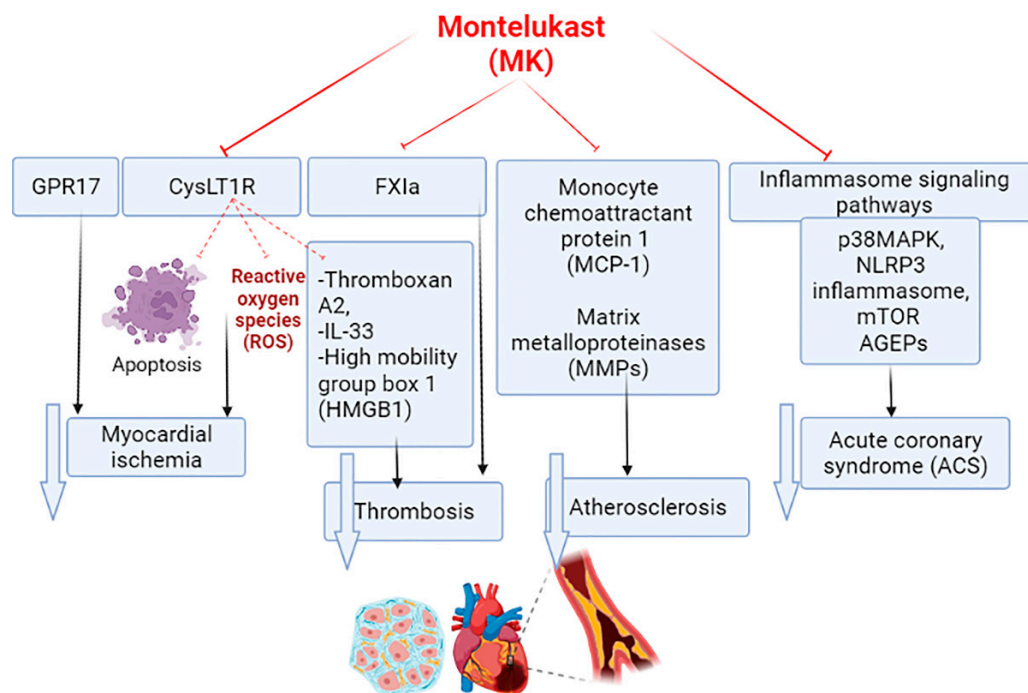


Figure 3. Mechanisms of montelukast in different cardiac and vascular diseases: Montelukast (MK) inhibits inflammasome signaling pathways, including p38 mitogen-activated protein kinase (p38MAPK), nod-like receptor pyrin 3 (NLRP3) inflammasome, mechanistic target of rapamycin (mTOR), and advanced glycation end-products (AGEPs). In addition, MK attenuates activation of cysteinileukotrine (CysLT) and activated factorXI with subsequent inhibition of inflammation and thrombosis-induced myocardial infarction and acute coronary syndrome (ACS).

In the atherosclerotic lesions, the expression of 5-LO and FLAP and their metabolites, including LTB₄, LTA₄, and LTC₄ and LTD₄ and LTC₄ are increased with the augmentation of the expression of CysLTR [41]. These findings suggest close relationships between the development of atherosclerosis and the activation of LT pathway, so targeting this pathway could limit the progression of the atherosclerotic process. In atherosclerotic lesions, the activated macrophages are augmented, which are the major source of 5-LO [41]. A clinical study observed that long-term treatment with MK reduced serum level of C-reactive protein (CRP) in the asthmatic patients [42]. Long-term follow-up studies proposed that MK by its anti-inflammatory effect may inhibit the advancement of atherosclerosis [43]. It has been shown that LT signaling pathway is implicated in the pathogenesis of atherosclerosis through suppression of lipid retention, accumulation of foam cells, and intimal hyperplasia with subsequent propagation of atherosclerosis [43].

Furthermore, LT increases the risk for rupture of atherosclerotic plaque through degradation of extracellular matrix [43]. For these reasons, augmentation of LTs may be correlated with the development of ischemic changes. In addition, over-expression of CysLTR in the atherosclerotic lesions increases the risk of atherosclerotic complications such as myocardial infarction, ischemic stroke, and aortic aneurysm [44]. Deletion of 5-LO in experimental mice reduced the development of atherosclerotic lesions by about 25-fold [45]. Likewise, an experimental study demonstrated that atorvastatin inhibits the

advancement of atherosclerotic lesions by inhibiting the expression of CysLT1R and FLAP as well as regulating the expression of 5-LO [46].

Regarding clinical studies, Revan et al. found that genetic variation in the LT pathway was linked with the development of early atherosclerotic lesions [47]. Genetic variations in the expression of 5-LO are associated with increasing intima-media thickness and progression of atherosclerosis in a cohort study [48]. These verdicts implicated the LT pathway in the pathogenesis of atherosclerosis; hence, LT pathway inhibitors or CysLTR antagonists could be effective in preventing the development of atherosclerosis. Different studies revealed that CysLT1R antagonists such as MK and FLAP inhibitors decrease the development of atherosclerosis in mice [49,50].

In the atherosclerotic lesions, LTC₄ expression and its mediator, known as multi-drug resistance protein 1 (MRP-1), were increased, causing oxidative stress injury in humans [51]. Of interest, LTC₄ and LTD₄ increase the expression of adhesion molecules such as P-selectin in the CysLT₂R-dependent pathway, and thus, MK may be ineffective in the suppression of this pathway, as it merely blocks CysLT₁R [52]. Furthermore, the induction release of pro-inflammatory cytokines and chemokines by LTD₄ was not suppressed by the action of MK and other CysLT₁R antagonists, as the release of inflammatory cytokines are mainly mediated by the CysLT₂R [53]. Remarkably, LTD₄ can induce the release of anti-inflammatory cytokine IL-10, which has a protective immunomodulatory effect against the development of atherosclerosis [53].

Moreover, overexpression of GPR99 is associated with the development of cardiac hypertrophy [54]. Nguyen et al. illustrated that MK can inhibit the pharmacological activity of GPR99, thereby limiting inflammatory progression and development of cardiac hypertrophy [55]. Further, changes in the expression of GPR107 and GPR99 could be the possible cause for the propagation and pathophysiology of hypertension in rats [21]. Moreover, the development and progression of chronic heart failure are associated with the escalation of the LT pathway [56]. Inhibition of LTB₄ may reduce the inflammatory changes and progression of heart failure [56]. Suppression of LTB₄ and AA metabolites decreases the risk for the development of heart failure [56].

These observations suggest that the LT pathway is implicated in the pathogenesis of atherosclerosis, cardiac hypertrophy, and heart failure. Inhibition of the LT pathway and CysLT₁R by MK might be effective in preventing cardiovascular complications.

5. Montelukast and Acute Coronary Syndrome

Notably, soluble epoxide hydrolase increases the expression of fatty acid synthase in the mononuclear cells in patients with ACS [57]. A prospective study involving 65 patients with ACS compared to 65 healthy controls illustrated that serum levels of soluble epoxide hydrolase and fatty acid synthase were increased in ACS patients [57]. A study including 667 ACS patients revealed that high-soluble epoxide hydrolase serum level is correlated with the severity of ACS [58]. Suppression of soluble epoxide hydrolase attenuates cardiac ischemic-reperfusion injury in vivo [59]. In addition, inhibition of soluble epoxide hydrolase attenuates the progression of hypertension, which is the major risk factor for the development of ACS [59]. These findings suggest that inhibition of soluble epoxide hydrolase by MK could be a therapeutic strategy for preventing the development of ACS.

On the other hand, PPAR- γ agonists were reported to be effective in the reduction of ACS [60]. PPAR- γ agonists reduce the expression of pro-inflammatory cytokines and macrophage activation in the coronary bed [60]. In a case-control study, PPAR- γ serum level was reduced in ACS patients and inversely correlated with cardiac troponin compared with controls [61]. Therefore, PPAR- γ serum level could be a predictive biomarker for the development of ACS mainly myocardial infarction [61]. Single-gene polymorphism in PPAR- γ predisposes the development of ACS in the Turkish population [62]. Remarkably, expression of LTC₄ and CysLT₁R are increased during adipogenesis through upregulation of PPAR- γ [63]. Therefore, inhibition of CysLT₁R by MK might be effective in the management of obesity-induced inflammatory disorders.

Of note, LTC₄ increases intima-media thickness and calcium contents in the coronary atherosclerotic plaques [64]. In addition, hypoxic stress in ACS increases the expression of CysLT₁R and LTC₄ in mice [49]. Within coronary atherosclerotic lesions, CysLTs are generated under hypoxic and ischemic conditions causing augmentation of vascular reactivity and propagation of coronary vasoconstriction [49,65]. Thus, CysLTs lead to noteworthy coronary hemodynamic changes. For example, administration of LTD₄ in patients with significant coronary stenosis leads to an augmentation of coronary resistance measured by coronary angiogram [66]. A previous *in vitro* study demonstrated that LTD₄- and LTC₄-induced coronary vasospasm during angiography was inhibited by MK [66]. It has been shown that the sensitivity of CysLTs to induce coronary vasospasm was increased due to the release of binding molecules, which increase the binding of CysLTs to the coronary atherosclerotic lesions [67]. Moreover, CysLT₁Rs expression is increased in response to the released pro-inflammatory cytokines from coronary atherosclerotic lesions [67]. Interestingly, CysLT₁R expression is higher than CysLT₂R in the carotid and coronary intima [68]. In an experimental study, CysLTs promote the release of pro-inflammatory cytokines with induction expression of adhesion molecules [68].

These findings proposed that CysLTs play a critical role in the pathogenesis of ACS, and the use of CysLT₁R antagonists such as MK might be of value in attenuating the severity of ACS (Figure 3).

6. Montelukast and Myocardial Infarction

It has been shown that LTs are involved in the pathogenesis of myocardial infarction. Additionally, genetic variation in 5-LO increases the risk of myocardial infarction in both animals and humans [32]. Production of CysLTs is augmented in ischemic-reperfusion injury following the development of myocardial infarction [32]. Different experimental studies illustrated that levels of CysLTs are increased after acute myocardial infarction. CysLTs induce coronary vascular resistance and increase infarct size and coronary endothelial dysfunction [69,70].

During the development of myocardial infarction, several inflammatory cells invade myocardial tissues that release LTs, which also recruit inflammatory cells and induce vascular smooth contractility, the proliferation of smooth muscle cells, and the increase of vascular permeability [71]. Gross et al.'s experimental study illustrated that production of LTs were augmented due to over-expression of 5-LO, which increases the release of pro-inflammatory cytokines [72]. A cohort study illustrated that overexpression of 5-LO increases the risk for the development of myocardial infarction [73]. Therefore, MK via inhibition of CysLT₁R can reduce the severity of myocardial ischemia by inhibiting apoptosis and the production of reactive oxygen species (ROS) (Figure 3) [74].

Moreover, LTD₄ plays an integral role in the development of myocardial ischemia through the induction of coronary constriction. LTD₄ has been reported to be a potent coronary vasoconstrictor in the isolated heart [75]. Surprisingly, zafirlukast was shown to be ineffective against experimental myocardial ischemia since myocardial ischemia is mainly mediated by CysLT₂R [76]. Notably, activation of CysLT₂R results in progressive myocardial ischemia with more inflammatory changes [76]. In addition, upregulation of endothelial CysLT₂R increases the risk of myocardial ischemia and apoptosis [77].

Moreover, GPR17, which is a receptor for LTD₄ and LTC₄, increases the risk of cardiac ischemia through interaction with cardiac CysLT₁R [78]. Inhibition of GPR17 by MK reduces myocardial ischemia and fibrosis [79]. Furthermore, urinary levels of CysLTs were increased following myocardial infarction and coronary bypass surgery [79]. Another study found that rupture of coronary plaque was associated with increased urinary LTs in patients with ACS [80]. However, urinary LTs as a potential biomarker of plaque rupture or inflammatory changes during myocardial infarction need to be elucidated through further study. Many studies observed that inhibition of the LT pathway increases recovery following myocardial infarction [81,82].

Despite these findings, there are controversial studies concerning the protective effects of CysLT1R antagonists in myocardial ischemia. For example, Shekher et al. found that CysLT1R antagonists did not alter infarct size and reperfusion injury following myocardial infarction [83]. Further, coronary ligation with the development of myocardial infarction and ischemic reperfusion injury in experimental mice was not ameliorated by 5-LO inhibitors or CysLT1R antagonists [77]. However, CysLT1R antagonists may produce neutral effects [84]. Genetic variation in LTA4 leads to ethnic risk factors for the development of myocardial infarction [85]. Recently, Hori et al.'s upregulation of LTB4 receptors exacerbated inflammatory changes following myocardial infarction [86].

A systematic review involving 28 studies illustrated that CysLT1R antagonists reduce the risk of myocardial infarction, ischemic stroke, and other cardiovascular complications [87]. In addition, CysLT1R antagonists could be effective for the secondary prevention of cardiovascular diseases. The reduction of recurrent myocardial infarction in male subjects on MK therapy was significant as compared to male subjects not on MK treatment (HR = 0.65, 95% CI = 0.43–0.99) in a three-year follow-up study [87]. These findings suggest that MK may affect the expression and activity of CysLT2R, which is chiefly engaged with myocardial ischemia.

Moreover, MK attenuates lipopolysaccharide (LPS)-induced myocardial injury in a dose-dependent manner [88]. Herein, MK could have cardioprotective effects against endotoxemia-induced inflammatory changes during acute myocardial injury. This effect of MK can be attributed to its anti-inflammatory and antioxidant effects [88]. A single-center, prospective, observational study showed that patients with previous myocardial infarction had higher urinary LTs as compared with unstable angina [89].

7. Effects of Montelukast on the Inflammatory Signaling Pathway in Acute Coronary Syndrome

Different inflammatory signaling pathways are activated during the development of ACS that correlated with the severity of ACS. Mitogen-activated protein kinase (MAPK) is activated during ACS [90]. MAPK family includes extracellular signal-regulated kinase (ERK), c-Jun N-terminal kinase (JNK), and p38. MAPK family and toll-like receptor 4 (TLR4) are increased in patients with ACS compared to the controls [91]. Indolfi et al. confirmed that p38MAPK was increased in the T lymphocytes in patients with ACS [92]. In a comparative study, p38MAPK activities in T lymphocytes were not significant in patients with unstable angina compared to the healthy controls. However, it was higher in patients with ST-elevation myocardial infarction [92]. These findings confirmed that p38MAPK could be a potential biomarker to differentiate subtypes of ACS.

Likewise, nod-like receptor pyrin 3 (NLRP3) inflammasome is an inflammatory signaling pathway involved in the release of pro-inflammatory cytokines with the induction of pyroptosis [19]. It has been shown that NLRP3 inflammasome is activated in ACS mainly in acute myocardial infarction, causing hyperinflammation and cardiac fibrosis [93]. Mauro et al. revealed that the NLRP3 inflammasome is highly activated in acute myocardial infarction [94]. NLRP3 inflammasome senses intracellular signaling during tissue ischemia and provokes immune response during acute myocardial infarction [94]. In addition, coronary ischemia induces activation of NLRP3 inflammasome through ROS [95]. Activated NLRP3 inflammasome in ACS triggers inflammatory changes and cardiac dysfunction. Moreover, the mechanistic target of the rapamycin (mTOR) signaling pathway is activated during heart failure following acute myocardial infarction through the autophagic process [96]. Likewise, advanced glycation end-products (AGEPs) are triggered during ACS and contribute to the development of post-myocardial remodeling and dysfunction [97].

Therefore, targeting p38MAPK, NLRP3 inflammasome, mTOR, and AGEPs may reduce the risk of complications in patients with ACS (Figure 3). It has been illustrated that MK can inhibit both p38MAPK and NLRP3 inflammasome in different studies. It has been reported that MK inhibits microglial activation through suppression of p38MAPK signaling [98]. Further, Zhou et al. demonstrated that MK attenuated neuropathic pain

by inhibiting p38MAPK and NF- κ B in a rat model [99]. Updated studies showed that MK alleviates hepatotoxicity by inhibiting the NLRP3 inflammasome signaling pathway in an animal model [100,101]. Furthermore, MK and other CysLT1R antagonists have abilities to block the activation of mTOR and AGEPs in various experimental studies [102,103].

These verdicts suggest that MK may reduce the severity of ACS by reducing the activity of p38MAPK, NLRP3 inflammasome, mTOR, and AGEPs. This pleiotropic effect of MK may not be mediated by blocking CysLT1R; thus, extensive researches are warranted in this regard.

Taken together, MK could be an effective novel therapy in the management of ACS through inhibition of pro-inflammatory CysLT1R and modulation of inflammasome signaling pathways. Experimental, preclinical, and clinical studies are recommended to confirm the potential therapeutic of MK in the management of ACS.

8. Montelukast and Thrombosis in Acute Coronary Syndrome

Thrombosis plays an integral role in the pathogenesis of ACS since disruption and rupture of coronary atherosclerotic plaque expose subendothelial collagen with the release of tissue factors and pro-coagulant molecules, which activate platelet activations and fibrin formation [104]. These changes induce endothelial dysfunction and immunothrombosis with the development of coronary obstruction and subsequent ischemic-reperfusion injury in ACS [104,105]. Platelet activations trigger atheromatous complications after rupture of the coronary atheromatous plaque [104,105].

Moreover, inflammation through activation of monocytes, macrophages, and mast and T cells contributes to the pathogenesis of ACS via rupture of atherosclerotic plaque and release of pro-inflammatory cytokines [106]. Pro-inflammatory cytokines induce the expression of endothelial adhesion molecules with succeeding thrombus formation [106–108].

It has been shown through *in silico* and *in vitro* experiments that MK had an antithrombotic effect by inhibiting factor XI (FXIa) [109]. *In vivo* study has also confirmed the inhibitory effect of MK on the activation of FXIa [110]. Houard and colleagues illustrated that expression of 5-LO and LTB₄ are increased in the intramural thrombus [110]. Thus, LTs are engaged with the propagation of coronary thrombosis. A previous study revealed that peptido-LTs are potent agonists of clotting factors including the von Willebrand factor with induction expression of P-selectin from endothelial cells [111]. Notably, LTC₄ induces the release of thromboxane A₂, IL-33, and high-mobility group box 1 (HMGB1) from activated platelets, causing immunothrombosis [112–114]. Tang et al. observed that stimulation of GRP91 increases platelet aggregation and release of LT4 [115]. Therefore, CysLT1R inhibitors such as MK may reduce platelet activation and aggregation during the propagation of ACS and thrombotic events.

Taken together, MK has the ability to attenuate thrombotic events by inhibiting platelet activation and clotting factors, which are activated during the development of ACS (Figure 3).

The present review had many limitations, including a paucity of clinical studies that evaluate the clinical effectiveness of MK in patients with ACS. Moreover, sequential levels of LTs were not assessed in most of the clinical studies to find the association with severity and recovery.

9. Conclusions

ACS is associated with activation of LT pathway with subsequent releases of various LTs, including LTB₄, LTC₄, and LTD₄, which cause inflammatory changes and the induction of immunothrombosis. LTs through CysLTRs induce activation of platelets and clotting factors with succeeding coronary thrombosis. CysLTR antagonists such as MK may reduce risk development of ACS and associated complications through suppression activation of platelet and clotting factors. Taken together, MK could be an effective agent in reducing severity of ACS and associated complications. Experimental and clinical studies are warranted in this regard.

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References

1. Al-Kuraishy, H.M.; Al-Gareeb, A.I.; Al-Buhadilly, A.K. Rosuvastatin improves vaspin serum levels in obese patients with acute coronary syndrome. *Diseases* **2018**, *6*, 9. [[CrossRef](#)] [[PubMed](#)]
2. Al-Kuraishy, H.M.; Al-Gareeb, A.I.; Samy, O.M. Statin therapy improves serum Annexin A1 levels in patients with acute coronary syndrome: A case-controlled study. *Int. J. Crit. Illn. Inj. Sci.* **2021**, *11*, 4. [[CrossRef](#)] [[PubMed](#)]
3. Al-Kuraishy, H.M.; Al-Gareeb, A.I.; Awad, M.S.; Alrifai, S.B. Assessment of serum prolactin levels in acute myocardial infarction: The role of pharmacotherapy. *Indian J. Endocrinol. Metab.* **2016**, *20*, 72. [[CrossRef](#)] [[PubMed](#)]
4. Al-Kuraishy, H.M.; Al-Gareeb, A.I.; Welson, N.N.; Batiha, G.E.-S. Trimetazidine and COVID-19-induced acute cardiac injury: A missed key. *Int. J. Clin. Pharm.* **2022**, *44*, 832–833. [[CrossRef](#)] [[PubMed](#)]
5. Sami, O.M.; Al-kuraishy, H.M.; Al-Gareeb, A.I. Differential effects of statins on annexin A1 serum level in patients with acute coronary syndrome: A pleiotropic update. *Ann. Trop. Med. Public Health* **2021**, *24*, 394–402. [[CrossRef](#)]
6. Mason, P.J.; Shah, B.; Tamis-Holland, J.E.; Bittl, J.A.; Cohen, M.G.; Safirstein, J.; Drachman, D.E.; Valle, J.A.; Rhodes, D.; Gilchrist, I.C. An update on radial artery access and best practices for transradial coronary angiography and intervention in acute coronary syndrome: A scientific statement from the American Heart Association. *Circ. Cardiovasc. Interv.* **2018**, *11*, e000035. [[CrossRef](#)]
7. Schwartz, G.G.; Steg, P.G.; Szarek, M.; Bhatt, D.L.; Bittner, V.A.; Diaz, R.; Edelberg, J.M.; Goodman, S.G.; Hanotin, C.; Harrington, R.A. Alirocumab and cardiovascular outcomes after acute coronary syndrome. *N. Engl. J. Med.* **2018**, *379*, 2097–2107. [[CrossRef](#)]
8. Hoxha, M.; Rovati, G.E.; Cavanillas, A.B. The leukotriene receptor antagonist montelukast and its possible role in the cardiovascular field. *Eur. J. Clin. Pharmacol.* **2017**, *73*, 799–809. [[CrossRef](#)]
9. Kellaway, C.; Trethewie, E. The liberation of a slow-reacting smooth muscle-stimulating substance in anaphylaxis. *Q. J. Exp. Physiol. Cogn. Med. Sci. Transl. Integr.* **1940**, *30*, 121–145. [[CrossRef](#)]
10. Al-Kuraishy, H.M.; Al-Gareeb, A.I.; Almulaiky, Y.Q.; Cruz-Martins, N.; Batiha, G.E.-S. Role of leukotriene pathway and montelukast in pulmonary and extrapulmonary manifestations of Covid-19: The enigmatic entity. *Eur. J. Pharmacol.* **2021**, *904*, 174196. [[CrossRef](#)]
11. Bankova, L.G.; Dwyer, D.F.; Yoshimoto, E.; Ualiyeva, S.; McGinty, J.W.; Raff, H.; von Moltke, J.; Kanaoka, Y.; Frank Austen, K.; Barrett, N.A. The cysteinyl leukotriene 3 receptor regulates expansion of IL-25-producing airway brush cells leading to type 2 inflammation. *Sci. Immunol.* **2018**, *3*, eaat9453. [[CrossRef](#)] [[PubMed](#)]
12. He, R.; Chen, Y.; Cai, Q. The role of the LTB4-BLT1 axis in health and disease. *Pharmacol. Res.* **2020**, *158*, 104857. [[CrossRef](#)] [[PubMed](#)]
13. Alorabi, M.; Cavalu, S.; Al-Kuraishy, H.M.; Al-Gareeb, A.I.; Mostafa-Hedeab, G.; Negm, W.A.; Youssef, A.; El-Kadem, A.H.; Saad, H.M.; Batiha, G.E.-S. Pentoxifylline and berberine mitigate diclofenac-induced acute nephrotoxicity in male rats via modulation of inflammation and oxidative stress. *Biomed. Pharmacother.* **2022**, *152*, 113225. [[CrossRef](#)] [[PubMed](#)]
14. Hayashi, H.; Fukutomi, Y.; Mitsui, C.; Kajiwara, K.; Watai, K.; Kamide, Y.; Nakamura, Y.; Hamada, Y.; Tomita, Y.; Sekiya, K. Omalizumab for aspirin hypersensitivity and leukotriene overproduction in aspirin-exacerbated respiratory disease. A randomized controlled trial. *Am. J. Respir. Crit. Care Med.* **2020**, *201*, 1488–1498. [[CrossRef](#)] [[PubMed](#)]
15. Batiha, G.E.-S.; Al-Gareeb, A.I.; Saad, H.M.; Al-Kuraishy, H.M. COVID-19 and corticosteroids: A narrative review. *Inflammopharmacology* **2022**, *30*, 1189–1205. [[CrossRef](#)]
16. Möller, I.; Murali, R.; Müller, H.; Wiesner, T.; Jackett, L.A.; Scholz, S.L.; Cosgarea, I.; van de Nes, J.A.; Sucker, A.; Hillen, U. Activating cysteinyl leukotriene receptor 2 (CYSLTR2) mutations in blue nevi. *Mod. Pathol.* **2017**, *30*, 350–356. [[CrossRef](#)]
17. Tantisira, K.G.; Drazen, J.M. Genetics and pharmacogenetics of the leukotriene pathway. *J. Allergy Clin. Immunol.* **2009**, *124*, 422–427. [[CrossRef](#)]

18. Park, J.S.; Chang, H.S.; Park, C.-S.; Lee, J.-H.; Lee, Y.M.; Choi, J.H.; Park, H.-S.; Kim, L.H.; Park, B.L.; Choi, Y.H. Association analysis of cysteinyl-leukotriene receptor 2 (CYSLTR2) polymorphisms with aspirin intolerance in asthmatics. *Pharmacogenet. Genom.* **2005**, *15*, 483–492. [[CrossRef](#)]
19. Mostafa-Hedeab, G.; Al-Kuraishy, H.M.; Al-Gareeb, A.I.; Jeandet, P.; Saad, H.M.; Batiha, G.E.-S. A raising dawn of pentoxifylline in management of inflammatory disorders in Covid-19. *Inflammopharmacology* **2022**, *30*, 799–809. [[CrossRef](#)]
20. Itadani, S.; Yashiro, K.; Aratani, Y.; Sekiguchi, T.; Kinoshita, A.; Moriguchi, H.; Ohta, N.; Takahashi, S.; Ishida, A.; Tajima, Y. Discovery of gemilukast (ONO-6950), a dual CysLT1 and CysLT2 antagonist as a therapeutic agent for asthma. *J. Med. Chem.* **2015**, *58*, 6093–6113. [[CrossRef](#)]
21. Calderón-Zamora, L.; Canizalez-Román, A.; León-Sicauros, N.; Aguilera-Mendez, A.; Huang, F.; Hong, E.; Villafaña, S. Changes in expression of orphan receptors GPR99 and GPR107 during the development and establishment of hypertension in spontaneously hypertensive rats. *J. Recept. Signal Transduct.* **2021**, *41*, 558–565. [[CrossRef](#)] [[PubMed](#)]
22. Lecca, D.; Raffaele, S.; Abbracchio, M.P.; Fumagalli, M. Regulation and signaling of the GPR17 receptor in oligodendroglial cells. *Glia* **2020**, *68*, 1957–1967. [[CrossRef](#)] [[PubMed](#)]
23. De Luca, L.; Steg, P.G.; Bhatt, D.L.; Capodanno, D.; Angiolillo, D.J. Cangrelor: Clinical data, contemporary use, and future perspectives. *J. Am. Heart Assoc.* **2021**, *10*, e022125. [[CrossRef](#)] [[PubMed](#)]
24. Mansour, A.; Bachelot-Loza, C.; Nessler, N.; Gaussem, P.; Gouin-Thibault, I. P2Y12 inhibition beyond thrombosis: Effects on inflammation. *Int. J. Mol. Sci.* **2020**, *21*, 1391. [[CrossRef](#)]
25. Kang, J.H.; Lim, H.; Lee, D.S.; Yim, M. Montelukast inhibits RANKL-induced osteoclast formation and bone loss via CysLTR1 and P2Y12. *Mol. Med. Rep.* **2018**, *18*, 2387–2398. [[CrossRef](#)]
26. Göbel, T.; Diehl, O.; Heering, J.; Merk, D.; Angioni, C.; Wittmann, S.K.; Buscato, E.I.; Kottke, R.; Weizel, L.; Schader, T. Zafirlukast is a dual modulator of human soluble epoxide hydrolase and peroxisome proliferator-activated receptor γ . *Front. Pharmacol.* **2019**, *10*, 263. [[CrossRef](#)]
27. Fuse809 (talk)—Own work, Public Domain. Available online: <https://commons.wikimedia.org/w/index.php?curid=31613539> (accessed on 18 August 2022).
28. Sansing-Foster, V.; Haug, N.; Mosholder, A.; Cocoros, N.M.; Bradley, M.; Ma, Y.; Pennap, D.; Dee, E.C.; Toh, S.; Pestine, E. Risk of psychiatric adverse events among montelukast users. *J. Allergy Clin. Immunol. Pract.* **2021**, *9*, 385–393.e312. [[CrossRef](#)]
29. Glockler-Lauf, S.D.; Finkelstein, Y.; Zhu, J.; Feldman, L.Y.; To, T. Montelukast and neuropsychiatric events in children with asthma: A nested case–control study. *J. Pediatr.* **2019**, *209*, 176–182.e174. [[CrossRef](#)]
30. Fox, C.W.; Khaw, C.L.; Gerke, A.K.; Lund, B.C. Montelukast and neuropsychiatric events—a sequence symmetry analysis. *J. Asthma* **2021**, 1–7. [[CrossRef](#)]
31. Hirvensalo, P.; Tornio, A.; Neuvonen, M.; Tapaninen, T.; Paile-Hyvärinen, M.; Kärjä, V.; Männistö, V.T.; Pihlajamäki, J.; Backman, J.T.; Niemi, M. Comprehensive pharmacogenomic study reveals an important role of UGT1A3 in montelukast pharmacokinetics. *Clin. Pharmacol. Ther.* **2018**, *104*, 158–168. [[CrossRef](#)]
32. Colazzo, F.; Gelosa, P.; Tremoli, E.; Sironi, L.; Castiglioni, L. Role of the cysteinyl leukotrienes in the pathogenesis and progression of cardiovascular diseases. *Mediat. Inflamm.* **2017**, *2017*, 2432958. [[CrossRef](#)] [[PubMed](#)]
33. Folco, G.; Rossoni, G.; Buccellati, C.; Berti, F.; Maclouf, J.; Sala, A. Leukotrienes in cardiovascular diseases. *Am. J. Respir. Crit. Care Med.* **2000**, *161*, S112–S116. [[CrossRef](#)] [[PubMed](#)]
34. Ibrahim, M.A.; Amin, E.F.; Ibrahim, S.A.; Abdelzaher, W.Y.; Abdelrahman, A.M. Montelukast and irbesartan ameliorate metabolic and hepatic disorders in fructose-induced metabolic syndrome in rats. *Eur. J. Pharmacol.* **2014**, *724*, 204–210. [[CrossRef](#)]
35. Imig, J.D.; Hammock, B.D. Soluble epoxide hydrolase as a therapeutic target for cardiovascular diseases. *Nat. Rev. Drug Discov.* **2009**, *8*, 794–805. [[CrossRef](#)] [[PubMed](#)]
36. Fleming, I. The pharmacology of the cytochrome P450 epoxygenase/soluble epoxide hydrolase axis in the vasculature and cardiovascular disease. *Pharmacol. Rev.* **2014**, *66*, 1106–1140. [[CrossRef](#)]
37. Şener, G.; Şehirli, Ö.; Velioğlu-Öğünç, A.; Çetinel, Ş.; Gedik, N.; Caner, M.; Sakarcı, A.; Yeğen, B.Ç. Montelukast protects against renal ischemia/reperfusion injury in rats. *Pharmacol. Res.* **2006**, *54*, 65–71. [[CrossRef](#)] [[PubMed](#)]
38. Ge, S.; Zhou, G.; Cheng, S.; Liu, D.; Xu, J.; Xu, G.; Liu, X. Anti-atherogenic effects of montelukast associated with reduced MCP-1 expression in a rabbit carotid balloon injury model. *Atherosclerosis* **2009**, *205*, 74–79. [[CrossRef](#)]
39. Liu, D.; Ge, S.; Zhou, G.; Xu, G.; Zhang, R.; Zhu, W.; Liu, Z.; Cheng, S.; Liu, X. Montelukast inhibits matrix metalloproteinases expression in atherosclerotic rabbits. *Cardiovasc. Drugs Ther.* **2009**, *23*, 431–437. [[CrossRef](#)]
40. Wang, Y.-X.J.; Ulu, A.; Zhang, L.-N.; Hammock, B. Soluble epoxide hydrolase in atherosclerosis. *Curr. Atheroscler. Rep.* **2010**, *12*, 174–183. [[CrossRef](#)]
41. Cipollone, F.; Mezzetti, A.; Fazio, M.L.; Cucurullo, C.; Iezzi, A.; Uchino, S.; Spigonardo, F.; Bucci, M.; Cucurullo, F.; Prescott, S.M. Association between 5-lipoxygenase expression and plaque instability in humans. *Arterioscler. Thromb. Vasc. Biol.* **2005**, *25*, 1665–1670. [[CrossRef](#)]
42. Allayee, H.; Hartiala, J.; Lee, W.; Mehrabian, M.; Irvin, C.G.; Conti, D.V.; Lima, J.J. The effect of montelukast and low-dose theophylline on cardiovascular disease risk factors in asthmatics. *Chest* **2007**, *132*, 868–874. [[CrossRef](#)] [[PubMed](#)]
43. Bäck, M. Leukotriene signaling in atherosclerosis and ischemia. *Cardiovasc. Drugs Ther.* **2009**, *23*, 41–48. [[CrossRef](#)] [[PubMed](#)]

44. Helgadottir, A.; Manolescu, A.; Thorleifsson, G.; Gretarsdottir, S.; Jonsdottir, H.; Thorsteinsdottir, U.; Samani, N.J.; Gudmundsson, G.; Grant, S.F.; Thorgeirsson, G. The gene encoding 5-lipoxygenase activating protein confers risk of myocardial infarction and stroke. *Nat. Genet.* **2004**, *36*, 233–239. [[CrossRef](#)] [[PubMed](#)]
45. Mehrabian, M.; Allayee, H.; Wong, J.; Shih, W.; Wang, X.-P.; Shaposhnik, Z.; Funk, C.D.; Lusis, A.J. Identification of 5-lipoxygenase as a major gene contributing to atherosclerosis susceptibility in mice. *Circ. Res.* **2002**, *91*, 120–126. [[CrossRef](#)]
46. Zhou, G.; Ge, S.; Liu, D.; Xu, G.; Zhang, R.; Yin, Q.; Zhu, W.; Chen, J.; Liu, X. Atorvastatin reduces plaque vulnerability in an atherosclerotic rabbit model by altering the 5-lipoxygenase pathway. *Cardiology* **2010**, *115*, 221–228. [[CrossRef](#)]
47. Bevan, S.; Lorenz, M.W.; Sitzer, M.; Markus, H.S. Genetic variation in the leukotriene pathway and carotid intima-media thickness: A 2-stage replication study. *Stroke* **2009**, *40*, 696–701. [[CrossRef](#)]
48. Dwyer, J.H.; Allayee, H.; Dwyer, K.M.; Fan, J.; Wu, H.; Mar, R.; Lusis, A.J.; Mehrabian, M. Arachidonate 5-lipoxygenase promoter genotype, dietary arachidonic acid, and atherosclerosis. *N. Engl. J. Med.* **2004**, *350*, 29–37. [[CrossRef](#)]
49. Nobili, E.; Salvado, M.D.; Folkersen, L.; Castiglioni, L.; Kastrup, J.; Wetterholm, A.; Tremoli, E.; Hansson, G.K.; Sironi, L.; Haeggström, J.Z. Cysteinyl leukotriene signaling aggravates myocardial hypoxia in experimental atherosclerotic heart disease. *PLoS ONE* **2012**, *7*, e41786. [[CrossRef](#)]
50. Duah, E.; Adapala, R.K.; Al-Azzam, N.; Kondeti, V.; Gombedza, F.; Thodeti, C.K.; Paruchuri, S. Cysteinyl leukotrienes regulate endothelial cell inflammatory and proliferative signals through CysLT2 and CysLT1 receptors. *Sci. Rep.* **2013**, *3*, 3274. [[CrossRef](#)]
51. Cole, S.P. Multidrug resistance protein 1 (MRP1, ABC1), a “multitasking” ATP-binding cassette (ABC) transporter. *J. Biol. Chem.* **2014**, *289*, 30880–30888. [[CrossRef](#)]
52. Papayianni, A.; Serhan, C.N.; Brady, H.R. Lipoxin A4 and B4 inhibit leukotriene-stimulated interactions of human neutrophils and endothelial cells. *J. Immunol.* **1996**, *156*, 2264–2272. [[PubMed](#)]
53. Uzonyi, B.; Lötzer, K.; Jahn, S.; Kramer, C.; Hildner, M.; Bretschneider, E.; Radke, D.; Beer, M.; Vollandt, R.; Evans, J.F. Cysteinyl leukotriene 2 receptor and protease-activated receptor 1 activate strongly correlated early genes in human endothelial cells. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 6326–6331. [[CrossRef](#)] [[PubMed](#)]
54. Omede, A. *Role of Alpha-Ketoglutarate Receptor G-Protein Coupled Receptor 99 (GPR99) in Cardiac Hypertrophy*; The University of Manchester: Manchester, UK, 2015.
55. Nguyen, A.V.; Bagood, M.D.; Wang, M.; Caryotakis, S.E.; Smith, G.; Yee, S.; Shen, H.; Isseroff, R.R.; Soulika, A.M. Montelukast, an Antagonist of Cysteinyl Leukotriene Signaling, Impairs Burn Wound Healing. *Plast. Reconstr. Surg.* **2022**, *150*, 92e–104e. [[CrossRef](#)]
56. Wang, Y.; Li, C.; Liu, Z.; Shi, T.; Wang, Q.; Li, D.; Wu, Y.; Han, J.; Guo, S.; Tang, B.; et al. DanQi Pill protects against heart failure through the arachidonic acid metabolism pathway by attenuating different cyclooxygenases and leukotrienes B4. *BMC Complement. Altern. Med.* **2014**, *14*, 67. [[CrossRef](#)]
57. Zhao, X.; Du, J.-Q.; Xu, D.-Y.; Zhao, S.-P. Effects of soluble epoxide hydrolase inhibitor on the expression of fatty acid synthase in peripheral blood mononuclear cell in patients with acute coronary syndrome. *Lipids Heal. Dis.* **2013**, *12*, 3. [[CrossRef](#)]
58. Oni-Orisan, A.; Cresci, S.; Jones, P.G.; Theken, K.N.; Spertus, J.A.; Lee, C.R. Association between the EPHX2 p. Lys55Arg polymorphism and prognosis following an acute coronary syndrome. *Prostaglandins Other Lipid Mediat.* **2018**, *138*, 15–22. [[CrossRef](#)] [[PubMed](#)]
59. Motoki, A.; Merkel, M.J.; Packwood, W.H.; Cao, Z.; Liu, L.; Iliff, J.; Alkayed, N.J.; Van Winkle, D.M. Soluble epoxide hydrolase inhibition and gene deletion are protective against myocardial ischemia-reperfusion injury in vivo. *Am. J. Physiol.-Heart Circ. Physiol.* **2008**, *295*, H2128–H2134. [[CrossRef](#)]
60. Libby, P. Current concepts of the pathogenesis of the acute coronary syndromes. *Circulation* **2001**, *104*, 365–372. [[CrossRef](#)]
61. Ali, S.A.-J.; Al-Shalah, H.; Al-Salihi, O. Assessment of circulating PPAR- γ Level as a risk and diagnostic biomarker in Acute Coronary Syndrome. *J. Univ. Babylon Pure Appl. Sci.* **2018**, *26*, 210–218. [[CrossRef](#)]
62. Arat, A.; Yılmaz, Ü.; Yılmaz, N.; Fazlıoğulları, O.; Çelik, F.; Başaran, C.; Zeybek, Ü. Effects of Leptin, Resistin, and PPAR-Gama Gene Variants on Obese Patients with Acute Coronary Syndrome in the Turkish Population. *JAREM. J. Acad. Res. Med.* **2020**, *10*, 166. [[CrossRef](#)]
63. Fujimori, K.; Uno, S.; Kuroda, K.; Matsumoto, C.; Maehara, T. Leukotriene C4 synthase is a novel PPAR γ target gene, and leukotriene C4 and D4 activate adipogenesis through cysteinyl LT1 receptors in adipocytes. *Biochim. Biophys. Acta (BBA)-Mol. Cell Res.* **2022**, *1869*, 119203. [[CrossRef](#)] [[PubMed](#)]
64. Iovannisci, D.M.; Lammer, E.J.; Steiner, L.; Cheng, S.; Mahoney, L.T.; Davis, P.H.; Lauer, R.M.; Burns, T.L. Association between a leukotriene C4 synthase gene promoter polymorphism and coronary artery calcium in young women: The Muscatine Study. *Arterioscler. Thromb. Vasc. Biol.* **2007**, *27*, 394–399. [[CrossRef](#)] [[PubMed](#)]
65. Piomelli, D.; Feinmark, S.J.; Cannon, P.J. Leukotriene biosynthesis by canine and human coronary arteries. *J. Pharmacol. Exp. Ther.* **1987**, *241*, 763–770. [[PubMed](#)]
66. Vigorito, C.; Giordano, A.; Cirillo, R.; Genovese, A.; Rengo, F.; Marone, G. Metabolic and hemodynamic effects of peptide leukotriene C4 and D4 in man. *Int. J. Clin. Lab. Res.* **1997**, *27*, 178–184. [[CrossRef](#)] [[PubMed](#)]
67. Eaton, A.; Nagy, E.; Pacault, M.; Fauconnier, J.; Bäck, M. Cysteinyl leukotriene signaling through perinuclear CysLT1 receptors on vascular smooth muscle cells transduces nuclear calcium signaling and alterations of gene expression. *J. Mol. Med.* **2012**, *90*, 1223–1231. [[CrossRef](#)]

68. Loötzer, K.; Spanbroek, R.; Hildner, M.; Urbach, A.; Heller, R.; Bretschneider, E.; Galczenski, H.; Evans, J.F.; Habenicht, A.J. Differential Leukotriene Receptor Expression and Calcium Responses in Endothelial Cells and Macrophages Indicate 5-Lipoxygenase-Dependent Circuits of Inflammation and Atherogenesis. *Arterioscler. Thromb. Vasc. Biol.* **2003**, *23*, e32–e36.
69. Peters-Golden, M.; Henderson, W.R., Jr. Leukotrienes. *N. Engl. J. Med.* **2007**, *357*, 1841–1854. [[CrossRef](#)]
70. Allen, S.P.; Sampson, A.P.; Piper, P.J.; Chester, A.H.; Ohri, S.K.; Yacoub, M.H. Enhanced excretion of urinary leukotriene E4 in coronary artery disease and after coronary artery bypass surgery. *Coron. Artery Dis.* **1993**, *4*, 899–904. [[CrossRef](#)]
71. Ni, N.C.; Ballantyne, L.L.; Mewburn, J.D.; Funk, C.D. Multiple-site activation of the cysteinyl leukotriene receptor 2 is required for exacerbation of ischemia/reperfusion injury. *Arterioscler. Thromb. Vasc. Biol.* **2014**, *34*, 321–330. [[CrossRef](#)]
72. Gross, G.J.; Falck, J.R.; Gross, E.R.; Isbell, M.; Moore, J.; Nithipatikom, K. Cytochrome P450 and arachidonic acid metabolites: Role in myocardial ischemia/reperfusion injury revisited. *Cardiovasc. Res.* **2005**, *68*, 18–25. [[CrossRef](#)]
73. Scott, W.; Pawlowski, N.; Andreach, M.; Cohn, Z. Resting macrophages produce distinct metabolites from exogenous arachidonic acid. *J. Exp. Med.* **1982**, *155*, 535–547. [[CrossRef](#)] [[PubMed](#)]
74. Becher, U.M.; Ghanem, A.; Tiyerili, V.; Fürst, D.O.; Nickenig, G.; Mueller, C.F. Inhibition of leukotriene C4 action reduces oxidative stress and apoptosis in cardiomyocytes and impedes remodeling after myocardial injury. *J. Mol. Cell. Cardiol.* **2011**, *50*, 570–577. [[CrossRef](#)] [[PubMed](#)]
75. Han, H.; Tian, R.; Neubauer, S.; Gaudron, P.; Hu, K.; Ertl, G. Effects of LTD4 and its specific antagonist L-660,711 in isolated rat hearts with chronic myocardial infarction. *Am. J. Physiol.-Heart Circ. Physiol.* **1994**, *266*, H2068–H2073. [[CrossRef](#)]
76. Ni, N.C.; Yan, D.; Ballantyne, L.L.; Barajas-Espinosa, A.; Amand, T.S.; Pratt, D.A.; Funk, C.D. A selective cysteinyl leukotriene receptor 2 antagonist blocks myocardial ischemia/reperfusion injury and vascular permeability in mice. *J. Pharmacol. Exp. Ther.* **2011**, *339*, 768–778. [[CrossRef](#)] [[PubMed](#)]
77. Jiang, W.; Hall, S.R.; Moos, M.P.; Cao, R.Y.; Ishii, S.; Ogunyankin, K.O.; Melo, L.G.; Funk, C.D. Endothelial cysteinyl leukotriene 2 receptor expression mediates myocardial ischemia-reperfusion injury. *Am. J. Pathol.* **2008**, *172*, 592–602. [[CrossRef](#)]
78. Qi, A.-D.; Harden, T.K.; Nicholas, R.A. Is GPR17 a P2Y/leukotriene receptor? examination of uracil nucleotides, nucleotide sugars, and cysteinyl leukotrienes as agonists of GPR17. *J. Pharmacol. Exp. Ther.* **2013**, *347*, 38–46. [[CrossRef](#)]
79. Carry, M.; Korley, V.; Willerson, J.T.; Weigelt, L.; Ford-Hutchinson, A.W.; Tagari, P. Increased urinary leukotriene excretion in patients with cardiac ischemia. In vivo evidence for 5-lipoxygenase activation. *Circulation* **1992**, *85*, 230–236. [[CrossRef](#)]
80. Prescott, E.; Pernow, J.; Saraste, A.; Åkerblom, A.; Angerås, O.; Erlinge, D.; Grove, E.L.; Hedman, M.; Jensen, L.O.; Svedlund, S. Design and rationale of FLAVOUR: A phase IIa efficacy study of the 5-lipoxygenase activating protein antagonist AZD5718 in patients with recent myocardial infarction. *Contemp. Clin. Trials Commun.* **2020**, *19*, 100629. [[CrossRef](#)]
81. Lee, C.C.; Appleyard, R.F.; Byrne, J.G.; Cohn, L.H. Leukotrienes D4 and E4 produced in myocardium impair coronary flow and ventricular function after two hours of global ischaemia in rat heart. *Cardiovasc. Res.* **1993**, *27*, 770–773. [[CrossRef](#)]
82. Rossoni, G.; Sala, A.; Berti, F.; Testa, T.; Buccellati, C.; Molta, C.; Muller-Peddinghaus, R.; Maclouf, J.; Folco, G. Myocardial protection by the leukotriene synthesis inhibitor BAY X1005: Importance of transcellular biosynthesis of cysteinyl-leukotrienes. *J. Pharmacol. Exp. Ther.* **1996**, *276*, 335–341.
83. Shekher, A.; Singh, M. Role of eicosanoid inhibition of ischemia reperfusion injury: Intact and isolated rat heart studies. *Methods Find. Exp. Clin. Pharmacol.* **1997**, *19*, 223–229. [[PubMed](#)]
84. Hahn, R.A.; MacDonald, B.R.; Simpson, P.J.; Potts, B.D.; Parli, C.J. Antagonism of leukotriene B4 receptors does not limit canine myocardial infarct size. *J. Pharmacol. Exp. Ther.* **1990**, *253*, 58–66. [[PubMed](#)]
85. Helgadottir, A.; Manolescu, A.; Helgason, A.; Thorleifsson, G.; Thorsteinsdottir, U.; Gudbjartsson, D.F.; Gretarsdottir, S.; Magnusson, K.P.; Gudmundsson, G.; Hicks, A. A variant of the gene encoding leukotriene A4 hydrolase confers ethnicity-specific risk of myocardial infarction. *Nat. Genet.* **2006**, *38*, 68–74. [[CrossRef](#)] [[PubMed](#)]
86. Horii, Y.; Nakaya, M.; Ohara, H.; Nishihara, H.; Watari, K.; Nagasaka, A.; Nakaya, T.; Sugiura, Y.; Okuno, T.; Koga, T. Leukotriene B4 receptor 1 exacerbates inflammation following myocardial infarction. *FASEB J.* **2020**, *34*, 8749–8763. [[CrossRef](#)] [[PubMed](#)]
87. Hoxha, M.; Lewis-Mikhael, A.-M.; Bueno-Cavanillas, A. Potential role of leukotriene receptor antagonists in reducing cardiovascular and cerebrovascular risk: A systematic review of human clinical trials and in vivo animal studies. *Biomed. Pharmacother.* **2018**, *106*, 956–965. [[CrossRef](#)]
88. Khodir, A.; Ghoneim, H.; Rahim, M.; Suddek, G. Montelukast attenuates lipopolysaccharide-induced cardiac injury in rats. *Hum. Exp. Toxicol.* **2016**, *35*, 388–397. [[CrossRef](#)]
89. Stodółkiewicz, E.; Sokołowska, B.; Rzeszutko, M.; Tomala, M.; Chrustowicz, A.; Żmudka, K.; Sanak, M.; Szczeklik, W. Leukotriene biosynthesis in coronary artery disease: Results of the Leukotrienes and Thromboxane In Myocardial Infarction (LTIMI) study. *Pol. Arch. Med. Wewnętrznej Pol. Arch. Intern. Med.* **2018**, *128*, 43–51. [[CrossRef](#)]
90. Vroegindewey, M.M.; Buljubasic, N.; Oemrawsingh, R.M.; Kardys, I.; Asselbergs, F.W.; van der Harst, P.; Umans, V.A.; Kietselaer, B.; Lenderink, T.; Liem, A. MAPK-cascade stimulating biomarkers in relation to recurrent coronary events following an acute coronary syndrome. In *The Prognostic Value of Coronary Imaging and Biomarkers in Ischemic Heart Disease*; Gildeprint: Enschede, The Netherlands, 2019; p. 123.
91. Wang, L.; Li, D.; Yang, K.; Hu, Y.; Zeng, Q. Toll-like receptor-4 and mitogen-activated protein kinase signal system are involved in activation of dendritic cells in patients with acute coronary syndrome. *Immunology* **2008**, *125*, 122–130. [[CrossRef](#)]

92. Indolfi, C.; Gasparri, C.; Vicinanza, C.; De Serio, D.; Boncompagni, D.; Mongiardo, A.; Spaccarotella, C.; Agosti, V.; Torella, D.; Curcio, A. Mitogen-activated protein kinases activation in T lymphocytes of patients with acute coronary syndromes. *Basic Res. Cardiol.* **2011**, *106*, 667–679. [[CrossRef](#)]
93. Wang, S.; Su, X.; Xu, L.; Chang, C.; Yao, Y.; Komal, S.; Cha, X.; Zang, M.; Ouyang, X.; Zhang, L. Glycogen synthase kinase-3 β inhibition alleviates activation of the NLRP3 inflammasome in myocardial infarction. *J. Mol. Cell. Cardiol.* **2020**, *149*, 82–94. [[CrossRef](#)]
94. Mauro, A.G.; Bonaventura, A.; Mezzaroma, E.; Quader, M.; Toldo, S. NLRP3 inflammasome in acute myocardial infarction. *J. Cardiovasc. Pharmacol.* **2019**, *74*, 175–187. [[CrossRef](#)] [[PubMed](#)]
95. Yin, Y.; Zhou, Z.; Liu, W.; Chang, Q.; Sun, G.; Dai, Y. Vascular endothelial cells senescence is associated with NOD-like receptor family pyrin domain-containing 3 (NLRP3) inflammasome activation via reactive oxygen species (ROS)/thioredoxin-interacting protein (TXNIP) pathway. *Int. J. Biochem. Cell Biol.* **2017**, *84*, 22–34. [[CrossRef](#)] [[PubMed](#)]
96. Suhara, T.; Baba, Y.; Shimada, B.K.; Higa, J.K.; Matsui, T. The mTOR signaling pathway in myocardial dysfunction in type 2 diabetes mellitus. *Curr. Diab. Rep.* **2017**, *17*, 38. [[CrossRef](#)]
97. Blackburn, N.J.R.; Vulesevic, B.; McNeill, B.; Cimenci, C.E.; Ahmadi, A.; Gonzalez-Gomez, M.; Ostojic, A.; Zhong, Z.; Brownlee, M.; Beisswenger, P.J.; et al. Methylglyoxal-derived advanced glycation end products contribute to negative cardiac remodeling and dysfunction post-myocardial infarction. *Basic Res. Cardiol.* **2017**, *112*, 57. [[CrossRef](#)] [[PubMed](#)]
98. Mansour, R.M.; Ahmed, M.A.; El-Sahar, A.E.; El Sayed, N.S. Montelukast attenuates rotenone-induced microglial activation/p38 MAPK expression in rats: Possible role of its antioxidant, anti-inflammatory and antiapoptotic effects. *Toxicol. Appl. Pharmacol.* **2018**, *358*, 76–85. [[CrossRef](#)] [[PubMed](#)]
99. Zhou, C.; Shi, X.; Huang, H.; Zhu, Y.; Wu, Y. Montelukast attenuates neuropathic pain through inhibiting p38 mitogen-activated protein kinase and nuclear factor-kappa B in a rat model of chronic constriction injury. *Anesth. Analg.* **2014**, *118*, 1090–1096. [[CrossRef](#)]
100. Fei, Z.; Zhang, L.; Wang, L.; Jiang, H.; Peng, A. Montelukast ameliorated pemetrexed-induced cytotoxicity in hepatocytes by mitigating endoplasmic reticulum (ER) stress and nucleotide oligomerization domain-like receptor protein 3 (NLRP3) activation. *Bioengineered* **2022**, *13*, 7894–7903. [[CrossRef](#)]
101. El-Kashef, D.H.; Zaghoul, R.A. Ameliorative effect of montelukast against carbon tetrachloride-induced hepatotoxicity: Targeting NLRP3 inflammasome pathway. *Life Sci.* **2022**, *304*, 120707. [[CrossRef](#)]
102. Tong, J.; Yu, Q.; Xu, W.; Yu, W.; Wu, C.; Wu, Y.; Yan, H. Montelukast enhances cytotoxic effects of carfilzomib in multiple myeloma by inhibiting mTOR pathway. *Cancer Biol. Ther.* **2019**, *20*, 381–390. [[CrossRef](#)] [[PubMed](#)]
103. Lei, C.; Wu, S.; Wen, C.; Li, Y.; Liu, N.; Huang, J.; Li, L.; Fu, M.; Liu, J. Zafirlukast attenuates advanced glycation end-products (AGEs)-induced degradation of articular extracellular matrix (ECM). *Int. Immunopharmacol.* **2019**, *68*, 68–73. [[CrossRef](#)]
104. Abbate, R.; Cioni, G.; Ricci, I.; Miranda, M.; Gori, A.M. Thrombosis and acute coronary syndrome. *Thromb. Res.* **2012**, *129*, 235–240. [[CrossRef](#)] [[PubMed](#)]
105. Carter, A.M. Inflammation, thrombosis and acute coronary syndromes. *Diabetes Vasc. Dis. Res.* **2005**, *2*, 113–121. [[CrossRef](#)]
106. Duman, H.; Çinier, G.; Bakırcı, E.M.; Duman, H.; Şimşek, Z.; Hamur, H.; Değirmenci, H.; Emlek, N. Relationship between C-reactive protein to albumin ratio and thrombus burden in patients with acute coronary syndrome. *Clin. Appl. Thromb. Hemost.* **2019**, *25*, 1076029618824418. [[CrossRef](#)] [[PubMed](#)]
107. Gorog, D.A.; Price, S.; Sibbing, D.; Baumbach, A.; Capodanno, D.; Gigante, B.; Halvorsen, S.; Huber, K.; Lettino, M.; Leonardi, S.; et al. Antithrombotic therapy in patients with acute coronary syndrome complicated by cardiogenic shock or out-of-hospital cardiac arrest: A joint position paper from the European Society of Cardiology (ESC) Working Group on Thrombosis, in association with the Acute Cardiovascular Care Association (ACCA) and European Association of Percutaneous Cardiovascular Interventions (EAPCI). *Eur. Heart J. Cardiovasc. Pharmacother.* **2021**, *7*, 125–140. [[CrossRef](#)] [[PubMed](#)]
108. Babalghith, A.O.; Al-kuraishy, H.M.; Al-Gareeb, A.I.; De Waard, M.; Sabatier, J.-M.; Saad, H.M.; Batiha, G.E.-S. The Potential Role of Growth Differentiation Factor 15 in COVID-19: A Corollary Subjective Effect or Not? *Diagnostics* **2022**, *12*, 2051. [[CrossRef](#)]
109. Wang, D.; Zhou, Y.; Qi, Y.; Song, M.; Yao, H.; Liao, C.; Lin, H.; Huang, M.; Zhuo, D.; Jiang, L. Antithrombotic Effects of Montelukast by Targeting Coagulation Factor XIa. Preprint. 2021.
110. Houard, X.; Ollivier, V.; Louedec, L.; Michel, J.B.; Back, M. Differential inflammatory activity across human abdominal aortic aneurysms reveals neutrophil-derived leukotriene B4 as a major chemotactic factor released from the intraluminal thrombus. *FASEB J.* **2009**, *23*, 1376–1383. [[CrossRef](#)]
111. Datta, Y.H.; Romano, M.; Jacobson, B.C.; Golan, D.E.; Serhan, C.N.; Ewenstein, B.M. Peptido-leukotrienes are potent agonists of von Willebrand factor secretion and P-selectin surface expression in human umbilical vein endothelial cells. *Circulation* **1995**, *92*, 3304–3311. [[CrossRef](#)]
112. Chan, S.J.; Ng, M.P.; Zhao, H.; Ng, G.J.; De Foo, C.; Wong, P.T.-H.; Seet, R. Early and sustained increases in leukotriene B4 levels are associated with poor clinical outcome in ischemic stroke patients. *Neurotherapeutics* **2020**, *17*, 282–293. [[CrossRef](#)]
113. Nichols, W.W.; Mehta, J.L.; Thompson, L.; Donnelly, W.H. Synergistic effects of LTC4 and TxA2 on coronary flow and myocardial function. *Am. J. Physiol.-Heart Circ. Physiol.* **1988**, *255*, H153–H159. [[CrossRef](#)]

114. Al-kuraishy, H.M.; Batiha, G.E.-S.; Faidah, H.; Al-Gareeb, A.I.; Saad, H.M.; Simal-Gandara, J. Pirfenidone and post-Covid-19 pulmonary fibrosis: Invoked again for realistic goals. *Inflammopharmacology* **2022**, 1–10. [[CrossRef](#)]
115. Tang, X.; Fuchs, D.; Tan, S.; Trauelsen, M.; Schwartz, T.W.; Wheelock, C.E.; Li, N.; Haeggström, J.Z. Activation of metabolite receptor GPR91 promotes platelet aggregation and transcellular biosynthesis of leukotriene C4. *J. Thromb. Haemost.* **2020**, *18*, 976–984. [[CrossRef](#)] [[PubMed](#)]