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21 Abstract

Mangroves are ecosystems interfacing terrestrial and marine environments submitted to extreme 22 23 abiotic factors (e.g. anoxia, flooding, salinity) producing stress on vegetation. Due to these stresses, we 24 hypothesized that mangroves potentially emit biogenic volatile organic compound (BVOC), particularly isoprenoids as they are defense compounds. Despite mangroves cover only about 5% of 25 26 the forest areas of the world, their emissions could impact air quality at the continental-ocean 27 interface. As a result, it is important to fill the gap in the knowledge about BVOC emissions from the canopy of the major mangrove trees. The aim of this study was thus to screen isoprenoid emissions of 28 29 the mangrove species. In this study, we sampled isoprenoid emissions of 15 species foliage among the 38 core species existing in the Indo-West Pacific (IWP) and the Atlantic Est Pacific (AEP) regions. 30 31 Sampling was performed using a branch-bag dynamic enclosure system and analyzed with gas chromatography coupled to mass spectrometry. Our analysis showed that mangrove tree species are 32 very low emitters suggesting that mangrove ecosystems would not strongly influence atmospheric 33 chemistry and air quality. 34

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36 Keywords

37 BVOC emissions; Isoprenoids; Mangroves; Flooded ecosystems; extreme environments

Mangrove forest weakly contribute to biosphere-atmosphere exchanges through BVOC.

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39 Highlights

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- Isoprenoid emissions of 15 mangrove species were investigated.
- Mangrove species studied are very low isoprenoid emitters.
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51 1. Introduction

Vegetation (croplands, shrublands, woodlands etc) is the most important source of volatile organic 52 compound (VOC) as it accounts for 1015 TgC.yr⁻¹, about 90% of total biogenic emissions (Guenther 53 et al. 1995). VOCs gather a wide variety of chemical compounds having a high vapor pressure (at 54 least 0.01 KPa) at ambient temperature. Among them, isoprenoids (including isoprene but also 55 monoterpenes and, to a lesser extent, sesquiterpenes), as well as highly volatile compounds such as 56 57 methanol, acetaldehyde, or formaldehyde, are mainly emitted (e.g. Saunier et al., 2017a). These compounds play a major environmental role on air quality and indirectly modulate climate conditions. 58 Indeed, in the presence of sunlight and nitrogen oxides (NOx), biogenic VOCs (BVOCs) contribute to 59 60 the formation of tropospheric ozone (Fehsenfeld et al. 1992) and secondary organic aerosols (Claeys et al. 2004; Ng et al. 2006), which in turn act on cloud formation and albedo (Makkonen et al. 2012). 61 Among BVOCs, isoprenoids are therefore key compounds for chemistry-model-transport and climate-62 63 change models that aim to forecast photo-chemical pollution episodes and their contribution in the future climate change (Makkonen et al. 2012). These two types of models rely on thousands of 64 65 publications that describe, since the 1980s, BVOCs emissions from plant species (Keenan et al. 2009), their dependence on environmental factors such as light and temperature (Guenther et al. 1993). Since 66 isoprenoid emissions are species-specific, it is crucial to investigate isoprenoid emissions from the 67 68 main vegetal formations at the global scale. For this purpose, temperate environments (e.g. 69 Mediterranean, continental, oceanic) have been widely studied in terms of BVOCs emissions (Owen et al. 2001; Ormeno et al. 2007; Saunier et al. 2017b), but plant species from extreme environments such 70 71 as mangroves have been clearly neglected (Rinnan et al. 2014; Exton et al. 2015).

Mangroves are extreme ecosystems interfacing terrestrial and marine environments. These forests are key elements of tropical coastal ecosystems as they provide many ecosystem services such as limiting erosion (e.g. cyclone, wave) and saline intrusion (Brander *et al.* 2012, Hilmi *et al.*, 2017), providing raw materials, food and habitat for fauna (e.g. migration, breeding, spawning), or being important carbon (C) sinks (Jakovac et al. 2020).They are found in two distinct regions: the Atlantic Est Pacific (AEP) and the Indo-West Pacific (IWP) (Spalding 2010) and account for 147,359 km2

78 (127,925–168,895) in 2020 (Bunting et al., 2022). In both geographical areas, there are several common genera but no common species. To handle their particular changing environmental conditions 79 80 (e.g. high salinity, flooding, anoxia), mangrove species have developed several specific defense mechanisms, including physical barriers (e.g. waxes, He et al. 2016) or chemical protections (e.g. 81 secondary metabolites, Naskar & Palit 2015). Indeed, specialized compounds can be produced and/or 82 emitted by plants to cope flooding and salinity as demonstrated for other species (Copolovici & 83 84 Niinemets 2016; Yang et al. 2018). However, to our knowledge, only a few studies have investigated 85 BVOC emissions from mangrove despite the large distribution of this unique ecosystem over the 86 world. For instance, Barr et al. (2003) focused on volatile compounds emitted by foliage of Red mangroves (Rhizophora mangle L.) in Florida, and found very low emissions of isoprene (< 0.1 87 $\mu g.g_{DW}^{-1}.h^{-1}$). Despite these reported low emissions, mangrove trees could have a non-negligible effect 88 89 on atmospheric processes since mangroves represent one of the most biomass-productive biomes on 90 our planet and cover about 75% of the coasts and estuaries of tropical regions (Ellison and Farnworth, 2001; FAO 2007). 91

The aim of this study was to perform for the first time a screening of isoprenoid emissions from 15 mangrove trees located in two geographical areas: the IWP (Vietnam) and the AEP (French Guyana) regions. We hypothesized that some mangrove species could be isoprenoid emitters in response to extreme conditions in mangrove ecosystems.

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97 2. Materials and methods

98 2.1 Species

99 From the 73 species recognized as "true" mangroves, we investigated 15 of the dominant mangrove 100 species among the 38 core species (Spalding 2010): Avicennia alba Blume, Avicennia germinans 101 (L.)L., Avicennia marina (Forsk) Vierh (Acanthaceae), Aegiceras corniculatum (L.) Blanco, Aegiceras 102 floridum Roem. & Schult. (Myrsinaceae), Bruguiera gymnorrhiza Lam., Ceriops zippeliana Blume, 103 Rhizophora apiculate Salvoza, Rhizophora racemose (Hochr.) Salvoza, Rhizophora stylosa Griff. 104 (Rhizophoraceae), Laguncularia racemose (L.) C.F. Gaertn., Lumnitzera littorea (Jack) Voigt, 105 Lumnitzera racemosa Wild. (Combretaceae), Sonneratia alba Sm. and Sonneratia caseolaris (L.) Engl. (Lythraceae). Due to the large surface area they occupy, these species could be the mostimportant contributors to the biosphere atmosphere relationship.

108 The 15 species were sampled in two distribution areas: 11 in the IWP (Vietnam) and 3 in the AEP 109 (French Guyana) regions. In Vietnam, the experimental site was located in the botanical garden of Can Gio District (CGBG), Ho Chi Minh City in the South of Vietnam (10°30'N 106°51'E). The Can Gio 110 mangrove is formed by the deltaic confluence of the Saigon, Dong Nai and Vam Co Rivers and it is 111 112 the first mangrove biosphere reserve in Vietnam (UNESCO/MAB Project 2000). This estuary is subject to semi-diurnal tides, and the tidal elevation ranges from 2 to 4 m. The climate is monsoonal, 113 with a dry season lasting from December to April and a wet season from May to November. In this 114 site, the annual mean temperature is 27.4°C with the highest (37.6°C) occurring in October and the 115 lowest (21.7°C) in February. The annual average rainfall ranges from 13000-14000mm with highest 116 monthly average in September (300-400mm). To the sampling site, not directly connected to the to the 117 118 main estuarine channel, the salinity ranges from 16 to 23 (Vinh et al. 2020).

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The second experimental site was located in French Guyana, along the Mahury River (04° 51.4429'N 052°15.8035'W). In this site, the climate is humid equatorial climate with 2 wet seasons and 2 dry seasons. French Guiana coast is mesotidal with semi-diurnal tides (Ray et al. 2020) with a tide elevation ranged from 0.92 m and 3.06 m in this site. The average annual temperature is 26.5 °C with a thermal amplitude of 3 °C at the most between the hottest and the coldest month and the average annual rainfall ranges from 2500 to 3000 mm y⁻¹. In the site the salinity ranges from 16 to 26 (Marchand et al. 2004).

Samplings were performed in the dry season during 2 field campaigns in April 2019 in Vietnam and
October 2019 in French Guyana. The environmental conditions in terms of temperature (°C), relative
humidity (%) occurring during the field campaigns were recorded using a HOBO device (Onset,
USA). Photosynthetically Active Radiation (PAR) was measured using a quantum sensor (PAR- SA
190[®], LI- COR, USA) (Table 1).

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133 2.2 Isoprenoid sampling

134 For each species, five mature trees (2-5m) were sampled during each field campaign and sampling site (only exception: 6 Avicenia alba were sampled), for a total of 76 trees sampled (see Table 2 for 135 136 details). The sampling procedure was described in detail in Saunier et al. (2017). On each sampled tree, one branch sun-exposed and without flower and having about 6 leaves was enclosed in a 137 polytetrafluoroethynele (PTFE) chamber of about 10 L. A continuous atmospheric air flow, filtered 138 with VOC and ozone scrubbers, flowed through the chamber at 1 L.min⁻¹. After a 30 min system 139 stabilization period, air from the enclosure was collected during 30 min at 0.2 L.min⁻¹ on cartridges 140 141 (two beds of adsorbants: tenax and cartotrap B), one for isoprene and one for other isoprenoids. Inlet 142 and outlet air flows were controlled by mass flow controllers (Porter Instrument, USA). Cartridges were then stored at -20°C until analysis (portable freezer in the field and classical one in the 143 144 laboratory). Samplings were performed between 10 h and 15 h (local time) on fully developed leaves. 145 Twice a day, blanks were performed to control inlet air quality.

One test was performed in April 2019 to check the reliability of our protocol of transport and storage.
According to this test, no significant difference was detected between an analysis right after the
sampling or after a period of transport/storage (Fig. S1 in supplementary files).

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150 2.3 GC-MS analyses

151 Cartridge analyses were conducted by gas chromatography (model 6890N, Agilent technologies, 152 USA) coupled to mass spectrometry (model 5973, Agilent Technologies, USA). VOC were extracted 153 using thermodesorption system (TDS3/CIS, Gerstel, Germany) using helium as carrier gas (50 154 mL.min⁻¹) at 250 °C during 10 min. The extracted compounds were concentrated on-line by cryo-155 trapping at -50 °C using liquid nitrogen. The sample was then injected by rapid heating to 250°C at 156 10°C/min. Initial oven temperature was 40 degrees hold for 5 minutes and then increasing by 3 157 degrees per minute to 250 °C maintained for 15 min.

Two distinct chromatographic methods were used, one to detect and quantify isoprene (C5) and another one to analyze C10-C15 compounds. Isoprene was analysed using a capillary Al/KCl column (Agilent Technologies, USA; 30 m x 0.25 mm x 5 μ m) as described in Saunier *et al.* (2017b) whereas other isoprenoids were separated on a HP-5MS column (Agilent Technology, USA; 30 m x 0.25 mm x 162 $0.25 \ \mu\text{m}$) as shown in Ormeño *et al.* (2007). For both analyses helium was the carrier gas at a constant 163 flow rate of 1 mL.min⁻¹. Isoprenoids were identified based on the co-injection of analytical standards 164 (Sigma-Aldrich, Germany), the Kovàts' index calculation through the injection of alkanes (C₈-C₂₀) as 165 well as the comparison of mass spectra to the NIST library (2011). BVOC emission rate (ER) were 166 calculated according to Saunier *et al.* (2017).

167 2.4 Statistical analyses

The statistical analyses were performed with R software (version 4.2.1). After having checked the normality of the data set, one way-ANOVA or Kruskal-Wallis tests were performed for isoprene emissions and other BVOC emissions according to the species. Dunn post-hoc tests were performed to detect the species with statical differences in their emissions.

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173 **3. Results**

The 15 studied mangrove species are low isoprenoid emitters (< 1.200 ng.gDW⁻¹.h⁻¹, Table 2). In details, species from the IWP region are the lower isoprene emitters with emissions rates ranging between 0.3 and 1.6 ng.gDW⁻¹.h⁻¹ while species from the AEP region present a larger range of isoprene emission rate with values ranging between 0.1 and 2.1 ng.g_{DW}⁻¹.h⁻¹. Isoprene emissions represent 0.3, 0.5 and 2.1% of total emissions for *A. germinans, L. racemosa* and *R. racemosa*, respectively.

Besides, those species are also low emitters of other BVOC ($200 - 1.2 \text{ ng. g}^{-1}\text{DW.h}^{-1}$, Table 2 and 3) and the main BVOC detected are monoterpenes but also benzenoids as well as methyl salicylate. Sesquiterpene emissions are very low, closed to the detection limit of the analyzers.

Other BVOC such as benzaldehyde, benzeneacetaldehyde, acetophenone or methyl salicylate
contribute (Table 3) significantly to the total BVOC emission specifically for *A. germinans* and *L. racemosa* with 76 and 82% of the total emissions, respectively.

186 *Rhizophora racemosa* is the highest emitter of the mangrove species with an average emission rate of 187 about 1200 ng.g⁻¹DW.h⁻¹. Contrary to the two other species from AEP region, *R. racemosa* emissions 188 are dominated by monoterpenes which represent about 93% of its total emissions. The three dominant 189 emitted monoterpenes of *R. racemosa* are α -pinene, sabinene and β -pinene.

191 4. Discussion

192 The low isoprenoids emissions observed for the 15 mangrove species sampled in the two geographical 193 regions (IWP and AEP) are in agreement with previous findings (Barr et al. 2003). In addition, Guenther et al. (1994) reported values below 100 ngC.g⁻¹.h⁻¹ for emissions from the genera Avicennia, 194 Laguncularia and Rhizophora in the United States of America, referring to the works of Zimmernan 195 196 (1979) and Evans *et al.* (1982). Moreover, the dominant monoterpenes α -pinene and β -pinene detected 197 here are also present in the study of Barr et al. (2003). Sabinene was detected in the essential oil of 198 Melaleuca cajuputi, a species not directly related to the mangrove but often found in the back of 199 mangroves (Sharif et al. 2009). The presence of monoterpenes but also benzenoids as well as methyl 200 salicylate in emissions from mangrove species is not surprising as they are observed in many other 201 biogenic emissions (Rodriguez-Saona et al. 2011; Gentner et al. 2014).

The two geographic areas of mangroves have no species in common but share *Avicennia* L. and *Rhizophora* L., the most characteristic genera of mangrove systems worldwide (Robert *et al.*, 2009). In our study, several species belonging to these two genera were sampled and show qualitatively different but always small emissions. These results agree with those of Lorto (2002) who showed that within the genus *Quercus*, that have a wide geographical distribution, non-emitting species and emitting species of isoprenoids (isoprene and/or terpenes) can be found.

Although they share a common ecological niche and must adapt to similar environmental conditions, mangroves are nonetheless a genetically diverse group of flowering plants (Duke, 2002). This may explain the observed differences in their emission spectra, which are also driven by internal factors such as genetics (Peñuelas and Llusià, 2001).

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This emission rate from mangroves is low compared to the emission rates of most other tropical plant species. For instance, Geron *et al.* (2002) performed a screening of BVOC from twenty species in a lowland wet forest located in Costa Rica. Half of these common species emitted isoprene at a rate ranging from 10 to 126 μ g.gDW⁻¹.h⁻¹ or about 100 times higher than the isoprene emissions of the 15 mangrove species under study. The same trend is observed for the other BVOCs that are generally emitted by tropical species at higher rates than mangrove species (Singh *et al.* 2011; Bracho-Nunez *et al.* 2013).

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221 We can confirm that mangroves invest in isoprenoid emissions as chemical defences against the extreme environmental conditions that typically occur in their environment (e.g. flooding, salinity) but 222 to a small extent. Indeed, mangroves have other strategies that ensure very good adaptation to these 223 224 extreme conditions. They possess strategies such as osmolyte production which absorb salt as previously observed in Avicennia genus and other halophytes (Khan et al. 2016) (Slama et al. 2015). 225 226 However, samplings were performed only on a short time. As a result, it cannot be excluded that we 227 missed plant response to salinity and or flooding that could have caused emission puffs. For example, 228 Eucalyptus globulus, a non-halophyte plant, exhibits puffs of isoprene only when salt stress is relieved 229 (Loreto & Delfine 2000). Velikova et al. (2012) also observed important emissions of volatile 230 isoprenoids from *Citrus* leaves, mainly limonene, during flooding with saline water.

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232 Moreover, it would be worth investigating the emission of highly volatile organic compounds such as 233 acetaldehyde and methanol since mangroves face anoxic conditions that could lead to alcoholic 234 fermentation processes and, as a result, ethanol could be produced and transformed to acetaldehyde 235 and to a lesser extent to acetic acid (Oikawa & Lerdau 2013). More generally, plants can also emit 236 methanol through growth process (Hüve et al. 2007) and acetone through acetoacetate decarboxylation (Fall 2003). Acetaldehyde and acetic acid impact air quality by contributing to the atmospheric 237 oxidative capacity and forming toxic secondary pollutants such as formaldehyde (Oikawa & Lerdau 238 2013). In addition, other halophytes have been shown to be a source of methyl halides and 239 240 dimethylsulphoniopropionate (DMSP) (Otte et al. 2004; Rhew & Mazéas 2010). One experimental 241 study conducted under greenhouse conditions also demonstrated that mangrove trees emit methyl halides with possible atmospheric impact (Manley et al. 2007). However, this study emphasizes the 242 need for field experiments to check for the occurrence of these emissions from mangroves. Further 243 investigations are thus required to explore if mangroves are a source of halide VOC. 244

246 **4.** Conclusion

Our results show that mangrove trees are very low isoprenoid emitters. Only *R. racemosa* can be regarded as a non-negligeable source though monoterpene emissions still low (~ 1000 ng.gDW.h⁻¹). Such results suggest that mangrove does not strongly affect local or regional atmospheric chemistry through BVOC emissions despite the large distribution of this ecosystem over the globe. However, despite the low-rate emission, the data from this study can be used in atmospheric chemistry models to study the impact of biogenic VOC emissions on air quality in areas where mangroves are present.

It is still worthy to note that we only made punctual samplings and that continuous monitoring of isoprenoid emissions should be carried out to check for potential seasonal variations. Moreover, as far as our results showed important differences between plant species, it could be interesting to broaden the sampling design including other sites (African west coast and Caribbean Islands) as they include mangrove species that have not been investigated.

It could be also interesting to focus on other compounds than isoprenoids such as highly volatile or halogenated compounds which might be tackled in future studies focused on BVOC emissions from mangroves.

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262 **5.** Acknowledgements

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270 **6.** Author contributions:

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 review & editing Amélie Saunier: Investigation, Methodology, Formal analysis, Data curation,

273 Writing - original draft Henri Wortham: Conceptualization, Investigation, Methodology, Visualization, Writing - review & editing Elena Ormeño: Conceptualization, Methodology, Writing 274 - review & editing Magali Proffit: Conceptualization, Methodology, Writing - review & editing 275 Caroline Lecareux: Resources, Methodology, Writing - review & editing Stéphane Greff: 276 Resources, Methodology, Writing - review & editing Dao Van Tan: Investigation, Resources, 277 278 Methodology Mai Sy Tuan: Resources, Huynh Duc Hoan: Resources, Bui Nguyen The Kiet: Resources Dounia Dhaou: Investigation, Methodology Virginie Baldy: Investigation, Methodology, 279 Writing - review & editing Anne Bousquet-Mélou: Funding acquisition, Project administration, 280 Supervision, Conceptualization, Investigation, Methodology, Visualization, Writing - review & 281 editing 282

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- **Table 1:** Means (± SE) of temperatures (T, °C), relative humidity (RH, %) and Photosynthetic Active
- 422 Radiation (PAR, μ mol.m⁻².s⁻¹) between 10 h and 15 h (sampling time for BVOC, local time) during 423 each field campaign. PAR in March 2019 could not be calculated due to technical problems with the 424 HOBO sensor.
 - Site
 Sampling
 T
 RH
 PAR

 Vietnam
 April 2019
 33.0 ± 0.2
 66.0 ± 0.5
 301 ± 23

 French Guyana
 October 2019
 35.0 ± 0.3
 60.0 ± 0.6
 603 ± 55

Table 2: Emission rates (ER, ng.gDW⁻¹.h⁻¹) of 15 mangrove species in terms of isoprene and other

426 BVOC (see Table 3 for details) detected with GC-MS, n.d.: no compound detected. Values are means

427 \pm SE. Statistical test are presented for each BVOC type and site. Lower case letters denote post-hoc

428 test differences at 95% confidence level (Due to a problem during the analysis, we lost a series of 429 samples from French Guyana and therefore only have 4 samples with results)

Sites	Species	ER isoprene*	n	ER other BVOC	n
		C5		$(C_{10} - C_{15})$	
Vietnam	Aegiceras corniculatum	$1.5 \pm 1.0^{\mathrm{a}}$	5	n.d.	5
	Aegiceras floridum	$1.0 \pm 0.4^{\mathrm{a}}$	5	n.d.	5
	Avicennia alba	0.3 ± 0.1^{a}	6	n.d.	6
	Avicennia marina	0.2 ± 0.1^{a}	5	n.d.	5
	Bruguiera gymnorrhiza	n.d.	5	n.d.	5
	Ceriops zippeliana	$0.8\pm0.5^{\mathrm{a}}$	5	n.d.	5
	Lumnitzera racemosa	$1.6\pm0.8^{ m b}$	5	n.d.	5
	Lumnitzera littorea	0.1 ± 0.1^{a}	5	n.d.	5
	Rhizophora apiculata	$0.3 \pm 0.2^{\mathrm{a}}$	5	n.d.	5
	Rhizophora stylosa	$0.7\pm0.4^{\mathrm{a}}$	5	n.d.	5
	Sonneratia alba	0.3 ± 0.2^{a}	5	n.d.	5
	Sonneratia caseolaris	1.3 ± 0.4^{a}	5	n.d.	5
		Kruskall-Wallis test	p=0.02		
French	Avicennia germinans	0.5 ± 0.3^{a}	5	212 ± 38	4
Guyana	Laguncularia racemosa	$0.3 \pm 0.3^{\mathrm{a}}$	5	222 ± 97	4
	Rhizophora racemosa	$2.1\pm0.8^{\rm b}$	4	1176 ± 626	4
	_	Kruskall-Wallis test p=0.04		Anova test p=0.09	

430

432	Table 3: Emissions rates (ng.gDW ⁻¹ .h ⁻¹) of Avicennia germinans, Laguncularia racemosa and
433	Rhizophora racemosa. Asterisks indicate the co-injection of analytical standards to perform compound
434	identifications and quantifications. Means \pm SE.

Compounds	A. germinans	L. racemosa	R. racemosa
Monoterpenes			
α-pinene [*]	18.5 ± 3.1	3.9 ± 2.1	393.6 ± 220.0
camphene	0.1 ± 0.1	n.d.	32.8 ± 25.4
sabinene	0.6 ± 0.6	n.d.	393.3 ± 268.0
β-pinene*	0.4 ± 0.4	0.1 ± 0.1	102.5 ± 68.2
β-myrcene	19.9 ± 6.2	7.3 ± 4.1	13.0 ± 2.7
α-terpinene	n.d.	n.d.	4.9 ± 2.9
p-cymene*	3.5 ± 2.0	3.4 ± 1.1	8.4 ± 3.6
Limonene*	2.8 ± 1.7	4.7 ± 1.9	72.8 ± 43.9
β -ocimene (Z)	n.d.	2.1 ± 1.6	n.d.
β -ocimene (E)	n.d.	10.9 ± 4.6	0.4 ± 0.4
γ-terpinene	n.d.	n.d.	12.3 ± 6.8
sabinene hydrate	n.d.	n.d.	0.4 ± 0.3
E-DMNT	0.5 ± 0.5	2.0 ± 1.8	n.d.
linalool	n.d.	3.4 ± 3.4	3.4 ± 2.4
allo-ocimene	n.d.	0.3 ± 0.3	0.2 ± 0.2
camphor	n.d.	n.d.	2.1 ± 1.3
pinocarvone	n.d.	n.d.	5.3 ± 3.6
borneol	0.4 ± 0.4	0.6 ± 0.6	40.1 ± 20.9
terpinen-4-ol	n.d.	n.d.	1.1 ± 0.5
myrtenal	n.d.	n.d.	4.9 ± 3.1
verbenone	n.d.	n.d.	0.9 ± 0.5
Sesquiterpenes			
β-caryophyllene	3.9 ± 3.9	n.d.	n.d.
α -bergamotene (Z)	0.1 ± 0.1	n.d.	0.7 ± 0.7
Other BVOC			
benzaldehyde	73.6 ± 11.3	89.1 ± 49.3	42.2 ± 8.3
benzeneacetaldehyde	2.1 ± 0.4	1.2 ± 1.1	0.8 ± 0.2
acetophenone	83.6 ± 14.2	90.2 ± 36.0	38.1 ± 5.7
methyl salicylate	2.6 ± 2.6	2.7 ± 2.5	0.4 ± 0.4

435 n.d. = not detected, E-DMNT = (E)-4,8-Dimethyl-1,3,7-nonatriene