



**HAL**  
open science

## Therapeutic advances in metastatic pancreatic cancer: a focus on targeted therapies

Anthony Turpin, Cindy Neuzillet, Elise Colle, Nelson Dusetti, Remy Nicolle, Jerome Cros, Louis De Mestier, Jean-Baptiste Bachet, Pascal Hammel

### ► To cite this version:

Anthony Turpin, Cindy Neuzillet, Elise Colle, Nelson Dusetti, Remy Nicolle, et al.. Therapeutic advances in metastatic pancreatic cancer: a focus on targeted therapies. *Therapeutic Advances in Medical Oncology*, 2022, 14, 10.1177/17588359221118019 . hal-04084980

**HAL Id: hal-04084980**

**<https://amu.hal.science/hal-04084980>**

Submitted on 6 Jun 2023

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

# Therapeutic advances in metastatic pancreatic cancer: a focus on targeted therapies

Anthony Turpin<sup>1</sup>, Cindy Neuzillet, Elise Colle, Nelson Dusetti, Rémy Nicolle, Jérôme Cros, Louis de Mestier, Jean-Baptiste Bachet and Pascal Hammel

**Abstract:** Mortality from pancreatic ductal adenocarcinoma (PDAC) is increasing worldwide and effective new treatments are urgently needed. The current treatment of metastatic PDAC in fit patients is based on two chemotherapy combinations (FOLFIRINOX and gemcitabine plus nab-paclitaxel) which were validated more than 8 years ago. Although almost all treatments targeting specific molecular alterations have failed so far when administered to unselected patients, encouraging results were observed in the small subpopulations of patients with germline *BRCA 1/2* mutations, and somatic gene fusions (*neurotrophic tyrosine receptor kinase*, *Neuregulin 1*, which are enriched in *KRAS* wild-type PDAC), *KRAS* G12C mutations, or microsatellite instability. While targeted tumor metabolism therapies and immunotherapy have been disappointing, they are still under investigation in combination with other drugs. Optimizing pharmacokinetics and adapting available chemotherapies based on molecular signatures are other promising avenues of research. This review evaluates the current expectations and limits of available treatments and analyses the existing trials. A permanent search for actionable vulnerabilities in PDAC tumor cells and microenvironments will probably result in a more personalized therapeutic approach, keeping in mind that supportive care must also play a major role if real clinical efficacy is to be achieved in these patients.

**Keywords:** immunotherapy, *KRAS*, metastatic, *NRG1*, *NTRK*, pancreatic cancer, PARP, precision medicine

Received: 17 April 2022; revised manuscript accepted: 18 July 2022.

## Introduction

Pancreatic ductal adenocarcinoma (PDAC) is one of a small category of cancers whose mortality is increasing and which will become the second leading cause of cancer death by 2030 in the United States and Europe.<sup>1</sup>

In the early 2010s, the use of the 5-fluorouracil, irinotecan, and oxaliplatin (FOLFIRINOX), and gemcitabine plus nab-paclitaxel combinations improved the prognosis in selected patients.<sup>2,3</sup> However, targeted therapies and immunotherapies, which have resulted in significant progress in patients with other tumor types, have so far failed in PDAC due to the biological characteristics of this entity.

Thus, the development of new, effective, and personalized treatments for PDAC is as urgent as

ever. At the same time, there have been recent, encouraging results with the poly (ADP-ribose) polymerase (PARP) inhibitor, olaparib to treat PDAC in patients with *BRCA1* or *BRCA2* gene germline mutations (gBRCAm)<sup>4</sup> and in the development of various molecules targeting specific somatic genetic alterations.<sup>5–7</sup> Nevertheless, the proportion of patients who can actually benefit from these treatments is still low either because these alterations are rare and can only be identified by technical means not available in all centers, or because validation to administer these treatments has not been obtained based on large randomized clinical trials.

This review discusses the hopes and limitations of recent advances in personalized treatment in patients with PDAC.

*Ther Adv Med Oncol*

2022, Vol. 14: 1–27

DOI: 10.1177/  
17588359221118019

© The Author(s), 2022.  
Article reuse guidelines:  
sagepub.com/journals-  
permissions

Correspondence to:

**Pascal Hammel**  
Department of Digestive  
and Medical Oncology,  
Hôpital Paul Brousse  
(AP-HP), 12 Avenue Paul  
Vaillant-Couturier, Villejuif  
94800, University of Paris  
Saclay, France  
[pascal.hammel@aphp.fr](mailto:pascal.hammel@aphp.fr)

**Anthony Turpin**  
Department of Medical  
Oncology, CNRS  
UMR9020, Inserm UMR-S  
1277-Canther-Cancer  
Heterogeneity, Plasticity  
and Resistance to  
Therapies, University Lille,  
CHU Lille, Lille, France

**Cindy Neuzillet**  
Department of Medical  
Oncology, Curie Institute,  
Versailles Saint-Quentin  
University, Paris-Saclay  
University, Saint-Cloud,  
France

**Elise Colle**  
Department of Digestive  
and Medical Oncology,  
Hospital Paul Brousse  
(AP-HP), Villejuif,  
University of Paris Saclay,  
France

**Nelson Dusetti**  
Cancer Research Center of  
Marseille, CRCM, Inserm,  
CNRS, Paoli-Calmettes  
Institut, Aix-Marseille  
University, Marseille, France

**Rémy Nicolle**  
Centre de Recherche sur  
l'Inflammation, INSERM,  
U1149, CNRS, ERL 8252,  
Université de Paris Cité,  
Paris, France

**Jérôme Cros**  
Department of Pathology,  
University of Paris Cité,  
Hospital Beaujon (AP-HP),  
Clichy, France

**Louis de Mestier**  
Department of  
Gastroenterology and  
Pancreatology, University of  
Paris Cité, Hospital Beaujon  
(AP-HP), Clichy, France

**Jean-Baptiste Bachet**  
Department of  
Gastroenterology and  
Digestive Oncology,  
Pitié-Salpêtrière Hospital,  
Sorbonne University, UPMC  
University, Paris, France

### Therapeutic targets of interest for standard of care

#### HRD/BRCAness tumors and PARP inhibitors

Both *BRCA1* and *BRCA2* proteins are critical to the repair of double-strand DNA breaks by homologous recombination repair (HRR), a form of DNA repair using a homologous DNA sequence to guide this process at the double strand break (DSB; Table 1 and Figure 1). This conservative mechanism restores the original DNA sequence at the site of DNA damage (Lord and Ashworth).<sup>8</sup> Although targeted therapies have failed in PDAC in molecularly unselected patients,<sup>9</sup> progress has been made in recent years identifying small molecular subgroups of patients with targetable alterations.

While the *BRCA* germline mutations are the most notable recent example, they are not alone. The concept of synthetic lethality was applied clinically using PARP inhibitors (PARPi) as single agent for maintenance therapy in patients with metastatic PDAC and the germline *BRCA1* or *BRCA2* mutation (gBRCAm) following platinum-based induction chemotherapy. For the first time, the POLO phase III trial confirmed the efficacy of a biomarker-driven approach in metastatic PDAC patients with the PARPi, olaparib. A significant benefit to progression-free survival (PFS) was demonstrated with maintenance olaparib *versus* placebo [median PFS: 7.4 months *versus* 3.8 months; hazard ratio (HR): 0.53; 95% confidence interval (CI): 0.35–0.82;  $p = 0.004$ ].<sup>4</sup>

**Table 1.** Progress and current limitations of medicine precision in PDAC treatment.

Target	ESMO-ESCAT evidence	Progress/promising	Current limitations	Questions	References
HRD/BRCAness	I-A	- Olaparib: positive phase III for tumor control (PFS) - Long duration of efficacy in patients responders	- 40% of primary resistance - Access and delay for screening	- Prediction of failure - Role in non-core mutations - Efficacy in somatic mutations - Management of resistance - Role of ATR inhibitors - Combination with chemo/CPI	POLO trial phase III Golan <i>et al.</i> <sup>4</sup>
dMMR/MSI-H	I-C	CPI (pembrolizumab): tumor response/control after failure of chemotherapies	- Efficacy in only ~20% of patients	- Efficacy germinal <i>versus</i> hypermethylation - Combination with CPI	KEYNOTE-158 (phase II) Aguirre <i>et al.</i> <sup>10</sup>
TMB-high	III-A	CPI (pembrolizumab): tumor response/control after failure of chemotherapies	- No data in PDAC	- Prediction of efficacy in PDAC	KEYNOTE-158 (phase II) Wang <i>et al.</i> <sup>11</sup>
NTRK fusion	I-C	Larotrectinib: tumor response/control after failure of chemotherapies	- Prevalence <1%	- Rarity of <i>NRG1</i> fusions - Only in KRAS wild-type tumors	Phase II Cocco <i>et al.</i> <sup>12</sup>
NRG1	II-B	Zenocutuzumab: tumor response/control after failure of chemotherapies	- Prevalence <1%	- Rarity of <i>NRG1</i> fusions - Only in KRAS wild-type tumors - Diversity of fusion partners	Schram (phase II) Boeck <i>et al.</i> <sup>13</sup> NCT02912949
KRAS G12C	II-B	- Encouraging results with sotorasib and adagrasib - RMC-6291 - Newly synthesized KRASG12C through increased EGFR and AURKA signaling	- Low prevalence [1%] - Accessibility to routine analysis and delay for results	- Combination with anti-EGFR - Timing of use (frontline ? Maintenance after chemo ? As second line ?)	KRYSTAL-1 study (phase II) Lito <i>et al.</i> <sup>14</sup> NCT03785249

(Continued)

**Table 1.** (Continued)

Target	ESMO-ESCAT evidence	Progress/promising	Current limitations	Questions	References
KRAS G12D	III-A	<ul style="list-style-type: none"> <li>- Encouraging results in other tumor type (lung cancer), ~40% of PDAC might be concerned</li> <li>- Extensive structure-based drug design high-resolution X-ray crystal structures</li> <li>- MRTX1133 binds switch II pocket, inhibits proteins interactions needed for activation of KRAS pathway</li> <li>- RMC-6236: tri-complex inhibitors of the oncogenic, GTP-bound form of KRASG12C overcome RTK-mediated escape mechanisms</li> <li>- mRNA-5671/V941) alone or combined with anti-PD-1</li> <li>- Short interfering RNA packaged within mesenchymal stem cell-derived exosomes</li> </ul>	<ul style="list-style-type: none"> <li>- Accessibility to routine analysis and delay for results</li> <li>- Lacks a reactive residue adjacent to switch II pocket</li> <li>- Activation of feedback networks</li> <li>- Decreased dependence to glycolysis, acquisition of mesenchymal phenotype</li> </ul>	<ul style="list-style-type: none"> <li>- Prediction of efficacy in PDAC</li> </ul>	<p>Amodio <i>et al.</i><sup>15</sup> NCT03608631 NCT04111458 NCT03948763 NCT03608631</p>
BRAFV600E	III-A	Dabrafenib + trametinib: sustained response reported	<ul style="list-style-type: none"> <li>- Prevalence 3% of PDAC but 10% of RASwt tumors</li> <li>- Accessibility to routine analysis and delay for results</li> </ul>	<ul style="list-style-type: none"> <li>- Combination with anti-EGFR</li> <li>- Combination with CPI</li> </ul>	<p>Retrospective series Case reports Wang <i>et al.</i>,<sup>16</sup> Hofmann <i>et al.</i><sup>17</sup></p>
MEK	III-A/V	Combination with CPI (e.g. selumetinib) or SOS1	No positive trials with EGFR inhibitors in advanced PDAC in phase II	Trials testing combinations with CPI are ongoing	<p>Phase II trials Moore <i>et al.</i>,<sup>18</sup> Huang <i>et al.</i><sup>19</sup></p>
Metabolism	V	Mitochondrial Asparagine: positive signals in phase II or phase IIb	<ul style="list-style-type: none"> <li>- Devimistat (mitochondrial enzymes) and encapsulated L-Asparaginase : negative phase III trial</li> </ul>	Further areas of development: to be defined	<p>Phase III trials - Marabelle <i>et al.</i><sup>20</sup> - Ying <i>et al.</i>,<sup>21</sup> Weinstein <i>et al.</i><sup>22</sup></p>
ECM	V	HA	Failure of PEGPH20 (HA) in phase III	Unexplained detrimental effect with mFOLFIRINOX Further areas of development?	<p>Phase III trials Yamamoto <i>et al.</i>,<sup>23</sup> Hindson <i>et al.</i><sup>24</sup></p>
	V	FAK inhibitors	Negative results in phase II with GSK2256098 and trametinib (NCT02428270)	Defactinib currently tested in combination with pembrolizumab and gemcitabine	<p>Phase II trial Melisi <i>et al.</i><sup>25</sup></p>
	V	BTK inhibitors	Negative phase III trial Limited efficacy with combination to CPI	Trials testing combinations with CPI are ongoing	<p>Phase I-II trials Rhim <i>et al.</i><sup>26</sup> Özdemir <i>et al.</i><sup>27</sup> Phase III trial Chen <i>et al.</i><sup>28</sup></p>
	IV-A	PI3K inhibitors	No data from clinical trials in PDAC	Trials testing combinations of PI3K/mTOR inhibitor with the CDK4/6 inhibitor trials are ongoing	<p>Trials ongoing NCT03065062</p>
	IV-A	CTGF inhibitors/high response rate and surgical resection with gemcitabine-nab-paclitaxel	No phase III trials	Combination of pamrevlumab plus gemcitabine-nab-paclitaxel or FOLFIRINOX is currently tested	<p>Phase I-II trials Dawson <i>et al.</i><sup>29</sup> Trials ongoing NCT03941093, NCT04229004</p>

(Continued)

Table 1. (Continued)

Target	ESMO-ESCAT evidence	Progress/promising	Current limitations	Questions	References
Stromal signatures	IV-B	GemPred signatures to predict chemotherapy efficacy (gemcitabine or FOLFIRINOX-based)	No data from clinical trials in PDAC	Feasibility	Trials ongoing: PACSign
ATR	III-A	ATR inhibitors/association with gemcitabine	- Preclinical data - Data in other tumors (ovarian cancer) but no data in PDAC	- Prediction of efficacy in PDAC according to <i>TP53/KRAS</i> mutations	
CAR-T cell	II-B	CAR-T cells with specific biomarkers - Mesothelin - CEA, etc.	- Only data from early trials - Toxicity	Feasibility	Phase I–II trials Foote <i>et al.</i> , <sup>30</sup> Henze <i>et al.</i> , <sup>31</sup> Trials ongoing
Vaccines	IV-A	Whole tumor cell vaccine (GVAX) and peptide-based vaccines	- Discordant results - No phase III trials	- mRNA vaccines, combinations with immunotherapy, selection according HLA-A2	NCT03806309
Microbiota	IV-A	Manipulation of the intratumoral microbiota is emerging as a new potential therapeutic approach in PDAC	Preclinical data	- Many bacteria involved - Feasibility	Trials ongoing: antibio-PAC

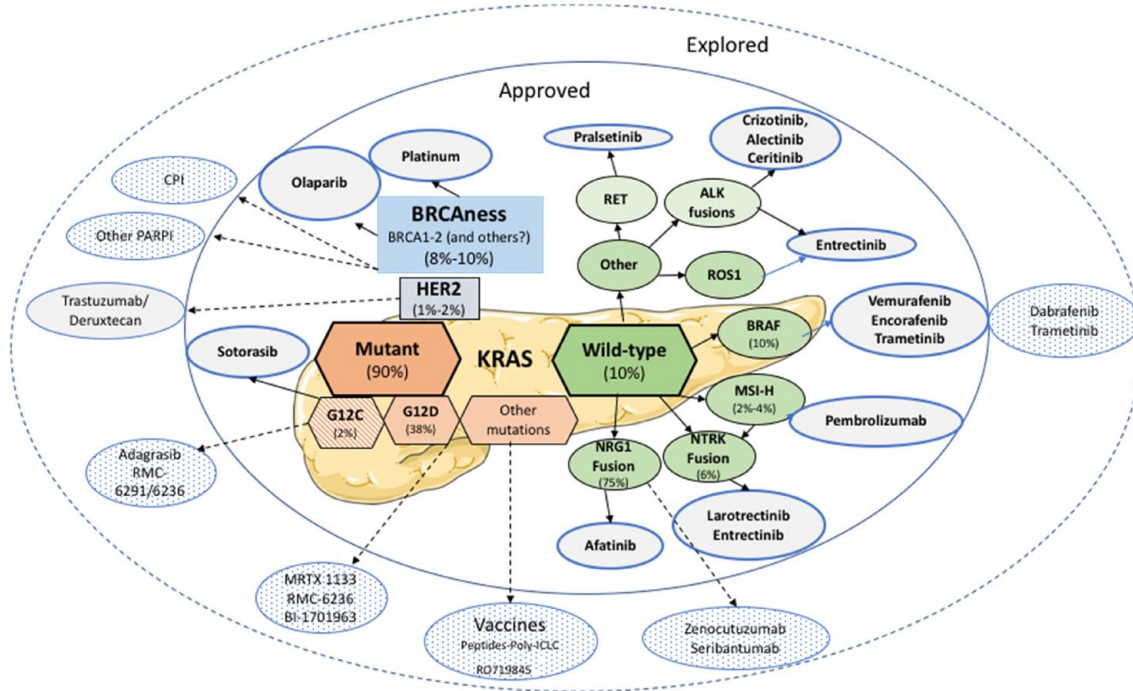
ATR, Rad3-related; BTK, bruton tyrosine kinase; CEA, carcinoembryonic antigen; CPI, checkpoint inhibitor; CTGF, connective tissue growth factor; ECM, extracellular matrix; EGFR, epidermal growth factor receptor; HA, hyaluronic acid; HLA, human leukocyte antigen; NTRK, neurotrophic tyrosine receptor kinase PAC, paclitaxel; PDAC, pancreatic ductal adenocarcinoma; PFS, progression-free survival; TMB, tumor mutational burden.

The impressive median overall survival (OS) in these patients with metastatic PDAC (19 months from the beginning of maintenance therapy) confirmed that gBRCAm confers sensitivity to both platinum-based chemotherapies and PARPi, resulting in favorable outcomes. In addition, Park *et al.*<sup>32</sup> later showed that the maximal therapeutic benefit with first-line platinum-based chemotherapy can be obtained in case of biallelic mutation in core HRR genes (*BRCA1*, *BRCA2*, and *PALB2*) compared to non-core genes (*ATM*, *BAP1*, *BARD1*, etc.). However, the POLO study did not show an advantage to OS in the olaparib arm compared to the placebo arm (median 19.0 *versus* 19.2 months; HR: 0.83; 95% CI: 0.56–1.22; *p* = 0.3487).<sup>33</sup> One explanation could be the extensive use of effective therapies following discontinuation of the study, resulting in tumor control in the placebo arm. This included reintroduction of platinum chemotherapy as well as the use of PARPi in 27.1% of patients, which was not allowed in the protocol. Despite this, the secondary endpoints (time to second disease progression or death, first and second subsequent cancer therapies or death and discontinuation), and estimated 3-year OS after randomization

(33.9% *versus* 17.8%) were in favor of the olaparib arm.<sup>33</sup> Olaparib was approved by the Food and Drug Administration and European Medicines Agency at the end of 2019 for maintenance therapy in patients with germline *BRCA1* or *BRCA2* mutations (5–7% of PDAC patients, with *BRCA2* representing about two-thirds of the gBRCAm) and metastatic PDAC, which has been controlled after at least 16 weeks of first-line platinum-based chemotherapy.<sup>34</sup> Systematic gBRCAm screening in PDAC patients is recommended in the National Comprehensive Cancer Network clinical guidelines.<sup>35</sup>

Thus, for the first time, targeted therapy has been shown to be effective in the treatment of metastatic PDAC in a phase III trial, although these results have raised several questions that will be discussed below.

Moreover, in a randomized phase II trial, O'Reilly *et al.*<sup>36</sup> showed that there was no advantage to survival with a combination of upfront platinum-based chemotherapy and PARPi in gBRCAm PDAC patients. This suggests that patient selection based on tumor control during platinum



**Figure 1.** Potential active drugs (approved and explored) in PDAC according to actionable targets.

chemotherapy can help identify patients who will benefit the most from PARPi. Indeed, about 20% of PDAC patients with gBRCAm are not platinum sensitive.<sup>37</sup>

**Resistance to PARPi.** Approximately 40% of patients have a primary resistance to olaparib and a secondary resistance often occurs after several months of PARPi administration. There are numerous mechanisms of resistance to PARPi: (i) restoration of the HRR pathway through *BRCA* reversion mutations and/or epigenetic upregulation of the gene, (ii) loss of *53BP1* with rewiring of the non-homologous end-joint repair pathway, (iii) replication fork protection, (iv) upregulation of cellular drug efflux pumps, (v) reduction in PARP1 activity, or (vi) changes in the tumor microenvironment (TME).<sup>38,39</sup> Integrating transcriptomic data with compound perturbation profiles could help identify drugs that can exploit induced vulnerability and thus overcome therapeutic resistance in multiple resistant clones.<sup>40</sup> Finally, another important approach would be detecting *BRCA* reversion mutations in circulating tumor DNA to anticipate the loss of PARPi efficacy.<sup>41</sup>

Rad3-related (*ATR*), a key protein DNA for damage response, senses replication stress and induces signaling into the S and G2/M

checkpoints to facilitate DNA break repair. Its activation is involved in resistance to PARPi. *ATR* inhibitors, such as VE-821, are a promising option to overcome these mechanisms of resistance to talazoparib and olaparib, mediated by inactivation of *SLFN11*<sup>42</sup> or to resensitize PARPi-resistant *BRCA1*-deficient cells which have restored HRR function to olaparib.<sup>43,44</sup> Finally, epigenetic resensitization through DNA methyltransferase inhibition, cell cycle checkpoint inhibition, drug combinations with antiangiogenic agents, BET inhibition or G quadruplex stabilization or combining drugs upfront to prevent resistance, should also be explored.<sup>40,42</sup>

**Efficacy of PARPi in non-core genes mutations?** There is very little information in the literature on the sensitivity to PARPi in tumors harboring a DNA damage repair mutation other than *BRCA1* or *BRCA2* (e.g. *ATM*, *BAP1*, *BARD1*, etc.) or on the activity of the different PARPi on somatic *versus* germline ‘BRCAness’ mutations.<sup>45,46</sup> Somatic ‘BRCAness’ mutations could be present in up to 15–17% of PDAC patients.<sup>47</sup> The activity of PARPi has been reported to be heterogeneous in other cancers depending on the presence of a germline or somatic mutation, and the gene involved (e.g. in prostate or ovarian cancer).<sup>48,49</sup> However, there is

not enough existing data to extend the use of olaparib to germline mutations other than *BRCA1* or *BRCA2* or to somatic ‘BRCAness’ mutations, and prospective trials are needed. The MAZEPPA French trial (NCT04348045) is an open-label, phase II study to assess the efficacy of genomic-driven maintenance therapy for PFS in metastatic PDAC patients whose tumor has been controlled after 4 months of mFOLFIRINOX chemotherapy. In this trial, patients with a BRCAness somatic profile receive olaparib while those with no BRCAness profile and with a *KRAS* mutation are randomized to receive durvalumab plus selumetinib or FOLFIRI.

*Which patients with PDAC should be tested for BRCAness mutations?* Ideally, all patients with PDAC should be tested<sup>50</sup> to identify whether a gBRCAm has immediate and individual therapeutic value. Moreover, diagnosis of a germline mutation can directly influence the patient’s relatives and requires an oncogenetic consultation with a longer delay. However, it should be remembered that the primary goal of gBRCAm testing is therapeutic because it was not broadly indicated before the POLO trial results, except in patients with a family history suggesting a potential genetic predisposition.

It has been shown that about 40–60% of patients with gBRCAm do not declare a family history of *BRCA*-related cancers (PDAC, breast, ovarian or prostate cancer)<sup>51</sup> either because they are not asked this question by the doctor, or the patient does not know or has forgotten the family history, or the family history is non-informative (incomplete penetrance).<sup>52</sup> Published data on referral rates in PDAC are limited. A US study reported that one-third of patients referred for genetic counseling did not complete germline testing for various reasons (worsening of disease severity, lack of follow-up, etc.).<sup>53</sup> An Italian real-life study also showed that germinal *BRCA 1/2* pathogenic variants were found in 8.1% of the population, and 29% of patients had a family history of potentially *BRCA*-associated tumors (breast ovarian, prostate, or pancreas). However, in the same study, 45% of patients with a gBRCA mutation had no or unknown family history of BRCAness.<sup>54</sup>

Thus, a family history of cancers from the *BRCA*-spectrum should not be considered necessary to select patients for gBRCAm screening. It would seem that the gastroenterology/oncology medical community needs to be more aware of the

importance of collecting family information and patient screening at the diagnosis of PDAC. However, it may not be cost-effective to screen all PDAC patients. For example, the gBRCAm might only be searched for in patients who are potential olaparib candidates, that is, who are fit for platinum-based chemotherapy.

*When should patients be tested for BRCAness mutations?* In US guidelines, early testing for *BRCA* mutations (excluding variants of unknown significance) in all PDAC patients is now recommended with both somatic and germline analyses.<sup>50</sup> Thus, testing can be proposed at diagnosis when planning for biologically adapted treatment (i.e. platinum-based chemotherapy and PARPi maintenance) whenever possible. In practice however, proposing this test during the first meeting with patients who are worried about this life-threatening disease can be difficult. Presenting testing as a potential opportunity to have access to additional therapeutic options can minimize a patient’s feeling of guilt for her/his relatives about transmitting the mutation to her/his children if a germline mutation is discovered. Another problem is the potential failure of upfront induction chemotherapy (17% of patients in the POLO study) which prevents the use of maintenance treatment such as PARPi.<sup>4</sup> Thus, it seems to be acceptable to wait until the first tumor evaluation to select patients for mBRCAg testing.

This explains why somatic testing was performed in the MAZEPPA trial after 2 months (four cycles) of mFOLFIRINOX to ensure tumor control. Finally, testing can encounter organizational difficulties to obtain the result within a sufficiently short period of time in relation to the therapeutic aim, or reimbursement depending on the country and center. Moreover, there is also the risk of poor quality material and thus to non-informative results with somatic testing.

#### *Molecular alterations in KRAS wild-type PDAC*

A hallmark of PDAC is the prevalence of the oncogenic mutation in the *KRAS* gene.<sup>55</sup> Activating *KRAS* mutations occur in approximately 90% of cases. They are an early and major event in pancreatic carcinogenesis. They result in permanent activation of the *KRAS* small GTPase protein, which acts as a molecular switch to activate various intracellular signaling pathways (including the MAP kinases and mTOR pathway) and transcription factors inducing

cell proliferation, invasion, and survival.<sup>56–58</sup> *KRAS*-driven tumors are largely refractory to anticancer therapies.

*KRAS* wild-type PDAC in 10% of PDACs,<sup>59</sup> an entity with a good prognosis in the literature,<sup>60</sup> offers therapeutic opportunities due to an enrichment in alternative, potentially actionable, oncogenic drivers which are (almost always) mutually exclusive with the *KRAS* mutation: (i) fusion genes and (ii) *BRAF* alterations.<sup>58</sup>

In a study by Philip *et al.*<sup>59</sup> based on whole exome sequencing of 2483 PDACs, MSI-H/dMMR tumors were found in 4.7% of the cases *versus* 0.7%, tumor mutational burden (TMB)-high in 4.5% *versus* 1% and a larger infiltration of CD8+ T cells, natural killers, and dendritic cells (DCs) which could make these *KRAS* wild-type tumors more sensitive to immune checkpoint inhibitors (CPIs).

Windon *et al.*<sup>61</sup> reported that patients with *KRAS* wild-type PDACs have a longer median OS (*KRAS* wild-type, 957 days *versus* *KRAS* mutant, 531 days;  $p=0.026$ ) than those without. Similarly, Ogura *et al.*<sup>62</sup> found significantly better OS in patients with *KRAS* wild-type PDAC (479 *versus* 255 days,  $p=0.03$ ).

In addition, *KRAS* wild-type PDAC appears to be sensitive to epidermal growth factor receptor (EGFR) inhibition.<sup>63</sup> Accordingly, the NOTABLE phase III trial, evaluating gemcitabine plus nimotuzumab (a humanized anti-EGFR monoclonal antibody) combination *versus* gemcitabine plus placebo have shown good results in the experimental arm with a significant improvement of OS [10.9 *versus* 8.5 months, HR: 0.50 (0.06–0.94),  $p=0.024$ ] in *KRAS* wild-type advanced PDACs.<sup>64</sup>

Previous studies examining the genomic profiles of *KRAS* wild-type PDAC have shown considerable heterogeneity, with *GNAS*, *BRAF*, and *CTNNB1* alterations and additional RAS pathway genes identified as oncogenic drivers and kinase fusions in *FGFR2*, *RAF*, *ALK*, *RET*, *MET*, neurotrophic tyrosine receptor kinase 1 (*NTRK1*), *ERBB4*, and *FGFR3*.<sup>47,59,65</sup> In particular, actionable somatic alterations were frequently identified in early onset pancreatic cancer, enriched in the *RAS* wild-type subgroup. For example, in a study performed at Sloan Kettering Memorial hospital in 132 early-onset PDAC, 15.9% had *RAS* wild-type PDAC with actionable alterations, including

*ETV6-NTRK3*, *TPR-NTRK1*, *SCLA5-Neuregulin 1 (NRG1)*, as well as *ATP1B1-NRG1* fusions, *IDH1 R132C* mutations, or mismatch repair deficiencies.<sup>66</sup> Another whole genome and transcriptome sequencing study that was performed to identify clinically actionable genomic alterations in young adults with PDAC alterations identified recurrent *NRG1* rearrangements that are supposed to drive PDAC development through aberrant ERBB receptor-mediated signaling.<sup>67</sup>

*NRG1* fusions. *NRG1* is a ligand that binds to human epidermal growth factor receptor 3 (*HER3*), promoting *HER2/HER3* heterodimerization and activation of PI3K/AKT/mTOR signaling. Chromosomal rearrangements involving *NRG1* are rare oncogenic drivers in solid tumors, enriched in *KRAS*<sup>wt</sup> PDAC and lung invasive mucinous adenocarcinoma.<sup>68,69</sup> *NRG1* fusion positive (*NRG1+*) *in vitro* and *in vivo* models are sensitive to *HER2/HER3* directed therapy.<sup>63,70–73</sup> There are over 20 partners of *NRG1* fusions in solid tumors and some reports reveal common partners according to cancer type. *CD74* has been shown to be the most frequent and *ATP1B1*, *CDH1*, and *VTCN1* were detected as a partner of *NRG1* fusion in PDAC.<sup>68,69</sup>

In a study by Jones *et al.*<sup>74</sup> in 47 patients with PDAC who underwent comprehensive whole genome and transcriptome sequencing and analysis, two of the three patients with *NRG1* fusion-positive tumors were treated with afatinib (tyrosine kinase inhibitor targeting *HER2* and *HER3*) and had a rapid, significant response on therapy.

A recent phase II basket trial with zenocutuzumab (*MCLA-128*), a bispecific antibody targeting *NRG1* fusion signaling, was launched in patients with *NRG1* fusion-positive PDAC, non-small-cell lung cancer (NSCLC), and other solid tumors (NCT02912949). Schram *et al.*<sup>75</sup> reported a partial response in 42% of patients with *MCLA-128* in a series of PDAC patients in this ongoing trial. Zenocutuzumab induced rapid and major radiological tumor regression and biomarker responses in heavily pretreated metastatic *KRAS* wild-type, *NRG1*-positive pancreatic cancer, with a good tolerance.

The rarity of *NRG1* fusions and the diversity of fusion partners may make their detection challenging. It is important to note that DNA sequencing is not sufficiently sensitive to detect these fusions. Thus, RNA sequencing is considered to be the gold standard for this purpose, and offers the greatest



likelihood of identifying *NRG1* gene fusions when the identity of the fusion partner is unknown.<sup>76–78</sup>

***NTRK* fusion.** The genes *NTRK1*, *NTRK2*, and *NTRK3*, respectively, encode tyrosine kinase receptors tropomyosin-related kinases A, B, and C (TRKA, TRKB, and TRKC). Fusion of a variety of different partners with *NTRK1*, *NTRK2*, or *NTRK3* results in oncogenic proteins that act for constitutive activation of the *RAS*-*MEK*-extracellular signal-regulated kinase (*ERK*), diacylglycerol-inositol-1,4,5-trisphosphate, and *PI3K*-*AKT* signaling pathways.<sup>69,79</sup>

However, these *NTRK* fusions are uncommon (<0.5% of PDAC), and mainly found in rare pediatric tumors, although they can also be encountered in common digestive cancers such as colorectal cancer, especially in case of *RAS*/*BRAF* wild-type dMMR/MSI-H tumors (with an enrichment in up to 40% of tumors), or in aggressive cancers, such as PDAC.<sup>12,80,81</sup> In a study by Allen *et al.*<sup>82</sup>, whole genome sequencing and RNAseq were performed in a series of 400 patients with resected or locally advanced/metastatic PDAC. The prevalence of *NTRK* fusions was 0.8% (3/400), but it was 6.25% (2/32) in *KRAS* wild-type tumors. Since the results of the NAVIGATE trial, these fusions represent a histagnostic alteration that can be targeted. In this phase II clinical trial that explores the efficacy and long-term safety of larotrectinib, 121 (79%) patients with *NTRK* fusion-positive solid tumors had an objective tumor response. One of the two patients with PDAC were responders.<sup>83,84</sup> In another study, two patients with PDAC and *NTRK* fusions achieved a partial response to entrectinib.<sup>7</sup>

Other targetable fusions have also been described but are uncommon (each <0.5%: RET, enriched in acinar pancreatic cancer, ALK, FGFR2, etc.).

#### *KRAS* mutations

Direct *KRAS* blockade remains a challenge, and successful inhibition of a key downstream effector pathway, the *RAF*-*MEK*-*ERK* cascade has been limited because of activation of feedback networks that keep the pathway in check (Table 2).<sup>85,86</sup> Overall, the unsuccessful clinical trials to prevent localization of the *RAS* protein to the plasma membrane and inhibit downstream oncogenic signaling by targeting *KRAS* effectors in *MEK1/2*, *ERK1/2*, or *AKT* alone or in combination have led

to the conclusion that *KRAS* was an ‘undruggable’ target.<sup>73</sup>

*G12D* (38%) and *G12V* (28%) are the most common mutations in PDAC.<sup>87–89</sup> In recent years, inhibitors have been developed to target the *KRASG12C* mutation (adagrasib, sotorasib), based on the physicochemical particularities of the cysteine amino acid (G12C). The mutated cysteine resides next to a pocket (P2) of the switch II region. The P2 pocket is only present in the inactive GDP-bound conformation of *KRAS* and has been exploited to establish covalent inhibitors of *KRASG12C*.<sup>14,90,91</sup> They bind to *KRAS*-GDP-bound and lock it in its ‘off’ form.<sup>90,92</sup> The targeting of this mutation was initially developed in lung and colorectal cancers. Early clinical trials of these inhibitors were promising in the *KRASG12C* mutant PDAC, which is only found in 1–2% of PDAC, in early clinical trials.<sup>93–95</sup>

Preclinical studies showed that sotorasib (AMG510) inhibited nearly all detectable phosphorylation of *ERK*, a key downstream effector of *KRAS*, leading to durable complete tumor regression in mice bearing *KRASG12C* tumors. Preclinically, AMG510 selectively targeted *KRASG12C* tumors, caused durable regression as a monotherapy, and could be combined with cytotoxic and targeted agents to synergistically kill tumor cells. Moreover, AMG510 treatment led to an inflamed TME that was highly responsive to immune checkpoint inhibition.<sup>96</sup>

In the phase I study by Hong *et al.*<sup>93</sup> in advanced solid tumors, no dose-limiting toxic effects were observed with sotorasib and no treatment-related adverse events resulted in death. Four patients with tumor types other than lung or colorectal cancers had a confirmed partial response, including one with PDAC.<sup>93</sup> In all, 12 patients with *KRASG12C* PDAC were enrolled in the next phase I/II study with sotorasib (NCT03600883). One out of 11 patients who could be evaluated had a partial response, 8 patients had stable disease, and 2 patients had progressive disease.<sup>93</sup> Recently, the combined data from the phase I/II CodeBreaK100 trial in 38 PDAC patients treated with sotorasib, the objective response rate (ORR) was 21% and disease control rate (DCR) 84.2% among patients who had already received at least one therapy for PDAC.<sup>94,97</sup>

Adagrasib (MRTX849) is another *KRAS G12C* inhibitor. At the ASCO GI 2022, Bekaii-Saab

**Table 2.** Trials with drugs targeting KRAS mutations.

KRAS	Cyclophosphamide-fludarabine then mutant KRAS G12V-specific TCR transduced autologous T cells ± anti-PD1	I/II	NCT04146298
ERK	Ulixertinib (BVD-523) is an oral, first-in-class ERK1/2 inhibitor	I	NCT04566393
KRAS	HBI-2376 = SHP2 inhibitor in advanced malignant solid tumors harboring KRAS or EGFR mutations	I	NCT05163028
KRAS	NBF-006: lyophilized lipid nanoparticle with encapsulated siRNA	I	NCT03819387
KRAS	Mesenchymal stromal cells-derived exosomes with KrasG12D siRNA	I	NCT03608631
KRAS	ELI-002 2P (lipid-conjugated immune-stimulatory oligonucleotide [Amph-CpG-7909] plus a mixture)	I	NCT04853017
KRAS-ERK	JAB-3312: inhibitor of Src homology 2 domain-containing phosphatase (SHP2)	I	NCT04121286
KRASG12C	LY3537982 combined with several agent in KRAS G12C-mutant tumors	I	NCT04956640
KRASG12C	Adagrasib in combination with BI 1701963 (SOS1 inhibitor) (KRYSTAL 14)	I	NCT04975256
KRASG12C	GDC-6036 combined with atezolizumab, cetuximab, bevacizumab, erlotinib	Ia/Ib	NCT04449874
KRASG12C	JDQ443 monotherapy and combined with TNO155 and spartalizumab	Ib/II	NCT04699188
KRASG12D	PBL transduced with a murine TCR against G12D variant of mutated RAS in HLA-A*11:01 patients	I/II	NCT03745326
KRASG12V	PBL transduced with a murine TCR against G12V variant of mutated RAS in HLA-A*11:01 patients	I/II	NCT03190941

ERK, extracellular signal-regulated kinase; HLA, human leukocyte antigen; TCRs, T-cell receptors.

*et al.*<sup>95</sup> presented the updated results of the KRYSTAL-1 study (NCT03785249) about the activity and safety of adagrasib as monotherapy in 30 patients with previously treated, inoperable or metastatic *KRASG12C*-mutant GI tumors other than colorectal cancer. Among the 12 patients with PDAC in this series (median prior lines of therapy:  $n=3$ ; median follow-up 8.1 months), a partial response was obtained in 5 out of 10 patients who could be evaluated. All patients achieved a DCR and the PFS was 6.6 months (95% CI 1.0–9.7). Adverse events were diarrhea (43%), vomiting (43%), and fatigue (29%) (grade 3/4: 21%).

More effective combinations of various targeted therapies have been studied preclinically with *KRAS G12C* inhibitors in colorectal cancer (e.g.

combination of adagrasib with anti-EGFR cetuximab in the KRYSTAL-1 study) and NSCLC.<sup>15,98</sup> Multiple combinations are now in clinical trials (Table 2), such as new combination arms in the CodeBreak 101 trial combining sotorasib with a *MEK* inhibitor, *PD-1* inhibitor, *SHP2* inhibitor, pan-*ERBB* inhibitor, *PD-L1* inhibitor, *CDK* inhibitor, or *mTOR* inhibitor (NCT04185883). The KRYSTAL-2 trial of adagrasib explores the role of the *SHP2* inhibitor TNO155 (NCT04330664). Multiple additional promising *G12C* inhibitors are currently in clinical development as well, including JNJ-74699157 (NCT04006301), GDC-6036 (NCT04449874), and JDQ443 (NCT04699188)<sup>99</sup> in PDAC patients. Whether the depth and duration of response will be comparable to that obtained in

lung cancer (up to 40% response rate) remains to be determined.

*KRASG12D* is a much more common alteration in PDAC and it represents a third of *KRAS* mutations. As shown in recent attempts, it lacks a reactive residue adjacent to the switch II pocket<sup>16,100</sup> and thus new selected drugs must be designed. The *G12D* inhibitor MRTX1133 was discovered through an intensive structure-based search during adagrasib synthesis and high-resolution X-ray crystal structures. Its efficacy has been found to be promising in mutant Panc 03-04 xenograft mouse tumor models,<sup>16</sup> which is important because the *KRAS* missense mutation *G12D* is the most predominant variant in human malignancies (35%), and 39% of PDAC patients harbor this mutation (Table 2).

Other inhibitors, such as SOS1:KRAS interaction inhibitors, could be effective on a larger spectrum of *KRAS* mutations. BI-3406, a potent and selective SOS1:KRAS interaction inhibitor, has been shown to decrease the formation of GTP-loaded RAS and reduce cell proliferation of a large fraction of *KRASG12C* and non-*G12C*-driven cancers *in vitro* and *in vivo*. In mouse models, BI-3406 attenuates feedback reactivation by MEK inhibitors and enhances sensitivity of *KRAS*-dependent cancers to MEK inhibition, resulting in tumor regression at well-tolerated doses.<sup>17</sup>

Mutant *KRAS* drives immune evasion in cancer cells by the upregulation of inhibitory immune checkpoint expression, the downregulation of MHC-I expression on tumor cells, the enhancement of cytokines and chemokines, and the recruitment of suppressive immune cells.<sup>19</sup> Mutant *KRAS* neoantigens presented on human HLA and T cells can recognize mutant *KRAS* cells and paved the way of strategies for targeting *KRAS* immunologically with vaccines alone or in combination.<sup>101</sup> A highly promising approach using T-cell engineering targeting the *KRAS G12D* hotspot mutation will be described further on in the chimeric antigen receptor (CAR)-T cells in section “CAR-T-cell development.”<sup>102</sup>

#### Targetable BRAF mutations

*BRAF* mutations, which are generally mutually exclusive with *KRAS*, are present in 3% of PDAC but up to 10% of *KRAS wild-type* PDAC.<sup>103</sup> *V600E BRAF* mutations are prominent. Other alterations, such as non-*V600E BRAF* in-frame

insertions or deletions or gene fusions, have been reported in 1% of patients with PDAC.<sup>10</sup>

A combination of the *BRAFV600E* inhibitor vemurafenib and the *MEK* inhibitor cobimetinib resulted in an objective tumor response in a patient with poorly differentiated, *V600E* mutant PDAC.<sup>104</sup> In another report, six patients, five with a *V600E mutation* and one with *CUX1-BRAF* fusion, received vemurafenib single agent.<sup>105</sup> In a series of the ‘Know Your Tumor’ program, two of 18 PDAC patients with *BRAF-mutant* PDAC received the dabrafenib/trametinib (*BRAF/MEK*-inhibitor) combination: one patient with tumor *V600E* mutation had a sustained response, while treatment failed in the other with *KRASG12A/BRAF K601N*.<sup>106</sup>

#### Immunotherapy in PDAC: microsatellite instability and high tumor mutational burden

The resistance of PDAC to immunotherapies could be due in part to a paucity of neoantigen-reactive tumor-infiltrating lymphocytes, which can mainly be explained by the low immunogenicity and non-inflamed phenotype of PDAC. The abundant stroma generates a hypoxic microenvironment and drives the recruitment of immunosuppressive cells through cancer-associated-fibroblast activation and transforming growth factor  $\beta$  (TGF $\beta$ ) secretion.<sup>107</sup> Nevertheless, we have begun to identify subgroups of responders to immunotherapies.

*dMMR/MSI-H tumors.* The deficient deficient mismatch repair (dMMR)/ microsatellite instability-high (MSI-H) phenotype could be an interesting target even in the presence of sporadic or Lynch syndrome (LS) tumors.<sup>108,109</sup> In a multicenter study of immunohistochemical analyses of 445 PDAC specimens, 1.6% of tumor samples were dMMR/MSI-H.<sup>110</sup> This rate was 2.61% in a meta-analysis of 34 studies pooling 8323 PDAC patients.<sup>60</sup> However, the dMMR/MSI-H PDAC rate in the latter study may have been overestimated, since some studies focused on PDAC subtypes enriched by this molecular alteration. dMMR/MSI-H in PDAC is strongly associated with the medullary and mucinous/colloid histological phenotype. This is associated with a *KRAS/TP53* wild-type molecular background and a higher rate of *JAK* (*JAK1* and *JAK3*) and *KMT2* (*KMT2C* and *KMT2D*) gene mutations, with no benefit to survival, like in colorectal cancer. The genomes of dMMR/MSI-H tumors harbor hundreds to thousands of somatic mutations that

encode potential neoantigens. Thus, often immunogenic, triggering upregulation of immune checkpoint proteins. In the phase II KEYNOTE-158 study testing the anti-programmed death-1 therapy pembrolizumab in pretreated patients with advanced non-colorectal dMMR/MSI-H tumors, the primary endpoint was the ORR according to Response Evaluation Criteria in Solid Tumors version 1.1, as assessed by an independent central radiologic review.<sup>111</sup> While there was a clear clinical benefit using this single immunotherapy agent, the ORR in the subgroup of patients with PDAC was only 18.2%, which is low compared to endometrial (57.1%), gastric (45.8%), cholangiocarcinoma (40.9%), small intestine (42.1%), or ovarian (33.3%) cancers.<sup>111</sup> This could be related to the specific unfavorable immune microenvironment or the misdiagnosis of PDAC or MSI-H.<sup>11</sup>

dMMR/MSI-H PDACs has been shown to have a higher density of CD8+ T cells at the invasive front ( $p=0.08$ ) and expression of the CD274 molecule more often than other PDACs ( $p=0.05$ ).<sup>110</sup> Nearly 10% of PDAC patients harbor germline variants, although most lack somatic second hits and its therapeutic significance has not been defined.<sup>112</sup> In a multicenter study of 289 patients with resected PDACs who were not pre-selected for age or personal/family cancer history, targeted germline analysis of 24 genes related to an inherited cancer predisposition identified pathogenic/probably pathogenic germline variants in nearly 10% of patients, including 7.3% with variants in genes related to dMMR genes. Patients with a LS have an 8.6-fold greater risk of developing PDAC than the general population<sup>113</sup> with a cumulated risk of PDAC of 3.7% and tumors exhibiting a medullary appearance with prominent lymphocytic infiltration.<sup>114</sup>

The literature on somatic-only dMMR status is more scarce. hMLH1 promoter methylation was detected in 60.0% of a series of 90 operated patients.<sup>115</sup> Finally, dMMR/MSI-H is often a genetic alteration enriched in *KRAS* wild-type PDAC.<sup>60</sup> These observations are the reason for the recent guidelines for dMMR somatic testing in PDAC and universal germline testing of dMMR tumors.<sup>35,116</sup>

*High tumor mutational burden.* Except for MSI tumors, immunotherapy has thus far failed in PDAC, even with ICI combinations<sup>117</sup> or chemotherapy.<sup>118</sup> TMB in PDAC is about one mutation per megabase (mut/Mb), which is 10-fold less

than that of melanoma or lung carcinoma. In addition, PDACs are characterized by low T-cell recruitment and dense desmoplastic stroma resulting in high interstitial pressure, poor tumor perfusion, and hypoxia. Activation of fibroblasts then releases cytokines [TGF $\beta$ , interleukin (IL)-6, colony-stimulating factor 1 (CSF1) etc.] leading to immunosuppressive cell recruitment (M2 macrophages, regulatory T cells (TREG), myeloid derived suppressor cells (MDSC), as well as exclusion and effector T-cell anergy, which may explain the failure of immune CPI treatments.<sup>107</sup>

Response rates with pembrolizumab have been shown to be significant on tissue TMB (tTMB) data. The rate in the KEYNOTE-158 study was approximately 30% for tTMB-high tumors (at least 10 mutations per megabase) (Table 1).<sup>20</sup> This response rate could mainly be due to the potential of tumor mutations to generate immunogenic neoantigens. There was no benefit to OS in the recent PA.7 randomized phase II trial evaluating gemcitabine-nab-paclitaxel with durvalumab and tremelimumab. However, tTMB and ATM were identified as predictive biomarkers of response.<sup>118</sup> Nevertheless, results are contradictory. In another study, data from over 10,000 tumors from patients included in The Cancer Genome Atlas were analyzed to evaluate the association of t-TMB with CPI treatment outcomes. This analysis did not support the use of TMB-H as a biomarker for treatment outcomes of several cancer types with CPI.<sup>119</sup> Further studies are needed to evaluate other tumor-specific types especially in PDAC for which data are lacking.<sup>20,119</sup>

*Immunotherapy proficient MMR tumors.* Except for dMMR/MSI-H PDAC, single-agent immunotherapy has not been found to be effective in PDAC because this entity combines poor tumor immunogenicity with a low mutation rate (Table 3).<sup>107</sup> Nevertheless, immunotherapy agents are being tested in numerous combinations with various molecules including tyrosine kinase inhibitors or classic gemcitabine-nab-paclitaxel (NCT 04498689, NCT04181645, and NCT03989310) or FOLFIRINOX schemas (NCT04324307, NCT03977272, and NCT04181645) (Table 3). It is interesting to note that T-cell reactivity to high-quality tumor neoantigens has been shown to be a characteristic in long-term survivors thus could lead to reposition immunotherapy in a subset of patients with MMR PDAC. To illustrate this, a promising approach using T-cell engineering will be described further on in this review (cf section "CAR-T-cell development").

**Table 3.** Combination of immunotherapy with other drugs in PDAC patients: trials in progress.

Combined agent	Immunotherapy	Type	Phase	NCT
Plerixafor	Cemiplimab	Anti-PD-1	II	NCT04177810
G-nab-P	Camrelizumab	Anti-PD-1	II	NCT04498689
G-nab-P or FOLFIRINOX	BsAb	PD-L1/CTLA4	I/II	NCT04324307
Single agent, 2d line	Retifanlimab	Anti-PD-1	II	NCT04116073
FOLFIRINOX	Not precised	Anti-PD-1	III	NCT03977272
Manganese, G-nab-P	Not precised	Anti-PD-1	I/II	NCT03989310
mFOLFIRINOX	Not precised	Anti-PD-1	III	NCT03977272
G-nab-P	SHR-120	Anti-PD-1	I	NCT04181645
SX-682 (CXCR1/2)	Nivolumab	Anti-PD-1	I	NCT04477343
CXCR4 antagonist,G-nab-P	Cemiplimab	Anti-PD-1	I	NCT04543071
Tadalafil (PDESi), vancomycin	Nivolumab	Anti-PD-1	II	NCT03785210
Autologous TIL	Pembrolizumab	Anti-PD-1	II	NCT01174121
ABBV-927 (anti-CD40), FOLFIRINOX	Budigalimab	Anti-PD-1	Ib/II	NCT04807972
LYT-200 (targets galectin-9)	Not precised	Anti-PD-1	I/II	NCT04666688
Epacadostat (IDO1i, CRS-207 (listeria monocytogenes), CY/GVAX)	Pembrolizumab	Anti-PD-1	II	NCT03006302
Siltuximab (anti-IL-6)	Spartalizumab	Anti-PD-1	I/II	NCT04191421
Aldoxorubicin, cyclophosphamide	Not precised	PD-L1 t-haNK*	II	NCT04390399
CDX-1140 (CD40Ab), CDX-301 (FLT3L)	Pembrolizumab	Anti-PD-1	I	NCT03329950
Olaparib	Pembrolizumab	Anti-PD-1	II	NCT04548752
Niraparib	TSR-042	Anti-PD-1	Ib	NCT04673448
Anlotinib (TKI)	Toripalimab	Anti-PD-1	II	NCT04718701
Cabozantinib (TKI)	Pembrolizumab	Anti-PD-1	II	NCT04820179
Anlotinib (TKI), 2d line	Pembrolizumab	Anti-PD-1	II	NCT05218629
Cabozantinib (TKI)	Pembrolizumab	Anti-PD-1	II	NCT05052723
Vaccine, imiquimod, sotigalimab (APX005M) (CD40 agonist)	Pembrolizumab	Anti-PD-1	I	NCT02600949

IL, interleukin; PDAC, pancreatic ductal adenocarcinoma.

### Emerging targets of interest

#### Metabolic pathways

The harsh PDAC microenvironment explains why tumor cells must find alternative nutrient sources.<sup>120</sup> Tumor cells become dependent upon

the activity of an oncogene for survival and proliferation *via* the mitochondrial metabolism and ATP production. This is mainly fueled by glutamine.<sup>121</sup> and *KRAS* signaling, which promotes extracellular glucose avidity and capture *via* upregulation of the glucose transporter GLUT1

and hexokinase.<sup>21</sup> This oncogene addiction may be the ‘Achilles’ heel’ of tumor cells and represent a potential therapeutic target.<sup>22</sup>

Devimistat selectively inhibits the tricarboxylic acid cycle in the mitochondria of tumor cells by impairing pyruvate dehydrogenase and  $\alpha$ -ketoglutarate dehydrogenase enzyme activity. Production of the anabolic intermediates required for effective DNA damage repair is decreased as a result of the impact on mitochondrial metabolism. In a phase I study, the combination of devimistat and modified FOLFIRINOX (mFOLFIRINOX) in 20 patients led to a high tumor response rate (61%).<sup>122</sup> However, the phase III study did not show any improved OS in the mFOLFIRINOX plus devimistat arm compared to FOLFIRINOX alone (HR = 0.95, median OS: 11.1 *versus* 11.7 months).<sup>123</sup>

Constitutive KRAS signaling leads to addiction to metabolites such as glutamine and asparagine, used by non-canonical metabolic pathways.<sup>121,124</sup> Glutamine deprivation and/or inhibition of enzymes downstream of KRAS result in suppression of PDAC cell growth. As PDAC are hypovascular tumors, enhanced asparagine synthetase expression is a key mechanism of adaptation to hypoxia and glucose deprivation.<sup>125</sup> Thus, modulation of glutamine and asparagine levels may be critical in these cells. L-Asparaginase hydrolyzes the amino acid asparagine into aspartic acid and ammonia. Its systemic administration decreases the amino acid in proliferating cancer cells that use exogenous asparagine.<sup>126</sup> The results of erythrocyte-encapsulated L-ASNase (eryaspase) tested in a phase IIb study combined with chemotherapy as a second line of treatment in patients with PDAC were positive.<sup>127</sup> However, it failed to improve survival in the phase III study.<sup>128</sup>

These findings add to the negative results from other trials targeting metabolism in PDAC with metformin,<sup>129</sup> evofosfamide,<sup>130</sup> or hydroxychloroquine (HCQ).<sup>131</sup> Nevertheless, other metabolic strategies are still being explored. Autophagy involves the degradation and recycling of damaged organelles and protein aggregates after autophagosome and lysosome fusion, which promote damaged cancer cell survival.<sup>132</sup> Autophagy is elevated in PDAC cells.<sup>133</sup> It promotes immune evasion by degrading the major histocompatibility complex class I (MHC-I) and decreasing tumor infiltration by CD8+ T cells.<sup>23</sup> MHC-I molecules are located in autophagosomes and lysosomes

rather than on the cell surface and are targeted for degradation by an autophagy-dependent mechanism. Thus, inhibiting autophagy with drugs such as HCQ could sensitize PDAC cells to immune checkpoint blockade.<sup>24</sup> The *ERK* inhibitor LY3214996, which has shown some activity in BRAF- and KRAS-mutant models, administered as a single-agent or combined with HCQ (NCT04386057), and the paricalcitol, HCQ and gemcitabine-nab-paclitaxel combination (NCT 04524702) have also been tested in two phase II trials in advanced PDAC patients.

The alpha-hydroxylated polyunsaturated fatty acid ABTL0812 induces sustained endoplasmic reticulum stress *via* induction of tribbles-3 pseudokinase expression, which results in inhibition of *AKT-MTORC1* and dihydroceramide accumulation in human PDAC cell lines leading to autophagic activation and cell death.<sup>134,135</sup> The combination of ABTL0812 with FOLFIRINOX is being tested for the first-line treatment of metastatic PDAC in a phase I-II trial (NCT04431258).

#### Stroma

*Recombinant human hyaluronidase.* PDACs have a dense and abundant stroma, which is known to be a critical mediator of disease progression through direct effects on cancer cells and indirect effects on the immune microenvironment of the tumor. The three major entities in the PDAC stroma are the extracellular matrix (ECM), vasculature (poorly developed in PDAC which are highly hypoxic), immune cells (mainly, protumoral, inflammatory M2 and MDSC and low number of cytotoxic T cells), and cancer-associated fibroblasts (CAFs).<sup>136,137</sup>

PDAC is a highly chemoresistant cancer which may be because desmoplasia impedes drug delivery. This suggests that stromal modifying agents, such as pegvorhyaluronidase alfa (PEGPH20), an enzyme that temporarily degrades hyaluronic acid (HA), could decrease tumor pressure and vascular compression, thus penetrating the ‘halo’ surrounding the tumor cells. PEGPH20 has been developed and investigated in phase I-III studies.<sup>138</sup> This drug was assessed in a phase Ib/II clinical trial in combination with modified FOLFIRINOX (mFOLFIRINOX). A unexplained detrimental effect was reported in the PEGPH20 arm with a median OS that was nearly 50% lower (7.7 months *versus* 14.4 months in the chemotherapy alone group) and an increase in the rate of severe

treatment-related adverse events (45% versus 9% in the mFOLFIRINOX alone arm).<sup>139</sup> Otherwise, in a phase III trial, the addition of PEGPH20 to gemcitabine plus nab-paclitaxel backbone chemotherapy did not improve survival.<sup>140</sup>

Other ECM targets are currently under clinical investigation.<sup>136</sup> For example, the normalization of tumor vasculature by blocking the vascular endothelial growth factor (*VEGF*)–*VEGF receptor 2* axis can improve pericyte coverage and tumor perfusion, leading to decreased hypoxia, increased drug delivery and CD8+ T-cell trafficking into the tumor through the action of the leukocyte adhesion molecules, intercellular adhesion molecule and vascular adhesion molecule.<sup>141</sup>

Moreover, the PDAC TME has distinct CAFs populations.<sup>142</sup> There is an overwhelming body of evidence showing that CAFs are not mere cellular bystanders in cancer, but active players in the process of cancer initiation, progression, and metastasization.<sup>143</sup> CAFs have long been considered to be pro-tumoral cancer cell partners in PDAC.

These data have led to a paradigm suggesting that stroma and in particular CAFs promote PDAC tumor growth and metastases and contribute to the extensive resistance to therapies. This mechanistic model promoted the clinical development of anti-tumor strategies to deplete the stroma by disrupting the signaling pathways involved in intercellular communication between PDAC tumor cells and CAFs. These include the Sonic Hedgehog (SHH) or TGFβ pathways, or direct targeting of the ECM, such as HA – the main ECM component responsible for the elevated interstitial fluid pressure, or collagen I. Preclinical studies in murine models showed that SHH inhibition reduced stromal abundance, increased tumor perfusion, and thus, improved chemotherapy delivery.<sup>144</sup> However, phase II studies with SHH inhibitors in PDAC were negative.<sup>145</sup> Similarly, although several clinical trials, including the phase III MAESTRO study, have targeted TGFβ in a variety of cancers including PDAC, and which is overexpressed in 35% of PDAC, patient OS was not improved.<sup>146</sup> Melisi *et al.*<sup>25</sup> have presented preliminary results of a phase Ib study (NCT02734160) testing the TGFβ receptor I kinase inhibitor galunisertib plus the anti-PD-L1 antibody durvalumab. DCR was of 25%.

Other ongoing trials with other agents such as bintrafusp (M7824) in a phase I/II trial plus

stereotaxic body radiation therapy (NCT04327986) or LGK974 (NCT01351103) are being tested. As mentioned above, PEGPH20 has also been evaluated with negative results.<sup>140</sup> Preclinical studies further highlighted the deleterious effects of stromal depletion. Non-selective suppression of CAFs from the stroma, either by genetic αSMA-positive cell depletion<sup>147</sup> or by pharmacological inhibition of the SHH pathway,<sup>26</sup> adversely modulated immunity (increase in immunosuppressive regulatory T cells) and angiogenesis (increased tumor vascularization) in PDAC TME and led to undifferentiated, more invasive tumors.<sup>26,27</sup> Moreover, deletion of the *Col1a1* gene in αSMA-positive CAFs led to a significant decrease in total stromal collagen type I deposition and acceleration of disease due to the suppression of CD8+ T cells.<sup>28</sup> Overall, these negative results (i) highlighted the limited predictability of preclinical models and (ii) suggested that some CAF subpopulations or ECM components may play a protective role in blocking the progression of PDAC. Despite certain disappointing results in this poorly vascularized, hypoxic microenvironment, clinical trials targeting angiogenesis and CAFs subtypes are needed.<sup>9</sup>

**FAK.** Nuclear focal adhesion kinase (FAK) regulates the transcription of chemokines that drive recruitment of tumor-associated regulatory T cells (Tregs), which contribute to an immunosuppressive TME in PDAC by inhibiting cytotoxic CD8+ T-cell activity.<sup>148</sup> Thus, FAK inhibition may modulate the cellular and molecular composition of immuno-suppressive TME.<sup>29</sup> On the other hand, stromal depletion by FAK inhibitor therapy may provoke treatment resistance through *STAT3* signaling activation.<sup>149</sup> The results of a phase II trial combining GSK2256098 and trametinib (NCT02428270) in advanced PDAC patients were disappointing.<sup>150</sup> The FAK inhibitor defactinib is currently being tested in combination with the anti-*PD-1*, pembrolizumab and gemcitabine (NCT02546531, NCT02758857, and NCT03727880), or trametinib (NCT02428270).

**BTK.** Bruton tyrosine kinase (BTK) is a non-receptor enzyme from the Tec kinase family expressed in hematopoietic cells (B, myeloid and mast cells, and platelets). BTK-dependent signaling in mast cells and myeloid cells plays an important role in peri-tumoral inflammatory stroma. Ibrutinib specifically inhibits the release of IL-8, MPC-1, and tumor necrosis factor α from mast cells leading to decreased fibrosis and extended

survival, and improves the response to clinical standard-of-care therapy in mouse models.<sup>151</sup> In the phase III RESOLVE trial, the addition of ibrutinib to nab-paclitaxel plus gemcitabine failed to improve OS (9.7 *versus* 10.8 months;  $p = 0.3225$ ) and provided an even shorter PFS (5.3 *versus* 6.0 months;  $p < 0.0001$ ) in the ibrutinib arm than in the placebo arm.<sup>152</sup> The efficacy of the combination of ibrutinib with the anti-PDL-1 durvalumab,<sup>153</sup> and acalabrutinib alone or combined with pembrolizumab<sup>154</sup> was found to be limited in PDAC patients.

**CTGF.** Connective tissue growth factor is strongly expressed in CAFs and tumor cells. Pamrevlumab (FG-3019) enhances the chemotherapy response without increasing drug delivery in murine ductal pancreas cancer by decreasing expression of the X-linked mammalian inhibitor of apoptosis protein gene.<sup>155</sup> In a phase I/II trial, patients with locally advanced PDAC who received a combination of gemcitabine/nab-paclitaxel plus pamrevlumab achieved surgical resection in 33% of cases *versus* 8% in those with gemcitabine-nab-paclitaxel alone.<sup>156</sup> The combination of pamrevlumab plus gemcitabine-nab-paclitaxel or FOLFIRINOX has been evaluated in patients with advanced PDAC in a phase III trial and the results are pending (NCT03941093 and NCT04229004).

#### *Tumoral and stromal signatures*

The choice of systemic therapy using combined, potentially toxic drugs is mainly based on the patient's performance status and biological/liver tests. Thus, diagnostic tests should be used to guide the choice of individualized therapy for patients.<sup>157</sup> Combined genomic, transcriptomic, and therapeutic profiling using patient-derived organoids (pharmacotyping) could help identify molecular and functional subtypes of PDAC for this purpose.<sup>158</sup> ORGANOTREAT-01 (NCT05267912) is a clinical trial establishing organoid models from the patient's biopsy to test the chemosensitivity of available drugs ('chemogram'). Even if pharmacotyping is promising as surrogate tumor models for personalized oncology, this strategy has several limitations for a highly aggressive disease such as PDAC: (1) because the delay required for pharmacotyping is long (usually several weeks), patients may receive treatments that modify tumor sensitivity; 2) in non-experienced centers the quality of the organoid cultures may be suboptimal (<60%) because this approach is technically challenging. On the other hand, the

predictive multigene signatures based on RNA expression measurement can be obtained rapidly from patient biopsies in breast, prostate, and pancreatic cancers.<sup>159–163</sup> In practice prediction of drug sensitivity despite the lack of defined molecular targets seems to be more useful than predicting the risk of recurrence. The predictive value of transcriptomic signatures in PDAC, in particular the basal-like and classic epithelial subtypes, is of great interest.<sup>164</sup> In the COMPASS trial (NCT02750657), Aung *et al.*<sup>165</sup> identified 30% of actionable mutations (*ARID1A*, *BRAF*, *CDK4/6*, *PIK3CA*, *PTEN*, and *RNF*) in their series of patients with PDAC. The efficacy of modified (m) FOLFIRINOX and gemcitabine-nab-paclitaxel combinations was assessed according to the molecular subtype of the tumor on RNA-sequencing. 'Basal-like' and 'classic' modified Moffitt subtypes were identified in 20% and 80% of cases, respectively. The basal-like subtype was associated with a lower objective response rate than the classic phenotype in patients who received mFOLFIRINOX (10% *versus* 33%, respectively,  $p = 0.02$ ) and median OS (5.9 *versus* 9.3 months, respectively, HR = 0.47; 95% CI: 0.32–0.69;  $p = 0.0001$ ).<sup>166</sup> Nevertheless, no alternative to mFOLFIRINOX has been found to be more effective for the basal-like subtype. Another transcriptomic signature is the Pancreatic Adenocarcinoma Molecular Gradient. Unlike all other stratification schemas, this signature classifies PDAC along a continuous gradient with a prognostic value that also predicts the response to mFOLFIRINOX.<sup>162,163</sup> GATA6 RNA expression is lower in basal-like tumors. GATA6 acts as a tumor suppressor by blocking *KRAS* G12V-driven pancreatic tumorigenesis and enforcing differentiation. GATA6-low cell lines derived from patient-derived xenografts were resistant to 5-fluorouracil but not gemcitabine patient-derived organoids.<sup>158,167</sup>

Nicolle *et al.*<sup>162,163</sup> have identified a RNA-based whole transcriptome signature that predicts sensitivity to gemcitabine ('GemPred') in the adjuvant setting after surgical resection of PDAC.<sup>168</sup> Among gemcitabine-treated patients, OS was significantly higher in GemPred-positive patients than in GemPred-negative patients [91.3 months (95% CI: 61.2–not reached) *versus* 33 months (95% CI: 24–35.2); HR 0.403 (95% CI: 0.221–0.735,  $p = 0.00216$ ), respectively]. Genome-wide RNA profiles were also obtained from routine formalin-fixed, paraffin-embedded samples of fine-needle aspirate material. This is valuable for routine use, as pancreatic biopsies are performed



using fine-needle aspiration under endoscopic ultrasound control.<sup>168</sup> Finally, this signature could help reposition gemcitabine in frail PDAC patients who are not eligible for FOLFIRINOX, whatever the indication.

#### High replicative stress

Genomic instability is a key hallmark of cancer, occurring due to defects in the DNA damage response and/or increased replication stress.<sup>169,170</sup> *ATR* and *ATM* are key components to the DNA damage response and are prime targets for DNA damage response inhibitors, because of their central regulatory function in activating the response to both single-strand breaks and DSBs. Both of these proteins work through distinct but overlapping pathways to halt the cell cycle and initiate DNA damage response pathways.<sup>169</sup>

There is an interest in using ataxia-telangiectasia and *ATR* protein kinase inhibitors in cancer, especially in high replicative stress cancers. *ATR* inhibition leads to replication fork collapse as well as the loss of the G2-M checkpoint in cancers with increased levels of replication stress (defined by slowing or stalling of replication forks, e.g. with *TP53* loss or *CCNE1* amplification), allowing cells with damaged DNA to progress prematurely into the M phase, leading to mitotic catastrophe and cell death.<sup>30,171–173</sup> Thus, *ATR* inhibition causes unresolved replication stress that leads to the induction of DNA DSBs causing cell death.<sup>174</sup> *ATR* inhibitors such as berzosertib have already been evaluated in preclinical studies.<sup>175</sup> Most PDAC display high replicative stress. A recent study based on whole exome sequencing datasets reported that the ‘squamous’ transcriptomic subtype of PDACs shows signs of high replicative stress, which may make them sensitive to *ATR* inhibitors. The authors of this study showed that a novel DNA replication stress transcriptomic signature predicts the response to new *ATR* and *WEE1* inhibitors. High replication stress and the presence of DNA repair gene deficiencies appear to be distinct entities, which may exist independently and could be targeted by different drugs.<sup>176</sup>

In particular, this ‘squamous’ subtype is enriched in *TP53* and *KRAS* mutations. Indeed, tumor cells with *TP53* mutations (a major cell cycle checkpoint) are subject to DNA damage, while those with *KRAS* mutations are exposed to replicative stress.<sup>171,177</sup> Thus, replicative stress may be increased in the presence of a double *KRAS* and

*TP53* mutation, which is found in more than 50% of pancreatic cancers.

#### Others

*CAR-T-cell development.* Adoptive T-cell therapy, which redirects a patient’s own T cells against a tumor antigen using CAR gene transfer technology, has been remarkably successful in hematological tumors in the past several years.<sup>178</sup>

The complex TME of PDAC, as well as the stromal obstacle limiting the immune response, and the expression of checkpoint blockade on T cells are obstacles to this approach.<sup>31</sup> CAR-T cells face major difficulties due to the aggressiveness of the disease and the usually late stage diagnosis with extensive tumor dissemination. CAF-depletion, remodeling approaches using CAR-T cells or pharmacological substances such as ATRA or nab-paclitaxel might help limit the influence on the TME.<sup>31</sup>

Some of these obstacles may be overcome or turned into a specific targeting strategy, and clinical trials to overcome the physical and environmental barriers in the TME are ongoing.<sup>179</sup>

Target antigens in these trials include mesothelin, prostate stem cell antigen, carcinoembryonic antigen (CEA), HER2, MUC1, and CD133. The best results from clinical trials in CAR-T cells in PDAC have been obtained from mesothelin<sup>180</sup> and CEA targets.<sup>181</sup>

Mesothelin is expressed in 80% of PDACs,<sup>182,183</sup> and 25%–100% of the cells in positive tumors express the antigen.<sup>184</sup> Positive results in a phase I study in chemotherapy-refractory metastatic PDAC provide evidence for the potential antitumor activity of autologous mesothelin-specific CAR-T cells, as well as PDAC resistance to the immune response.<sup>180</sup> A more recent phase I trial investigated the safety and activity of lentiviral-transduced CAR-T-meso in 15 patients with malignant pleural mesothelioma, ovarian carcinoma, and PDAC. Patients with PDAC showed the greatest percentage of lesions with a disease progression within 28 days.<sup>185</sup> Other results in patient-derived xenograft models from patients with stage IV PDAC suggest that a switchable CAR-T system to target the antigen HER2, upregulated on tumor cells is effective against aggressive, disseminated PDAC.<sup>186</sup> This strategy is ambitious because historically, when a CAR-T HER2/NEU was tested, the patient died from

cardiopulmonary toxicity within days.<sup>187</sup> The CEA is a glycoprotein expressed in nearly 75% of pancreatic cancers<sup>188</sup> and it can also be targeted in PDAC. There are ongoing trials evaluating the role of CAR-T cells activated with the CEA for this indication.<sup>189</sup> While there are no reports of pancreatic cancer-specific CAR T-cell trials targeting CEA, there are preliminary results from a phase I study evaluating the feasibility of delivering first-generation CAR-T-cell therapy to patients with advanced CEACAM5+ tumors. No objective clinical responses were observed and this trial was stopped due to pulmonary toxicity.<sup>190</sup> A recent trial identified CEACAM7 (CGM2), a member of the CEA family of proteins expressed in a large subset of PDAC tumors, as a potential CAR T-cell target for PDAC. CAR-T cells targeting CEACAM7 can target antigen-expressing tumor cells, and mediate remission in patient-derived xenograft tumors.<sup>181</sup> Thus, clinical trials are ongoing. Recently, Leidner *et al.* reported the results of a patient who was treated with her own autologous T cells that were genetically engineered to clonally express two allogeneic HLA-C\*08:02-restricted T-cell receptors (TCRs) targeting the mutant KRAS G12D expressed in her tumor. This resulted in an impressive regression of PDAC metastases. These very promising results suggest that TCR gene therapy targeting the KRAS G12D hotspot mutation merits further study in prospective clinical trials.<sup>102</sup>

*Restoring the immunity by vaccines.* Although cancer vaccines have been evaluated in clinical settings and an antitumor immune response has been shown in PDAC,<sup>191</sup> there is no clear proof that they will be used in clinical practice in the near future.<sup>192</sup>

GVAX, an irradiated allogeneic whole tumor cell vaccine in which PDAC cells are engineered to express granulocyte-macrophage colony stimulating factor (GM-CSF), was evaluated in a small study including 30 previously treated PDAC patients.<sup>193</sup> Disease control with the combination of GVAX and CPIs was promising in that study.<sup>193</sup> In another more recent study, GVAX and ipilimumab maintenance therapy did not improve OS compared to the continuation of chemotherapy and resulted in poorer survival in metastatic PDAC. However, clinical responses and biological effects on immune cells were observed.<sup>194</sup> Therapeutic cancer vaccines also include DC, DNA, and peptide vaccines that provoke the presentation of immunogenic cancer antigens to the immune system, resulting in activation of cancer

antigen-specific CTLs *in vivo* (and a subsequent anti-cancer immune response).<sup>191</sup> In this context, the results of mesothelin, Mucin-1 (MUC1) and telomerase-based vaccine trials were disappointing.<sup>192</sup> For example, tolerance was good and some efficacy was shown in a phase I/II trial of a combination of telomerase peptide GV1001 with GM-CSF for nonresectable PDAC.<sup>195</sup> However, after these promising results, no benefit was found in a phase III trial evaluating a combination of chemotherapy with a telomerase vaccination (GV1001 or TeloVac).<sup>196</sup> In another recent study, a new concept was proposed involving screening of PDAC antigens to develop an mRNA vaccine. A series of targetable antigens including ADAM9, EFN2, MET, TPX2, TMOD3, and WNT7A were found to be promising mRNA vaccine candidates.<sup>197</sup> Therapeutic cancer vaccines targeting *KRAS* were also developed to elicit a *KRAS*-specific immunological response and seem to be promising and safe with some clinical benefit.<sup>198</sup> Although results are not highly encouraging several vaccine trials are still ongoing to evaluate non dMMR/MSI-H PDAC immune responders, either whole cell or peptide-vaccines, according to HLA-A2 status (TEDOPAM-PRODIGE63 trial, NCT03806309).

*Role of microbiota.* Oral and gut microbiota, the intratumoral microbiome, that is, the bacterial and fungal microorganisms present in the tumor, has recently been shown to be a new partner of PDAC TME, modulating pancreatic carcinogenesis, intratumoral immune infiltrates, and the response to chemotherapy.<sup>199</sup> Geller *et al.*<sup>200</sup> showed that intratumoral gammaproteobacteria (e.g. *Klebsiella pneumoniae*, *Escherichia coli* K12, *Mycoplasma Hyorhinis*) expressing long isoforms of CDA, an enzyme involved in the metabolism of gemcitabine, were found in 76% of resected PDAC samples. These bacteria could degrade gemcitabine *in vitro*, and induced therapeutic resistance to this drug in a mouse colorectal cancer model. Resistance to gemcitabine was suppressed when this enzyme was deleted or by co-treatment with ciprofloxacin. Thus, bacteria might be responsible for the poor clinical response to gemcitabine in PDAC.<sup>200</sup> In addition, Nejman *et al.*<sup>201</sup> found that intratumoral bacteria are predominantly intracellular and present in both cancerous and immune cells (macrophages and others). In tumor tissue, the correlation between bacterial species or their predicted functions and response to immune therapy have been established in a large spectrum of tumors. A retrospective study of 211 patients with PDAC

supported this hypothesis showing that adjuvant treatment with gemcitabine improved PFS in patients without *Klebsiella pneumoniae* in the bile culture. Similarly, quinolone administration improved OS in patients with quinolone-sensitive *Klebsiella pneumoniae*.<sup>202</sup>

Based on this evidence, manipulation of the intratumoral microbiota is emerging as a potential new therapeutic approach to PDAC. To enhance their efficacy, these strategies are usually developed in combination with other therapeutic options such as chemotherapy or immunotherapy. Among them, fecal microbiota transplantation, diet-based interventions, antibiotics, and probiotics are under investigation.<sup>203</sup>

Two studies are currently recruiting to explore the role of gut microbiota (PDAC patients or healthy controls) on anti-MSLN (mesothelin protein) CAR-T treatment (NCT04203459) or fecal microbiota transplantation (in patients with resectable PDAC) (NCT04975217).

### Conclusion

The number of actionable molecular targets for PDAC is significantly increasing, offering promising potential treatments and emphasizing the importance of continued intensive clinical and basic research in this field. Olaparib paved the way in the subset of patients with *BRCA 1/2* germline mutations. Somatic analysis of tumors by next generation sequencing is now more accessible allowing a theragnostic approach to this rapidly progressing disease. The effective targeting of *KRAS* mutations is clearly a major challenge because this gene drives up to 90% of PDAC and explains most cases of therapeutic resistance. Although targeting metabolic pathways is theoretically promising, results have been disappointing until now. The stroma signatures may help improve selection of patients for a given chemotherapy in the near future. While immunotherapies are not effective as single agent, new combinations with other drugs or T-cell engineering may bring them to the forefront once again. Finally, these promising trends in the therapeutic landscape are indissociable from all aspects supportive care.

### Declarations

*Ethics approval and consent to participate*  
Not applicable.

### Consent for publication

Not applicable.

### Author contribution(s)

**Anthony Turpin:** Conceptualization; Formal analysis; Writing – original draft; Writing – review & editing.

**Cindy Neuzillet:** Formal analysis; Validation; Writing – review & editing.

**Elise Colle:** Formal analysis; Validation; Writing – review & editing.

**Nelson Dusetti:** Formal analysis; Validation; Writing – review & editing.

**Remy Nicolle:** Formal analysis; Validation; Visualization.

**Jérôme Cros:** Formal analysis; Validation; Visualization.

**Louis de Mestier:** Formal analysis; Validation; Visualization.

**Jean-Baptiste Bachet:** Formal analysis; Visualization; Writing – review & editing.

**Pascal Hammel:** Conceptualization; Formal analysis; Supervision; Validation; Visualization; Writing – original draft; Writing – review & editing.

### Acknowledgements

Dale Roche-Lebrec for the manuscript English language editing.

### Funding

The authors received no financial support for the research, authorship, and/or publication of this article.

### Competing interests

The authors declare that there is no conflict of interest.

### Availability of data and materials

Not applicable.

### ORCID iD

Anthony Turpin  <https://orcid.org/0000-0002-2282-0101>

### References

1. Rahib L, Wehner MR, Matrisian LM, *et al.* Estimated projection of US cancer incidence

- and death to 2040. *JAMA Netw Open* 2021; 4: e214708.
2. Conroy T, Desseigne F, Ychou M, *et al.* FOLFIRINOX versus gemcitabine for metastatic pancreatic cancer. *N Engl J Med* 2011; 364: 1817–1825.
  3. Von Hoff DD, Ervin T, Arena FP, *et al.* Increased survival in pancreatic cancer with nab-paclitaxel plus gemcitabine. *N Engl J Med* 2013; 369: 1691–1703.
  4. Golan T, Hammel P, Reni M, *et al.* Maintenance olaparib for germline BRCA-mutated metastatic pancreatic cancer. *N Engl J Med* 2019; 381: 317–327.
  5. Nevala-Plagemann C, Hidalgo M and Garrido-Laguna I. From state-of-the-art treatments to novel therapies for advanced-stage pancreatic cancer. *Nat Rev Clin Oncol* 2020; 17: 108–123.
  6. Singh RR and O'Reilly EM. New treatment strategies for metastatic pancreatic ductal adenocarcinoma. *Drugs* 2020; 80: 647–669.
  7. Pishvaian MJ, Bender RJ, Halverson D, *et al.* Molecular profiling of patients with pancreatic cancer: initial results from the know your tumor initiative. *Clin Cancer Res* 2018; 24: 5018–5027.
  8. Lord CJ and Ashworth A. PARP inhibitors: synthetic lethality in the clinic. *Science* 2017; 355: 1152–1158.
  9. Neuzillet C, Rousseau B, Kocher H, *et al.* Unravelling the pharmacologic opportunities and future directions for targeted therapies in gastro-intestinal cancers Part 1: GI carcinomas. *Pharmacol Ther* 2017; 174: 145–172.
  10. Aguirre AJ, Nowak JA, Camarda ND, *et al.* Real-time genomic characterization of advanced pancreatic cancer to enable precision medicine. *Cancer Discov* 2018; 8: 1096–1111.
  11. Wang Y, Cuggia A, Pacis A, *et al.* Pancreatic cancer progression in a patient with lynch syndrome receiving immunotherapy: a cautionary tale. *J Natl Compr Cancer Netw* 2021; 19: 883–887.
  12. Cocco E, Benhamida J, Middha S, *et al.* Colorectal carcinomas containing hypermethylated MLH1 promoter and wild-type BRAF/KRAS are enriched for targetable kinase fusions. *Cancer Res* 2019; 79: 1047–1053.
  13. Boeck S, Jung A, Laubender RP, *et al.* KRAS mutation status is not predictive for objective response to anti-EGFR treatment with erlotinib in patients with advanced pancreatic cancer. *J Gastroenterol* 2013; 48: 544–548.
  14. Lito P, Solomon M, Li LS, *et al.* Allele-specific inhibitors inactivate mutant KRAS G12C by a trapping mechanism. *Science* 2016; 351: 604–608.
  15. Amodio V, Yaeger R, Arcella P, *et al.* EGFR blockade reverts resistance to KRASG12C inhibition in colorectal cancer. *Cancer Discov* 2020; 10: 1129–1139.
  16. Wang X, Allen S, Blake JF, *et al.* Identification of MRTX1133, a noncovalent, potent, and selective KRASG12D inhibitor. *J Med Chem* 2022; 65: 3123–3133.
  17. Hofmann MH, Gmachl M, Ramharter J, *et al.* BI-3406, a potent and selective SOS1-KRAS interaction inhibitor, is effective in KRAS-driven cancers through combined MEK inhibition. *Cancer Discov* 2021; 11: 142–157.
  18. Moore MJ, Goldstein D, Hamm J, *et al.* Erlotinib plus gemcitabine compared with gemcitabine alone in patients with advanced pancreatic cancer: a phase III trial of the National Cancer Institute of Canada Clinical Trials Group. *J Clin Oncol* 2007; 25: 1960–1966.
  19. Huang L, Guo Z, Wang F, *et al.* KRAS mutation: from undruggable to druggable in cancer. *Signal Transduct Target Ther* 2021; 6: 386.
  20. Marabelle A, Fakih M, Lopez J, *et al.* Association of tumour mutational burden with outcomes in patients with advanced solid tumours treated with pembrolizumab: prospective biomarker analysis of the multicohort, open-label, phase II KEYNOTE-158 study. *Lancet Oncol* 2020; 21: 1353–1365.
  21. Ying H, Kimmelman AC, Lyssiotis CA, *et al.* Oncogenic Kras maintains pancreatic tumors through regulation of anabolic glucose metabolism. *Cell* 2012; 149: 656–670.
  22. Weinstein IB and Joe A. Oncogene addiction. *Cancer Res* 2008; 68: 3077–3080.
  23. Yamamoto K, Venida A, Yano J, *et al.* Autophagy promotes immune evasion of pancreatic cancer by degrading MHC-I. *Nature* 2020; 581: 100–105.
  24. Hindson J. PDAC resistance to immunotherapy—a role for autophagy? *Nat Rev Gastroenterol Hepatol* 2020; 17: 382.
  25. Melisi D, Hollebecque A, Oh DY, *et al.* A phase Ib dose-escalation and cohort-expansion study of safety and activity of the transforming growth factor (TGF)  $\beta$  receptor I kinase inhibitor galunisertib plus the anti-PD-L1 antibody durvalumab in metastatic pancreatic cancer. *J Clin Oncol* 2019; 37: 4124.
  26. Rhim AD, Oberstein PE, Thomas DH, *et al.* Stromal elements act to restrain, rather than support, pancreatic ductal adenocarcinoma. *Cancer Cell* 2014; 25: 735–747.
  27. Özdemir BC, Hensel J, Secondini C, *et al.* The molecular signature of the stroma response

- in prostate cancer-induced osteoblastic bone metastasis highlights expansion of hematopoietic and prostate epithelial stem cell niches. *PLoS One* 2014; 9: e114530.
28. Chen Y, Kim J, Yang S, *et al.* Type I collagen deletion in  $\alpha$ SMA+ myofibroblasts augments immune suppression and accelerates progression of pancreatic cancer. *Cancer Cell* 2021; 39: 548.e6–565.e6.
  29. Dawson JC, Serrels A, Stupack DG, *et al.* Targeting FAK in anticancer combination therapies. *Nat Rev Cancer* 2021; 21: 313–324.
  30. Foote KM, Blades K, Cronin A, *et al.* Discovery of 4-{4-[(3R)-3-methylmorpholin-4-yl]-6-[1-(methylsulfonyl)cyclopropyl]pyrimidin-2-yl}-1H-indole (AZ20): a potent and selective inhibitor of ATR protein kinase with monotherapy in vivo antitumor activity. *J Med Chem* 2013; 56: 2125–2138.
  31. Henze J, Tacke F, Hardt O, *et al.* Enhancing the efficacy of CAR T cells in the tumor microenvironment of pancreatic cancer. *Cancers* 2020; 12: E1389.
  32. Park W, Chen J, Chou JF, *et al.* Genomic methods identify homologous recombination deficiency in pancreas adenocarcinoma and optimize treatment selection. *Clin Cancer Res* 2020; 26: 3239–3247.
  33. Golan T, Hammel P, Reni M, *et al.* Overall survival from the phase III POLO trial: maintenance olaparib for germline BRCA-mutated metastatic pancreatic cancer. *J Clin Oncol* 2021; 39: 378.
  34. US Food and Drug Administration. FDA briefing document: NDA 208558/S-010 (Lynparza (olaparib)), <https://www.fda.gov/media/133539/download>
  35. National Comprehensive Cancer Network (NCCN). Clinical practice guidelines in oncology: NCCN guidelines for pancreatic adenocarcinoma V.1.2021.
  36. O'Reilly EM, Lee JW, Zalupski M, *et al.* Randomized, multicenter, phase II trial of gemcitabine and cisplatin with or without veliparib in patients with pancreas adenocarcinoma and a germline BRCA/PALB2 mutation. *J Clin Oncol* 2020; 38: 1378–1388.
  37. Golan T, Atias D, Stossel C, *et al.* Patient-derived xenograft models of BRCA-associated pancreatic cancers. *Adv Drug Deliv Rev* 2021; 171: 257–265.
  38. Fojo T and Bates S. Mechanisms of resistance to PARP inhibitors—three and counting. *Cancer Discov* 2013; 3: 20–23.
  39. McMullen M, Karakasis K, Madariaga A, *et al.* Overcoming platinum and PARP-inhibitor resistance in ovarian cancer. *Cancers* 2020; 12: E1607.
  40. Shih DJH, Chen MK, Yin J, *et al.* Exploiting induced vulnerability to overcome PARPi resistance and clonal heterogeneity in BRCA mutant triple-negative inflammatory breast cancer. *Am J Cancer Res* 2022; 12: 337–354.
  41. Lin KK, Harrell MI, Oza AM, *et al.* BRCA reversion mutations in circulating tumor DNA predict primary and acquired resistance to the PARP inhibitor rucaparib in high-grade ovarian carcinoma. *Cancer Discov* 2019; 9: 210–219.
  42. Murai J, Feng Y, Yu GK, *et al.* Resistance to PARP inhibitors by SLFN11 inactivation can be overcome by ATR inhibition. *Oncotarget* 2016; 7: 76534–76550.
  43. Yazinski SA, Comaills V, Buisson R, *et al.* ATR inhibition disrupts rewired homologous recombination and fork protection pathways in PARP inhibitor-resistant BRCA-deficient cancer cells. *Genes Dev* 2017; 31: 318–332.
  44. Bradbury A, Hall S, Curtin N, *et al.* Targeting ATR as cancer therapy: a new era for synthetic lethality and synergistic combinations? *Pharmacol Ther* 2020; 207: 107450.
  45. O'Reilly EM, Lee JW, Zalupski M, *et al.* Randomized, multicenter, phase II trial of gemcitabine and cisplatin with or without veliparib in patients with pancreas adenocarcinoma and a germline BRCA/PALB2 mutation. *J Clin Oncol* 2020; 38: 1378–1388.
  46. Reiss KA, Mick R, O'Hara MH, *et al.* Phase II study of maintenance rucaparib in patients with platinum-sensitive advanced pancreatic cancer and a pathogenic germline or somatic variant in BRCA1, BRCA2, or PALB2. *J Clin Oncol* 2021; 39: 2497–2505.
  47. Singhi AD, George B, Greenbowe JR, *et al.* Real-time targeted genome profile analysis of pancreatic ductal adenocarcinomas identifies genetic alterations that might be targeted with existing drugs or used as biomarkers. *Gastroenterology* 2019; 156: 2242.e4–2253.e4.
  48. Mirza MR, Coleman RL, González-Martín A, *et al.* The forefront of ovarian cancer therapy: update on PARP inhibitors. *Ann Oncol* 2020; 31: 1148–1159.
  49. Díaz-Mejía N, García-Illescas D, Morales-Barrera R, *et al.* PARP inhibitors in advanced prostate cancer: when to use them? *Endocr Relat Cancer* 2021; 28: T79–T93.

50. Sohal DPS, Kennedy EB, Cinar P, *et al.* Metastatic pancreatic cancer: ASCO guideline update. *J Clin Oncol*. Epub ahead of print 5 August 2020. DOI: 10.1200/JCO.20.01364.
51. Lowery MA, Kelsen DP, Capanu M, *et al.* Phase II trial of veliparib in patients with previously treated BRCA-mutated pancreas ductal adenocarcinoma. *Eur J Cancer* 2018; 89: 19–26.
52. Holter S, Borgida A, Dodd A, *et al.* Germline BRCA mutations in a large clinic-based cohort of patients with pancreatic adenocarcinoma. *J Clin Oncol* 2015; 33: 3124–3129.
53. Walker EJ, Carnevale J, Pedley C, *et al.* Referral frequency, attrition rate, and outcomes of germline testing in patients with pancreatic adenocarcinoma. *Fam Cancer* 2019; 18: 241–251.
54. Peretti U, Cavaliere A, Niger M, *et al.* Germinal BRCA1-2 pathogenic variants (gBRCA1-2pv) and pancreatic cancer: epidemiology of an Italian patient cohort. *ESMO Open* 2021; 6: 100032.
55. Luo J. KRAS mutation in pancreatic cancer. *Semin Oncol* 2021; 48: 10–18.
56. Barbacid M. Ras genes. *Annu Rev Biochem* 1987; 56: 779–827.
57. Scheffzek K, Ahmadian MR, Kabsch W, *et al.* The Ras-RasGAP complex: structural basis for GTPase activation and its loss in oncogenic Ras mutants. *Science* 1997; 277: 333–338.
58. Simanshu DK, Nissley DV and McCormick F. RAS proteins and their regulators in human disease. *Cell* 2017; 170: 17–33.
59. Philip PA, Azar I, Xiu J, *et al.* Molecular characterization of **KRAS** wild-type tumors in patients with pancreatic adenocarcinoma. *Clin Cancer Res* 2022; 28: 2704–2714.
60. Luchini C, Brosens LAA, Wood LD, *et al.* Comprehensive characterisation of pancreatic ductal adenocarcinoma with microsatellite instability: histology, molecular pathology and clinical implications. *Gut* 2021; 70: 148–156.
61. Windon AL, Loaiza-Bonilla A, Jensen CE, *et al.* A KRAS wild type mutational status confers a survival advantage in pancreatic ductal adenocarcinoma. *J Gastrointest Oncol* 2018; 9: 1–10.
62. Ogura T, Yamao K, Hara K, *et al.* Prognostic value of K-ras mutation status and subtypes in endoscopic ultrasound-guided fine-needle aspiration specimens from patients with unresectable pancreatic cancer. *J Gastroenterol* 2013; 48: 640–646.
63. Schultheis B, Reuter D, Ebert MP, *et al.* Gemcitabine combined with the monoclonal antibody nimotuzumab is an active first-line regimen in KRAS wildtype patients with locally advanced or metastatic pancreatic cancer: a multicenter, randomized phase IIb study. *Ann Oncol* 2017; 28: 2429–2435.
64. Qin S, Bai Y, Wang Z, *et al.* Nimotuzumab combined with gemcitabine versus gemcitabine in K-RAS wild-type locally advanced or metastatic pancreatic cancer: a prospective, randomized-controlled, double-blinded, multicenter, and phase III clinical trial. *J Clin Oncol* 2022; 40: LBA4011.
65. Australian Pancreatic Cancer Genome Initiative, Bailey P, Chang DK, *et al.* Genomic analyses identify molecular subtypes of pancreatic cancer. *Nature* 2016; 531: 47–52.
66. Varghese AM, Singh I, Singh R, *et al.* Early-onset pancreas cancer: clinical descriptors, genomics, and outcomes. *J Natl Cancer Inst* 2021; 113: 1194–1202.
67. Heining C, Horak P, Uhrig S, *et al.* NRG1 fusions in KRAS wild-type pancreatic cancer. *Cancer Discov* 2018; 8: 1087–1095.
68. Jonna S, Feldman RA, Swensen J, *et al.* Detection of NRG1 gene fusions in solid tumors. *Clin Cancer Res* 2019; 25: 4966–4972.
69. Umemoto K and Sunakawa Y. The potential targeted drugs for fusion genes including NRG1 in pancreatic cancer. *Crit Rev Oncol Hematol* 2021; 166: 103465.
70. Safran H, Iannitti D, Ramanathan R, *et al.* Herceptin and gemcitabine for metastatic pancreatic cancers that overexpress HER-2/neu. *Cancer Invest* 2004; 22: 706–712.
71. Safran H, Miner T, Bahary N, *et al.* Lapatinib and gemcitabine for metastatic pancreatic cancer. A phase II study. *Am J Clin Oncol* 2011; 34: 50–52.
72. Boeck S, Jung A, Laubender RP, *et al.* EGFR pathway biomarkers in erlotinib-treated patients with advanced pancreatic cancer: translational results from the randomised, crossover phase III trial AIO-PK0104. *Br J Cancer* 2013; 108: 469–476.
73. Choi M, Bien H, Mofunanya A, *et al.* Challenges in Ras therapeutics in pancreatic cancer. *Semin Cancer Biol* 2019; 54: 101–108.
74. Jones MR, Williamson LM, Topham JT, *et al.* NRG1 gene fusions are recurrent, clinically actionable gene rearrangements in KRAS wild-type pancreatic ductal adenocarcinoma. *Clin Cancer Res* 2019; 25: 4674–4681.
75. Schram AM, O'Reilly EM, O'Kane GM, *et al.* Efficacy and safety of zenocutuzumab in advanced pancreas cancer and other solid tumors harboring NRG1 fusions. *J Clin Oncol* 2021; 39: 3003.

76. Russo A, Lopes AR, Scilla K, *et al.* NTRK and NRG1 gene fusions in advanced non-small cell lung cancer (NSCLC). *Precis Cancer Med* 2020; 3: 14.
77. Laskin J, Liu SV, Tolba K, *et al.* NRG1 fusion-driven tumors: biology, detection, and the therapeutic role of afatinib and other ErbB-targeting agents. *Ann Oncol* 2020; 31: 1693–1703.
78. Nagasaka M and Ou SHI. Neuregulin 1 fusion-positive NSCLC. *J Thorac Oncol* 2019; 14: 1354–1359.
79. Kheder ES and Hong DS. Emerging targeted therapy for tumors with NTRK fusion proteins. *Clin Cancer Res* 2018; 24: 5807–5814.
80. Sato K, Kawazu M, Yamamoto Y, *et al.* Fusion kinases identified by genomic analyses of sporadic microsatellite instability-high colorectal cancers. *Clin Cancer Res* 2019; 25: 378–389.
81. Wang J, Yi Y, Xiao Y, *et al.* Prevalence of recurrent oncogenic fusion in mismatch repair-deficient colorectal carcinoma with hypermethylated MLH1 and wild-type BRAF and KRAS. *Mod Pathol* 2019; 32: 1053–1064.
82. Allen MJ, Zhang A, Bavi P, *et al.* Molecular characterisation of pancreatic ductal adenocarcinoma with NTRK fusions and review of the literature. *J Clin Pathol*. Epub ahead of print 28 September 2021. DOI: 10.1136/jclinpath-2021-207781.
83. Hong DS, DuBois SG, Kummar S, *et al.* Larotrectinib in patients with TRK fusion-positive solid tumours: a pooled analysis of three phase I/II clinical trials. *Lancet Oncol* 2020; 21: 531–540.
84. Hong DS, Shen L, van Tilburg CM, *et al.* Long-term efficacy and safety of larotrectinib in an integrated dataset of patients with TRK fusion cancer. *J Clin Oncol* 2021; 39: 3108.
85. McCormick F. Progress in targeting RAS with small molecule drugs. *Biochem J* 2019; 476: 365–374.
86. Cox AD, Fesik SW, Kimmelman AC, *et al.* Drugging the undruggable RAS: mission possible? *Nat Rev Drug Discov* 2014; 13: 828–851.
87. Haigis KM. KRAS alleles: the devil is in the detail. *Trends Cancer* 2017; 3: 686–697.
88. Buscaïl L, Bournet B and Cordelier P. Role of oncogenic KRAS in the diagnosis, prognosis and treatment of pancreatic cancer. *Nat Rev Gastroenterol Hepatol* 2020; 17: 153–168.
89. Kargbo RB. Targeting KRAS G12D mutant for the potential treatment of pancreatic cancer. *ACS Med Chem Lett* 2021; 12: 1643–1645.
90. Ostrem JM, Peters U, Sos ML, *et al.* K-Ras(G12C) inhibitors allosterically control GTP affinity and effector interactions. *Nature* 2013; 503: 548–551.
91. Patricelli MP, Janes MR, Li LS, *et al.* Selective inhibition of oncogenic KRAS output with small molecules targeting the inactive state. *Cancer Discov* 2016; 6: 316–329.
92. Kargbo RB. Inhibitors of G12C mutant ras proteins for the treatment of cancers. *ACS Med Chem Lett* 2019; 10: 10–11.
93. Hong DS, Fakih MG, Strickler JH, *et al.* KRASG12C inhibition with sotorasib in advanced solid tumors. *N Engl J Med* 2020; 383: 1207–1217.
94. Strickler JH, Satake H, Hollebecque A, *et al.* First data for sotorasib in patients with pancreatic cancer with **KRAS** p.G12C mutation: a phase I/II study evaluating efficacy and safety. *J Clin Oncol* 2022; 40: 360490.
95. Bekaii-Saab TS, Spira AI, Yaeger R, *et al.* KRYSTAL-1: updated activity and safety of adagrasib (MRTX849) in patients (Pts) with unresectable or metastatic pancreatic cancer (PDAC) and other gastrointestinal (GI) tumors harboring a KRAS<sup>G12C</sup> mutation. *J Clin Oncol* 2022; 40: 519.
96. Canon J, Rex K, Saiki AY, *et al.* The clinical KRAS(G12C) inhibitor AMG 510 drives anti-tumour immunity. *Nature* 2019; 575: 217–223.
97. Sotorasib tackles KRAS G12C-mutated pancreatic cancer. *Cancer Discov* 2022; 12: 878–879.
98. Molina-Arcas M, Moore C, Rana S, *et al.* Development of combination therapies to maximize the impact of KRAS-G12C inhibitors in lung cancer. *Sci Transl Med* 2019; 11: eaaw7999.
99. Lee MS and Pant S. Personalizing medicine with germline and somatic sequencing in advanced pancreatic cancer: current treatments and novel opportunities. *Am Soc Clin Oncol Educ Book* 2021; 41: 1–13.
100. McGregor LM, Jenkins ML, Kerwin C, *et al.* Expanding the scope of electrophiles capable of targeting K-Ras oncogenes. *Biochemistry* 2017; 56: 3178–3183.
101. Yarchoan M, Johnson BA, Lutz ER, *et al.* Targeting neoantigens to augment antitumour immunity. *Nat Rev Cancer* 2017; 17: 209–222.
102. Leidner R, Sanjuan Silva N, Huang H, *et al.* Neoantigen T-cell receptor gene therapy in

- pancreatic cancer. *N Engl J Med* 2022; 386: 2112–2119.
103. Witkiewicz AK, McMillan EA, Balaji U, *et al.* Whole-exome sequencing of pancreatic cancer defines genetic diversity and therapeutic targets. *Nat Commun* 2015; 6: 6744.
  104. Seghers AK, Cuyle PJ and Van Cutsem E. Molecular targeting of a BRAF mutation in pancreatic ductal adenocarcinoma: case report and literature review. *Target Oncol* 2020; 15: 407–410.
  105. Hainsworth JD, Meric-Bernstam F, Swanton C, *et al.* Targeted therapy for advanced solid tumors on the basis of molecular profiles: results from mypathway, an open-label, phase IIa multiple basket study. *J Clin Oncol* 2018; 36: 536–542.
  106. Guan M, Bender RJ, Pishvaian MJ, *et al.* Molecular and clinical characterization of BRAF mutations in pancreatic ductal adenocarcinomas (PDACs). *J Clin Oncol* 2018; 36: 214.
  107. Hilmi M, Bartholin L and Neuzillet C. Immune therapies in pancreatic ductal adenocarcinoma: where are we now? *World J Gastroenterol* 2018; 24: 2137–2151.
  108. Hu ZI, Shia J, Stadler ZK, *et al.* Evaluating mismatch repair deficiency in pancreatic adenocarcinoma: challenges and recommendations. *Clin Cancer Res* 2018; 24: 1326–1336.
  109. Latham A, Srinivasan P, Kemel Y, *et al.* Microsatellite instability is associated with the presence of lynch syndrome pan-cancer. *J Clin Oncol* 2019; 37: 286–295.
  110. Lupinacci RM, Goloudina A, Buhard O, *et al.* Prevalence of microsatellite instability in intraductal papillary mucinous neoplasms of the pancreas. *Gastroenterology* 2018; 154: 1061–1065.
  111. Marabelle A, Le DT, Ascierto PA, *et al.* Efficacy of pembrolizumab in patients with noncolorectal high microsatellite instability/mismatch repair-deficient cancer: results from the phase II KEYNOTE-158 study. *J Clin Oncol* 2020; 38: 1–10.
  112. Yurgelun MB, Chittenden AB, Morales-Oyarvide V, *et al.* Germline cancer susceptibility gene variants, somatic second hits, and survival outcomes in patients with resected pancreatic cancer. *Genet Med* 2019; 21: 213–223.
  113. Kastrinos F. Risk of pancreatic cancer in families with lynch syndrome. *JAMA* 2009; 302: 1790.
  114. Bujanda L and Herreros-Villanueva M. Pancreatic cancer in lynch syndrome patients. *J Cancer* 2017; 8: 3667–3674.
  115. Li M and Zhao ZW. Clinical implications of mismatched repair gene promoter methylation in pancreatic cancer. *Med Oncol Northwood Lond Engl* 2012; 29: 970–976.
  116. Taieb J and Abdallah R. How I treat pancreatic cancer. *ESMO Open* 2020; 4: e000818.
  117. O'Reilly EM, Oh DY, Dhani N, *et al.* Durvalumab with or without tremelimumab for patients with metastatic pancreatic ductal adenocarcinoma: a phase II randomized clinical trial. *JAMA Oncol* 2019; 5: 1431–1438.
  118. Renouf DJ, Knox JJ, Kavan P, *et al.* LBA65 The canadian cancer trials group PA.7 trial: results of a randomized phase II study of gemcitabine (GEM) and nab-paclitaxel (Nab-P) vs GEM, nab-P, durvalumab (D) and tremelimumab (T) as first line therapy in metastatic pancreatic ductal adenocarcinoma (mPDAC). *Ann Oncol* 2020; 31: S1195.
  119. McGrail DJ, Pilié PG, Rashid NU, *et al.* High tumor mutation burden fails to predict immune checkpoint blockade response across all cancer types. *Ann Oncol* 2021; 32: 661–672.
  120. Halbrook CJ and Lyssiotis CA. Employing metabolism to improve the diagnosis and treatment of pancreatic cancer. *Cancer Cell* 2017; 31: 5–19.
  121. Son J, Lyssiotis CA, Ying H, *et al.* Glutamine supports pancreatic cancer growth through a KRAS-regulated metabolic pathway. *Nature* 2013; 496: 101–105.
  122. Alistar A, Morris BB, Desnoyer R, *et al.* Safety and tolerability of the first-in-class agent CPI-613 in combination with modified FOLFIRINOX in patients with metastatic pancreatic cancer: a single-centre, open-label, dose-escalation, phase I trial. *Lancet Oncol* 2017; 18: 770–778.
  123. Philip PA, Buyse ME, Alistar AT, *et al.* Avenger 500, a phase III open-label randomized trial of the combination of CPI-613 with modified FOLFIRINOX (mFFX) versus FOLFIRINOX (FFX) in patients with metastatic adenocarcinoma of the pancreas. *J Clin Oncol* 2019; 37: TPS479.
  124. Liu J, Ji S, Liang C, *et al.* Critical role of oncogenic KRAS in pancreatic cancer (review). *Mol Med Rep*; 13: 4943–4949.
  125. Cui H, Darmanin S, Natsuisaka M, *et al.* Enhanced expression of asparagine synthetase under glucose-deprived conditions protects pancreatic cancer cells from apoptosis induced by glucose deprivation and cisplatin. *Cancer Res* 2007; 67: 3345–3355.



126. Van Trimpont M, Peeters E, De Visser Y, *et al.* Novel insights on the use of L-asparaginase as an efficient and safe anti-cancer therapy. *Cancers* 2022; 14: 902.
127. Hammel P, Fabienne P, Mineur L, *et al.* Erythrocyte-encapsulated asparaginase (eryaspase) combined with chemotherapy in second-line treatment of advanced pancreatic cancer: an open-label, randomized phase IIb trial. *Eur J Cancer* 2020; 124: 91–101.
128. Hammel P, El-Hariry I, Macarulla T, *et al.* Trybeca-1: a randomized, phase III study of eryaspase in combination with chemotherapy versus chemotherapy alone as second-line treatment in patients with advanced pancreatic adenocarcinoma (NCT03665441). *J Clin Oncol* 2022; 40: 518–518.
129. Kordes S, Pollak MN, Zwinderman AH, *et al.* Metformin in patients with advanced pancreatic cancer: a double-blind, randomised, placebo-controlled phase II trial. *Lancet Oncol* 2015; 16: 839–847.
130. Van Cutsem E, Lenz HJ, Furuse J, *et al.* MAESTRO: a randomized, double-blind phase III study of evofosfamide (Evo) in combination with gemcitabine (Gem) in previously untreated patients (pts) with metastatic or locally advanced unresectable pancreatic ductal adenocarcinoma (PDAC). *J Clin Oncol* 2016; 34: 4007–4007.
131. Karasic TB, O'Hara MH, Loaiza-Bonilla A, *et al.* Effect of gemcitabine and nab-paclitaxel with or without hydroxychloroquine on patients with advanced pancreatic cancer: a phase II randomized clinical trial. *JAMA Oncol* 2019; 5: 993–998.
132. Kocak M, Ezazi Erdi S, Jorba G, *et al.* Targeting autophagy in disease: established and new strategies. *Autophagy* 2022; 18: 473–495.
133. Perera RM, Stoykova S, Nicolay BN, *et al.* Transcriptional control of autophagy-lysosome function drives pancreatic cancer metabolism. *Nature* 2015; 524: 361–365.
134. Erazo T, Lorente M, López-Plana A, *et al.* The new antitumor drug ABTL0812 inhibits the Akt/mTORC1 axis by upregulating tribbles-3 pseudokinase. *Clin Cancer Res* 2016; 22: 2508–2519.
135. Muñoz-Guardiola P, Casas J, Megías-Roda E, *et al.* The anti-cancer drug ABTL0812 induces ER stress-mediated cytotoxic autophagy by increasing dihydroceramide levels in cancer cells. *Autophagy* 2021; 17: 1349–1366.
136. Hosein AN, Brekken RA and Maitra A. Pancreatic cancer stroma: an update on therapeutic targeting strategies. *Nat Rev Gastroenterol Hepatol* 2020; 17: 487–505.
137. Hosein AN, Dougan SK, Aguirre AJ, *et al.* Translational advances in pancreatic ductal adenocarcinoma therapy. *Nat Cancer* 2022; 3: 272–286.
138. Hakim N, Patel R, Devoe C, *et al.* Why HALO 301 failed and implications for treatment of pancreatic cancer. *Pancreas (Fairfax)* 2019; 3: e1–e4.
139. Ramanathan RK, McDonough SL, Philip PA, *et al.* Phase IB/II randomized study of FOLFIRINOX plus pegylated recombinant human hyaluronidase versus FOLFIRINOX alone in patients with metastatic pancreatic adenocarcinoma: SWOG S1313. *J Clin Oncol* 2019; 37: 1062–1069.
140. Van Cutsem E, Tempero MA, Sigal D, *et al.* Randomized phase III trial of pegvorhyaluronidase alfa with Nab-Paclitaxel plus gemcitabine for patients with hyaluronan-high metastatic pancreatic adenocarcinoma. *J Clin Oncol* 2020; 38: 31853194.
141. Huang Y, Yuan J, Righi E, *et al.* Vascular normalizing doses of antiangiogenic treatment reprogram the immunosuppressive tumor microenvironment and enhance immunotherapy. *Proc Natl Acad Sci U S A* 2012; 109: 17561–17566.
142. Neuzillet C, Tijeras-Raballand A, Ragulan C, *et al.* Inter- and intra-tumoural heterogeneity in cancer-associated fibroblasts of human pancreatic ductal adenocarcinoma. *J Pathol* 2019; 248: 51–65.
143. Kalluri R. The biology and function of fibroblasts in cancer. *Nat Rev Cancer* 2016; 16: 582–598.
144. Olive KP, Jacobetz MA, Davidson CJ, *et al.* Inhibition of hedgehog signaling enhances delivery of chemotherapy in a mouse model of pancreatic cancer. *Science* 2009; 324: 1457–1461.
145. Catenacci DVT, Junttila MR, Karrison T, *et al.* Randomized phase Ib/II study of gemcitabine plus placebo or vismodegib, a hedgehog pathway inhibitor, in patients with metastatic pancreatic cancer. *J Clin Oncol* 2015; 33: 4284–4292.
146. Teicher BA. TGFβ-directed therapeutics: 2020. *Pharmacol Ther* 2021; 217: 107666.
147. Özdemir BC, Pentcheva-Hoang T, Carstens JL, *et al.* Depletion of carcinoma-associated fibroblasts and fibrosis induces immunosuppression and accelerates pancreas

- cancer with reduced survival. *Cancer Cell* 2014; 25: 719–734.
148. Serrels A, Lund T, Serrels B, *et al.* Nuclear FAK controls chemokine transcription, Tregs, and evasion of anti-tumor immunity. *Cell* 2015; 163: 160–173.
  149. Jiang H, Liu X, Knolhoff BL, *et al.* Development of resistance to FAK inhibition in pancreatic cancer is linked to stromal depletion. *Gut* 2020; 69: 122–132.
  150. Aung KL, McWhirter E, Welch S, *et al.* A phase II trial of GSK2256098 and trametinib in patients with advanced pancreatic ductal adenocarcinoma (PDAC) (MOBILITY-002 Trial, NCT02428270). *J Clin Oncol* 2018; 36: 409–409.
  151. Massó-Vallés D, Jauset T, Serrano E, *et al.* Ibrutinib exerts potent antifibrotic and antitumor activities in mouse models of pancreatic adenocarcinoma. *Cancer Res* 2015; 75: 1675–1681.
  152. Tempero M, Oh DY, Tabernero J, *et al.* Ibrutinib in combination with nab-paclitaxel and gemcitabine for first-line treatment of patients with metastatic pancreatic adenocarcinoma: phase III RESOLVE study. *Ann Oncol* 2021; 32: 600–608.
  153. Hong D, Rasco D, Veeder M, *et al.* A phase Ib/II study of the bruton tyrosine kinase inhibitor ibrutinib and the PD-L1 inhibitor durvalumab in patients with pretreated solid tumors. *Oncology* 2019; 97: 102–111.
  154. Overman M, Javle M, Davis RE, *et al.* Randomized phase II study of the bruton tyrosine kinase inhibitor acalabrutinib, alone or with pembrolizumab in patients with advanced pancreatic cancer. *J Immunother Cancer* 2020; 8: e000587.
  155. Neesse A, Frese KK, Bapiro TE, *et al.* CTGF antagonism with mAb FG-3019 enhances chemotherapy response without increasing drug delivery in murine ductal pancreas cancer. *Proc Natl Acad Sci U S A* 2013; 110: 12325–12330.
  156. Picozzi V, Alseidi A, Winter J, *et al.* Gemcitabine/nab-paclitaxel with pamrevlumab: a novel drug combination and trial design for the treatment of locally advanced pancreatic cancer. *ESMO Open* 2020; 5: e000668.
  157. Costello E, Greenhalf W and Neoptolemos JP. New biomarkers and targets in pancreatic cancer and their application to treatment. *Nat Rev Gastroenterol Hepatol* 2012; 9: 435–444.
  158. Tiriác H, Belleau P, Engle DD, *et al.* Organoid profiling identifies common responders to chemotherapy in pancreatic cancer. *Cancer Discov* 2018; 8: 1112–1129.
  159. Sparano JA, Gray RJ, Makower DF, *et al.* Adjuvant chemotherapy guided by a 21-gene expression assay in breast cancer. *N Engl J Med* 2018; 379: 111–121.
  160. Cardoso F, van't Veer LJ, Bogaerts J, *et al.* 70-Gene signature as an aid to treatment decisions in early-stage breast cancer. *N Engl J Med* 2016; 375: 717–729.
  161. Zhao SG, Chang SL, Spratt DE, *et al.* Development and validation of a 24-gene predictor of response to postoperative radiotherapy in prostate cancer: a matched, retrospective analysis. *Lancet Oncol* 2016; 17: 1612–1620.
  162. Nicolle R, Gayet O, Bigonnet M, *et al.* Relevance of biopsy-derived pancreatic organoids in the development of efficient transcriptomic signatures to predict adjuvant chemosensitivity in pancreatic cancer. *Transl Oncol* 2022; 16: 101315.
  163. Nicolle R, Gayet O, Duconseil P, *et al.* A transcriptomic signature to predict adjuvant gemcitabine sensitivity in pancreatic adenocarcinoma. *Ann Oncol* 2021; 32: 250–260.
  164. Puleo F, Nicolle R, Blum Y, *et al.* Stratification of pancreatic ductal adenocarcinomas based on tumor and microenvironment features. *Gastroenterology* 2018; 155: 1999.e3–2013.e3.
  165. Aung KL, Fischer SE, Denroche RE, *et al.* Genomics-driven precision medicine for advanced pancreatic cancer: early results from the COMPASS trial. *Clin Cancer Res* 2018; 24: 1344–1354.
  166. O'Kane GM, Grünwald BT, Jang GH, *et al.* GATA6 expression distinguishes classical and basal-like subtypes in advanced pancreatic cancer. *Clin Cancer Res* 2020; 26: 4901–4910.
  167. Martinelli P, Carrillo-de Santa Pau E, Cox T, *et al.* GATA6 regulates EMT and tumour dissemination, and is a marker of response to adjuvant chemotherapy in pancreatic cancer. *Gut* 2017; 66: 1665–1676.
  168. Nicolle R, Blum Y, Duconseil P, *et al.* Establishment of a pancreatic adenocarcinoma molecular gradient (PAMG) that predicts the clinical outcome of pancreatic cancer. *EBioMedicine* 2020; 57: 102858.
  169. Pilié PG, Tang C, Mills GB, *et al.* State-of-the-art strategies for targeting the DNA damage response in cancer. *Nat Rev Clin Oncol* 2019; 16: 81–104.

170. Hanahan D. Hallmarks of cancer: new dimensions. *Cancer Discov* 2022; 12: 31–46.
171. Gilad O, Nabet BY, Ragland RL, *et al.* Combining ATR suppression with oncogenic Ras synergistically increases genomic instability, causing synthetic lethality or tumorigenesis in a dosage-dependent manner. *Cancer Res* 2010; 70: 9693–9702.
172. Huntoon CJ, Flatten KS, Wahner Hendrickson AE, *et al.* ATR inhibition broadly sensitizes ovarian cancer cells to chemotherapy independent of BRCA status. *Cancer Res* 2013; 73: 3683–3691.
173. Kim H, George E, Ragland R, *et al.* Targeting the ATR/CHK1 axis with PARP inhibition results in tumor regression in BRCA-mutant ovarian cancer models. *Clin Cancer Res* 2017; 23: 3097–3108.
174. Terranova N, Jansen M, Falk M, *et al.* Population pharmacokinetics of ATR inhibitor berzosertib in phase I studies for different cancer types. *Cancer Chemother Pharmacol* 2021; 87: 185–196.
175. Fokas E, Prevo R, Hammond EM, *et al.* Targeting ATR in DNA damage response and cancer therapeutics. *Cancer Treat Rev* 2014; 40: 109–117.
176. Dreyer SB, Upstill-Goddard R, Paulus-Hock V, *et al.* Targeting DNA damage response and replication stress in pancreatic cancer. *Gastroenterology* 2021; 160: 362–377
177. Al Zubaidi T, Gehrisch OHF, Genois MM, *et al.* Targeting the DNA replication stress phenotype of KRAS mutant cancer cells. *Sci Rep* 2021; 11: 3656.
178. June CH, O'Connor RS, Kawalekar OU, *et al.* CAR T cell immunotherapy for human cancer. *Science* 2018; 359: 1361–1365.
179. D'Aloia MM, Zizzari IG, Sacchetti B, *et al.* CAR-T cells: the long and winding road to solid tumors. *Cell Death Dis* 2018; 9: 282.
180. Beatty GL, O'Hara MH, Lacey SF, *et al.* Activity of mesothelin-specific chimeric antigen receptor T cells against pancreatic carcinoma metastases in a phase I trial. *Gastroenterology* 2018; 155: 29–32.
181. Raj D, Nikolaidi M, Garcés I, *et al.* CEACAM7 is an effective target for CAR T-cell therapy of pancreatic ductal adenocarcinoma. *Clin Cancer Res* 2021; 27: 1538–1552.
182. Argani P, Iacobuzio-Donahue C, Ryu B, *et al.* Mesothelin is overexpressed in the vast majority of ductal adenocarcinomas of the pancreas: identification of a new pancreatic cancer marker by serial analysis of gene expression (SAGE). *Clin Cancer Res* 2001; 7: 3862–3868.
183. Weidemann S, Perez D, Izbicki JR, *et al.* Mesothelin is commonly expressed in pancreatic adenocarcinoma but unrelated to cancer aggressiveness. *Cancer Invest* 2021; 39: 711–720.
184. Hassan R, Laszik ZG, Lerner M, *et al.* Mesothelin is overexpressed in pancreaticobiliary adenocarcinomas but not in normal pancreas and chronic pancreatitis. *Am J Clin Pathol* 2005; 124: 838–845.
185. Haas AR, Tanyi JL, O'Hara MH, *et al.* Phase I study of lentiviral-transduced chimeric antigen receptor-modified T cells recognizing mesothelin in advanced solid cancers. *Mol Ther* 2019; 27: 1919–1929.
186. Raj D, Yang MH, Rodgers D, *et al.* Switchable CAR-T cells mediate remission in metastatic pancreatic ductal adenocarcinoma. *Gut* 2019; 68: 1052–1064.
187. Morgan RA, Yang JC, Kitano M, *et al.* Case report of a serious adverse event following the administration of T cells transduced with a chimeric antigen receptor recognizing ERBB2. *Mol Ther* 2010; 18: 843–851.
188. Allum WH, Stokes HJ, Macdonald F, *et al.* Demonstration of carcinoembryonic antigen (CEA) expression in normal, chronically inflamed, and malignant pancreatic tissue by immunohistochemistry. *J Clin Pathol* 1986; 39: 610–614.
189. Chmielewski M, Hahn O, Rappl G, *et al.* T cells that target carcinoembryonic antigen eradicate orthotopic pancreatic carcinomas without inducing autoimmune colitis in mice. *Gastroenterology* 2012; 143: 1095.e2–1107.e2.
190. Thistlethwaite FC, Gilham DE, Guest RD, *et al.* The clinical efficacy of first-generation carcinoembryonic antigen (CEACAM5)-specific CAR T cells is limited by poor persistence and transient pre-conditioning-dependent respiratory toxicity. *Cancer Immunol Immunother* 2017; 66: 1425–1436.
191. Schizas D, Charalampakis N, Kole C, *et al.* Immunotherapy for pancreatic cancer: a 2020 update. *Cancer Treat Rev* 2020; 86: 102016.
192. Ducreux M, Seufferlein T, Van Laethem JL, *et al.* Systemic treatment of pancreatic cancer revisited. *Semin Oncol* 2019; 46: 28–38.
193. Le DT, Lutz E, Uram JN, *et al.* Evaluation of ipilimumab in combination with allogeneic pancreatic tumor cells transfected with a

- GM-CSF gene in previously treated pancreatic cancer. *J Immunother* 2013; 36: 382–389.
194. Wu AA, Bever KM, Ho WJ, *et al.* A phase II study of allogeneic GM-CSF-transfected pancreatic tumor vaccine (GVAX) with ipilimumab as maintenance treatment for metastatic pancreatic cancer. *Clin Cancer Res* 2020; 26: 5129–5139.
195. Bernhardt SL, Gjertsen MK, Trachsel S, *et al.* Telomerase peptide vaccination of patients with non-resectable pancreatic cancer: a dose escalating phase I/II study. *Br J Cancer* 2006; 95: 1474–1482.
196. Middleton G, Silcocks P, Cox T, *et al.* Gemcitabine and capecitabine with or without telomerase peptide vaccine GV1001 in patients with locally advanced or metastatic pancreatic cancer (TeloVac): an open-label, randomised, phase III trial. *Lancet Oncol* 2014; 15: 829–840.
197. Huang X, Zhang G, Tang T, *et al.* Identification of tumor antigens and immune subtypes of pancreatic adenocarcinoma for mRNA vaccine development. *Mol Cancer* 2021; 20: 44.
198. Zhang Y, Ma JA, Zhang HX, *et al.* Cancer vaccines: targeting KRAS-driven cancers. *Expert Rev Vaccines* 2020; 19: 163–173.
199. Tijeras-Raballand A, Hilmi M, Astorgues-Xerri L, *et al.* Microbiome and pancreatic ductal adenocarcinoma. *Clin Res Hepatol Gastroenterol* 2021; 45: 101589.
200. Geller LT, Barzily-Rokni M, Danino T, *et al.* Potential role of intratumor bacteria in mediating tumor resistance to the chemotherapeutic drug gemcitabine. *Science* 2017; 357: 1156–1160.
201. Nejman D, Livyatan I, Fuks G, *et al.* The human tumor microbiome is composed of tumor type-specific intracellular bacteria. *Science* 2020; 368: 973–980.
202. Weniger M, Hank T, Qadan M, *et al.* Influence of *Klebsiella pneumoniae* and quinolone treatment on prognosis in patients with pancreatic cancer. *Br J Surg* 2021; 108: 709–716.
203. Arsenijevic T, Nicolle R, Bouchart C, *et al.* Pancreatic cancer meets human microbiota: close encounters of the third kind. *Cancers* 2021; 13: 1231.

Visit SAGE journals online  
[journals.sagepub.com/  
 home/tam](https://journals.sagepub.com/home/tam)

 SAGE journals