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REVIEW

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Microorganisms associated with hedgehog arthropods

Linda Benkacimi^{1,2}, Adama Zan Diarra^{1,2}, Jean-Michel Bompar³, Jean-Michel Bérenger^{1,2} and Philippe Parola^{1,2*}

Abstract

Hedgehogs are small synanthropic mammals that live in rural areas as well as in urban and suburban areas. They can be reservoirs of several microorganisms, including certain pathogenic agents that cause human and animal public health issues. Hedgehogs are often parasitized by blood-sucking arthropods, mainly hard ticks and fleas, which in turn can also carry various vector-borne microorganisms of zoonotic importance. Many biotic factors, such as urbanization and agricultural mechanization, have resulted in the destruction of the hedgehog's natural habitats, leading these animals to take refuge near human dwellings, seeking food and shelter in parks and gardens and exposing humans to zoonotic agents that can be transmitted either directly by them or indirectly by their ectoparasites. In this review, we focus on the microorganisms detected in arthropods sampled from hedgehogs worldwide. Several microorganisms have been reported in ticks collected from these animals, including various *Borrelia* spp., *Anaplasma* spp., *Ehrlichia* spp., and *Rickettsia* spp. species as well as *Coxiella burnetii* and *Leptospira* spp. As for fleas, *C. burnetii*, *Rickettsia* spp., *Wolbachia* spp., *Mycobacterium* spp. and various *Bartonella* species have been reported. The detection of these microorganisms in arthropods does not necessarily mean that they can be transmitted to humans and animals. While the vector capacity and competence of fleas and ticks for some of these microorganisms has been proven, in other cases the microorganisms may have simply been ingested with blood taken from an infected host. Further investigations are needed to clarify this issue. As hedgehogs are protected animals, handling them is highly regulated, making it difficult to conduct epidemiological studies on them. Their ectoparasites represent a very interesting source of information on microorganisms circulating in populations of these animals, especially vector-borne ones.

Keywords Hedgehogs, Ticks, Fleas, Microorganisms, Zoonoses

Background

Hedgehogs are small synanthropic nocturnal omnivorous mammals whose dorsal part of the skin is wholly covered with spines [1, 2]. They are hibernating animals, dig their own burrows, and live in rural areas as well as in urban and suburban areas [1]. Hedgehogs belong to the

Erinaceidae family, and include up to 17 known species divided into five genera (*Erinaceus*, *Atelerix*, *Hemiechinus*, *Paraechinus* and *Mesechinus*) distributed worldwide [3, 4]. Hedgehogs are found in Europe, Africa and Asia and were introduced by importation into New Zealand and the Americas [1, 4, 5]. Hedgehog populations have been continuously decreasing, leading the authorities of several countries to provide protection by classifying them in their lists of species threatened with extinction. It is therefore prohibited to kill them or to retain them in captivity [3].

Interest in these animals has increased over the years. The first studies, dating from the late nineteenth century, were mainly focused on the histology, physiology and anatomy of hedgehogs [6]. Researchers have been

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very interested in their hibernation process, mainly in the physiological changes allowing them to go from an homeothermic animal status to a poikilothermic animal status and vice versa [7].

In the 1940s, laboratory-reared populations of *Erinaceus europaeus* hedgehogs were established as a means to meet research needs [8]. However, it was only in the 1950s that researchers began to show greater interest in their role as reservoirs for pathogens involved in human and animal public health issues and possibly responsible for zoonoses [5, 9, 10]. Laboratory-reared hedgehog specimens were used in experimental models which aimed to determine their sensitivity to different microorganisms, including *Borrelia* strains (*B. duttoni*, *B. crocidurae*, *B. hispanica* and *B. persica*) [9]. The experimental infection of a hedgehog by *Trypanosoma gambiense* showed that this parasite caused a chronic infection in the animal leading to its death [11]. Different microorganisms have already been reported in hedgehogs, including bacteria (*Yersinia pseudotuberculosis*, *Coxiella burnetii*, *Mycobacterium* spp., *Corynebacterium* spp., methicillin-resistant *Staphylococcus aureus*, *Salmonella* spp., *Leptospira* spp., *Chlamydia psittaci* and *Streptococcus* spp.), protozoa (*Cryptosporidium* spp., *Toxoplasma gondii*), fungus (*Candida albicans*, *Microsporium* spp., *Trichophyton erinacei*) and viruses (Coronaviruses, *Herpesvirus*, rabies virus, *Paramyxovirus*) [2, 3]. Similarly, several vector-borne microorganisms have also been detected, including *Tick-borne encephalitis virus* (TBEV), *Crimean-Congo hemorrhagic fever orthonairovirus* (CCHFV), Severe Fever with Thrombocytopenia Syndrome Virus (SFTSV), *Bhanja bandavirus*, *Yersinia pestis*, *Rickettsia* spp., *Bartonella* spp., *Babesia* spp., *Candidatus* *Neorhlichia mikurensis* and *Leishmania* spp. [2, 3, 12–15]. In addition, several studies have reported hedgehog infection by *Borrelia burgdorferi sensu stricto* (*B. burgdorferi* s.s.), the agent of Lyme disease [3, 16], and *Anaplasma phagocytophilum*, the agent of granulocytic anaplasmosis [3, 12].

Hedgehogs are often parasitized by mites and blood-sucking arthropods (Fig. 1), mainly hard ticks and fleas (Fig. 2) [12, 13]. *Ixodes hexagonus* and *Ixodes ricinus* are the most frequently collected ticks from hedgehogs [3, 13, 17, 18], but other tick species have also been reported, including *Rhipicephalus sanguineus sensu lato* (*Rh. sanguineus* s.l.) (Fig. 2), *Rhipicephalus (Boophilus) annulatus*, *Rhipicephalus camicasi*, *Haemaphysalis erinacei* (Fig. 2), *Haemaphysalis flava*, *Haemaphysalis longicornis*, *Haemaphysalis colesbergensis*, *Haemaphysalis norvali*, *Amblyomma marmoreum*, *Dermacentor nuttalli*, *Hyalomma aegyptium*, *Hyalomma excavatum*, *Hyalomma*



Fig. 1 Hedgehogs parasitized by ticks

asiaticum, *Hyalomma dromedarii*, *Hyalomma schulzei*, *Ixodes acuminatus*, *Rhipicephalus nuttalli* and the soft tick *Ornithodoros* [4, 13, 16–30]. All developmental stages (larva, nymph and male/female adults) have been found feeding on these animals [27, 31]. Moreover, one study has shown that odors related to animal health status seems to have an impact on the tick infestation rate [4]. The most common species of flea collected from hedgehogs is *Archaeopsylla erinacei* (Fig. 2) [4, 16, 18, 21, 32]. Other flea species have also been reported (*Leptopsylla segnis*, *Ctenocephalides canis* and *Ctenocephalides felis*) [17, 18, 33, 34].

Fleas and ticks are also known to be vectors of different pathogens, thereby presenting a risk to human and animal health, causing such diseases as bartonellosis, flea-borne spotted fever rickettsiosis, plague, murine typhus for fleas and borreliosis, anaplasmosis, tick-borne spotted fever rickettsiosis, ehrlichiosis and several arboviruses for ticks [13, 16, 21, 35, 36]. Some of these flea and tick species have a weak host specificity and can bite different hosts, including wild, domestic and peri-domestic animals as well as humans; as such, they can thus participate in the exchange of microorganisms between their different hosts. [16, 18, 27, 37]. In addition, some tick species feed on a different host at each developmental stage and can thus actively participate in the spread of pathogens that could be responsible for zoonoses [38].

Many biotic factors, such as urbanization, land consolidation and large-scale mechanized agriculture,

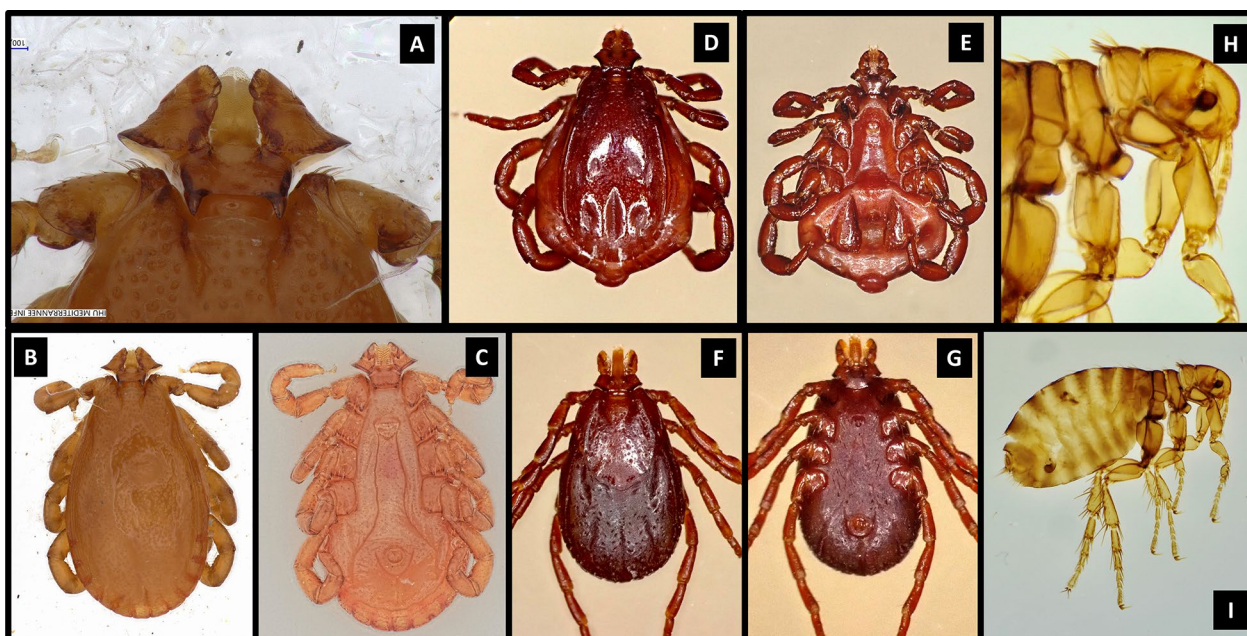


Fig. 2 Photographs of arthropods sampled from *Aterlix algirus* hedgehogs taken with a Zeiss Axio Zoom V16 stereomicroscope (Carl Zeiss Meditec France S.A.S., Marly le Roi, France) and a VHX-7000 digital microscope (Keyence Corp., Osaka, Japan). **a–c** *Haemaphysalis erinacei* capitulum, dorsal and ventral views, **d, e** *Rhipicephalus sanguineus* sensu lato male dorsal and ventral views, **f, g** *Rhipicephalus sanguineus* sensu lato female dorsal and ventral views, **h, i** *Archaeopsylla erinacei* female flea

have resulted in the destruction of the hedgehog's natural habitats, leading them to take refuge near human dwellings where they seek food and shelters in gardens and parks [2, 3]. In some countries and populations, hedgehogs can be used in traditional medicine and also eaten [3, 4]. In addition, the European hedgehog *E. europaeus* and the African pygmy hedgehog *Aterlix albiventris* are frequently taken as pets and are often transported with their parasites and pathogens from one region to another, exposing humans to several zoonotic agents, including those carried by their ectoparasites [2, 13, 18].

Two interesting reviews on hedgehog zoonoses have been published. However, the authors of these reviews mainly focused on the microorganisms detected in the hedgehogs themselves, and only partially addressed those detected in the ectoparasites collected from them by mentioning some of the pathogens associated with ticks but not those associated with fleas [2, 3].

The aim of the present review is to collate and list the information available in the literature on the microorganisms depicting a zoonotic threat that have been detected in arthropods sampled from different hedgehog species worldwide. We do not detail here the microorganisms detected in the hedgehogs themselves because this topic has been adequately covered in the two previous reviews [2, 3].

Microorganisms detected in ticks collected from hedgehogs

Coxiella burnetii

Coxiella burnetii is a Gram-negative obligate, intracellular bacterium [39] that is the causative agent of Q fever [20, 39]. This bacterium is widely distributed in the environment and affects different domestic and wild animal species as well as humans and arthropods [20, 27, 39], but livestock are considered to be the principal reservoir for this zoonotic disease [3, 20]. This bacterium can be transmitted to humans and animals through direct contact with infected animals or indirectly via aerosols, infected urine, feces, birth products, milk and other dairy products [3, 20, 28].

Coxiella burnetii has been reported in over 40 tick species collected from several animals, including hedgehogs [27]. It has been detected in *Hyalomma excavatum* collected from *Hemiechinus auritus* (the long-eared hedgehog) in Egypt (Alexandria) [28], in 3.2% (4/125) of *Ha. flava* ticks collected from *Erinaceus amurensis* (the Amur hedgehogs) captured in forests near the cesspools in Xianning City, Hubei Province, in Central China [20] and in *Ha. erinacei* sampled from *Aterlix algirus* (North African hedgehog) collected dead in Bouira Province of northern Algeria [21]. Previous studies reported *C. burnetii* in hedgehog tissues, suggesting that these animals as well as their associated ectoparasites could be considered

as reservoir hosts for this pathogen [3, 20, 21]. Ticks have been suspected of playing a role in the epidemiology and transmission of *C. burnetii*. However, to our knowledge, only one experimental study has demonstrated the vectorial capacity of *Hyalomma aegyptium* ticks for *C. burnetii* [27, 40]. Moreover, ticks have been shown to transmit *C. burnetii* horizontally through their saliva and feces and also vertically through transstadial and transovarial routes [27].

***Borrelia* spp.**

Borrelia burgdorferi sensu lato (*B. burgdorferi* s.l.) is a spirochete bacteria and the causative agent of Lyme borreliosis, the most common vector-borne disease in Europe and USA, infecting various vertebrate reservoir hosts [3, 41–43]. Ticks, mainly the endophilic *I. hexagonus* and those of the exophilic *I. ricinus* complex, are the recognized vectors, and transmission occurs during blood meals through their infected saliva [18, 41–44]. Humans are considered as dead-end hosts and do not play a role in the epidemiology of *B. burgdorferi* s.l. [43]. The *B. burgdorferi* s.l. group contains three *Candidatus* genospecies and at least 20 validated species, including *B. burgdorferi* s.s., *B. afzelii*, *B. garinii*, *B. bavariensis* and *B. spielmanii* [41, 42, 45].

Preliminary results of a study conducted by Skuballa et al. showed that an *I. hexagonus* nymph and an *I. ricinus* female tick sampled from European hedgehogs from Karlsruhe (Germany) were infected with *B. afzelii* and *B. spielmanii* [41]. Another study conducted in Germany on *I. hexagonus* ticks collected from hedgehogs showed that two specimens were infected with *B. afzelii*, one was infected with *B. spielmanii* and one was co-infected with *B. garinii* and *B. afzelii* [42]. Furthermore, the DNA of *B. miyamotoi*, the spirochete bacterium belonging to the relapsing fever group, causing human non-specific febrile illness, and *B. bavariensis*, have been detected in both *I. hexagonus* and *I. ricinus* collected from hedgehogs [3]. These *Borrelia* species, which are either confirmed or suspected pathogens for humans, have also been previously detected in hedgehog tissue, suggesting that they may be reservoir hosts. However, further investigation is needed in order to determine the precise role that hedgehogs play in the epidemiology of these bacteria [3, 18, 41].

Anaplasma phagocytophilum

Anaplasma phagocytophilum is an obligate intracellular Gram-negative bacterium [31, 46]. It is an emerging tick-borne zoonotic pathogen responsible for granulocytic anaplasmosis, an acute febrile disease of public health importance that affects humans, domestic and wild animals distributed worldwide [3, 12, 46]. *Ixodes* ticks, mainly *I. ricinus* in Europe, *I. scapularis*

and *I. pacificus* in the Americas and *I. persulcatus* in Asia, have been reported to be the main vectors of *A. phagocytophilum*, and wild rodents are considered as natural reservoir hosts. Infection of humans is accidental, and humans are considered to be dead-end hosts [3, 12, 31, 46].

Anaplasma phagocytophilum was detected in 15/38 (39.47%) engorged *I. hexagonus* female ticks collected from 7/14 (50%) *E. europaeus* hedgehogs from Germany [12], in 74/277 (27%) *I. hexagonus* and 6/25 (24%) *I. ricinus* ticks sampled from hedgehogs captured alive in the Netherlands, in 413/563 (73.36%) *I. ricinus* and 90/338 (26.63%) *I. hexagonus* sampled from a captive hedgehog population in Baden-Württemberg (Germany) [31] and in the same two tick species from hedgehogs in a study conducted in Belgium [3]. In addition, 4/13 (30.8%) pools of *Dermacentor nuttalli* ticks collected from hedgehogs in Hebei Province (China) were positive for *A. phagocytophilum*. *Dermacentor* ticks are unusual hosts, and their vectorial capacity should therefore be investigated [46].

Hedgehogs are often parasitized by ticks and thus can be involved in the circulation of this bacterium [3]. Indeed, *A. phagocytophilum* has also been detected in *E. europaeus* and *Erinaceus roumanicus* hedgehog blood and tissue [3, 12, 31]. The sequences obtained by Skuballa et al. from hedgehog tissue were almost identical to those they obtained from the ticks collected from the hedgehogs in a study they conducted in Germany, indicating that transmission can probably occur from one of these hosts to the other [12]. Only some of the different *A. phagocytophilum* strains have been confirmed to be human pathogens, and the reservoirs of the pathogenic strains involved in human disease remain unknown [12, 46]. Sequencing has revealed that the most frequent variant (variant 'A') found in the study conducted by Silaghi et al. [31] was similar to that reported previously in human, canine and equine granulocytic anaplasmosis [31]. The role of the strains found in hedgehogs and their ticks in terms of public health issues remains unclear, but the findings provided by Silaghi et al. [31] suggest they may have some involvement. Further studies are necessary to clarify this.

Anaplasma marginale

Anaplasma marginale is a tick-borne pathogen causing a hemolytic disease that can sometimes be severe [26]. About 20 tick species have been reported as potential vectors in which the bacteria multiply in the midgut and salivary glands. Nevertheless, *Dermacentor* spp., *Rhipicephalus annulatus* and *Rhipicephalus bursa* are considered to be the main vectors [26, 47]. Transmission can occur between some tick species through the transovarian, transstadial and intra-stadial routes, and it has been

previously proven that transmission to another host is possible [26].

Using PCR and sequencing, *A. marginale* has been detected in three *Rhipicephalus turanicus* tick pools and in the blood of 2/53 (3.8%) live long-eared hedgehogs (*H. auritus*) from which the ticks were sampled. These hedgehogs were captured in urban and suburban areas around Zabol in south-eastern Iran [26]. In addition, 40 *I. hexagonus* sampled from *E. europaeus* hedgehogs in the Netherlands tested positive for this bacterium [26]. The infection in hedgehogs seems to be subject to cyclical variations, raising questions as to whether the infection is persistent, making this animal a suitable reservoir, or whether it is rapidly eliminated by the immune system [26]. Further investigations are needed to clarify the involvement of hedgehogs in the epidemiological cycle of this bacterium.

Anaplasma bovis

Anaplasma bovis is an Anaplasmataceae agent of bovine anaplasmosis, a tick-borne infectious disease, which parasitizes monocytes and macrophages. It causes animal body weight loss, abortions and decreased milk production, and frequently leads to death of the infected animal [48, 49]. *Anaplasma bovis* has been detected in 4% (5/125) of *Ha. flava* ticks sampled from *E. amurensis* hedgehogs captured in forests near the cesspools in Xianning City, Hubei Province, in central China [20].

Ehrlichia spp.

The DNA of two potential new uncultivated *Ehrlichia* species has been reported in ticks collected from hedgehogs. The first was phylogenetically close to *Ehrlichia ewingii* and was detected in 9.6%, (12/125) of *Ha. flava* ticks sampled from *E. amurensis* hedgehogs captured in forests near the cesspools in Xianning City, Hubei Province, in central China [20]. The second was detected in *Rh. sanguineus* s.l. and *Ha. erinacei* ticks sampled from *A. algirus* hedgehogs in Bouira Province in north-central Algeria [21]. Phylogenetic analysis revealed that the detected *Ehrlichia* spp. clustered together with another uncultured *Ehrlichia* in a clade separate from other known species [21]. These two *Ehrlichia* spp. have not been cultivated to allow formal description of the species. Their potential pathogenicity for animals and humans also remains unknown and requires investigation.

Rickettsia

Rickettsia are obligate intracellular bacteria causing emerging and re-emerging diseases worldwide [50]. The principal vectors are hematophagous arthropods, and they are mainly associated with ticks, fleas and lice [50].

Rickettsia massiliae

Several studies conducted in Algeria reported detecting *R. massiliae* in hedgehog ticks. Indeed, this bacterium has been detected in *Rh. sanguineus* s.l. ticks sampled from hedgehogs in Algiers [51], in north-east Algeria [4] and in 40/59 (67,80%) *Rh. sanguineus* s.l. ticks sampled from dead *A. algirus* hedgehogs in the Bouira province [21]. The DNA of *R. massiliae* has also been found in 22/212 (10.38%) *Rh. sanguineus* s.l. collected from five European hedgehogs (*E. europaeus*) captured in seven districts in the north and center of Portugal [13] and in 11/12 (91.7%) *Rh. sanguineus* s.l. ticks collected from a female *E. europaeus* hedgehog that was captured in the city center of Marseille, France [16]. *Rhipicephalus sanguineus* s.l. ticks represent the main vectors of *R. massiliae*, and this bacterium has been detected in their saliva [51]. Transovarial and transstadial transmission has also been reported among the tick population, suggesting that these ticks might be potentially reservoirs [16, 52]. Cases of human infection by *R. massiliae* have been reported in the literature [13].

Rickettsia conorii

Rickettsia conorii is the etiological agent of Mediterranean spotted fever (MSF) rickettsiosis, an endemic zoonosis in the Mediterranean basin transmitted mainly by the brown dog tick *Rh. sanguineus* s.l. [50]. *Rickettsia conorii conorii* strain Malish has been detected in a *Rh. sanguineus* s.l. specimen collected from a hedgehog in Algiers [51].

Other Rickettsia species

Other *Rickettsia* species, including *R. japonica* and *R. raoultii*, have been detected in 4/125 (3.2%) and 3/125 (2.4%) *Ha. flava* ticks, respectively, collected from *E. amurensis* hedgehogs captured in forests near the cesspools in Xianning City, Hubei Province in central China [20]. *Rickettsia heilongjiangensis* (*R. heilongjiangensis* XY-1) was reported for the first time in 79/110 (71.8%) *Ha. flava* ticks and in 7/45 (15.6%) *E. amurensis* hedgehogs from which the positive ticks were collected in Xuyi County, southeast China [25]. *Rickettsia helvetica* has been reported in *I. hexagonus* and *I. ricinus* ticks sampled from hedgehogs in Belgium [3, 13]. The agent of Siberian tick typhus, *Rickettsia sibirica sibirica*, has been detected in ticks from hedgehogs collected in a suburb of Beijing, China, and this bacterium has been also isolated from the same hedgehog which provided the positive tick sample [4].

In the same study conducted in Xuyi County, southeast China mentioned above, a potential new species, *Candidatus Rickettsia xuyiensis* XY-2 was detected in 8/110

(7,3%) *Ha. flava* ticks collected from *E. amurensis* hedgehogs [25]. The hedgehog from which the positive ticks were collected tested also positive [25].

A potential new uncultivated *Rickettsia* species of the spotted fever group has been reported in 11.25% of *Rh. sanguineus* s.l. and in 77% of *Ha. erinacei* ticks sampled from *A. algirus* and *Paraechinus aethiopicus* hedgehogs in north-east Algeria [4], and another potential new uncultivated *Rickettsia* species close to the previous one has been detected in two *Ha. erinacei* ticks collected from *A. algirus* hedgehogs in the Bouira province in Algeria [21].

Rickettsia species have already been detected in hedgehogs, which suggests that they are suitable reservoir hosts. However, their precise role in the epidemiological cycle of these rickettsial agents remain poorly known [4, 13].

***Leptospira* spp.**

Leptospira spp. are obligate aerobic spirochete bacteria belonging to the Leptospiraceae family and are the agent of leptospirosis, a widespread zoonotic disease. Humans can become infected after direct contact with infected urine or indirectly through a contaminated environment [3, 53].

Leptospira spp. was detected in two *Rh. sanguineus* s.l. ticks collected from *A. algirus* hedgehogs for the first time in Algeria. However, sequencing did not allow the identification of the species [21].

Several studies have reported *Leptospira* species in hedgehog kidneys worldwide [3]. *Leptospira interrogans* and *L. borgpetersenii* have been reported in wild hedgehogs in France, *L. interrogans* has been reported in wild *E. amurensis* (Amur hedgehogs) from China and in *A. algirus* hedgehogs from the Bouira Province, Algeria [3, 21] and *L. ballum* has also been isolated from hedgehogs [3]. These animals are strongly suspected to play a role in the epidemiology of this zoonotic disease and its transmission to humans [3]. The involvement of ticks in the cycle should be further investigated to find out whether the specimens in the study by Aouadi et al. mentioned above were positive simply because they had ingested blood from an infected host or whether they have real vectorial capacity and competence for *Leptospira* species.

All of the microorganisms reported in the different tick species sampled from hedgehogs are summarized in Fig. 3 and in Table 1.

Microorganisms detected in fleas collected from hedgehogs

Rickettsia

Rickettsia felis

Rickettsia felis, the agent of flea-borne spotted fever rickettsiosis, is an emerging Gram-negative obligate

intracellular bacterium reported worldwide which affects humans [54]. Although fleas, mainly cat fleas *Ctenocephalides felis*, are the recognized vectors, this bacterium has been also reported in other flea and arthropod species, such as booklice [36, 54, 55]. In addition, mosquitos have been reported to be a competent vector under experimental conditions [56] (Fig. 4)

Rickettsia felis have been detected in *A. erinacei* fleas collected from different hedgehog species, including *A. algirus* hedgehogs in a western region of Algeria (the Oran district) and in a north-central region of Algeria (the Bouira province) [21, 57]; in 33.3% of the fleas sampled from *E. europaeus* hedgehogs from Spain [32]; and with a high infection rate (99.2% and 95.5%) in *A. erinacei* fleas collected, respectively, from *E. europaeus* hedgehogs captured in the city center of Marseille in France, and in *A. algirus* and *P. aethiopicus* hedgehogs in Algeria [4, 16]. Furthermore, *R. felis* has been detected in *A. erinacei maura* collected from *E. europaeus* in Mértola, a southern region of mainland Portugal [35] and in *A. erinacei* and *C. felis* fleas collected from *A. algirus* hedgehogs in Chebat El Balout (Ouled Driss, Souk Ahras) in Algeria [17]. *Rickettsia felis* transovarial transmission has been reported in fleas, suggesting that they might be potentially reservoirs. Nevertheless, the role of hedgehogs in the epidemiological cycle remains unclear, and further investigations are necessary [57].

***Rickettsia felis*-like organisms**

A very high infection rate of a *R. felis*-like organism has been detected in *A. erinacei* fleas collected from hedgehogs in Bavaria (Germany). The authors concluded that this may be a strain variant of *R. felis* as the sequences are closely related to it and it appears to be associated with hedgehogs and their fleas [58]. Moreover, *Rickettsia asemonensis* DNA was identified in 55/117 (47%) *A. erinacei* fleas sampled from European hedgehogs (*E. europaeus*) captured in seven districts of north and center Portugal [13]. Further investigations are needed to determine the involvement of hedgehogs and their fleas in the epidemiology and circulation of *R. felis*-like organisms. Although a *R. felis*-like organism has previously been reported in a human (blood sample from a Malaysian patient), the pathogenicity of these bacteria for humans and animals remains to be determined [13, 58].

Rickettsia typhi

Rickettsia typhi, the etiological agent of murine typhus, has been detected in one *A. erinacei* flea sampled from an *E. europaeus* hedgehog in Spain [32]. This bacterium is usually associated with rodents and *Xenopsylla* fleas, which are considered to be the main vectors [32]. Further investigations and experimental models should be

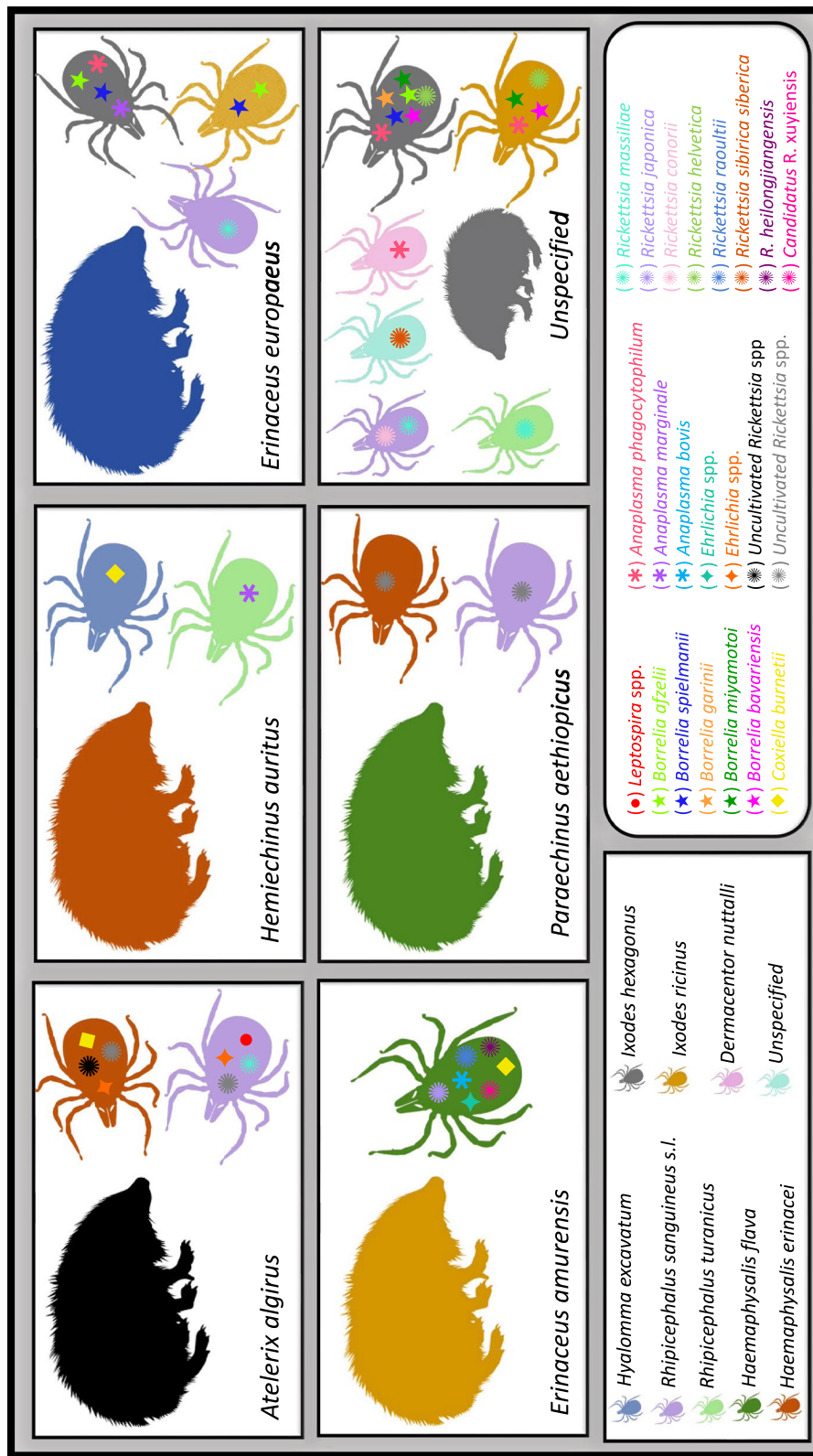


Fig. 3 Microorganisms detected in different tick species sampled from hedgehogs worldwide

Table 1 Summary table of microorganisms detected in ticks and fleas collected from different hedgehog species worldwide

Microorganism	Arthropod	Arthropod species	hedgehog host species	Location	References	
<i>Coxiella burnetii</i>	Tick	<i>Hyalomma excavatum</i>	<i>Hemiechinus auritus</i>	Alexandria, Egypt	[28]	
		<i>Haemaphysalis flava</i>	<i>Erinaceus amurensis</i>	Xianning City, Hubei Province, central China	[20]	
		<i>Haemaphysalis erinacei</i>	<i>Atelerix algirus</i>	Bouira province, northern Algeria	[21]	
	Flea	<i>Archaeopsylla erinacei</i>	<i>Atelerix algirus</i>	Bouira province, northern Algeria	[21]	
<i>Borrelia afzelii</i>	Tick	<i>Ixodes hexagonus</i>	<i>Erinaceus europaeus</i>	Karlsruhe, Germany	[41]	
			Unspecified	Germany	[42]	
<i>Borrelia spielmanii</i>	Tick	<i>Ixodes ricinus</i>	<i>Erinaceus europaeus</i>	Karlsruhe, Germany	[41]	
			<i>Ixodes hexagonus</i>	<i>Erinaceus europaeus</i>	Karlsruhe, Germany	[41]
<i>Borrelia garinii</i>	Tick	<i>Ixodes hexagonus</i>	Unspecified	Germany	[42]	
			<i>Ixodes ricinus</i>	<i>Erinaceus europaeus</i>	Karlsruhe, Germany	[41]
<i>Borrelia miyamotoi</i>	Tick	<i>Ixodes hexagonus</i>	Unspecified	-	[3]	
<i>Borrelia bavariensis</i>	Tick	<i>Ixodes hexagonus</i>	Unspecified	-	[3]	
			<i>Ixodes ricinus</i>	Unspecified	-	[3]
<i>Anaplasma phagocytophilum</i>	Tick	<i>Ixodes hexagonus</i>	<i>Erinaceus europaeus</i>	Germany	[12]	
			unspecified	Netherlands	[31]	
			Unspecified	Baden-Württemberg, Germany	[31]	
			Unspecified	Belgium	[3]	
			<i>Ixodes ricinus</i>	Unspecified	Netherlands	[31]
			Unspecified	Baden-Württemberg, Germany	[31]	
			Unspecified	Belgium	[3]	
<i>Anaplasma marginale</i>	Tick	<i>Dermacentor nuttalli</i>	Unspecified	Hebei Province, China	[46]	
			<i>Rhipicephalus turanicus</i>	<i>Hemiechinus auritus</i>	Zabol, south-eastern Iran	[26]
			<i>Ixodes hexagonus</i>	<i>Erinaceus europaeus</i>	Netherlands	[26]
<i>Anaplasma bovis</i>	Tick	<i>Haemaphysalis flava</i>	<i>Erinaceus amurensis</i>	Xianning City, Hubei Province, central China	[20]	
<i>Ehrlichia</i> spp. (phylogenetically close to <i>Ehrlichia ewingii</i>)	Tick	<i>Haemaphysalis flava</i>	<i>Erinaceus amurensis</i>	Xianning City, Hubei Province, central China	[20]	
<i>Ehrlichia</i> spp.	Tick	<i>Rhipicephalus sanguineus sensu lato</i>	<i>Atelerix algirus</i>	Bouira province, northern Algeria	[21]	
			<i>Haemaphysalis erinacei</i>	<i>Atelerix algirus</i>	Bouira province, northern Algeria	[21]
<i>Wolbachia</i> spp.	Flea	<i>Archaeopsylla erinacei</i>	<i>Atelerix algirus</i>	Bouira province, northern Algeria	[21]	
			<i>Erinaceus europaeus</i>	Spain	[32]	
<i>Rickettsia massiliae</i>	Tick	<i>Rhipicephalus sanguineus sensu lato</i>	Unspecified	north-east Algeria	[4]	
			Unspecified	Algiers	[51]	
			<i>Atelerix algirus</i>	Bouira province, northern Algeria	[21]	
			<i>Erinaceus europaeus</i>	Marseille, France	[16]	
<i>Rickettsia conorii</i>	Tick	<i>Rhipicephalus sanguineus sensu lato</i>	<i>Erinaceus europaeus</i>	north and centre of Portugal	[13]	
			Unspecified	Algiers	[51]	
			Unspecified	Algiers	[51]	
<i>Rickettsia japonica</i>	Tick	<i>Haemaphysalis flava</i>	<i>Erinaceus amurensis</i>	Xianning City, Hubei Province, central China	[20]	
<i>Rickettsia raoultii</i>	Tick	<i>Haemaphysalis flava</i>	<i>Erinaceus amurensis</i>	Xianning City, Hubei Province, central China	[20]	

Table 1 (continued)

Microorganism	Arthropod	Arthropod species	hedgehog host species	Location	References	
<i>Rickettsia helvetica</i>	Tick	<i>Ixodes hexagonus</i>	Unspecified	Belgium	[3]	
		<i>Ixodes ricinus</i>	Unspecified	Belgium	[3]	
<i>Rickettsia sibirica sibirica</i>	Tick	unspecified	Unspecified	suburb of Beijing	[4]	
<i>Rickettsia heilongjiangensis</i>	Tick	<i>Haemaphysalis flava</i>	<i>Erinaceus amurensis</i>	Xuyi County, Southeast China	[25]	
<i>Rickettsia felis</i>	Flea	<i>Archaeopsylla erinacei</i>	<i>Atelerix algirus</i>	Oran district, western Algeria	[51]	
				Bouira province, northern Algeria	[21]	
				north-east Algeria	[4]	
			<i>Erinaceus europaeus</i>	Ouled Driss, Souk Ahras, Algeria	[17]	
				Spain	[32]	
				Marseille, France	[16]	
				North-east Algeria	[4]	
<i>Paraechinus aethiopicus</i>	<i>Archaeopsylla erinacei maura</i>	<i>Erinaceus europaeus</i>	Mértola, southern mainland Portugal	[35]		
			Ouled Driss, Souk Ahras, Algeria	[17]		
<i>Rickettsia felis</i> -like organisms	Flea	<i>Archaeopsylla erinacei</i>	Unspecified	Bavaria, Germany	[58]	
<i>Rickettsia asemonensis</i>	Flea	<i>Archaeopsylla erinacei</i>	<i>Erinaceus europaeus</i>	North and center Portugal	[13]	
<i>Rickettsia typhi</i>	Flea	<i>Archaeopsylla erinacei</i>	<i>Erinaceus europaeus</i>	Spain	[32]	
<i>Candidatus Rickettsia xuyiensis</i>	Tick	<i>Haemaphysalis flava</i>	<i>Erinaceus amurensis</i>	Xuyi County, Southeast China	[25]	
Uncultivated <i>Rickettsia</i> spp.	Tick	<i>Rhipicephalus sanguineus sensu lato</i>	<i>Atelerix algirus</i>	North-east Algeria	[4]	
				<i>Paraechinus aethiopicus</i>	North-east Algeria	[4]
			<i>Haemaphysalis erinacei</i>	<i>Atelerix algirus</i>	North-east Algeria	[4]
					<i>Paraechinus aethiopicus</i>	North-east Algeria
Uncultivated <i>Rickettsia</i> spp.	Tick	<i>Haemaphysalis erinacei</i>	<i>Atelerix algirus</i>	Bouira province, northern Algeria	[21]	
<i>Leptospira</i> spp.	Tick	<i>Rhipicephalus sanguineus sensu lato</i>	<i>Atelerix algirus</i>	Bouira province, northern Algeria	[21]	
<i>Bartonella henselae</i>	Flea	<i>Archaeopsylla erinacei</i>	<i>Erinaceus roumanicus</i>	Budapest, Hungary	[61]	
<i>Bartonella elizabethae</i>	Flea	<i>Archaeopsylla erinacei</i>	Unspecified	Western Algeria	[60]	
<i>Bartonella clarridgeiae</i>	Flea	<i>Archaeopsylla erinacei</i>	Unspecified	Western Algeria	[60]	
Uncultivated <i>Bartonella</i> spp. (phylogenetically close to <i>B. clarridgeiae</i>)	Flea	<i>Archaeopsylla erinacei</i>	<i>Atelerix algirus</i>	Bouira province, northern Algeria	[21]	
<i>Mycobacterium</i> spp.	Flea	<i>Archaeopsylla erinacei</i>	<i>Erinaceus europaeus</i>	Seville, Spain	[32]	

performed to find out whether this flea species could be a suitable reservoir and vector for *R. typhi*.

Bartonella spp.

Bartonella are emerging Gram-negative fastidious oxidase-negative bacteria [59, 60]. Eleven species have been reported to be involved in various human diseases worldwide [59]. They are associated with different arthropod species, mainly fleas [35]. Transmission usually occurs through infected feces [60].

Bartonella henselae, the etiological agent of cat scratch disease in humans, has been detected in an *A. erinacei* flea collected on northern white-breasted hedgehogs (*E. roumanicus*) in Budapest, Hungary [61]. In addition, *Bartonella elizabethae* and *Bartonella clarridgeiae* have been detected in *A. erinacei* fleas collected from hedgehogs in

Algeria (Ouled Ben Aouali and Tafraoui districts) [60]. A potential new uncultivated *Bartonella* spp. which is phylogenetically close to *B. clarridgeiae* has also been detected in the same flea species sampled from *A. algirus* hedgehogs in Algeria (Bouira province) [21].

Different *Bartonella* species have also been reported in hedgehogs. *Atelerix algirus* hedgehog spleens from Algeria have tested positive for *B. elizabethae* and *B. tribocorum* [59, 60]. *Bartonella washoensis* and *B. melophagi* have been reported in *E. europaeus* hedgehogs in urban and suburban areas of the Czech Republic [62]. Furthermore, several potential new *Bartonella* species have been reported, including clones which are phylogenetically close to *B. taylorii*, *B. clarridgeiae*, and *B. rochalimae* detected in the northern white-breasted hedgehog (*E. roumanicus*), the European hedgehog (*E. europaeus*), the

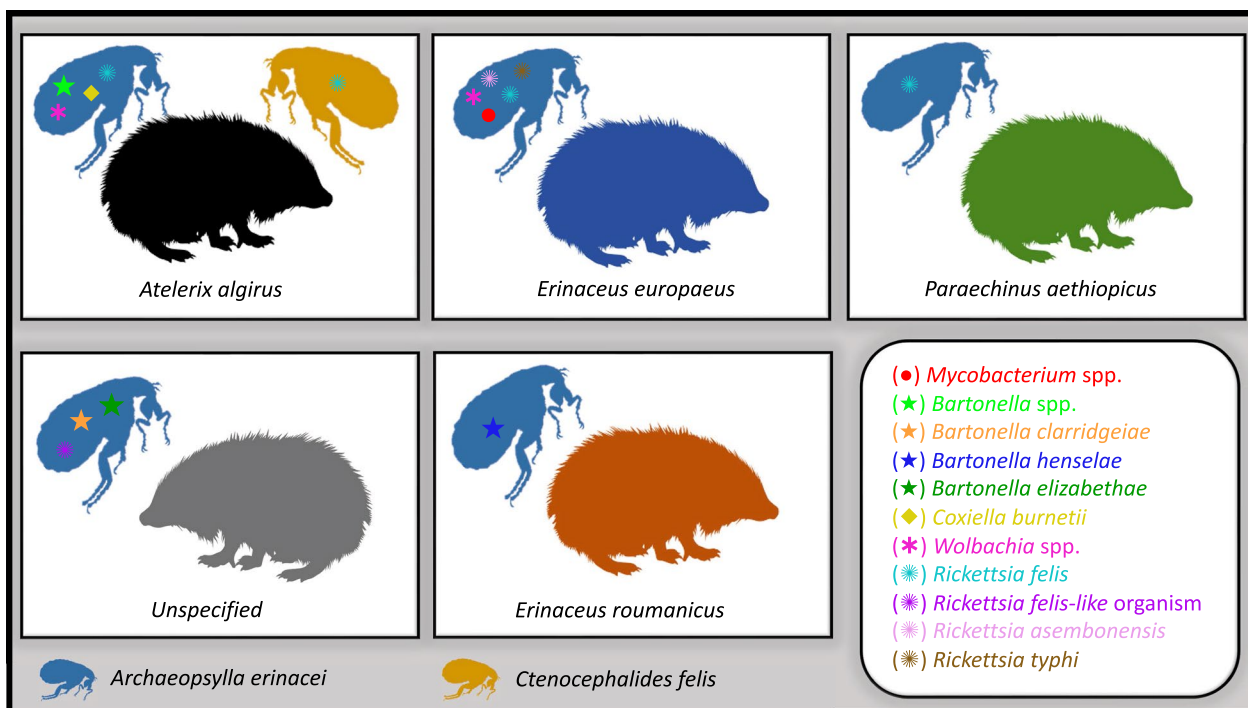


Fig. 4 Microorganisms detected in different flea species sampled from hedgehogs worldwide

southern white-breasted hedgehog (*E. concolor*) and the north African hedgehog (*A. algirus*) from different countries [21, 62–65]. The aforementioned studies highlight that *Bartonella* species have been reported in hedgehogs as well as in the fleas collected from them, suggesting that hedgehogs maybe potential reservoirs. The role of fleas as vectors of these different *Bartonella* species is deserving of further investigation through experimental models.

***Wolbachia* spp.**

Wolbachia are endosymbiotic bacteria of the Anaplasmataceae family which have been associated with various arthropod species [66]. *Wolbachia* spp. has been detected in *A. erinacei* fleas sampled from *A. algirus* and *E. europaeus* hedgehogs from Algeria and Spain, respectively [21, 32]. These bacteria are known to be transmitted through transovarian and transstadial routes in arthropod populations [67]. However, they have no direct impact on human and animal health.

***Mycobacterium* spp.**

Mycobacteria are Gram-positive bacilli bacteria. The diseases and symptoms that they cause in humans and animals depend on the bacterial species involved [3]. Only one published study has reported *Mycobacterium* spp. in 1/18 (5.5%) of *A. erinacei* fleas sampled from *E. europaeus* hedgehogs in Seville, Spain, but sequencing

failed to identify this bacteria to the species level [32]. Different mycobacterial species, such as the causative agent of paratuberculosis in ruminants (Johne's disease) *Mycobacterium avium paratuberculosis*, the agent of bovine tuberculosis *M. bovis* and *M. avium*, have been already reported in hedgehogs [3]. Two other studies reported *M. marinum* in both an African pygmy hedgehog (*A. albiventris*) in Japan and a European hedgehog [3]. These animals are, therefore, strongly suspected to be involved in the propagation of these bacteria. However, the involvement of fleas is still unknown, and further epidemiological investigations, together with experimental models, could help reveal whether the fleas have a vectorial capacity and competence, or whether the *A. erinacei* fleas in the study of Zurita et al. were positive only because they had ingested infected blood [3].

Coxiella burnetii

As described above, *C. burnetii* is a pathogenic agent distributed worldwide [39]. Only one study has reported the presence of this bacterium in four *A. erinacei* fleas sampled from *A. algirus* hedgehogs in the Bouira province in Algeria [21]. Organs as well as ticks sampled from the same hedgehogs that provided the fleas also tested positive for this bacterium [21]. Fleas are unusual hosts for *C. burnetii*, and it can be assumed that they acquired this pathogen by co-feeding with

infected ticks or by ingesting blood from an infected host [21]. Further studies are needed to determine the role of fleas in the transmission cycle of Q fever.

All of the microorganisms reported in the different flea species sampled from hedgehogs are summarized in Fig. 4 and in Table 1.

Conclusions

This review provides information available in the literature on microorganisms detected in arthropods sampled from hedgehogs worldwide. Several microorganisms have been reported in ticks and fleas collected from these animals, including pathogenic agents which are responsible for zoonosis in animals and humans. The detection of these microorganisms in arthropods does not necessarily mean that they are vectors and that they can transmit the microorganisms to humans and animals. While the vector capacity and competence of fleas and ticks for some of these microorganisms has been proven, for others, such as *Leptospira* spp. and *Mycobacterium* spp., their detection in ticks and fleas must be viewed with some caution. They may simply have been ingested together with blood taken from an infected host. For this reason, further field epidemiological investigations, together with experimental models, are needed to clarify whether or not arthropods play a role in the maintenance and transmission of these bacteria. Until such evidence is forthcoming, these microorganisms detected in arthropods, in particular pathogenic ones, should nevertheless be added to the repertoire of microorganisms to be considered in the context of the epidemiological surveillance and diagnostic of hedgehog zoonoses.

In the present state of our knowledge, we have only a blurred picture of the role of hedgehogs as a reservoir host. Epidemiological studies on hedgehogs are difficult to conduct since they are protected animals and handling them is, therefore, highly regulated. Accordingly, their ectoparasites represent a very interesting source of information about the microorganisms circulating in these animal populations, especially vector-borne ones, without endangering them. Hedgehogs should also be dewormed to limit the risk of transmission of these pathogens to humans and other animals.

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Author contributions

LB did the bibliography, wrote, and edited the original manuscript, constructed the figures and participated in taking the arthropod photographs. PP and AZD supervised, validated, reviewed, and edited the manuscript. JMB and

JMB took hedgehog pictures, reviewed and edited the manuscript. All authors reviewed and approved the final version of the manuscript.

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Data availability

All data regarding this study are included in the manuscript.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

- Symonds MRE. Phylogeny and life histories of the 'Insectivora': controversies and consequences. *Biol Rev.* 2005;80:93–128. <https://doi.org/10.1017/S1464793104006566>.
- Riley PY, Chomel BB. Hedgehog zoonoses. *Emerg Infect Dis.* 2005;11:1–5. <https://doi.org/10.3201/eid1101.040752>.
- Ruszkowski JJ, Hetman M, Turlewicz-podbielska H, Pomorska-mol M. Hedgehogs as a potential source of zoonotic pathogens—a review and an update of knowledge. *Animals.* 2021;11:1754. <https://doi.org/10.3390/ani11061754>.
- Khalidi M, Socolovschi C, Benyettou M, Barech G, Biche M, Kernif T, et al. *Rickettsiae* in arthropods collected from the North African hedgehog (*Atelerix algirus*) and the desert hedgehog (*Paraechinus aethiopicus*) in Algeria. *Comp Immunol Microbiol Infect Dis.* 2012;35:117–22. <https://doi.org/10.1016/j.cimid.2011.11.007>.
- Webster WM. Susceptibility of the hedgehog (*Erinaceus europaeus*) to infection with *Leptospira pomona*. *Nature.* 1957;80:1372. <https://doi.org/10.1038/1801372a0>.
- Carlier EW. Contributions to the histology of the hedgehog (*Erinaceus europaeus*)—part 1 the alimentary canal. *J Anat Physiol.* 1892;27:84–111.
- Carlier EW, Evans CAL. A chemical study of the hibernating gland of the hedgehog, together with the changes which it undergoes during winter sleep. *J Anat Physiol.* 1903;38:15–31.
- Ranson RM. New laboratory animals from wild species: breeding a laboratory stock of hedgehogs (*Erinaceus europaeus* L.). *J Hyg (Lond).* 1941;41(2):131–8. <https://doi.org/10.1017/S0022172400012365>.
- Lapierre J, Rousset JJ, Picot M. Sensitivity of the hedgehog (*Erinaceus europaeus*) to different species of *Borrelia*. *C R Seances Soc Biol Fil.* 1959;153:1718–20.
- Giroud P, Le Gac P, Dumas N, Colas-Belcour J. The hedgehog *Erinaceus europaeus*, reservoir or *Rickettsia* or of neo-rickettsia and even of agents of the ovine abortion group. *Bull Soc Pathol Exot Fil.* 1959;52:726–30.
- Lapierre J, Rousset JJ, Picot H. Receptivity of the hedgehog (*Erinaceus europaeus*) to *Trypanosoma gambiense*. *Ann Parasitol Hum Comp.* 1960;35:188–9. <https://www.parasite-journal.org/articles/parasite/pdf/1960/01/parasite1960351p188.pdf>
- Skuballa J, Petney T, Pfaffle M, Taraschewski H. Molecular detection of *Anaplasma phagocytophilum* in the European hedgehog (*Erinaceus europaeus*) and its ticks. *Vector Borne Zoonotic Dis.* 2010;10:1055–7.
- Barradas PF, Mesquita JR, Mateus TL, Ferreira P, Amorim I, Gärtner F, et al. Molecular detection of *Rickettsia* spp in ticks and fleas collected from

- rescued hedgehogs (*Erinaceus europaeus*). *Exp Appl Acarol.* 2021;83:449–60. <https://doi.org/10.1007/s10493-021-00600-y>.
14. Tomas-Pérez M, Khaldi M, Riera C, Mozo-Leon D, Ribas A, Hide M, et al. First report of natural infection in hedgehogs with *Leishmania major*, a possible reservoir of zoonotic cutaneous leishmaniasis in Algeria. *Acta Trop.* 2014;135:44–9. <https://doi.org/10.1016/j.actatropica.2014.03.018>.
 15. Földvári G, Jahfari S, Rigó K, Jablonszky M, Szekeres S, Majoros G, et al. *Candidatus neoehrlichia mikurensis* and *Anaplasma phagocytophilum* in urban hedgehogs. *Emerg Infect Dis.* 2014;20:496–8. <https://doi.org/10.3201/eid2003.130935>.
 16. Marié J, Davoust B, Socolovschi C, Raoult D, Parola P. Molecular detection of rickettsial agents in ticks and fleas collected from a european hedgehog (*Erinaceus europaeus*) in Marseilles, France. *Comp Immunol Microbiol Infect Dis.* 2012;35:77–9. <https://doi.org/10.1016/j.cimid.2011.11.005>.
 17. Leulmi H, Aouadi A, Bitam I, Bessas A, Benakhlia A, Raoult D, et al. Detection of *Bartonella taylorae*, *Coxiella burnetii* and rickettsiae in arthropods and tissues from wild and domestic animals in northeastern Algeria. *Parasit Vectors.* 2016;9:27. <https://doi.org/10.1186/s13071-016-1316-9>.
 18. Földvári G, Rigó K, Jablonszky M, Biró N, Majoros G. Ticks and the city: ectoparasites of the Northern white-breasted hedgehog (*Erinaceus roumanicus*) in an urban park. *Ticks Tick Borne Dis.* 2011;2:231–4. <https://doi.org/10.1016/j.ttbdis.2011.09.001>.
 19. Alghamdi SQ, Low VL, Alkathiry HA, Alagaili AN, McGarry JW, Makepeace BL. Automatic barcode gap discovery reveals diverse clades of *Rhipicephalus* spp. and *Haemaphysalis* spp. ticks from small mammals. *Parasit Vectors.* 2021;14:541. <https://doi.org/10.1186/s13071-021-05049-x>.
 20. Fang L, Lei S, Yan Z, Xiao X, Liu J, Gong X, et al. Detection of multiple intracellular bacterial pathogens in *Haemaphysalis flava* ticks collected from hedgehogs in Central China. *Pathogens.* 2021;10:115. <https://doi.org/10.3390/pathogens10020115>.
 21. Aouadi N, Benkacimi L, Diarra AZ, Laroche M, Bérenger J-M, Bitam I, et al. Microorganisms associated with the North African hedgehog *Atelerix algirus* and its parasitizing arthropods in Algeria. *Comp Immunol Microbiol Infect Dis.* 2022;80:1–18. <https://doi.org/10.1016/j.cimid.2021.101726>.
 22. Apanaskevich DA, Schuster AL, Horak IG. The genus *Hyalomma*. VII. Redescription of all parasitic stages of *H. (euhyalomma) dromedarii* and *H. (E) schulzei* (Acari: Ixodidae). *J Med Entomol.* 2008;45:817–31.
 23. Apanaskevich DA, Horak IG. The genus *Hyalomma*. XI. Redescription of all parasitic stages of *H. (euhyalomma) asiaticum* (Acari: Ixodidae) and notes on its biology. *Exp Appl Acarol.* 2010;52(2):207–20. <https://doi.org/10.1007/s10493-010-9361-0>.
 24. Horak IG, Welman S, Hallam SL, Lutermann H, Mzilikazi N. Ticks of four-toed elephant shrews and Southern African hedgehogs. *Onderstepoort J Vet Res.* 2011;78:243. <https://doi.org/10.4102/ojvr.v78i1.243>.
 25. Zhu C, Ai L, Qi Y, Liu Y, Li H, Ye F, et al. Molecular detection of spotted fever group *Rickettsiae* in hedgehogs (*Erinaceus amurensis*) and hedgehog—attached ticks in Xuyi County Southeast China. *Exp Appl Acarol.* 2022;88:97–111. <https://doi.org/10.1007/s10493-022-00721-y>.
 26. Khodadadi N, Nabavi R, Sarani A, Saadati D, Ganjali M, Mihalca AD, et al. Identification of *Anaplasma marginale* in long-eared hedgehogs (*Hemiechinus auritus*) and their *Rhipicephalus turanicus* ticks in Iran. *Ticks Tick Borne Dis.* 2021;12:101641. <https://doi.org/10.1016/j.ttbdis.2020.101641>.
 27. Pastiu AI, Matei IA, Mihalca AD, D'Amico G, Dumitrache MO, Kalmár Z, et al. Zoonotic pathogens associated with *Hyalomma aegyptium* in endangered tortoises: evidence for host-switching behaviour in ticks? *Parasit Vectors.* 2012;5:301. <https://doi.org/10.1186/1756-3305-5-301>.
 28. Loftis AD, Reeves WK, Szumlas DE, Abbassy MM, Helmy IM, Moriarity JR, et al. Rickettsial agents in Egyptian ticks collected from domestic animals. *Exp Appl Acarol.* 2006;40:67–81. <https://doi.org/10.1007/s10493-006-9025-2>.
 29. Zhou Q, Li J, Guo X, Liu J, Song Q, Gong X, et al. Records of three mammal tick species parasitizing an atypical host, the multi-ocellated racerunner lizard, in arid regions of Xinjiang China. *Parasit Vectors.* 14:2021;14:135. <https://doi.org/10.1186/s13071-021-04639-z>.
 30. Sang C, Yang M, Xu B, Liu G, Yang Y, Kairullayev K, et al. Tick distribution and detection of *Babesia* and *Theileria* species in Eastern and Southern Kazakhstan. *Ticks Tick Borne Dis.* 2021;12:101817. <https://doi.org/10.1016/j.ttbdis.2021.101817>.
 31. Silaghi C, Skuballa J, Thiel C, Pfister K, Petney T, Pfäffle M, et al. The European hedgehog (*Erinaceus europaeus*)—a suitable reservoir for variants of *Anaplasma phagocytophilum*? *Ticks Tick Borne Dis.* 2012;3:49–54. <https://doi.org/10.1016/j.ttbdis.2011.11.005>.
 32. Zurita A, Benkacimi L, El Karkouri K, Cutillas C, Parola P, Laroche M. New records of bacteria in different species of fleas from France and Spain. *Comp Immunol Microbiol Infect Dis.* 2021;76:1–9. <https://doi.org/10.1016/j.cimid.2021.101648>.
 33. Goz Y, Yilmaz AB, Aydın A, Dicle Y. Ticks and fleas infestation on east hedgehogs (*Erinaceus concolor*) in Van Province, Eastern Region of Turkey. *J Arthropod Borne Dis.* 2016;10:50–4.
 34. Nematollahi A, Helan JA, Golezardiy H, Zaboli N, Nouruzi M, Azari M. Parasitic fauna of East European hedgehog (*Erinaceus concolor*) and their pathological aspects in Iran. *Adv Zool Botany.* 2014;2:1–5. <https://doi.org/10.13189/azb.2014.020101>.
 35. De Sousa R, Fournier P-E, Santos-Silva M, Amaro F, Bacellar F, Raoult D. Molecular detection of *Rickettsia felis*, *Rickettsia typhi* and two genotypes closely related to *Bartonella elizabethae*. *Am J Trop Med Hyg.* 2006;75:727–31.
 36. Parola P, Davoust B, Raoult D. Tick- and flea-borne rickettsial emerging zoonoses. *Vet Res.* 2005;36:469–92. <https://doi.org/10.1051/vetres>.
 37. Greigert V, Brunet J, Ouarti B, Laroche M, Pfaff AW, Henon N, et al. The trick of the hedgehog: case report and short review about *Archaeopsylla erinacei* (Siphonaptera : Pulicidae) in human health. *J Med Entomol.* 2020;57:318–23. <https://doi.org/10.1093/jme/tjz157>.
 38. Pfäffle M, Petney T, Elgas M, Skuballa J, Taraschewski H. Tick-induced blood loss leads to regenerative anaemia in the European hedgehog (*Erinaceus europaeus*). *Parasitology.* 2009;136:443–52. <https://doi.org/10.1017/S0031182009005514>.
 39. Devaux CA, Omar OI, Million M, Raoult D. *Coxiella burnetii* in dromedary camels (*Camelus dromedarius*): a possible threat for humans and livestock in North Africa and the Near and Middle East? *Front Vet Sci.* 2020;7:558481. <https://doi.org/10.3389/fvets.2020.558481>.
 40. Široký P, Kubelová M, Modrý D, Erhart J, Literák I, Eva Š, et al. Tortoise tick *Hyalomma aegyptium* as long term carrier of q fever agent *Coxiella burnetii*—evidence from experimental infection. *Parasitol Res.* 2010;107:1515–20. <https://doi.org/10.1007/s00436-010-2037-1>.
 41. Skuballa J, Oehme R, Hartelt K, Petney T, Bücher T, Kimmig P, et al. European hedgehogs as hosts for *Borrelia* spp Germany. *Emerg Inf Dis.* 2007;3:952–3. <https://doi.org/10.3201/eid1306.070224>.
 42. Strube C, Montenegro VM, Epe C, Eckelt E, Schnieder T. Establishment of a minor groove binder-probe based quantitative real time PCR to detect *Borrelia burgdorferi* sensu lato and differentiation of *Borrelia spielmanii* by ospA-specific conventional PCR. *Parasit Vectors.* 2010;3:69. <https://doi.org/10.1186/1756-3305-3-69>.
 43. Gern L. Life cycle of *Borrelia burgdorferi* sensu lato and transmission to humans. *Curr Probl Dermatol.* 2009;37:18–30. <https://doi.org/10.1159/000213068>.
 44. Grubhoffer L, Golovchenko M, Vancová M, Zacharovová-slavi K, Rudenko N, Oliver JH Jr. Lyme borreliosis: insights into tick/host-*Borrelia* relations. *Folia Microbiol.* 2005;53:279–94. <https://doi.org/10.14411/fp.2005.039>.
 45. Wolcott KA, Margos G, Fingerle V, Becker NS. Host association of *Borrelia burgdorferi* sensu lato: a review. *Ticks Tick Borne Dis.* 2021;12:101766. <https://doi.org/10.1016/j.ttbdis.2021.101766>.
 46. Yaxue Z, Hongtao J, Qiuyue W, Zhixin F, Hongwei G, Pengpeng L, et al. Molecular detection of *Anaplasma phagocytophilum* in Ixodid ticks in Hebei Province, China. *Vector Borne Zoonotic Dis.* 2011;11:1323–7.
 47. Rodríguez SD, Ortiz MÁG, Ocampo RJ, Murguía CAV. Molecular epidemiology of bovine anaplasmosis with a particular focus in Mexico. *Infect Genet Evol.* 2009;9:1092–101. <https://doi.org/10.1016/j.meegid.2009.09.007>.
 48. André MR. Diversity of *Anaplasma* and *Ehrlichia/Neoehrlichia* agents in terrestrial wild carnivores worldwide: implications for human and domestic animal health and wildlife conservation. *Front Vet Sci.* 2018;5:293. <https://doi.org/10.3389/fvets.2018.00293>.
 49. Park J, Han DG, Ryu JH, Chae JB, Chae JS, Yu DH, et al. Molecular detection of *Anaplasma bovis* in holstein cattle in the Republic of Korea. *Acta Vet Scand.* 2018;60:1–5. <https://doi.org/10.1186/s13028-018-0370-z>.
 50. Parola P, Paddock CD, Socolovschi C, Labruna MB, Mediannikov O, Kernif T. Update on tick-borne rickettsioses around the world: a geographic approach. *Clin Microbiol Rev.* 2013;26:657–702. <https://doi.org/10.1128/CMR.00032-13>.

51. Bitam I, Parola P, Matsumoto K, Rolain JM, Baziz B, Boubidi SC, et al. First molecular detection of *R. conorii*, *R. aeschlimannii*, and *R. massiliae* in ticks from Algeria. *Ann New York Acad Sci.* 2006;1078:368–72. <https://doi.org/10.1196/annals.1374.073>.
52. Matsumoto K, Ogawa M, Brouqui P, Raoult D, Parola P. Transmission of *Rickettsia massiliae* in the tick, *Rhipicephalus turanicus*. *Med Vet Entomol.* 2005;19:263–70. <https://doi.org/10.1111/j.1365-2915.2005.00569.x>.
53. Adler B, de la Moctezuma A. *P. Leptospira* and leptospirosis. *Vet Microbiol.* 2010;140:287–96.
54. Angelakis E, Mediannikov O, Parola P, Raoult D. *Rickettsia felis*: the complex journey of an emergent human pathogen. *Trends Parasitol.* 2016;32:554–64. <https://doi.org/10.1016/j.pt.2016.04.009>.
55. Thepparit C, Sunyakumthorn P, Guillotte ML, Popov VL, Foil LD, Macaluso KR. Isolation of a rickettsial pathogen from a non-hematophagous arthropod. *PLoS ONE.* 2011. <https://doi.org/10.1371/journal.pone.0016396>.
56. Dieme C, Bechah Y, Socolovschi C, Audoly G, Berenger J, Faye O, et al. Transmission potential of *Rickettsia felis* infection by *Anopheles gambiae* mosquitoes. *Proc Natl Acad Sci USA.* 2015;112:8088–93. <https://doi.org/10.1073/pnas.1413835112>.
57. Bitam I, Parola P, De Dittmar La Cruz K, Matsumoto K, Baziz B, Rolain JM, et al. First molecular detection of *Rickettsia felis* in fleas from Algeria. *Am J Tropical Med Hyg.* 2006;74:532–5.
58. Gilles J, Silaghi C, Just FT, Pradel I, Pfister K. Polymerase chain reaction detection of *Rickettsia felis*—like organism in *Archaeopsylla erinacei* (Siphonaptera: Pulicidae) from Bavaria Germany. *J Med Entomol.* 2009;46:703–7. <https://doi.org/10.1603/033.046.0338>.
59. Bitam I, Rolain J-M, Kernif T, Baziz B, Parola P, Raoult D. *Bartonella* species detected in rodents and hedgehogs from Algeria. *Clin Microbiol Infect.* 2008;15:102–3. <https://doi.org/10.1111/j.1469-0691.2008.02180.x>.
60. Bitam I, Rolain JM, Nicolas V, Tsai Y, Parola P, Gundi VAKB, et al. A multi-gene analysis of diversity of *Bartonella* detected in fleas from Algeria. *Comp Immunol Microbiol Infect Dis.* 2012;35:71–6. <https://doi.org/10.1016/j.cimid.2011.11.002>.
61. Hornok S, Földvári G, Rigó K, Meli ML, Tóth M, Molnár V, et al. Vector-borne agents detected in fleas of the northern white-breasted hedgehog. *Vector Borne Zoonotic Dis.* 2014;14:74–6. <https://doi.org/10.1089/vbz.2013.1387>.
62. Majerova K, Gutiérrez R, Fonville M, Hönig V, Papežik P, Hofmannová L, et al. Hedgehogs and squirrels as hosts of zoonotic *Bartonella* species. *Pathogens.* 2021;10:1–16.
63. Szekeres S, van Leeuwen AD, Tóth E, Majoros G, Sprong H, Földvári G. Road-killed mammals provide insight into tick-borne bacterial pathogen communities within urban habitats. *Transbound Emerg Dis.* 2019;2019:277–86. <https://doi.org/10.1111/tbed.13019>.
64. Greco G, Zarea AAK, Sgroi G, Tempesta M, D'Alessio N, Lanave G, et al. Zoonotic *Bartonella* species in Eurasian wolves and other free-ranging wild mammals from Italy. *Zoonoses Public Health.* 2021;68:316–26. <https://doi.org/10.1111/zph.12827>.
65. Marciano O, Gutiérrez R, Morick D, King R, Nachum-Biala Y, Baneth G, et al. Detection of *Bartonella spp.* in wild carnivores, hyraxes, hedgehog and rodents from Israel. *Parasitology.* 2016;143:1232–42. <https://doi.org/10.1017/S0031182016000603>.
66. Pruneau L, Moumène A, Meyer FD, Marcelino I, Lefrançois T, Vachiéry N. Understanding Anaplasmataceae pathogenesis using “Omics” approaches. *Front Cell Infect Microbiol.* 2014;4:86. <https://doi.org/10.3389/fcimb.2014.00086>.
67. Landmann F. The *Wolbachia* endosymbionts. *Microbiol Spectrum.* 2019;7(2). <https://doi.org/10.1128/microbiolspec.BAI-0018-2019>.

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