

# **Spike timing-dependent plasticity and memory**

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#### **Abstract**

 Spike timing-dependent plasticity (STDP) is a bidirectional form of synaptic plasticity discovered about 30 years ago and based on the relative timing of pre- and post-synaptic spiking activity with a millisecond precision. STDP is thought to be involved in the formation of memory but the millisecond-precision spike-timing required for STDP is difficult to reconcile with the much slower timescales of behavioral learning. This review therefore aims to expose and discuss recent findings about i) the multiple STDP learning rules at both excitatory and inhibitory synapses *in vitro*, ii) the contribution of STDP-like synaptic plasticity in the formation of memory *in vivo* and iii) the implementation of STDP rules in artificial neural networks and memristive devices.

#### **Introduction: memory formation, associative learning and synaptic plasticity**

 Memory is usually defined as the ability to use past experience to adapt and optimize the behavior for present and future events. Formation of internal representation in the form of memory take places in several steps; acquisition, encoding and retrieval. The formation of the memory trace (i.e. the engram) and retrieval of memory has been reviewed recently [1] [2]. Encoding of memory is thought to be principally achieved through persistent modifications in both excitatory and inhibitory synaptic transmission [3] [4] in conjunction with modulation of intrinsic neuronal excitability [5]. In associative learning such as Pavlovian conditioning, a form of Hebbian plasticity [6], storage of new information relies on the occurrence of a specific event that can be an automatic behavioral response (salivation) to an unconditioned stimulus (meat in Pavlov experiment). In neurobiological terms, this stimulus is thought to produce a strong response, i.e. a neuronal discharge of action potentials (APs). The temporal association of this stimulus with a neutral stimulus (bell ring) causing initially no behavioral response leads after a few repetitions to the acquisition of a new behavior (salivation) in response to the neutral stimulus (bell ring). The temporal contiguity between the two stimuli is important as the dissociation or the random presentation of the two stimuli does not induce any learning.

 Temporal synchrony of neural events is critical for sensory perception of visual scenes or auditory stimuli [7]. This temporal binding is thought to be mediated by both network properties [8] and axonal sodium channels that read and transmit levels of input synchrony in 81 local brain circuits [9]•. But according to the Hebb law, repeated temporal synchrony between pre- and post-synaptic activity may also lead to reinforcement of excitatory inputs.

 Here we review recent findings devoted to spike timing-dependent synaptic plasticity (STDP) both *in vitro* and *in vivo*. In a first section, this review will establish the definition and history of STDP. Then, the multiplicity of STDP learning rules at excitatory synapses and inhibitory synapses will be discussed. Next, the mechanisms underlying STDP will be 87 dissected and the involvement of STDP-like plasticity in the formation of place field in the hippocampus *in vivo* will be debated. Finally, the implementation of STDP rule in artificial neural networks will be mentioned.

#### **Definition and brief history of STDP**

 STDP is a form of long-term synaptic modification that is induced by the relative order of pre- and post-synaptic spiking activity [10] [11] [12]. Originally derived from classical conditioning [13] and Hebbian synaptic plasticity [14] [15], the concept of STDP emerged during the 1990's with the discovery of the importance of timing in the polarity of long-lasting synaptic changes [16] [17] [18] [19] [20]. Following the Hebbian principle, timing-dependent long-term synaptic potentiation (t-LTP) results from the positive temporal correlation between an excitatory post- synaptic potential (EPSP) followed a few milliseconds later by one or more postsynaptic APs [15] [19] [20]. In contrast, timing-dependent long-term synaptic depression (t-LTD) results from negative temporal correlation between an EPSP preceded by one or more postsynaptic APs [16] [19] [20]. Thus, STDP is classically characterized by an asymmetric learning rule that defines a causality relation [21] (**Figure 1A**). STDP is a general learning rule as it has been reported in insects [22], and mammals including humans [23] [24,25]. It should be noted that the spike number used to induce synaptic plasticity determines the extent of the t-LTP and t- LTD windows, since the larger the number of spikes, the larger the temporal windows [16] [19] (**Figure 1B**). t-LTD requires a higher number of association than t-LTP. In fact, while 3-6 pairings are sufficient to induce t-LTP with positive delays, this number is not sufficient to induce t-LTD [26]. Similar conclusion has been drawn by another group using 20 pairings [27].

 While t-LTP is due to the activation of postsynaptic NMDA receptor (NMDAR) by the back-propagated AP in the dendrite [19] [20], some forms of t-LTD has been shown to involve presynaptic NMDAR and postsynaptic endocannabinoid (eCB) signaling [28] [29]. Experimentally, APs are triggered by a depolarizing step of current, naturally, however, they are produced by a strong excitatory input as in the case of Pavlovian conditioning. Post- synaptic spiking is not required in all the cases. Strong synaptic depolarization has been shown to be more effective than back-propagated action potentials during induction of t-LTP in CA1 neurons [30]. In addition, subthreshold mossy fiber EPSPs are able to provide sufficient depolarization to induce STDP at recurrent excitatory input in CA3 neurons [31]. Dendritic  NMDA spikes initially found in neocortical basal dendrites [32] has been shown to be critical in STDP induction at hippocampal CA3-CA3 synapses [33] [34] and at olfactory bulb inputs in the piriform cortex [35]. In addition, theoretical studies indicate that NMDA spikes promotes cooperativity among synapses and lowers plasticity threshold [36].

#### **Multiple STDP rules at excitatory synapses**

 While early papers reported classical asymmetric STDP curves at synapses formed by pairs of CA3-CA3 pyramidal neurons [19], recent findings indicate a symmetric STDP curve with t- LTP only (i.e. a lack of t-LTD) at recurrent excitatory synapses of CA3 pyramidal neurons [37] (**Figure 1C**). A similar symmetric STDP curve has been observed at Schaffer collateral inputs to CA1 pyramidal neurons in adult but not in young mice [38]●. Importantly, the absence of t- LTD for negative correlation (post- before pre-) in adults involves release from astrocytes since calcium chelation of the astrocyte cytosol prevents t-LTP and restores classical t-LTD.

 Concurrent STDP rules for thalamic and cortical inputs have been observed in layer 2- 3 neurons of the barrel cortex during postnatal development [39]. The emergence of t-LTD is related to the emergence of eCB signaling and may account for the transition from network formation to the emergence of the critical period [40]. In the striatum, the thalamo-striatal inputs that contribute to the initiation of movement, display a conventional asymmetric STDP curve whereas cortico-striatal inputs display an inverted asymmetric curve [41] (**Figure 1C**). In addition, cortico-striatal inputs express different learning rules as a function of the position in the striatum. Symmetric STDP curve in the dorsolateral striatum but a t-LTD-only curve in the dorsomedial striatum [42] (**Figure 1C**). Interestingly, t-LTP depends on eCB signaling in this brain region [43].

 STDP rule depends also on the cooperativity among dendritic spines in CA1 pyramidal neurons [44]. Whereas t-LTP and t-LTD are observed at single dendritic spines, t-LTP but not t-LTD occurs at clustered dendritic spines [45]●. The suppression of t-LTD has been recently reported in the anterior cingulate cortex following induction of neuropathic pain [46]. In this

 case, t-LTD impairment is mediated by the exclusion of GluN2B-containing NMDAR from the postsynaptic site.

#### **STDP in inhibitory circuits**

 STDP at glutamatergic synapses of GABAergic interneurons has been studied. For instance, eCB-dependent t-LTD has also been reported at excitatory synapses on hippocampal GABAergic interneurons [47] [48]. However, no t-LTP is found at excitatory inputs of fast spiking (FS) interneurons [49] except if adrenergic receptors are co-stimulated [50].

 Early studies showed that STDP at inhibitory synapses generally obeys anti-Hebbian rule (i.e., t-LTP for negative correlation and t-LTD for positive correlation, **Figure 2A**) [51]. At hippocampal inhibitory synapses, a symmetrical STDP curve with t-LTP only has been reported, however [52]. This discrepancy among studies results in fact from different developmental stages. In the auditory cortex, the GABAergic synapse established by FS interneurons onto layer 4 glutamatergic neurons undergoes asymmetric anti-Hebbian STDP during the critical period (**Figure 2A**) [53]●●. However, in mature animals the STDP rules becomes symmetric with time-dependent LTP of inhibition (t-LTPi) only (**Figure 2A**). t-LTPi depends of retrograde brain-derived factor (BDNF) signaling whereas time-dependent LTD of 162 inhibition (t-LTDi) involves GABA<sub>B</sub> receptors [53]●●. Interestingly, sound exposure during the 163 critical period converts t-LTDi into t-LTPi while genetic deletion of GABA<sub>B</sub>R prevents auditory critical period plasticity. The asymmetric anti-Hebbian STDP curve for inhibitory synapses during the critical period suggests that coordinated plasticity occurs between excitatory and inhibitory plasticity at least during development (**Figure 2B**). In computational networks, the symmetric plasticity rule with t-LTPi only described at inhibitory synapses of the auditory cortex [54] [53]●● or at glycinergic synapses of the medial superior olive [55] allows the generation of novelty responses [56].

 Inhibitory STDP plays an essential role in the formation and maintenance of functional cortical circuitry as its introduction in modelled recurrent networks provides a homeostatic

mechanism leading to asynchronous irregular network states [57]. In addition, inhibitory STDP

concurs to balance excitatory and inhibitory inputs in a model of feed-forward inhibition [58].

#### **Mechanisms of STDP: mitochondria, potassium channel & calcium levels**

176 Calcium ( $Ca^{2+}$ ) influx is an essential trigger of STDP since most if not all forms of STDP reported so far depends on NMDAR activation, voltage-gated calcium channels or the mobilization of intracellular calcium stores [12].

 Another player recently came into play: dendritic mitochondria. In fact, during period of 180 enhanced neuronal activity, cytosolic  $Ca^{2+}$  elevations propagate into the mitochondrial matrix that subsequently accelerate ATP production. Mitochondrial activity is critical for local protein 182 synthesis in the dendrites [59] and synaptic plasticity [60]. Using a mitochondria-specific Ca<sup>2+</sup> 183 probe, the group of Fleidervish recently showed that mitochondrial  $Ca^{2+}$  is elevated during coincident EPSP and AP, but not during an EPSP or an AP alone [61]●. Importantly, 185 mitochondrial  $Ca^{2+}$  also decodes spike frequency and spike number. This study thus points to the role of mitochondria in STDP induction.

 NMDAR and metabotropic glutamate receptor (mGluR) have been both implicated in synaptic plasticity. While postsynaptic NMDAR is necessary for the induction of t-LTP at many different synapses, the primary role of presynaptic NMDAR for the induction of t-LTD in the cortex [29] and hippocampus [62] has been revealed recently. mGluR acts as a coincidence detector at synapses where NMDAR is not involved in plasticity induction [63]. Hippocampal t- LTD and t-LTP induced by negative correlation at different ages require activation of mGluR5 [38,62]. Voltage-gated channels are also involved in STDP control. In fact, calcium-activated 194 small conductance K<sup>+</sup> (SK) channels shunt calcium influx through NMDARs [64] and mGluR has a negative action on SK channels [65] [66]. Moreover, large conductance calcium-196 activated K<sup>+</sup> (BK) channels activation reduces EPSP amplitude at neocortical synapses with small head volume [67], suggesting their possible involvement in the control of synaptic plasticity [68]. Other potassium channels gate induction of STDP. The adrenergic-gating of t-LTP in hippocampal CA1 neurons is mediated by Kv1 channels [69]. Mild stress down-

 regulates postsynaptic Kv4.2 channels through the PIK3, leading to the up-regulation of t-LTP in neurons from nucleus accumbens [70].

 The external calcium concentration varies in the brain between 1.1 and 1.5 mM [71,72]. However, all the *in vitro* work devoted to STDP uses much higher calcium concentration (i.e. 2.0 to 3.0 mM). The large variability of STDP curves reviewed here can be explained by fluctuations in postsynaptic calcium as showed theoretically by Graupner and Brunel [73]. Based on this theoretical work, Inglebert & co-workers showed indeed that physiological calcium has a profound impact on STDP curve [74]●●. While in high external calcium (3 mM), a conventional asymmetric curve is reported, in 1.8 mM only t-LTD is observed and in 1.3 mM, no synaptic plasticity is found (**Figure 3A**). In physiological calcium (1.3 mM), plasticity could 210 be restored by increasing the frequency of stimulation (t-LTP) or increasing the number of postsynaptic spikes used during induction (t-LTD) [74]●●. Similarly, in the neocortex, physiological calcium concentration reduces or abolishes t-LTP and t-LTD [75]●. Both plasticity can, however, be restored with higher frequency stimulation.

#### **STDP-based plasticity and memory formation**

 CA1 place fields can emerge from a initially unresponsive neuron when spontaneously occurring dendritic plateau potential is triggered during the exploration [76]●. In this case, a single postsynaptic depolarization triggering multiple spikes is paired with putative presynaptic activity may lead to the reinforcement of excitatory synapses that were initially silent or to the 220 silence of inhibitory inputs that were initially active. More recently, the same group showed that when the dendritic plateau potential occurs a few seconds before the place field-evoked a new place field appears at the detriment of the old place field [77]●● (**Figure 4**). This finding rehabilitates the use of a burst of APs that induces large LTD window found in early studies [16] [17] as well as the use of single dendritic events to induce synaptic plasticity [78] [76]●.

 Dopamine and serotonin are important neuromodulators of synaptic plasticity that are respectively linked to positive or negative reward during learning. In hippocampal and neocortical neurons, dopamine renders the STDP rule symmetric with t-LTP only [79,80] while

 serotonin promotes t-LTD only [81]. A mathematical model shows that the interplay of dopamine and serotonin improves hippocampal learning [82]. Similarly, noradrenaline facilitates induction of t-LTP in the visual cortex [81], and stimulation of noradrenergic or serotoninergic system during visual stimulation respectively promotes reinforcement or depression of visual cortical responses and can rescue deficits induced by monocular deprivation [83,84]●. A fundamental problem in neuroscience is how the brain associates and encodes events that are separated in time. Postsynaptic burst reactivation of hippocampal neurons during dopaminergic stimulation has recently shown to enable associative plasticity of temporally dis-contiguous inputs [85].

 Memory is particularly altered in Alzheimer disease (AD) with a higher predisposition to develop the disease for women. Classical LTP induced by presynaptic high-frequency stimulation is impaired in mouse models of AD (see [86] for recent review). Interestingly, STDP 240 is also altered in murine models of AD. t-LTP is impaired at CA1 pyramidal neurons that are proximal to the amyloid-β plaque but not at CA1 neurons that are distal (>200 µm) [87]. Similarly, t-LTP is altered at excitatory synapses on cortical layer 2/3 (L2/3) pyramidal cells because soluble amyloid-β protein reduces the opening of AMPAR [88]. At L5 cortical neurons, t-LTP is also impaired in a mouse model of AD [89]. STDP-like forms of plasticity induced by transcranial magnetic stimulation are impaired in AD patients [90]. Matrix metalloproteinases 246 (MMP) play a crucial role in the pathogenesis of AD and induction of t-LTP in the barrel cortex requires normal activity of MMP-9 [91]. In addition, the predisposition of females to AD has been confirmed experimentally by showing that in a mouse model of AD, t-LTP shows a faster decay in females compared to males [92].

 STDP-like mechanisms are also involved in neuropsychiatric diseases such as depression induced by chronic stress. The lateral habenula has recently emerged as a key brain region in the pathophysiology of depression. Elevated neuronal activity in the lateral habenula is a sign of depression [93]. Potentiating hypothalamic to lateral habenula inputs by an EPSP-burst pairing leads to lasting changes of emotional status similar to the depression [94]. Moreover, endocannabinoid-dependent t-LTD is reduced following induction of social

 defeat stress in neurons of the nucleus accumbens [95], another brain region involved in reward and depressive disorders. In the same brain region, chronic unpredictable mild stress enhances t-LTP [70], thus indicating that a general increase in synaptic excitation is induced by stress in nucleus accumbens.

#### **Implementation of STDP in artificial neural networks and memristive devices**

 Many synaptic plasticity rules found in natural circuits have been incorporated in artificial neural networks. For instance, incorporation in artificial neural networks of the back-propagating t- LTP found in hippocampal circuits [96], reduces the computational cost without affecting the accuracy of the artificial circuits [97]. This indicates that self-back-propagating plasticity produces coordinated modifications across neuronal layers. Temporally symmetric STDP rule reported in CA3 neurons [37] has been shown to be essential in a network model of area CA3 for proper sequence replay in both forward and backward directions [98].Sequential spiking activity reflecting previous experience is considered as a hallmark of learning. However, the way cortical circuits avoid the opposite problem, i.e. producing spurious sequences that are not relevant to learning, was not clearly established. This problem has been theoretically and experimentally solved recently. In fact, STDP in cortical circuits containing excitatory and inhibitory neurons eliminates spurious sequences of action potentials [99]. In addition, responses to artificial stimuli measured in the visual cortex of awake ferret initially includes spurious sequences which diminished over extended exposure [99].

 Incorporation of unsupervised STDP learning rule in artificial neural networks has revealed a remarkable ability for object and digit recognition [100●,101]. Moreover, in an artificial saccadic system under the control of a cerebellar-like artificial spiking network incorporating STDP rules at the parallel fiber-Purkinje cell synapse, accuracy and speed of simulated saccadic eye movements are improved through synaptic plasticity [102].

 Increasingly large computing needs at a low energy cost have led to the development of new nanometric memory media based on the properties of brain synapses. Memristors (or memristive devices) are memory resistors, i.e. electrical resistance switches that can retain a

 state of internal resistance based on the recently applied voltage and current. Functionally, memristors are able to mimic synapse function and thus correspond to tunable electronic synapses. Physically, they are based on ionic motion and are made of a layer of metal oxide sandwiched between two metal electrodes [103]. STDP rules have been successfully implemented in memristors [104] [105] [106] [107]. For instance, a network of memristors including STDP rules is able to robustly detect the coincidence by selectively increasing synaptic efficacy of synchronized inputs [104]. Furthermore, memristors are able to capture natural synapses properties as frequency dependent plasticity and temporal integration of plasticity induced by triplets of pre- and post-synaptic spikes [105]. However, memristors are not limited to mimicking real brain synapses, and self-powered, second-order memristors are able to emulate functional properties of visual cortical neurons such as orientation selectivity [108,109]. This research field is extremely dynamic and there are good reasons to believe that the implementation of STDP in artificial synapses will lead to tremendous improvements in computing power at a low energy cost.

#### **Conclusion and perspectives**

 Impressive progresses have been achieved these last 30 years in the understanding of STDP rules at excitatory and inhibitory synapses, in different cortical layers, in various brain regions and during learning. Initially reported *in vitro*, STDP is now clearly demonstrated *in vivo*. However, some questions remain unanswered. First, a large part of our knowledge on the role of STDP in learning and memory comes from modelling studies that present the advantage to be largely predictive but these models are often too simplistic to account for the complexity of the mechanisms occurring in the real brain.

 STDP rules at excitatory inputs described for ages with abnormally elevated external calcium are in fact profoundly altered in physiological extracellular calcium [74]●●. It is possible that the symmetric STDP curve (i.e., LTP only) observed at both CA3 recurrent excitatory synapses [37] and in mature CA3-CA1 synapses [38]● could become asymmetric (i.e., LTD/LTP) in physiological calcium.

 In addition, the STDP rules in physiological calcium are still unknown at inhibitory synapses on pyramidal neurons and on interneurons. In particular, the developmental transition from a t-LTPi/t-LTDi plasticity rule to a uniform t-LTPi rule [53]●● could perhaps not be preserved in physiological calcium. In addition, STDP at inhibitory synapses may obey different learning rules depending on the nature of the presynaptic interneuron and the projection sites. Therefore, the use of paired-recording (or optogenetic tools) must be generalized to unambiguously characterize the presynaptic interneuron [53]●● [110,111].

 Another set of questions are related to the induction of STDP *in vivo*: what triggers dendritic spikes in hippocampal pyramidal neurons during spatial exploration *in vivo* [76]● 321 [77]••? This may result from a selective and massive dendritic disinhibition together with a sudden and massive synaptic and intrinsic excitation. What triggers this abrupt disinhibition? Disinhibition is classically mediated by the selective activation of inhibitory interneuron- targeting GABAergic cells such as vasoactive intestinal polypeptide- (VIP) expressing interneurons. While to date the contribution of VIP interneurons to STDP remains unknown, a recent study shows that associative LTP is possible upon activation of VIP interneurons [112]●. Further studies will certainly provide answers to all these pending questions.

#### **Conflict of interest statement**

The authors declare no conflict of interest.

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 **Figure 1**. Spike timing-dependent plasticity (STDP) at excitatory inputs. A. Asymmetric STDP curve. Left, enhancement of synaptic transmission (t-LTP) following repeated EPSP-AP sequences and depression of synaptic transmission (t-LTD) following repeated AP-EPSP sequences. Middle, time-course of t-LTP (top) and t-LTD (bottom). Right, plasticity as a function of spike timing. For positive correlation (EPSP-AP, Δt>0) t-LTP is observed (red) whereas for negative correlation (AP-EPSP, Δt <0) induces t-LTD. B. Time windows for t-LTP and t-LTD depends on the spike number. For 1 postsynaptic AP, the time window is small (~0.1 s) whereas for a large number of postsynaptic AP (8-10), the time window is larger (~1 sec; adapted from [16,19]). C. Variability of STDP curves at excitatory inputs. Left, asymmetric STDP curve with a second t-LTD window. Middle, t-LTP only and t-LTD only. Right, anti-Hebbian asymmetric curve.



 **Figure 2**. STDP at inhibitory synapses. A. Anti-Hebbian asymmetric curve observed at fast- spiking interneuron-pyramidal neuron in young mice becomes symmetric with age (adapted from [53]●●). B. Synergy between plasticity at excitatory and inhibitory synapses. Simplified circuit constituted of three neurons (2 pyramidal cells and 1 FS interneuron). Top, pre-post spiking activity leads to enhanced synaptic excitation and reduced synaptic inhibition (t-LTDi). Bottom, post-pre spiking activity reduces synaptic excitation and enhance synaptic inhibition (t-LTPi).



 **Figure 3**. STDP under various external calcium concentrations in CA1 pyramidal neurons recorded in a whole-cell configuration. Pre-synaptic stimulation was evoked by a stimulation electrode placed in the Schaffer collaterals. These results were obtained with a simple 1:1 protocol (i.e. 1 presynaptic and 1 postsynaptic action potential) with a low pairing frequency of 0.3 Hz and 100 repetitions for t-LTP, and 150 repetitions for t-LTD. In 1.3 mM extracellular calcium, no plasticity is induced. In 1.8 mM extracellular calcium, pre-post and post-pre protocols leads to t-LTD. In 3 mM extracellular calcium, pre-post protocol leads to t-LTP and post-pre protocol leads to t-LTD. Adapted from [74]●●.



 **Figure 4**. STDP-like plasticity induced by dendritic calcium spike in mouse CA1 hippocampal neurons during spatial exploration. A. & B. Emergence of a place field in a silent neuron. A. Voltage responses of a CA1 pyramidal neuron before and after the spontaneous occurrence of a dendritic spike (red trace). B. Interpretation of the plasticity. In initial conditions (Before), the neuron is silent during the exploration of the environment (A) indicating that presynaptic CA3 synapses are weak (B). After dendritic calcium spike (After), the CA3 input spatially corresponding to the animal position when the dendritic calcium spike occurred is strongly potentiated (symbolized by a big red terminal, B). C & D. Plasticity of an existing place field induced by dendritic calcium spike. C. The dendritic calcium spike precedes the place-field evoked discharge by a few hundred milliseconds. D. Interpretation of the changes. In initial conditions (Before), the CA3 input corresponding to the animal position in the place field is strong (big and black terminal). After dendritic calcium spike, this input weakens (green and small terminal) and the neighbor input is potentiated (red and big terminal). Adapted from [76]● [77]●●.

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