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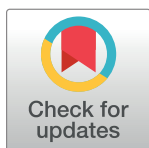
RESEARCH ARTICLE

Detection of porcine enteric viruses (Kobuvirus, Mamastrovirus and Sapelovirus) in domestic pigs in Corsica, France

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Abstract

Many enteric viruses are found in pig farms around the world and can cause death of animals or important production losses for breeders. Among the wide spectrum of enteric viral species, porcine Sapelovirus (PSV), porcine Kobuvirus (PKoV) and porcine Astrovirus (PAstV) are frequently found in pig feces. In this study we investigated sixteen pig farms in Corsica, France, to evaluate the circulation of three enteric viruses (PKoV, PAstV-1 and PSV). In addition to the three viruses studied by RT-qPCR (908 pig feces samples), 26 stool samples were tested using the Next Generation Sequencing method (NGS). Our results showed viral RNA detection rates (i) of 62.0% [58.7–65.1] (n = 563/908) for PSV, (ii) of 44.8% [41.5–48.1] (n = 407/908) for PKoV and (iii) of 8.6% [6.8–10.6] (n = 78/908) for PAstV-1. Significant differences were observed for all three viruses according to age (P-value = 2.4e–13 for PAstV-1; 2.4e–12 for PKoV and 0.005 for PSV). The type of breeding was significantly associated with RNA detection only for PAstV-1 (P-value = 9.6e–6). Among the 26 samples tested with NGS method, consensus sequences corresponding to 10 different species of virus were detected. This study provides first insight on the presence of three common porcine enteric viruses in France. We also showed that they are frequently encountered in pigs born and bred in Corsica, which demonstrates endemic local circulation.

Introduction

Pig farms make an important contribution to the economy of world agriculture and are an important source of food. Porcine diarrhea can cause mortality in animals, especially in piglets, and cause economic losses to the pig farmers; many of the pathogens responsible can also infect humans. A very broad spectrum of viruses that can cause porcine diarrhea has been found in pig feces, including porcine Sapelovirus (PSV), porcine Kobuvirus (PKoV), porcine Sapovirus, porcine Astrovirus (PAstV), porcine Bocavirus and porcine Rotavirus [1–7]. The

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most prevalent viruses detected in pig feces are PKoV, Mamastroviruses or Astrovirus 4 (PAstV), porcine Circovirus (PCV) and PSV [8, 9].

Kobuviruses belong to the *Picornaviridae* family. The genome is a single-stranded 8.2–8.3-kb RNA molecule that contains a large open reading frame coding for a polyprotein [10, 11]. Different species of kobuviruses have been found around the world in diverse animal species (pigs, cattle, sheep, goats, bats, rodents, felines, canines, etc.) and humans. It is suspected to be a pathogen that causes digestive disorders, particularly diarrhea in humans and animals, with transmission occurring via the fecal–oral route [12].

Porcine Sapelovirus (Family *Picornaviridae*, genus *Sapelovirus*) is a non-enveloped virus of 7.5–8.3 kb positive-polarity single-stranded RNA genome [13]. *Sapelovirus* genus is closely related to the genus Enterovirus and consists of three species: *Avian Sapelovirus*, *Sapelovirus A* (Porcine Sapelovirus [PSV]) and *Sapelovirus B* (simian Sapelovirus), with a single serotype [14]. PSV is transmitted via the fecal–oral route, and infection of pigs can be asymptomatic or associated with diarrhea, respiratory distress, encephalitis, skin lesions and reproductive tract disorders [15–17]. PSV is circulating in China, India, Korea, the United States, Brazil and Europe (Germany, the United Kingdom and Spain) [16–23]. Recently, PSV was detected and characterized for the first time in France, in Corsica. Importantly, the PSV-infected piglet from which the sequenced strain was isolated (PSV OPY-1-Corsica-2017; Genbank accession no. MH513612) was born and bred on the island of Corsica, suggesting local transmission [24].

Astroviruses are nonenveloped single-stranded RNA viruses with positive polarity, with an icosahedral capsid [25], that belong to the family *Astroviridae*, which includes two genera: *Mamastrovirus* (mammals) and *Avastrovirus* (avian) (ICTV, *Astroviridae*, 2019). Astroviruses can infect a large spectrum of animal species (pigs, deer, marine mammals, rodents, birds, pets, etc.) as well as humans [26]. Astrovirus infections are generally associated with more or less severe gastrointestinal signs in mammals (Mendez and Arias, 2007), but have also been detected in healthy individuals [27]. In humans, they cause intestinal disorders, particularly in children and immunocompromised individuals [28, 29].

The present study was conducted in Corsica, a French Mediterranean island, where livestock farming is a principal economic activity. In this region, more than 54,000 pigs, predominantly of the “Nustrale” breed, are bred using a traditional extensive farming system [30, 31]. Traditional extensive (or semi-extensive) outdoor system of pig farming is the main method of breeding. It favors contact with wild animals, which could result in sharing of pathogens such as hepatitis E virus (HEV) and Aujeszky’s disease agent [32, 33]. For HEV, we recently reported RNA detection in 9.2% of tested pig stool samples, with 75% of pig farms showing at least one positive sample [34]. Exploring the circulation of other enteric viruses in such pig farms could help to gain knowledge in the epidemiological cycle of HEV through the possible role of co-infection and super-infection. The main aim of this study was to detect and characterize three common porcine enteric viruses (PKoV, PAstV-1 and PSV) by molecular detection analysis of faeces collected within Corsican pigfarms.

Materials and methods

Study area, pig farms and sampling plan

Study area, samples/data collection, pig farms sampled, sampling plan and ethics statement are as described previously [34].

Briefly, (i) we collected fresh stool samples individually on the ground where pigs were pasturing and also intra-rectally using a glove with the help of a qualified technician (the individual level was control for each sample; feces were collected directly after defecation); (ii) three

types of breeding system operated in Corsica were included: seven outdoor extensive farms (E-farms), six outdoor semi-extensive farms (SE-farms) and three indoor closed farms (C-farms); (iii). for each stool sample, the township, anonymous breeder code, breeding type, age and breed of pig and nature of the sample (“rectal” or “on the ground” feces) were recorded; (iv). Four age categories were defined among the young pigs: 1–3 months, 3–4 months, 4–6 months and adults (older than 6 months). Samples from plots where post-weaning pigs were held together with older pigs (age mixed) were classified as the “Herd” group. Information on the individual health status of the pigs was not collected by a case report form. However, there is no apparent disease in the farms during the collect according to breeders. All the samples were collected between April and September 2017.

RNA extraction and reverse transcription–quantitative polymerase chain reaction (RT–qPCR)

One gram of fecal sample was resuspended in 9 mL of phosphate-buffered saline and then centrifuged at $5,000 \times g$ for 10 min. The resulting supernatant was collected and stored at -80°C until processed. Viral RNA was extracted from 200 μL of supernatant using QIAamp Cadore Pathogen on a QIAcube HT (Qiagen, Hilden, Germany) according to the manufacturer’s instructions. Samples were spiked with an internal control (T4 and MS2 phages) before extraction, to monitor the extraction and subsequent steps, as described previously. Each pool was spiked before extraction with a predefined amount of MS2 bacteriophage in order to monitor the subsequent steps (nucleic acid purification, reverse transcription and PCR amplification) and to detect the presence of inhibitors and enzymatic reactions as described previously [35]. All extractions and RT–qPCRs were checked with the presence of a curve for phage and positive and negative controls respectively. Nucleic acids were eluted in 100 μL of RE buffer and stored at -80°C .

Samples initially collected for the detection of HEV RNA were analyzed by RT–qPCR for PKoV, PSV and PAsV-1. Of the 919 samples initially collected, 908 were available for this study. Details of the three molecular assay [8, 36] are presented in Table 1 using an Applied Quant Studio 3 (Applied Biosystems, CA, USA). For PKoV RNA and PAsV-1 RNA detection, a RT–qPCR test was considered positive if negative controls were negative, positive controls were positive and an exponential curve was observed before a 35-Ct threshold.

For PSV RNA detection, the qPCR machine was programmed to perform a melt-curve analysis at the end of the run to ensure assay specificity; RT–qPCR results were considered positive if a melt curve was detected at between 83°C and 85°C and an amplification curve was observed before a 35-Ct threshold. The QuantiTect SYBR® Green PCR Kit was used for this biomolecular detection (Qiagen, Hilden, Germany).

Table 1. RT–qPCR detection assays used in the study.

Viruses	Name of primers and probes	Sequences	References
PSV	FW: SYBR-PSV1 primer	GGCAGTAGCGTGGCGAGC	[36]
	REV: SYBR-PSV2 primer	CTACTCTCCTGTAACCAGT	
PKoV	FW: T-248-F-PKoV	TCTCTGACCTCTGAAGTGCACT	[8]
	REV: T-249-R-PKoV	TGAAGAAGCCATGTGTCTTGTC	
	Probe: T-250-PKoV-FAM	GGTTGCGTGGCTGGGAATCCAC	
PAsV-1	FW: T217-F-PAsV-1	CCAAAACCAGCAATCCGTCAA	
	REV: T218-R-PAsV-1	GCCCCTAAAGCAACGATCGG	
	Probe: T-219-PAsV-1-VIC	TTCTTGTC AAGGATAATACGGGG	

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Statistical analyses

The detection rate of RNA viruses (PKoV, PAsTV-1 and PSV) in pig fecal samples was calculated at the individual level and the pig farm level. Positivity rate was also estimated in each subgroup, and a two-sided 95% confidence interval [95% CI] was calculated. Categorical variables were expressed as the number of cases (percentages). Frequencies were compared using the χ^2 test or Fisher's exact test ($P < 0.05$). A bivariate analysis was carried out to identify the variables that were related to infection with each virus. The multivariate logistic regression analysis included variables that were related to outcome variables in the bivariate analysis with a P -value < 0.2 or a possible association. Odds ratios (ORs), including their 95% CIs, were calculated for the logistic regression models. As in Capai, F. [34], samples with no associated age (Herd group) were excluded from the multivariate analysis, and previous results for the detection rate of HEV RNA among pig feces were included in the analysis to estimate a possible association between coinfections with different viruses [37]. All statistical analyses were performed using the R program (<http://www.r-project.org>).

Virus genome sequencing

Virus genome sequencing was performed for 26 stool samples as described previously [38]. Only, 26 analyses could be realized for financial reason and these 26 samples were randomly selected among the overall samples. A random RT-qPCR was performed using tagged random primers. A ProtoScript® II Reverse Transcriptase kit (New England Biolabs) was used for reverse transcription with random tagged primers, and Platinum® Taq High Fidelity polymerase enzyme (Thermo Fisher Scientific) with specific primers for amplification. After Qubit quantification using Qubit® dsDNA HS Assay Kit and Qubit 2.0 fluorometer (ThermoFisher Scientific), amplicons were fragmented (sonication) into fragments of 200 bp length. Libraries were built by adding barcodes for sample identification, and primers for amplification using the AB Library Builder System (ThermoFisher Scientific). To pool equimolar amounts of the barcoded samples, a quantification by quantitative PCR using an Ion Library TaqMan™ Quantitation Kit (Thermo Fisher Scientific) was performed. An automated Ion Chef instrument (ThermoFisher) was used for emulsion PCR of the pools and loading them on a 520 chip. Sequencing was performed using S5 Ion torrent technology (Thermo Fisher Scientific) following the manufacturer's instructions. Reads were trimmed (reads with quality score < 0.99 and length < 100 bp were removed, and the 30 first and 30 last nucleotides were removed from the reads), and de novo contigs were produced. These contigs were submitted to Blastn to determine the best reference sequences(s). A consensus sequence was obtained after mapping of the reads on the previously determined reference using CLC genomics workbench software 20.0.4 (Qiagen). The de novo contig was compared with the consensus sequence to ensure that the reference sequence did not affect the consensus sequence. Only sequences corresponding to enteric viruses found in pigs were selected for analysis.

Results

As suggested by Arya, Antonisamy [39], we previously calculated the minimum sample size required to achieve the objectives related to the HEV ($n = 176$ stool specimens) [34]. Overall, we collected 919 pig feces samples from 16 pig farms selected according to location and breeding system.

For PSV RNA detection, using the SYBR green A range of temperatures around 84°C (83–85°C) was tolerated. Indeed, a one-nucleotide difference within the amplified sequence can impact the melt-curve dissociation temperature [40, 41].

Viral RNA detection rate for the three viruses in feces from domestic pigs and univariate analysis

Overall, 908 samples were available for the detection of the three virus of interest, 310 were from E-farms, 396 from SE-farm and 201 from C-farm.

Our results showed viral RNA detection rates (i) of 62.0% [58.7–65.1] (n = 563/908) for PSV, (ii) of 44.8% [41.5–48.1] (n = 407/908) for PKoV and (iii) of 8.6% [6.8–10.6] (n = 78/908) for PAsV-1 (Table 2).

For PSV and PKoV, there was no statistical association with the type of breeding system; in contrast, PAsV-1 was detected more frequently in C-farms compared with SE- and E-farms (P-value = 9.6e–6) (Table 2).

Significant differences were observed for all three viruses according to age (P-value = 2.4e–13 for PAsV-1; 2.4e–12 for PKoV and 0.005 for PSV) (Table 2). RNA virus detection by age group showed a significant decrease in the rate of positive cases after three months for PAsV-1 (30.6% vs. 6.0%; P-value = 3.87e–7) and between 1 and 3 months (69.4%) and in adults (28.9%) for PKoV (P-value = 6.37e–12). For PSV, the detection rates by age group were between 61.0% and 77.5% (Fig 1). However, the positivity rate among pigs under six months of age was significantly lower than that in pigs older than 6 months (71.4% vs. 61.1%; P-value = 0.014; OR = 1.59, CI 95% 1.09–2.32).

Description of coinfections in samples of pig feces

Table 3 lists all infections and coinfections detected in pig feces samples. Of the 908 samples tested, 697 samples were positive for at least one virus (76.8%). A total of 344 samples contained at least two distinct viral RNA (37.9%), of which 259 specimens (28.5%) were coinfecting by two viruses, 78 specimens (8.6%) were coinfecting by three viruses and seven specimens (0.8%) were positive for all four viruses (Table 3).

Table 2. Viral RNA in stools of domestic pigs stratified by breeding system and age.

Factor	Condition	Number of samples		positive (n)	PKoV RNA detection (%)		P-value	PAsV-1 RNA detection (%)		P-value	PSV RNA detection (%)		P-value		
		N	%		%	[95% CI]		%	[95% CI]		%	[95% CI]			
Breeding systems	E-farm	310	34.1	136	43.9	[38.3–49.6]	0.91	17	5.5	[38.3–49.6]	9.6e–6	186	60.0	[54.3–65.5]	0.0094
	SE-farm	396	43.6	180	45.5	[40.5–50.5]		27	6.8	[4.5–9.8]		233	58.8	[53.8–63.7]	
	C-farm	201	22.1	91	45.3	[38.2–52.4]		34	16.9	[12–22.8]		143	71.1	[64.3–77.3]	
Age	1–3 months	111	12.2	77	69.4	[59.9–77.7]	2.4e–12	34	30.6	[22.2–40.1]	2.4e–13	86	77.5	[68.6–84.9]	0.005
	3–4 months	143	15.7	84	58.7	[50.2–66.9]		9	6.3	[2.9–11.6]		91	63.6	[55.2–71.5]	
	4–6 months	162	17.8	71	43.8	[36.0–51.8]		4	2.5	[0.7–6.2]		120	74.1	[66.6–80.6]	
	Adults (>6 months)*	190	20.9	54	28.4	[22.1–35.4]		17	8.9	[5.3–13.9]		116	61.1	[53.7–68.0]	
	Herds (age mixed)	302	33.3	121	40.1	[34.5–45.8]		14	4.6	[2.6–7.7]		150	49.7	[43.9–55.4]	
All pigs		908	100.0	407	44.8	[41.5–48.1]		78	8.6	[6.8–10.6]		563	62.0	[58.7–65.1]	

*6–8 months old (n = 47); Sow/Boar (n = 30); at least 6 months old (n = 108).

E-farm: Extensive farm; SE-farm: semi-extensive farm; C-farm: closed farm; Herds: pigs without associated age.

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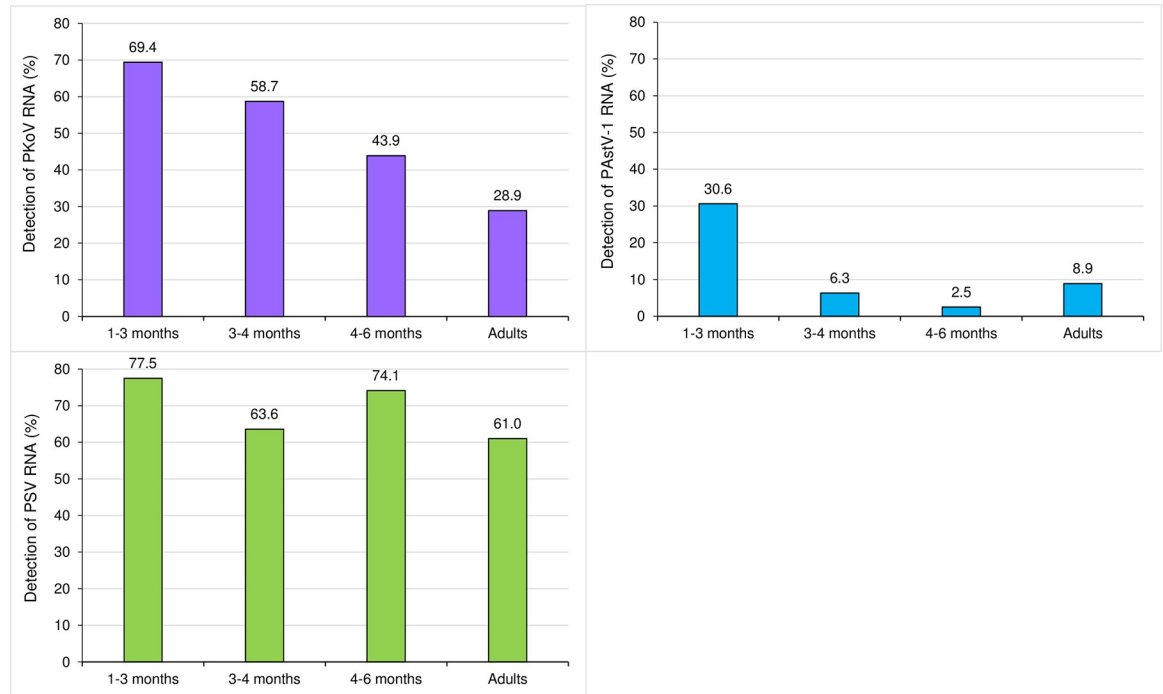


Fig 1. Detection rate of each RNA virus by age group.

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Detection rate of viral RNA at the farm level. At least one pig was detected positive for PKoV RNA in each of the 16 pig farms sampled (100%), 62.5% for PASTV-1 infection (n = 10/16) and 93.8% for PSV infection (n = 15/16). The positivity rate at farm level ranged between 22.5% and 80.8%, 0.0% and 34.0%, and 0.0% and 87.5% for PKoV, PASTV-1 and PSV, respectively. The detail of the detection rates of the three viruses for each farm were presented on the [S1 Table](#).

Multivariate analysis: Associated factors identified for each viral infection

A multivariate logistic regression analysis was performed, and the results showed associations depending on the three viruses ([Table 4](#)). A strong association was observed between PKoV and PSV detection (OR = 3.36 [2.44–4.67]; *P*-value = 9.1e–15). PKoV was also associated with PASTV-1 codetection (OR = 2.16 [1.29–3.70]; *P*-value = 0.015) and young pigs under 4 months of age (OR = 3.11 [2.31–4.20]; *P*-value = 4.3e–10). PASTV-1 detection was also associated with

Table 3. Coinfections in pig feces samples.

	N	Number for each coinfection					
No infection	211						
Single infection	353	PKoV	PSV	PASTV-1	HEV		
		102	229	9	13		
Double infection	259	PKoV/PSV	PKoV/PASTV-1	PKoV/HEV	PSV/HEV	PASTV-1/PSV	PASTV-1/HEV
		216	4	3	21	14	1
Triple infection	78	PKoV/PASTV-1/PSV	PKoV/HEV/PSV	PKoV/HEV/PASTV-1	HEV/PASTV-1/PSV		
		39	35	2	2		
All four viruses	7						

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Table 4. Statistical analysis of factors associated with each virus (multivariate logistic regression model with random effect at the farm level).

Viruses	Factor	Condition	OR	[95% CI]	P-value adjusted
PKoV	Coinfection	PSV	3.36	[2.44–4.67]	9.1e–15
	Coinfection	PAstV-1	2.16	[1.29–3.70]	0.015
	Age group	0–4 months	3.11	[2.31–4.20]	4.3e–10
PAstV-1	Coinfection	PKoV	2.25	[1.32–3.90]	0.014
	Age group	>3 months	0.18	[0.11–0.29]	2.8e–9
PSV	Coinfection	HEV	2.01	[1.17–3.64]	0.041
	Coinfection	PKoV	2.92	[2.13–4.04]	3.7e–8

Only factors with P -values < 0.05 are included.

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age, with pigs over 3 months of age significantly less frequently infected than younger pigs (OR = 0.18 [0.11–0.29]; P -value = 2.8e–9). PSV infection was not associated with age or type of breeding.

Next-generation sequencing (NGS). In addition to the three viruses studied by RT–qPCR, NGS analysis was performed in a second phase on 26 random selected samples. Overall, consensus sequences corresponding to 10 different species of virus were obtained. PAstVs were detected in ten samples (38.5%), porcine stool/serum-associated circular virus in six (23.1%), Bocavirus in five (19.2%), Sapelovirus and Posavirus in four (15.4%), Circovirus in three (11.5%), Pasivirus in two (7.7%) and Rotavirus and porcine Enterovirus G in one (3.8%). Among these samples, 14 (54%) had consensus sequences corresponding to at least two different viruses.

Porcine astroviruses strains detected among our samples were closed to pig strains from different countries: Hungary (JQ340310 90.81–92% of identity); United States (KJ495987 93.36% of identity) and New Zealand (KJ495990 92.8% of identity). The main astroviruses found, were *Mammastrovirus* 3, *Porcine Astrovirus* 2 and 4. Concerning Bocaviruses strains detected, identity of 97.08% was determined with pigs from: a Hungarian domestic pig (KF206167), 95–97.65% with a Chinese pigs (KX017193; KM402139 and HM053693), 94.13–96.48% with an American pigs (KF025394 and KF025484) and 95.28% with a Croatian pig (KF206161). Concerning, the sequences obtained for the Sapelovirus, the strongest homologies were observed when compared with strain MH513612 also isolated in Corsica (92–99.91% of identity) [24].

For the other results, Table 5 summarizes all the sequences obtained, their length, the reference sequence to which each corresponds and the percentage of nucleotide identity with this sequence. Six of the recovered sequences had a quality score required for submission to GenBank and the accession numbers range from OL739527 to OL739532.

Discussion

This study investigated the detection rate of three enteric viral infections, PKoV, Astrovirus-1 and Sapelovirus, in 908 pigs of different ages and breeding systems. To our knowledge, this work is the first to study factors associated with presence of viral RNA of PSV, PKoV and PAstV-1 in stools of domestic pigs in France.

RNA detection rate of the three enteric viruses

PSV was the most prevalent (62%) and observed rates are higher than those described in the US, Brazil and India (7–31%) [9, 17, 20]. However, the existing results for PSV are not all

Table 5. Diversity of sequences found in pig feces and percentage of identity with strains in the literature.

Samples	Accession number	Viruses	Sequence of reference	Accession number of the sequence of reference	Consensus length	% Cover	% Identity
1	/	/	/	/	/	/	/
2	OL739527	Sapelovirus	Sapelovirus A strain OPY-1-Corsica-2017 polyprotein gene, complete cds	MH513612	5,927	96	92.46
	/	Bocavirus	Porcine bocavirus strain G85-1AT-HU, complete genome	KF206167	3,507	100	97.08
	/	Bocavirus	Porcine bocavirus 3C isolate pig/ZJD/China/2006, complete genome	JN681175	3,683	99	91.61
	/	Porcine Astrovirus	Mamastrovirus 3 isolate PAsTV_GER_L00855-K14_14-04_2014 genome assembly, complete genome: monopartite	LT898434	1,508	52	90.88
3	/	Sapelovirus	Sapelovirus A strain OPY-1-Corsica-2017 polyprotein gene, complete cds	MH513612	492	100	93.09
4	/	Bocavirus	Porcine bocavirus strain CH/HNZM, complete genome	KX017193	2,105	99	97.22
	/	Porcine Astrovirus	Astrovirus wild boar/WBAstV-1/2011/HUN, complete genome	JQ340310	428	100	91.59
	/	Porcine serum-associated circular virus	Porcine serum-associated circular virus isolate BR3, complete genome	KU203353	531	86	86.33
	/	Enterovirus G	Enterovirus G isolate GER/F9-6/12-02-2013 polyprotein gene, partial cds	MF113376	956	100	86.72
5	/	Sapelovirus	Porcine Sapelovirus isolate PSV_P1-3-3_Contig(g12h12) polyprotein gene, partial cds	KF705647	498	98	92.77
	/	Sapelovirus	Sapelovirus A strain OPY-1-Corsica-2017 polyprotein gene, complete cds	MH513612	1,687	100	92.12
	/	Sapelovirus	Sapelovirus A isolate HuN21 polyprotein gene, complete cds	MF440649	1,230	100	86.59
	/	Sapelovirus	Sapelovirus A isolate PSV_GER_L00798-K11_14-02_2014 genome assembly, complete genome: monopartite	LT900497	1,894	100	87.08
	/	Bocavirus	Porcine bocavirus strain 644-1DI-HR, complete genome	KF206161	4,580	100	95.28
	/	Bocavirus	Porcine bocavirus isolate GD11, complete genome	KM402139	4,580	100	95.09
	/	Porcine Astrovirus	Porcine astrovirus 2 clone KDC-6 ORF1ab gene, partial cds; and ORF2 gene, complete cds	KJ495987	466	100	93.36
	/	Porcine Astrovirus	Porcine astrovirus 2 genes for ORF1ab, ORF1a, ORF2, complete cds, strain: PoAstV2/JPN/HgYa2-3/2015	LC201588	422	100	87
	/	Porcine Astrovirus	Astrovirus wild boar/WBAstV-1/2011/HUN, complete genome	JQ340310	533	100	90.81
	/	Porcine stool-associated circular virus	Porcine stool-associated circular virus 7 isolate EP2-B, complete genome	KJ577813	1,191	99	82.59
6	/	Posavirus	Posavirus 1 isolate PsaV_GER_L01017-K01_15-07_2015 genome assembly, complete genome: monopartite	LT898419	741	100	96.63
	/	Posavirus	Posavirus sp. isolate 12144_61, complete genome	KX673217	559	100	97.5
	/	Bocavirus	Porcine bocavirus 3 isolate IA159-3 NS1 and NP1 genes, complete cds; and VP1/VP2 gene, partial cds	KF025387	491	100	84.85
	OL739528	Porcine serum-associated circular virus	Porcine serum-associated circular virus isolate BR3, complete genome	KU203353	929	99	86.03
7	/	/	/	/	/	/	
8	/	/	/	/	/	/	

(Continued)

Table 5. (Continued)

Samples	Accession number	Viruses	Sequence of reference	Accession number of the sequence of reference	Consensus length	% Cover	% Identity
9	OL739529	Circovirus	Circovirus sp. isolate PoCirV_VIRES_JL01_C5 capsid protein gene, partial cds	MK377643	1,038	100	85.17
	/	Circovirus	Circovirus sp. isolate PoCirV_VIRES_GX05_C4 replicase gene, partial cds	MK377558	639	64	89.54
	/	Porcine Astrovirus	Porcine astrovirus 2 genes for ORF1ab, ORF1a, ORF2, complete cds, strain: PoAstV2/JPN/Ishi-Ya4/2015	LC201589	879	78	82.61
	OL739530	Porcine Astrovirus	Porcine astrovirus 4 genes for ORF1ab, ORF1a, ORF2, complete cds, strain: PoAstV4/JPN/Bu5-10-2/2014	LC201603	989	100	83.64
10	/	Bocavirus	Porcine bocavirus 1 pig/ZJD/China/2006, complete genome	HM053693	383	100	97.65
11	/	Porcine Astrovirus	Porcine astrovirus 4 isolate 15–12, complete genome	KU764486	1,624	99	87.98
	/	Porcine Astrovirus	Porcine astrovirus 4 isolate 15–13, complete genome	KU764484	1,624	99	88.21
	/	Porcine stool-associated circular virus	Porcine stool-associated circular virus 7 isolate EP3-C, complete genome	KJ577814	719	100	86.77
12	/	Posavirus	Posavirus sp. isolate 17668_12, complete genome	KX673279	1,606	93	85.53
13	/	Porcine Astrovirus	Porcine astrovirus 4 strain JXJA, complete genome	KX060808	1,187	100	89.9
	/	Bocavirus	Porcine bocavirus 3 isolate IL330 NS1 and NP1 genes, complete cds; and VP1/VP2 gene, partial cds	KF025394	483	100	96.48
14	/	Porcine Astrovirus	Porcine astrovirus 4 strain 35/USA, complete genome	JF713713	1,798	100	90.12
	/	Porcine Astrovirus	Porcine astrovirus 4 strain CH/JXZS/2014, complete genome	KX060809	1813	100	90.13
15	OL739531	Porcine Astrovirus	Mamastrovirus 3 isolate PoAstV_VIRES_HeB02_C4 ORF1ab and ORF1a genes, partial cds	MK378508	2,192	100	90.81
	/	Circovirus	Circovirus sp. isolate PoCirV_VIRES_SD02_C2 replicase gene, partial cds	MK377699	557	100	88.6
16	OL739532	Circovirus	Circovirus sp. isolate PoCirV_VIRES_HeB04_C3 capsid protein gene, partial cds	MK377607	777	100	90.27
17	/	Porcine Astrovirus	Porcine astrovirus 4 genes for ORF1ab, ORF1a, ORF2, complete cds, strain: PoAstV/JPN/Mo12-1-2/2015	LC201610	869	100	89.31
	/	Porcine Astrovirus	Porcine astrovirus 2 clone NZP-93_Subtype_2 ORF1ab gene, partial cds	KJ495990	375	100	92.8
	/	Bocavirus	Porcine bocavirus 3 isolate IA13-1 VP1/VP2 gene, complete cds	KF025484	931	100	94.13
	/	Porcine Astrovirus	Mamastrovirus 2 isolate U083, complete genome	KY940077	2,043	100	80.33
	/	Porcine Astrovirus	Mamastrovirus 3 isolate PoAstV_VIRES_GZ04_C10 ORF1ab and ORF1a genes, partial cds	MK378502	869	100	89.77
18	/	Porcine Astrovirus	Porcine astrovirus 4 genes for ORF1ab, ORF1a, ORF2, complete cds, strain: PoAstV4/JPN/Ishi-Ya7-1/2015	LC201613	1,970	91	89.22
	/	Porcine Astrovirus	Mamastrovirus 2 isolate U083, complete genome	KY940077	5,230	85	87.06
	/	Porcine Astrovirus	Mamastrovirus 3 isolate PAstV_GER_L00855-K14_14-04_2014 genome assembly, complete genome: monopartite	LT898434	4,050	100	85.87
19	/	/	/	/	/	/	/
20	/	Sapelovirus	Sapelovirus A strain OPY-1-Corsica-2017 polyprotein gene, complete cds	MH513612	2,360	100	99.91
	/	Pasivirus	Swine pasivirus SPaV1/US/17-50816IA60467-1/2001 polyprotein gene, complete cds	MG674090	1,186	100	77.88
	/	Porcine stool-associated circular virus	Porcine serum-associated circular virus isolate BR2, complete genome	KU203352	1,168	99	92.76

(Continued)

Table 5. (Continued)

Samples	Accession number	Viruses	Sequence of reference	Accession number of the sequence of reference	Consensus length	% Cover	% Identity
21	/	/	/	/	/	/	/
22	/	Rotavirus	Porcine rotavirus B isolate PoRVB_VP2_VIRES_NM01_C2 VP2 gene, partial cds	MK379346	55	100	98.18
23	/	/	/	/	/	/	/
24	/	Posavirus	Posavirus sp. isolate 17668_12, complete genome	KX673279	3,003	100	88.71
	/	Pasivirus	Pasivirus A isolate SPaV-A GER L01061-K07 15-03 2015 genome assembly, complete genome: monopartite	LT898422	451	100	83.44
	/	Porcine stool-associated circular virus	Porcine stool-associated circular virus 7 isolate EP3-C, complete genome	KJ577814	702	100	83.25
25	/	/	/	/	/	/	/
26	/	Posavirus	Posavirus sp. isolate 17668_13_2, complete genome	KX673281	1,282	99	89.31
	/	Posavirus	Posavirus 3 strain 10611 polyprotein gene, complete cds	KT833079	860	100	94.07
	/	Posavirus	Posavirus strain 7048 polyprotein gene, partial cds	KT833076	1.121	99	87.92

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performed with the same method (SYBR, classical RT-qPCR, NGS) so it is complicated to compare the results in the literature.

PKoV RNA was detected in almost half of samples, which is similar to data reported from pig farms in China (45.7%) [42] and Japan (45.4%) [43]. In five European countries, PKoV infections have been described previously as highly prevalent in both diarrheic and healthy pigs, 54.5% and 58.2%, respectively [8].

PAstV-1 RNA was detected in less than 10% of our samples, but comparison is difficult because other studies were performed at the genus level (*Mamastrovirus*: 52% positive) [9] or for all PAstVs combined (Thailand: 6.5%; MN, USA: 62%; Slovakia: 93.2%) [44–46].

Very early exposure of pigs

The analysis of the detection of viral RNA according to pig age showed that for all viruses tested, pigs < 4 months old were consistently the age group exhibiting the highest rate. This association between age and rate of infection was confirmed for PKoV and PAstV-1 in the multivariate analysis. It may correlate with high exposure after weaning (about 2 months after birth) but also suggest persisting presence of viruses in the breeding environment and loss of maternal immunity. Piglet passive immunity is derived from colostrum and not from breastfeeding [47], with a decrease in immunoglobulins A, G and M after farrowing [48]. This trend was also observed during our previous study of HEV [34]. Correlation of decreasing positivity rates and increasing age of pigs is described for Kobuviruses in Italy [49] and East Africa [50]. A higher PKoV detection rate in young piglets has also been reported in other studies [10, 51, 52]. However, the kinetics of infection were different in each study and may depend on the organization of each farm and other environmental characteristics. Study in different age-group 0-1months, 1-2months, etc. or follow-up of individual animals could help confirming this hypothesis.

In traditional Corsican farms, the average slaughter age is higher (12–18 months) compared with industrial farms. Therefore, at the time of slaughtering the majority of *Nustrale* Corsican pigs will have cleared replicative viral infection. The same finding was already reported for hepatitis E virus [34]. In contrast, the situation is different for Sapeloviruses with rates of replicating infection at 60% in adult pigs. Farming type does not seem to influence the detection rates of studied viruses but still need confirmation from future studies.

A potentially very broad spectrum of viruses in pig feces

In this study, 37.9% of pigs were coinfecting with at least two different viruses (including HEV). Strong trends were observed in the multivariate analysis regarding the associations between different viral infections (PKoV/PSV, PKoV/PAstV-1, PSV/HEV and PSV/PKoV). This wide variety of viruses in the feces was confirmed with the NGS method, which showed the presence of other viruses frequently found in pigs: Posavirus, other Astroviruses, Bocavirus, Enterovirus G, Circovirus, Pasivirus and Rotavirus. Coinfections were also evidenced in the NGS results, with more than half of the samples containing consensus sequences corresponding to at least two different viruses. These results are in line with previous studies reporting multiple coinfections of farmed pigs with porcine enteric viruses [6, 37], especially in pigs with diarrhea [53–55]. Our results are also interesting concerning the epidemiological cycle of HEV because co-infections can diminish the immune system of pigs, change the symptomatology of infections, [56, 57]. For example, experimental co-infection of HEV with Porcine Reproductive and Respiratory Syndrome Virus significantly prolonged HEV shedding [58].

Our study has several limitations. First, it was not initially designed to study these three viruses but to determine the prevalence of the HEV in pig farms in Corsica. Differences in detection could exist depending on criteria such as sampling time and place, age of pigs and clinical background of the tested animal population (diarrheic or healthy pigs). The lack of collection of clinical information (diarrhea or other symptoms) for the pigs included may have led to a bias in the analysis of the data; these data could have been essential for the improvement of knowledge concerning the studied viruses. The number of viruses studied could also have been larger to better assess the presence of major enteric viruses in Corsican pig farms. Finally, the lack of phylogenetic analysis of the different strains found is also a major limitation of our study. Indeed, the sequences obtained being in very varied zones of the genome of the various viruses, the phylogenetic analyses could not be realized.

In future studies, information about possible symptoms in pigs should be collected. It would also be useful to evaluate the phylogeny of the different strains found and to set up an RT-qPCR primer system to distinguish the different strains of Sapelovirus. Moreover, in view of the large variety of viruses present in the pig feces and the availability of microfluidic PCR technology, it would be helpful to set up microarrays that can detect all the principal known porcine enteric viruses. Moreover, the real impact of Kobuvirus, Astroviruses, Sapelovirus and other enteric viruses on animal health and breeding systems remains largely unknown and needs further epidemiological studies.

In conclusion, this study provides first insight on the presence of three common porcine enteric viruses in France and showed that they are frequently encountered in Corsica in pig farms using the traditional extensive breeding. The three viruses studied were found on almost all the farms, indicating widespread distribution. Moreover, the pigs tested were born and bred in Corsica, which demonstrates endemic local circulation. Whether such infections and co-infections can affect the productivity, impact the growth of pigs or cause immune weakness remains to be established. So far, this study has to be considered as a first step in the study of enteric viruses in Corsican pig farms.

Supporting information

S1 Table. Positivity rates for the three viruses by pig farm.

(DOCX)

S2 Table.

(DOCX)

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Ethical approval

The main part of the molecular analysis was carried out on fresh stools from the ground without contact with animals (98.4% of the samples). Fifteen of the 908 stools collected were intra-rectal samples, and were initially performed in a diagnostic setting (diarrheic pigs). We took an aliquot of these samples which have been collected by a qualified technician in the presence of the breeder. In this case, the study was exempt from ethical authorization because we did not handle animals and the intra-rectal samples were performed independently of the study with a diagnostic aim. All the breeders gave their explicit agreement for the collection of the samples, and all the data were anonymized.

Author Contributions

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References

1. Estes M.K., Palmer E.L., and Obijeski J.F. Rotaviruses: a review. *Curr Top Microbiol Immunol*. 1983. 105: p. 123–84 (1983). https://doi.org/10.1007/978-3-642-69159-1_3 PMID: 6313296
2. Meng X.J. Emerging and re-emerging swine viruses. *Transbound Emerg Dis*. 2012. 59 Suppl 1: p. 85–102 (2012). <https://doi.org/10.1111/j.1865-1682.2011.01291.x> PMID: 22225855
3. Shan T. et al. Genomic characterization and high prevalence of bocaviruses in swine. *PLoS One*. 2011. 6(4): p. e17292 (2011). <https://doi.org/10.1371/journal.pone.0017292> PMID: 21525999
4. Reuter G., Boldizsar A., Kiss I., and Pankovics P. Candidate new species of Kobuvirus in porcine hosts. *Emerg Infect Dis*. 2008. 14(12): p. 1968–70 (2008). <https://doi.org/10.3201/eid1412.080797> PMID: 19046542
5. Liu G.H. et al. RT-PCR test for detecting porcine sapovirus in weanling piglets in Hunan Province, China. *Trop Anim Health Prod*. 2012. 44(7): p. 1335–9 (2012). <https://doi.org/10.1007/s11250-012-0138-9> PMID: 22492394
6. Zhang B. et al. Viral metagenomics analysis demonstrates the diversity of viral flora in piglet diarrhoeic faeces in China. *J Gen Virol*. 2014. 95(Pt 7): p. 1603–1611 (2014). <https://doi.org/10.1099/vir.0.063743-0> PMID: 24718833
7. Yu J.M. et al. Identification of a novel picornavirus in healthy piglets and seroepidemiological evidence of its presence in humans. *PLoS One*. 2013. 8(8): p. e70137 (2013). <https://doi.org/10.1371/journal.pone.0070137> PMID: 23936384
8. Zhou W. et al. Molecular investigations on the prevalence and viral load of enteric viruses in pigs from five European countries. *Vet Microbiol*. 2016. 182: p. 75–81 (2016). <https://doi.org/10.1016/j.vetmic.2015.10.019> PMID: 26711031

9. Chen Q. et al. Metagenomic analysis of the RNA fraction of the fecal virome indicates high diversity in pigs infected by porcine endemic diarrhea virus in the United States. *Virology*. 2018. 15(1): p. 95 (2018). <https://doi.org/10.1186/s12985-018-1001-z> PMID: 29801460
10. Reuter G., Boldizsar A., and Pankovics P. Complete nucleotide and amino acid sequences and genetic organization of porcine kobuvirus, a member of a new species in the genus Kobuvirus, family Picornaviridae. *Arch Virol*. 2009. 154(1): p. 101–8 (2009). <https://doi.org/10.1007/s00705-008-0288-2> PMID: 19096904
11. Yu J.M. et al. Analysis and characterization of the complete genome of a member of a new species of kobuvirus associated with swine. *Arch Virol*. 2011. 156(5): p. 747–51 (2011). <https://doi.org/10.1007/s00705-010-0907-6> PMID: 21274731
12. Khamrin P., Maneekarn N., Okitsu S., and Ushijima H. Epidemiology of human and animal kobuviruses. *Virus Disease*. 2014. 25(2): p. 195–200 (2014). <https://doi.org/10.1007/s13337-014-0200-5> PMID: 25674585
13. Krumbholz A. et al. Sequencing of porcine enterovirus groups II and III reveals unique features of both virus groups. *J Virol*. 2002. 76(11): p. 5813–21 (2002). <https://doi.org/10.1128/jvi.76.11.5813-5821.2002> PMID: 11992011
14. ICTV, I.C.o.T.o.V. Genus: *Kobuvirus*. 2019; Available from: <https://talk.ictvonline.org/ictv-reports/ictv-online-report/positive-sense-rna-viruses/picornavirales/w/picornaviridae/686/genus-kobuvirus>.
15. Kim D.S. et al. Porcine Sapelovirus Uses alpha2,3-Linked Sialic Acid on GD1a Ganglioside as a Receptor. *J Virol*. 2016. 90(8): p. 4067–4077 (2016). <https://doi.org/10.1128/JVI.02449-15> PMID: 26865725
16. Lan D. et al. Isolation and characterization of the first Chinese porcine sapelovirus strain. *Arch Virol*. 2011. 156(9): p. 1567–74 (2011). <https://doi.org/10.1007/s00705-011-1035-7> PMID: 21618029
17. Ray P.K. et al. Porcine sapelovirus among diarrhoeic piglets in India. *Transbound Emerg Dis*. 2018. 65(1): p. 261–263 (2018). <https://doi.org/10.1111/tbed.12628> PMID: 28244229
18. Arruda P.H. et al. Detection of a novel sapelovirus in central nervous tissue of pigs with poliomyelitis in the USA. *Transbound Emerg Dis*. 2017. 64(2): p. 311–315 (2017). <https://doi.org/10.1111/tbed.12621> PMID: 28160432
19. Chen J. et al. Complete genome sequence of a novel porcine Sapelovirus strain YC2011 isolated from piglets with diarrhea. *J Virol*. 2012. 86(19): p. 10898 (2012). <https://doi.org/10.1128/JVI.01799-12> PMID: 22966190
20. Donin D.G. et al. First report of Porcine teschovirus (PTV), Porcine sapelovirus (PSV) and Enterovirus G (EV-G) in pig herds of Brazil. *Trop Anim Health Prod*. 2014. 46(3): p. 523–8 (2014). <https://doi.org/10.1007/s11250-013-0523-z> PMID: 24362793
21. Schock A. et al. Investigation into an outbreak of encephalomyelitis caused by a neuroinvasive porcine sapelovirus in the United Kingdom. *Vet Microbiol*. 2014. 172(3–4): p. 381–9 (2014). <https://doi.org/10.1016/j.vetmic.2014.06.001> PMID: 24984944
22. Buitrago D. et al. A survey of porcine picornaviruses and adenoviruses in fecal samples in Spain. *J Vet Diagn Invest*. 2010. 22(5): p. 763–6 (2010). <https://doi.org/10.1177/104063871002200519> PMID: 20807938
23. Son K.Y. et al. Full-length genomic analysis of Korean porcine Sapelovirus strains. *PLoS One*. 2014. 9(9): p. e107860 (2014). <https://doi.org/10.1371/journal.pone.0107860> PMID: 25229940
24. Piorkowski G. et al. First Identification and Genomic Characterization of a Porcine Sapelovirus from Corsica, France, 2017. *Microbiol Resour Announc*. 2018. 7(11) (2018).
25. Monroe S.S. et al. Subgenomic RNA sequence of human astrovirus supports classification of Astroviridae as a new family of RNA viruses. *J Virol*. 1993. 67(6): p. 3611–4 (1993). <https://doi.org/10.1128/JVI.67.6.3611-3614.1993> PMID: 8497068
26. Johnson C. et al. Astrovirus Pathogenesis. *Viruses*. 2017. 9(1) (2017). <https://doi.org/10.3390/v9010022> PMID: 28117758
27. Reuter G., Pankovics P., and Boros A. Identification of a novel astrovirus in a domestic pig in Hungary. *Arch Virol*. 2011. 156(1): p. 125–8 (2011). <https://doi.org/10.1007/s00705-010-0827-5> PMID: 20931250
28. Holtz L.R. et al. Astrovirus MLB2 viremia in febrile child. *Emerg Infect Dis*. 2011. 17(11): p. 2050–2 (2011). <https://doi.org/10.3201/eid1711.110496> PMID: 22099095
29. Wylie K.M. et al. Sequence analysis of the human virome in febrile and afebrile children. *PLoS One*. 2012. 7(6): p. e27735 (2012). <https://doi.org/10.1371/journal.pone.0027735> PMID: 22719819
30. Casabianca F, Poggioli A, Rossi JD, and Maestrini O, The beginning of collective management for the Corsican pig breed. Building up a standard and elaborating the breeders performance recording., in Serie A, Seminaires Mediterraneens, Mediterraneennes O, Editor. 2000. p. 23–34.

31. DRAAF., D.R.d.I.A.d.I.A.e.d.I.F., Panorama de l'agriculture en Corse. 2017.
32. Charrier F. et al. Aujeszky's Disease and Hepatitis E Viruses Transmission between Domestic Pigs and Wild Boars in Corsica: Evaluating the Importance of Wild/Domestic Interactions and the Efficacy of Management Measures. *Front Vet Sci*. 2018. 5: p. 1 (2018). <https://doi.org/10.3389/fvets.2018.00001> PMID: 29417054
33. Jori F. et al. Assessment of Domestic Pigs, Wild Boars and Feral Hybrid Pigs as Reservoirs of Hepatitis E Virus in Corsica, France. *Viruses*. 2016. 8(8) (2016). <https://doi.org/10.3390/v8080236> PMID: 27556478
34. Capai L. et al. Drastic decline of hepatitis E virus detection in domestic pigs after the age of 6 months, Corsica, France. *Transbound Emerg Dis*. 2019 (2019). <https://doi.org/10.1111/tbed.13304> PMID: 31336031
35. Ninove L. et al. RNA and DNA bacteriophages as molecular diagnosis controls in clinical virology: a comprehensive study of more than 45,000 routine PCR tests. *PLoS One*. 2011. 6(2): p. e16142 (2011). <https://doi.org/10.1371/journal.pone.0016142> PMID: 21347398
36. Kim D.S. et al. Pathogenesis of Korean SapelovirusA in piglets and chicks. *J Gen Virol*. 2016. 97(10): p. 2566–2574 (2016). <https://doi.org/10.1099/jgv.0.000571> PMID: 27487773
37. Xiao C.T. et al. Identification and characterization of novel porcine astroviruses (PAstVs) with high prevalence and frequent co-infection of individual pigs with multiple PAstV types. *J Gen Virol*. 2013. 94(Pt 3): p. 570–582 (2013). <https://doi.org/10.1099/vir.0.048744-0> PMID: 23223616
38. Stang A., Korn K., Wildner O., and Uberla K. Characterization of virus isolates by particle-associated nucleic acid PCR. *J Clin Microbiol*. 2005. 43(2): p. 716–20 (2005). <https://doi.org/10.1128/JCM.43.2.716-720.2005> PMID: 15695669
39. Arya R., Antonisamy B., and Kumar S. Sample size estimation in prevalence studies. *Indian J Pediatr*. 2012. 79(11): p. 1482–8 (2012). <https://doi.org/10.1007/s12098-012-0763-3> PMID: 22562262
40. Huang Q. et al. Multiplex fluorescence melting curve analysis for mutation detection with dual-labeled, self-quenched probes. *PLoS One*. 2011. 6(4): p. e19206 (2011). <https://doi.org/10.1371/journal.pone.0019206> PMID: 21552536
41. Westerman L.J. et al. Development of a real-time PCR for identification of brachyspira species in human colonic biopsies. *PLoS One*. 2012. 7(12): p. e52281 (2012). <https://doi.org/10.1371/journal.pone.0052281> PMID: 23284968
42. Yang Z. et al. Genetic characterization of porcine kobuvirus and detection of coinfecting pathogens in diarrheic pigs in Jiangsu Province, China. *Arch Virol*. 2014. 159(12): p. 3407–12 (2014). <https://doi.org/10.1007/s00705-014-2204-2> PMID: 25119679
43. Khamrin P. et al. Molecular detection of kobuvirus sequences in stool samples collected from healthy pigs in Japan. *Infect Genet Evol*. 2010. 10(7): p. 950–4 (2010). <https://doi.org/10.1016/j.meegid.2010.06.001> PMID: 20547246
44. Kumthip K. et al. Detection and genetic characterization of porcine astroviruses in piglets with and without diarrhea in Thailand. *Arch Virol*. 2018. 163(7): p. 1823–1829 (2018). <https://doi.org/10.1007/s00705-018-3806-x> PMID: 29569070
45. Salamunova S. et al. Molecular detection of enteric viruses and the genetic characterization of porcine astroviruses and sapoviruses in domestic pigs from Slovakian farms. *BMC Vet Res*. 2018. 14(1): p. 313 (2018). <https://doi.org/10.1186/s12917-018-1640-8> PMID: 30340595
46. Mor S.K. et al. Detection and molecular characterization of Porcine astrovirus strains associated with swine diarrhea. *J Vet Diagn Invest*. 2012. 24(6): p. 1064–7 (2012). <https://doi.org/10.1177/1040638712458781> PMID: 22956487
47. Bourne F.J. and Curtis J. The transfer of immunoglobins IgG, IgA and IgM from serum to colostrum and milk in the sow. *Immunology*. 1973. 24(1): p. 157–62 (1973). PMID: 4685370
48. Klobasa F. and Butler J.E. Absolute and relative concentrations of immunoglobulins G, M, and A, and albumin in the lacteal secretion of sows of different lactation numbers. *Am J Vet Res*. 1987. 48(2): p. 176–82 (1987). PMID: 3826853
49. Di Profio F. et al. Molecular detection of porcine kobuviruses in Italian swine. *Res Vet Sci*. 2013. 95(2): p. 782–5 (2013). <https://doi.org/10.1016/j.rvsc.2013.06.020> PMID: 23871420
50. Amimo J.O. et al. Molecular detection and genetic characterization of kobuviruses and astroviruses in asymptomatic local pigs in East Africa. *Arch Virol*. 2014. 159(6): p. 1313–9 (2014). <https://doi.org/10.1007/s00705-013-1942-x> PMID: 24327095
51. Park S.J. et al. Molecular detection of porcine kobuviruses in pigs in Korea and their association with diarrhea. *Arch Virol*. 2010. 155(11): p. 1803–11 (2010). <https://doi.org/10.1007/s00705-010-0774-1> PMID: 20820823

52. Barry A.F. et al. First detection of kobuvirus in farm animals in Brazil and the Netherlands. *Infect Genet Evol.* 2011. 11(7): p. 1811–4 (2011). <https://doi.org/10.1016/j.meegid.2011.06.020> PMID: 21763785
53. Sachsenroder J. et al. Simultaneous identification of DNA and RNA viruses present in pig faeces using process-controlled deep sequencing. *PLoS One.* 2012. 7(4): p. e34631 (2012). <https://doi.org/10.1371/journal.pone.0034631> PMID: 22514648
54. Shan T. et al. The fecal virome of pigs on a high-density farm. *J Virol.* 2011. 85(22): p. 11697–708 (2011). <https://doi.org/10.1128/JVI.05217-11> PMID: 21900163
55. Zhang Q. et al. Occurrence and investigation of enteric viral infections in pigs with diarrhea in China. *Arch Virol.* 2013. 158(8): p. 1631–6 (2013). <https://doi.org/10.1007/s00705-013-1659-x> PMID: 23494226
56. Ouyang T., Zhang X., Liu X., and Ren L. Co-Infection of Swine with Porcine Circovirus Type 2 and Other Swine Viruses. *Viruses.* 2019. 11(2) (2019). <https://doi.org/10.3390/v11020185> PMID: 30795620
57. Saade G. et al. Coinfections and their molecular consequences in the porcine respiratory tract. *Vet Res.* 2020. 51(1): p. 80 (2020). <https://doi.org/10.1186/s13567-020-00807-8> PMID: 32546263
58. Salines M. et al. Hepatitis E virus chronic infection of swine co-infected with Porcine Reproductive and Respiratory Syndrome Virus. *Vet Res.* 2015. 46: p. 55 (2015). <https://doi.org/10.1186/s13567-015-0207-y> PMID: 26048774