

Anti-Jo-1 autoantibodies: biomarkers of severity and evolution of the disease in antisynthetase syndrome

Robin Arcani, Louise Rey, Alice Mazziotto, Daniel Bertin, Gilles Kaplanski, Pierre-André Jarrot, Pierre Lafforgue, Geoffroy Venton, Xavier Heim, Patrick Villani, et al.

▶ To cite this version:

Robin Arcani, Louise Rey, Alice Mazziotto, Daniel Bertin, Gilles Kaplanski, et al.. Anti-Jo-1 autoantibodies: biomarkers of severity and evolution of the disease in antisynthetase syndrome. Arthritis Research & Therapy, 2023, 25 (1), pp.125. 10.1186/s13075-023-03116-5 . hal-04391186

HAL Id: hal-04391186 https://amu.hal.science/hal-04391186v1

Submitted on 11 Apr2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

RESEARCH

Arthritis Research & Therapy



Anti-Jo-1 autoantibodies: biomarkers of severity and evolution of the disease in antisynthetase syndrome



Robin Arcani^{1,2*}, Louise Rey³, Alice Mazziotto¹, Daniel Bertin³, Gilles Kaplanski^{2,4}, Pierre-André Jarrot^{2,4}, Pierre Lafforgue⁵, Geoffroy Venton⁶, Xavier Heim^{2,3}, Patrick Villani¹, Jean-Louis Mège³, Alexandre Brodovitch³ and Nathalie Bardin^{2,3}

Abstract

Background Anti-Jo-1 autoantibodies represent essential markers in the diagnosis of antisynthetase syndrome (ASS). In this retrospective study, we aimed to investigate whether their concentrations and fluctuations could both respectively reflect the severity and evolution of ASS.

Methods Between 2015 and 2020, clinical and biological features of ASS patients with at least one positive measure of anti-Jo-1 autoantibody were collected. At each serum sampling, we assessed myositis activity by using the Myositis Intention to Treat Activities Index (MITAX) and compared anti-Jo-1 concentrations with ASS severity, anti-Jo-1 concentrations between patients with and without active disease, and changes in anti-Jo-1 concentrations with disease activity.

Results Forty-eight patients with ASS had at least one positive determination of anti-Jo-1 concentration. Among them, twenty-nine patients had at least two determinations of anti-Jo-1 autoantibody in their follow-up. We showed that these autoantibody concentrations were significantly correlated with MITAX (r=0.4, p=0.03) and creatine kinase concentration (r=0.34, p=0.002) and that they were significantly higher in patients with active disease than in those with inactive disease (91.7 IU/L vs 44.4 IU/L, p=0.016). During follow-up, we found a significant correlation between fluctuations of anti-Jo-1 autoantibody concentrations and MITAX score (r=0.7, p<0.0001).

Conclusion Our results suggest that anti-Jo-1 autoantibody concentration could be a predictive marker of the severity and evolution of ASS and show that their quantification could represent a precious tool for disease monitoring and for improving the therapeutic management of ASS patients.

Keywords Anti-Jo-1 autoantibodies, Antisynthetase syndrome, Biomarkers

*Correspondence: Robin Arcani robin.arcani@ap-hm.fr Full list of author information is available at the end of the article



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Introduction

Inflammatory myopathies are a broad spectrum of autoimmune diseases [1]. Subgroups of myositis were recently defined [2]: dermatomyositis, inclusion body myositis, immune-mediated necrotizing myopathy, and antisynthetase syndrome (ASS). ASS was individualized in patients with clinical manifestations and myositis-specific autoantibodies: antisynthetase autoantibodies. Anti-Jo-1 autoantibodies, directed against the histidyl-tRNA synthetase, are routinely detected in a quantitative way (ELISA or laser bead immunoassays) or in a qualitative/ semi-quantitative way (immunoblots). They are the most commonly detected autoantibodies in patients' sera who have ASS [3].

The disease status of ASS can be assessed with different clinical scorings, e.g., the Myositis Intention to Treat Activities Index (MITAX), which is an index part of the Myositis Disease Activity Assessment Tool (MDAAT) [4, 5]. However, in clinical practice, it is complex to assess disease severity and to monitor disease activity with the available tools. Moreover, there are multiple scores to assess disease activity, without any gold standard [6]. There is a trend towards an association between myositis activity and creatine kinase (CK) concentration, but it is an insufficient surrogate to assess disease severity [7, 8], so other biomarkers must be investigated. There are only a few studies assessing whether their concentration might be predictive of severity or useful monitoring markers for predicting the progression of ASS [9]. In this study, we evaluated if the quantification of anti-Jo-1 autoantibody concentration and its monitoring is associated with ASS severity and evolution, respectively.

Materials and methods

Patient

We conducted a retrospective study from the database of the Immunology Department of the University Hospitals of Marseille, France. We selected all patients with at least one positive measurement of anti-Jo-1 antibody between 2015 and 2020, with a probable or definite myositis diagnosis, based on Bohan and Peter's criteria [10] and on the 2017 ACR/EULAR criteria for inflammatory myopathies [11]. Associated cancers were those declared from 3 years before to 3 years after ASS diagnosis [12].

Disease activity measurement

Along with each serum sampling, we retrospectively evaluated myositis activity from the medical records. We retrospectively applied the MITAX [4]. The MITAX includes the assessment of the physician's intention-to-treat active disease present during the preceding month in each of 7 organ systems (pulmonary, cardiovascular, muscular, constitutional symptoms, cutaneous, skeletal, and gastrointestinal) using a 4-point ordinal scale (0 points: no evidence or discount, 1 point: contentment, 3 points: beware, 9 points: active). To minimize the bias of a retrospective design, the MITAX was calculated by two independent physicians who experimented in myositis (RA and AM) blinded from the anti-Jo-1 antibody concentrations and the disease activity. The assessment of MITAX was used as a surrogate to estimate the disease severity at the time of anti-Jo1 measurement and to follow the activity of the disease over time (at each anti-Jo1 sampling). The anti-Jo-1 measurements were performed on the day of the clinical examination. A patient was classified with inactive disease when the following criteria were satisfied for 3 previous months: (1) no evidence of muscular, articular, cutaneous, or pulmonary activity; (2) serum CK concentration < 300 IU/L; (3) corticosteroid dosage < 10 mg/day with a stable dose of immunosuppressive therapy.

Immunological tests

Antinuclear autoantibodies (ANA) were detected by indirect immunofluorescence on HEp-2 cells (Bio-Rad Laboratories, Hercules, CA) at a screening dilution of 1/160. Anti-extractable nuclear antigen (ENA) antibodies including anti-SSA/Ro 60 kDa, anti-SSB/ La, anti-Ro52kDa/TRIM21, anti-RNP, anti-Sm, anti-Scl70, anti-centromere B, and anti-Jo-1 autoantibodies were quantified using commercially available EliATM kits (Phadia, Uppsala, Sweden; now part of Thermo Fisher Scientific, Waltham, MA). Cutoff values for anti-ENA were 10 IU/m. The sera were tested for anti-Jo-1, anti-PL7, anti-PL12, anti-EJ, anti-OJ, anti-MDA5, anti-Mi2, anti-TIF1y, anti-SRP, anti-SAE1, anti-SAE2, and anti-NXP2 autoantibodies with BlueDiver® Myositis Profile 12-OJ immunodots (Alphadia SA/SN, Wavre, Belgium) when prescribed by the physicians. We assessed the changes in anti-Jo-1 concentration according to the result of the $EliA^{TM}$ kits.

Ethical

This study was approved by the Institutional Review Board of Assistance Publique-Hôpitaux de Marseille (GDPR number PADS21-270). The study was conducted according to the Declaration of Helsinki.

Statistical analysis

Quantitative variables were described using means and standard deviations (SD): categorical variables using numbers and percentages. Quantitative data were compared using Student's t or Mann–Whitney U tests,

while qualitative data were compared with the chisquare or Fisher's exact test when appropriate. Spearman's rank correlations were calculated to quantify the relationships between disease activity and anti-Jo-1 antibody concentrations. Some relationships between disease activity and anti-Jo-1 antibody concentrations were calculated considering MITAX subscorings as ordinal variables using the Kruskall-Wallis and Dunn's multiple comparison tests. We defined correlations as follows: no correlation, weak, moderate, or strong correlations when *r* was < 0.25 0.25–0.5, 0.5–0.75, and > 0.75, respectively. Tests were two-sided, with *p*-values < 0.05 considered to be significant. All analyses were performed with the R software (R Foundation for Statistical Computing, Vienna, Austria) and GraphPad Prism 8.0.0 (GraphPad Software, San Diego, USA). The study was conducted in accordance with the recommendations of the STROBE initiative.

Results

Description of the cohort (Table 1)

Between 2015 and 2020, 48 patients (35 female patients, 72.9%) with at least one positivity of anti-Jo-1 antibody were retrospectively included. All patients had a probable or definite inflammatory myopathy according to Bohan

```
and Peter's criteria [10] and to the 2017 ACR/EULAR
criteria for inflammatory myopathies [11]. The mean age
at the beginning of symptoms was 52.1 \pm 14.4 years, and
the mean age at diagnosis of ASS was 53.9 \pm 14.8 years.
The mean follow-up was 93.6±89.3 months (range:
9-372). There was an associated autoimmune disease
in 10 patients (20.8%): rheumatoid arthritis (RA) (n=6,
12.5%), primary Sjögren's syndrome (n=2, 4.2\%), sys-
temic lupus erythematosus (SLE) (n=2, 4.2\%), Hashi-
moto's thyroiditis (n=1, 2.1%), systemic sclerosis (n=1,
2.1%), and antiphospholipid syndrome (n = 1, 2.1\%).
There was an associated neoplasm in 3 patients (6.3%): 2
colic adenocarcinomas and 1 breast cancer. CK concen-
tration was elevated (>170 IU/L) at least at one antibody
measurement in 14/44 patients (31.8%) during follow-
up. Four patients were under statins during at least one
point of the follow-up (atorvastatin n=3, simvastatin
n=1). Twenty-seven patients (56.3%) were negative for
ANA. Besides positivity for anti-Jo-1, screening for other
autoantibodies showed anti-ENA to be negative in 25
patients (52.1%). Sixteen patients (33.3%) presented an
anti-Ro52kDa/TRIM21 antibody, 5 (10.4%) an anti-SSA
60 kDa antibody, 4 (8.3%) an anti-centromere B, and 2
(4.2%) an anti-SSB antibody. All patients tested (29/29)
```

were positive for anti-Jo-1 screening with immunodot

Table 1	Demographic	and clinical	l features of the	patients
---------	-------------	--------------	-------------------	----------

Characteristics	Whole cohort (<i>n</i> =48)	Patients with one unique sampling of anti-Jo-1 (<i>n</i> = 19)	Patients with at least 2 samplings of anti-Jo-1 (<i>n</i> =29)	<i>p</i> -value
Age at diagnosis ^a	53.9±14.8	54.6±15.8	53.5±14.5	0.82
Female/male	35/13	12/7	23/6	0.22
Duration of follow-up in months ^a	93.6±89.3	94.7±106.8	93±78.6	0.95
Clinical feature ^b				
Myositis	39 (81.3)	15 (78.9)	24 (82.8)	0.74
Arthralgia/arthritis	39 (81.3)	16 (84.2)	23 (79.3)	0.67
Interstitial lung disease	35 (72.9)	12 (63.2)	23 (79.3)	0.22
Mechanic's hands	22 (45.8)	6 (31.6)	16 (55.2)	0.11
Raynaud's phenomenon	10 (20.8)	3 (15.8)	7 (24.1)	0.49
Rash	3 (6.3)	1 (5.3)	2 (6.9)	0.82
Digestive symptoms	12 (25)	1 (5.3)	11 (37.9)	0.011
Treatment lines ^b				
Corticosteroids	45 (93.8)	18 (94.7)	27 (93.1)	0.82
Methotrexate	21 (43.8)	8 (42.1)	13 (44.8)	0.85
Azathioprine	15 (31.3)	5 (26.3)	10 (34.5)	0.55
Rituximab	11 (22.9)	3 (15.8)	8 (27.6)	0.34
Intravenous immunoglobulins	9 (18.8)	5 (26.3)	4 (13.8)	0.28
Mycophenolate mofetil	7 (14.6)	1 (5.3)	6 (20.7)	0.14
Cyclophosphamide	4 (8.3)	1 (5.3)	3 (10.3)	0.54
Cyclosporin A	1 (2.1)	0 (0)	1 (3.4)	0.41
Number of treatment lines ^a	2.4 ± 1.2	2.2±1	2.5±1.3	0.29

 a Mean \pm SD

^b n (%)

assay. Two patients (6.9%) of 29 screened for myositisspecific or myositis-associated autoantibodies were positive for anti-PL-12 and anti-HMGCR, respectively. Six patients underwent muscular biopsy: in 5, typical patterns of inflammatory myopathy were found. All patients received treatment for ASS: 13 (27.1%), 14 (29.2%), 15 (31.3%), and 6 (12.5%) patients received one, two, three, or more than 3 lines of treatment, respectively. Corticosteroids were the most often used treatment (45 patients, 93.8%). Of 48 patients, 29 had at least two determinations of anti-Jo-1 antibody concentration. In these monitored patients, there was a mean of 3.2 ± 2.1 determinations of anti-Jo-1 antibody concentration per patient. Sixteen patients (55.2%) had 2 determinations of anti-Jo-1 concentration and 8 patients had at least 4 determinations (27.6%) of anti-Jo-1 concentration.

Anti-Jo-1 concentration is a good marker of ASS severity in contrast to CK concentration

The severity of ASS, assessed by CK concentration, MITAX (total scoring and subscorings in each organ system), and disease activity, was analyzed according to the anti-Jo-1 concentrations.

In the whole cohort of 48 patients with at least one measurement of anti-Jo-1 autoantibody concentration, we found a significant weak correlation between anti-Jo-1 and CK concentration (r=0.34, CI95% 0.044–0.59, p=0.002, Fig. 1A), whereas no correlation was found between CK concentration and disease activity as evaluated by MITAX (r=0.21, CI95%: -0.094-0.49, p=0.15, Fig. 1B).

In addition, we found a significant, but weak, association between anti-Jo-1 concentrations and MITAX (r=0.4, CI95%: 0.13–0.62 p=0.003, Fig. 2A). There was a significant association between anti-Jo-1 concentration and pulmonary MITAX (r=0.44, p=0.002, mean anti-Jo-1 concentration:

 50.5 ± 28.4 vs 108.9 ± 66.2 , p = 0.0062 between patients with MITAX: 0 vs MITAX: 3, Fig. 2B). We did not find any statistical association between anti-Jo-1 concentration and the muscular, articular, cutaneous, cardiological, gastrointestinal, or constitutional disease activity.

When comparing anti-Jo-1 concentrations to patients with active and inactive disease, it showed that anti-Jo-1 concentrations were higher in those with active disease (mean anti-Jo-1 concentration: 91.7 IU/L vs 44.4 IU/L, p=0.016, Fig. 3A). Similar results were obtained with MITAX (9.7 vs 2, p < 0.01, Fig. 3B), in contrast to CK concentration for which no significant difference was observed between patients (with active or inactive disease) (882 IU/L vs 257 IU/L, p > 0.05, Fig. 3C).

Anti-Jo-1 concentrations represent a follow-up marker in ASS evolution

In 29 patients with at least two measurements of anti-Jo-1, followed for 93 months on average, we investigated at each visit a potential association between the changes of anti-Jo-1 concentrations and those of MITAX, including subscorings in each organ system. As presented in Fig. 4, fluctuations of anti-Jo-1 concentrations were significantly and moderately correlated with those of MITAX (r=0.7, CI95%: 0.55-0.81, p<0.0001, Fig. 4A). Similarly, the anti-Jo-1 concentration variations were also weakly correlated with those of muscular (r=0.37, CI95%: 0.13-0.57, p=0.0024, Fig. 4B), pulmonary (r = 0.3, CI95%: 0.053-0.51, p = 0.015, Fig. 4C), and cutaneous (r = 0.27, CI95%: 0.019-0.49, p = 0.03, Fig. 4D) MITAX subscorings. We did not find any statistical association between anti-Jo-1 concentration changes and CK concentration changes (r=0.28, CI95%: -0.098-0.59, p = 0.13) or between CK concentration changes and MITAX changes (r = -0.05, CI95%; -0.64-0.58,p = 0.86).

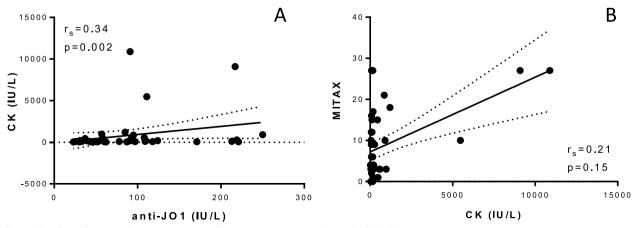


Fig. 1 Correlation between CK concentration and anti-Jo-1 concentration (A) and MITAX (B)

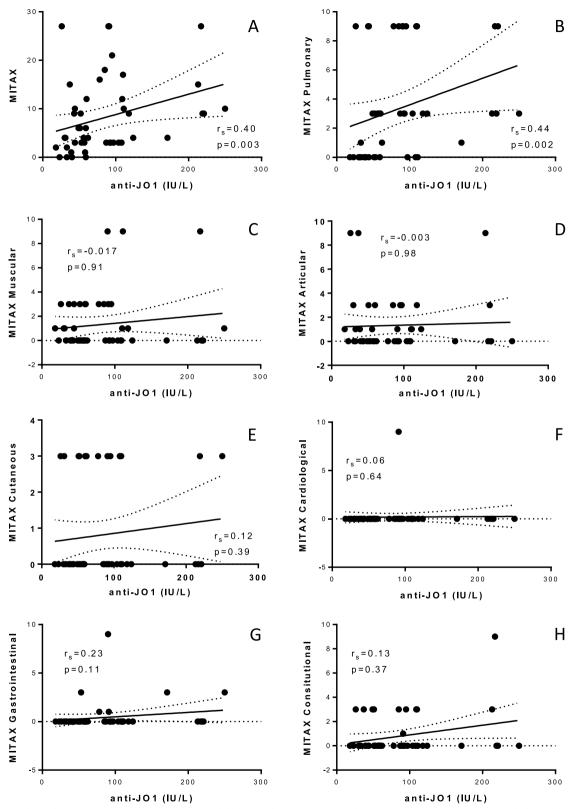
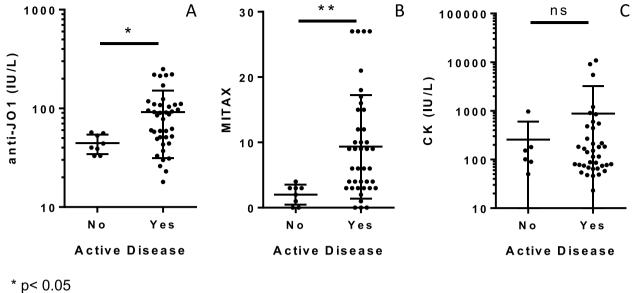


Fig. 2 Correlation between anti-Jo-1 concentration and global (A), pulmonary (B), muscular (C), articular (D), cutaneous (E), cardiological (F), gastrointestinal (G), and constitutional (H) disease activity



** p< 0.01

Fig. 3 Correlation between disease activity and anti-Jo-1 concentration (A), MITAX (B), and CK concentration (C)

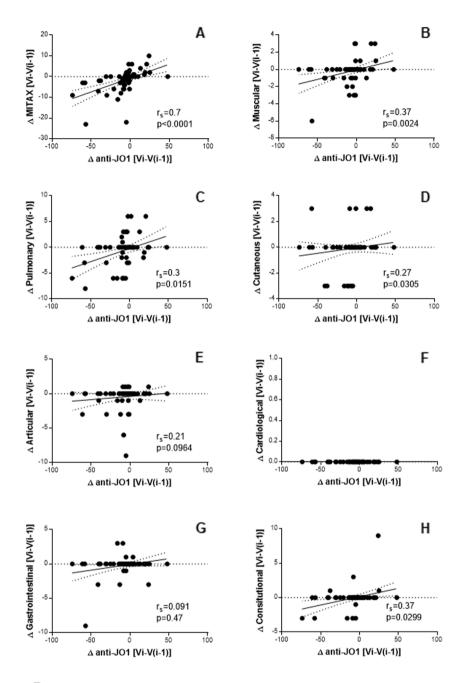
Discussion

Predicting ASS progression presents a crucial interest to adapt therapeutic management. Our results showed that quantification of anti-Jo-1 autoantibody concentration is of interest because it is associated with disease severity, and their monitoring could be useful to monitor ASS evolution. These autoantibodies are routinely searched and quantified in medical laboratories, making them available and reliable biomarkers in clinical practice.

In the literature, numerous data evidenced the major role of anti-Jo-1 autoantibodies as the most prevalent diagnostic marker for ASS patients [3], but few data investigated their potential usefulness in disease monitoring. In agreement with our results, Stone et al. showed a correlation between anti-Jo-1 concentrations and the global severity of the disease and between anti-Jo-1 concentration and muscular and articular activities [13]. They also reported 11 patients with several determinations of anti-Jo-1 concentration and concomitant clinical assessment based on MITAX evaluation, in which a correlation was found between anti-Jo-1 fluctuations and changes in ASS activity. Our study, conducted on 29 patients with at least two measurements of anti-Jo-1 concentration, represents the largest cohort of followed patients with this rare autoimmune disease in the literature. As such, Aggarwal et al. [14] found that clinical improvement post-rituximab was associated with a decrease in anti-Jo-1 concentration in a cohort of patients treated with rituximab for inflammatory myopathies. These data suggest that quantification of anti-Jo-1 autoantibodies can help in identifying patients with poor outcomes, who require more aggressive treatment.

These findings indicate that anti-Jo-1 autoantibodies, myositis-specific autoantibodies, could be involved in the pathogenicity of myositis as well as in the flare of the disease. Tissue modifications of the expression of the antigenic target, histidyl-tRNAsynthetase, may be responsible for the recruitment and activation of both innate and adaptive immune cells. Therefore, anti-Jo-1 autoantibodies, reflecting autoimmunization, could be correlated with the cellular response [15]. Some studies showed that anti-Jo-1 autoantibodies were found when T lymphocytes (against Jo-1) were present [16]. Moreover, the B-cells' expansion, and consequently the autoantibodies production, is amplified by the TCD4 cell response [17]. Taken together, our results and the beneficial effect of B-cells' depletion in myositis [14, 18] are an element for the role of anti-Jo-1 autoantibodies in the emergence of tissular lesions in myositis.

Our study had some biases. Our sample size was quite small. This is common when studying rare diseases like ASS. However, compared to the data reported in the literature, our cohort is one of the largest. One of the limitations of our study is due to its retrospective design. However, our results, in agreement with previous studies [13, 14], pave the way to a multicenter prospective study.



Footnotes:

 Δ anti-JO1 [Vi-Vi(i-1)] corresponds to the difference between the anti-Jo-1 level at the visit and the anti-Jo-1 level at the previous visit

△MITAX [Vi-Vi(i-1)] corresponds to the difference between the MITAX at the

visit and the MITAX at the previous visit

Fig. 4 Correlation between anti-Jo-1 concentration changes and disease activity evolution (global (A), muscular (B), pulmonary (C), cutaneous (D), articular (E), cardiological (F), gastrointestinal (G), constitutional (H) disease activity) in each patient (*n* = 29) during the follow-up

In conclusion, like the monitoring of anti-dsDNA in SLE or of C-reactive protein in RA [19, 20], the quantification of anti-Jo-1 autoantibodies represents an attractive biomarker to monitor the disease activity. Managing ASS is a major challenge, and there are no consensual guidelines to address this issue. The anti-Jo-1 antibody quantification can be another tool to assess the evolution of the disease. We could suggest their quantification at each clinical assessment to help the practitioners' decisions. Multicenter prospective studies will be necessary to validate our findings and to assess the timing and frequency of quantification of anti-Jo-1 autoantibodies according to the therapeutic strategies applied.

Acknowledgements

Not applicable.

Authors' contributions

RA, LR, AM, DB, JLM, AB and NB contributed to the conception and to the design of the study. RA, LR, AM, DB, GK, PAJ, PL, GV, XH and PV contributed to the acquisition of the data. All the authors contributed to the interpretation of the data. RA, LR, AB and NB have drafted the work. All the authors revised the drafted work. All the authors have approved the submitted version and have agreed both to be personally accountable for the author's own contributions and to ensure that questions related to the accuracy or integrity of any part of the work, even ones in which the author was not personally involved, are appropriately investigated, resolution documented in the literature.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Availability of data and materials

The data that support the findings of this study are available from the corresponding author, but restrictions apply to the availability of these data and so are not publicly available. Data are however available from the authors upon reasonable request and with permission from the corresponding author.

Declarations

Ethics approval and consent to participate

This study was approved by the Institutional Review Board of Assistance Publique-Hôpitaux de Marseille (GDPR number PADS21-270). The Institutional Review Board waived patient approval.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹ Internal Medicine and Therapeutics Department, CHU La Timone, Assistance Publique-Hôpitaux de Marseille (AP-HM), 264 Rue Saint-Pierre, 13005 Marseille, France. ²Center for Cardiovascular and Nutrition Research (C2VN), INRA, 1260, INSERM UMR_S 1263, Aix-Marseille University, Marseille, France. ³Biogénopôle, CHU La Timone, Assistance Publique-Hôpitaux de Marseille (AP-HM), Marseille, France. ⁴Department of Internal Medicine and Clinical Immunology, CHU La Conception, Assistance Publique-Hôpitaux de Marseille (AP-HM), Marseille, France. ⁵Department of Rheumatology, CHU Sainte-Marguerite, Assistance Publique-Hôpitaux de Marseille (AP-HM), Marseille, France. ⁶Hematology and Cellular Therapy Department, CHU La Conception, Assistance Publique-Hôpitaux de Marseille (AP-HM), Marseille, France.

Received: 14 May 2023 Accepted: 12 July 2023 Published online: 22 July 2023

References

- Selva-O'Callaghan A, Pinal-Fernandez I, Trallero-Araguás E, Milisenda JC, Grau-Junyent JM, Mammen AL. Classification and management of adult inflammatory myopathies. Lancet Neurol. 2018;17:816–28.
- Mariampillai K, Granger B, Amelin D, Guiguet M, Hachulla E, Maurier F, et al. Development of a new classification system for idiopathic inflammatory myopathies based on clinical manifestations and myositis-specific autoantibodies. JAMA Neurol. 2018;75:1528–37.
- Cavagna L, Trallero-Araguás E, Meloni F, Cavazzana I, Rojas-Serrano J, Feist E, et al. Influence of antisynthetase antibodies specificities on antisynthetase syndrome clinical spectrum time course. J Clin Med. 2019;8:2013.
- Sultan SM, Allen E, Oddis CV, Kiely P, Cooper RG, Lundberg IE, et al. Reliability and validity of the myositis disease activity assessment tool. Arthritis Rheum. 2008;58:3593–9.
- Isenberg DA, Allen E, Farewell V, Ehrenstein MR, Hanna MG, Lundberg IE, et al. International consensus outcome measures for patients with idiopathic inflammatory myopathies. Development and initial validation of myositis activity and damage indices in patients with adult onset disease. Rheumatology (Oxford). 2004;43:49–54.
- Rider LG, Werth VP, Huber AM, Alexanderson H, Rao AP, Ruperto N, et al. 6. Measures of adult and juvenile dermatomyositis, polymyositis, and inclusion body myositis: Physician and Patient/Parent Global Activity, Manual Muscle Testing (MMT), Health Assessment Questionnaire (HAQ)/Childhood Health Assessment Questionnaire (C-HAQ), Childhood Myositis Assessment Scale (CMAS), Myositis Disease Activity Assessment Tool (MDAAT), Disease Activity Score (DAS), Short Form 36 (SF-36), Child Health Questionnaire (CHQ), physician global damage, Myositis Damage Index (MDI), Quantitative Muscle Testing (QMT), Myositis Functional Index-2 (FI-2), Myositis Activities Profile (MAP), Inclusion Body Myositis Functional Rating Scale (IBMFRS), Cutaneous Dermatomyositis Disease Area and Severity Index (CDASI), Cutaneous Assessment Tool (CAT), Dermatomyositis Skin Severity Index (DSSI), Skindex, and Dermatology Life Quality Index (DLQI). Arthritis Care Res (Hoboken). 2011;63(Suppl 11):S118-157.
- Benveniste O, Drouot L, Jouen F, Charuel J-L, Bloch-Queyrat C, Behin A, et al. Correlation of anti-signal recognition particle autoantibody levels with creatine kinase activity in patients with necrotizing myopathy. Arthritis Rheum. 2011;63:1961–71.
- Werner JL, Christopher-Stine L, Ghazarian SR, Pak KS, Kus JE, Daya NR, et al. Antibody levels correlate with creatine kinase levels and strength in anti-3-hydroxy-3-methylglutaryl-coenzyme A reductase-associated autoimmune myopathy. Arthritis Rheum. 2012;64:4087–93.
- 9. Yoshida S, Akizuki M, Mimori T, Yamagata H, Inada S, Homma M. The precipitating antibody to an acidic nuclear protein antigen, the Jo-1, in connective tissue diseases. A marker for a subset of polymyositis with interstitial pulmonary fibrosis. Arthritis Rheum. 1983;26:604–11.
- Bohan A, Peter JB. Polymyositis and dermatomyositis (first of two parts). N Engl J Med. 1975;292:344–7.
- Lundberg IE, Tjärnlund A, Bottai M, Werth VP, Pilkington C, de Visser M, et al. 2017 European League Against Rheumatism/American College of Rheumatology classification criteria for adult and juvenile idiopathic inflammatory myopathies and their major subgroups. Ann Rheum Dis. 2017;76:1955–64.
- Dani L, Ian Che W, Lundberg IE, Hellgren K, Holmqvist M. Overall and site-specific cancer before and after diagnosis of idiopathic inflammatory myopathies: a nationwide study 2002–2016. Semin Arthritis Rheum. 2021;51:331–7.
- Stone KB, Oddis CV, Fertig N, Katsumata Y, Lucas M, Vogt M, et al. Anti-Jo-1 antibody levels correlate with disease activity in idiopathic inflammatory myopathy. Arthritis Rheum. 2007;56:3125–31.
- Aggarwal R, Oddis CV, Goudeau D, Koontz D, Qi Z, Reed AM, et al. Autoantibody levels in myositis patients correlate with clinical response during B cell depletion with rituximab. Rheumatology (Oxford). 2016;55:991–9.

- 15. Tsui FW, Siminovitch KA. Anti-Jo-1 autoantibodies and the immunopathogenesis of autoimmune myositis. Int Rev Immunol. 1991;7:225–35.
- Katsumata Y, Ridgway WM, Oriss T, Gu X, Chin D, Wu Y, et al. Species-specific immune responses generated by histidyl-tRNA synthetase immunization are associated with muscle and lung inflammation. J Autoimmun. 2007;29:174–86.
- 17. Craft J, Fatenejad S. Self antigens and epitope spreading in systemic autoimmunity. Arthritis Rheum. 1997;40:1374–82.
- Oddis CV, Reed AM, Aggarwal R, Rider LG, Ascherman DP, Levesque MC, et al. Rituximab in the treatment of refractory adult and juvenile dermatomyositis and adult polymyositis: a randomized, placebo-phase trial. Arthritis Rheum. 2013;65:314–24.
- Esdaile JM, Abrahamowicz M, Joseph L, MacKenzie T, Li Y, Danoff D. Laboratory tests as predictors of disease exacerbations in systemic lupus erythematosus. Why some tests fail Arthritis Rheum. 1996;39:370–8.
- Felson DT, Smolen JS, Wells G, Zhang B, van Tuyl LHD, Funovits J, et al. American College of Rheumatology/European League Against Rheumatism provisional definition of remission in rheumatoid arthritis for clinical trials. Arthritis Rheum. 2011;63:573–86.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

